IMPACT OF SUPPLEMENTING DIET WITH MORINGA OLEIFERA LEAVES AND VITAMIN C ON GROWTH PERFORMANCE, BLOOD CONSTITUENTS, ANTIOXIDANT INDICES AND HORMONE PROFILES OF JAPANESE QUAILS UNDER SUBTROPICAL CLIMATIC CONDITIONS

W. Fouad¹; A.Y. Kassab² and M.M. Embadir¹

¹Poultry Production Department, Faculty of Agriculture, New Valley University, Egypt. ²Animal Production Department, Faculty of Agriculture, New Valley University, Egypt. *Corresponding Author: Walid Fouad, walidfouad1971@agr.nvu.edu.eg

(Received 23/3/2022, accepted 7/5/2022)

SUMMARY

This experiment was conducted to evaluate the Impact of supplementing diet with Moringa oleifera leaves (MOL) and vitamin C on growth performance, blood constituents, antioxidant indices and hormone profiles of Japanese quails under subtropical climatic conditions. A total 180 unsexed Japanese quails chicks, 7day old were randomly distributed into 6 groups housed with 3 replicate, 10 chicks/ each. In the 1st group (control), chicks were fed on a basal diet, while those in the 2nd (T1), 3rd (T2), 4th (T3), 5th (T4) and 6th (T5) groups were fed on the same basal diet supplemented with 3g MOL/kg diet, 6g MOL/kg diet, 200ppm Vit. C, 3g MOL/kg diet + 200ppm Vit. C, and 6g MOL/kg diet + 200ppm Vit. C respectively. Average maximum room temp°C during the day between 40-45 °C and 50% relative humidity were recorded during the experimental period until 6 weeks age. The obtained results showed that the chicks supplemented with 6g MOL/kg diet + Vit. C 200ppm had significantly greater body weight, body weight gain, Feed consumption and percentages of dressing and internal organs and improved feed conversion ratio, followed by (T2), (T4) then (T3) and (T1) compared with control group. The hematological traits, immunity parameters, antioxidant indices for chicks in treated groups were significantly (P<0.01) improved compared with control group. Serum proteins, albumin, globulin, high density lipoproteins (HDL) and thyroid hormones were significantly improved, while total serum lipids, cholesterol, glucose concentration, low density lipoproteins (LDL), liver enzymes, creatinine and urea were significantly (P<0.01) decreased compared with control group. However, no significant effects were detected on white blood cells due to the addition of different treatments compared to the control. The bacterial count in chick intestine were remarkably decreased due to addition MOL and Vit.C at different levels compared to the control. It can be concluded that MOL supplementation in Japanese quail diets is a good natural source and can be used as a safe alternative to synthetic Vit.C under subtropical climatic conditions.

Keywords: Quail, Moringa leaves, vit. C, growth performance, subtropical climatic conditions, hormone.

INTRODUCTION

Certain cities in Egypt's tropical and subtropical regions are subject to heat stress conditions, with highs outside ambient temperature reaching 45°C and more frequent records of temperatures above the critical point of 35°C during the summer months of June through September (Faisal, 2008). One of the most frequent pressures impacting production parameters is temperature, and in poultry, a temperature above 30°C is indicative of heat stress. Heat stress in meat-type poultry flocks has been linked to high mortality rates, low feed intake, low body weight increase, and poor feeding efficiency (Sahin et al., 2002a, b). Heat stress causes a rise in free radical production, which in turn promotes the production of reactive oxygen species and oxidative stress, increasing lipid oxidation (Altan et al., 2003). As a result of heat stress, oxidative stress showed a two-fold increase in malondialdehyde, an indicator of lipid peroxidation, in skeletal muscle (Mujahid et al., 2009 and Wang et al., 2009). It also showed decreased concentrations of minerals and serum vitamins (Vit.), which are crucial for the antioxidant defense system (Sahin et al., 2002, 2009). Antioxidant vitamin supplements (Puthpongsiriporn et al., 2001; Franchini et al., 2002) or natural substances with antioxidant potential (Sahin et al., 2008 and Tuzcu et al., 2008) can both lower lipid peroxidation.

Vitamin C acts as an antioxidant with other antioxidants through synergistic effects. The antioxidant activity of Vit. E was increased in the presence of Vit. C by reducing tocopherol radicals to their active

form of Vit. E (Doba et al., 1985). Several studies have revealed that adult poultry is able to synthesize Vit. C to meet its requirements under normal conditions. However, their requirements have been found to increase during stress, and several studies have reported the beneficial effects of supplementing poultry feed with ascorbic acid (Abidin and Khatoon, 2013 and Maurice et al., 2004). Dietary supplementation with Vit. C alleviated stress metabolic, improved performance, enhanced immunological status, and reduced mortality. Several studies have revealed that the optimum response in terms of growth performance, feed efficiency, feed conversion ratio (FCR), carcass quality, and survival rate in broilers under heat stress appeared with average supplements of 200-250 mg/kg Vit. C. However, the required Vit. C in laying hens is about 200–500 mg/kg to enhance feed efficiency, egg production, and egg quality under heat stress (Abidin and Khatoon, 2013, Sahin et al., 2002 and Seven et al., 2010). It is known that natural antioxidants such as vitamin C., flavonoids, tocopherol, and other phenolic compounds are present in some plants such as the Moringa plant. Moringa oleifera leaves are rich in Vitamin A and C, considered to be useful in catarrhal afflictions, said to have purgative properties and to promote digestion, and used as an external application for wounds (The Wealth of India1962, (A Dictionary of Indian Raw Materials and Industrial Products)). Moringa oleifera has been cultivated for decades in North Sinai, Aswan, and the New Valley. Moringa leaves are very rich in large amounts of Vit. (A, B, C, E) and carotene, polyphenols, calcium, iron, phosphorous, and protein (Murro et al., 2003). Moreover, is one such plant that has been identified as containing natural antioxidants (Siddhuraju and Becker, 2003). The antioxidant effect of MOL was due to the presence of polyphenols, anthocyanins, tannins, glycosides, and thiocarbamates, which activate antioxidant enzymes, remove free radicals and inhibit oxidase (Luqmans et al., 2012). Using MOL as a feed additive in poultry feeding requires an investigation of its nutritional value, as well as its effect on blood parameters as a measure of both the nutritional and medicinal benefits of the leaves in broiler chicks (Ebenebe et al., 2012). Moringa is rich in nutrients that improve the digestion of other foods and reduce the activity of pathogenic bacteria and molds, helping chickens to display their natural genetic potential (Gaia, 2005). Atawodi (2010) noted that MOL contains polyphenols such as catechol, methyl gallate, ellagic acid, kaempferol quercetin, and gallate. Supplementation to the moringa diet for broilers was effective in enhancing the oxidative stability of chicken meat (Qwele et al., 2013). Moringa plant, when used in poultry feed, improves growth performance and immune response under Heat stress conditions in poultry (Abou Sekken, 2015). Fouad and El-Rayes (2019) reported that supplementation of Moringa leaves to the diet improved productive performance, carcass characteristics, blood components, hormones, antioxidant indices, and immune parameters of Japanese quail chicks.

Therefore, this study was conducted to study the effect of different levels of MOL as a new source of antioxidants, and Vit. C on growth performance and physiological parameters of Japanese quail under subtropical climatic conditions.

MATERIALS AND METHODS

The experiment was carried out on a private farm in El Kharga city, New Valley Governorate, Egypt from August to September.

Moringa source and preparation:

Moringa (Moringa Oleifera) leaves were obtained from a New Valley Governorate of Egypt. The harvested moringa leaves were air-dried in the shade under a shed until they were crispy to the touch. The leaves were then milled using a hammer mill of sieve size 3 mm, to obtain a product herein referred to as moringa leaf meal (MOLM) to be ready for mixing with the other diet ingredients. The ingredients and calculated analysis of the experimental diets illustrated in Table (1a) were formulated to cover the nutrient requirements of Japanese quail recommended by the National Research Council (NRC (1994). The chemical composition of MOL is shown in Table (1b). The activities of enzymatic antioxidants and the level of non-enzymatic antioxidants in Moringa oleifera leaves are shown in (Tables 2a and 2b) according to Sreelatha and Padma (2009).

Chemical properties of vitamin C:

Ascorbic acid (AA), generally referred to as vitamin C, is a powder or crystals that are white or slightly yellow, a little acid.m.p.190°C-192°C, readily soluble in water, a little soluble in alcohol, and uncomfortably soluble in ether and chloroform and other organic solvents (LeBlanc, 2019).

Ingredients	%
Ground yellow corn	53. 20
Soya bean meal (44%)	37.00
Fish meal (60.05%)	5.50
Vegetable oil	1.00
Oyster Shel	1.00
Mono Calcium Phosphate	1.50
DL-Methionine	0.15
Salt	0.15
Minerals and vitamins premix	0.50
Calculated analysis	
Crude protein (%)	24.00
ME (kcal/kg)	2900.00
Calorie/protein ratio (C/P	120.83
Calcium (%)	1.2
Lysine	1.47
Methionine%	0. 57
Phosphorus (%)	0.55

 Table (1a): Composition and calculated analysis of the experimental diet- through the growing period.

*Each 1 kg contains: Vit. A, 12000 IU; D3, 2000 IU; E, 20 mg; K3, 3 mg; B2, 7 mg; B3, 12 mg; B5, 3 mg; B12, 0.03 mg; Biotin, 0.1 mg; Choline chloride, 300 mg; Mn, 130 mg; Fe, 70 mg; Zn, 60 mg; Cu, 12 mg; I, 1 mg; Se, 0.2 mg.

	Table ((1b):	: Chemical	composition	from N	Aoringa	Oleifera	leaves of	experimental stud	v.
--	---------	----------------	------------	-------------	--------	---------	----------	-----------	-------------------	----

Parameter (%)	Composition%
Dry matter (DM)	94.25
Crude protein	23.80
Crude ether extract	5.50
Total ash	9.75
Nitrogen free extracts (NFE)	38.63
Crude fiber	16.57

Table (2a): Activities of enzymatic antioxidants in Moringa oleifera leaves.

Parameter	Matured leaves
SOD (Ua/g)	14.64
CAT (Ub/g)	106.84
GPx (Uc/g)	163.68
GST (U#/g)	0.30

^a 1 Unit = Amount of enzyme that gives 50% inhibition of the extent of NBT reduction in 1 min

^b I Unit = Amount of enzyme required to decrease the absorbance at 240 nm by 0.05 units

^c 1 Unit = Change of absorbance/minute at 430 nm d

1 Unit = µmol of CDNB conjugated/min

Table (2b): Levels of non-enzymatic antioxidants in Moringa oleifera leaves.

Parameter	Matured leaves
Ascorbic acid (mg/g)	6.60
Tocopherol ($\mu g/g$)	6.53
Total carotenoids (mg/g)	92.38

Experimental design:

One hundred and eighty unsexed Japanese quail chicks, aged seven days, were randomly assigned to six treatment groups, each groups housed in three replicate pens each consisting of ten chicks. The chicks' live weights were almost similar. Under comparable supervision and hygienic settings, chicks were grown. Water and food were given freely during the trial, which concluded after six weeks of age. In order to satisfy the nutritional needs of chicks, the basal diet (control) is designed to provide 24% CP and 2900 Kcal. Table (1a) lists the components of the basal diet, and Table (1b) lists the concentrations

of non-enzymatic antioxidants in MOL. The first group of chicks was fed a basal diet and was regarded as the control group. The other groups, which were T1, T2, T3, T4, T5, and T6, were fed the same basal diet supplemented with 3g MOL/kg diet, 6g MOL/kg diet, and 200 ppm of vitamin C, as well as 3g MOL/kg diet + Vit. C 200 ppm, and 6g MOL/kg diet + Vit. C 200 ppm , respectively. Each bird was weighed in grams both at the start of the experiment (1 week) and at the conclusion (6 weeks). Every bird was housed under identical environmental circumstances. Over the course of the trial, up until six weeks of age, the average highest room temperature C during the day was reported between 40 and 45 °C with 50% relative humidity. Body weight (BW) and Feed consumption, BWG for each replicate, and FCR as g feed/g gain were calculated.

Temperature and humidity:

Between August and September, four daily readings of mean the ambient temperature and relative humidity were made twice in the day and twice at night. From one to six weeks of age, the average maximum room temperature, measured indoors, is between 40 and 45 degrees Celsius during the day with 50% relative humidity (Tables 3).

Month		Average temp °C during the night	Average temp °C during the day	RH %
	Second week	27	45	50
August	Third week	25	43	48
	Fourth week	26	44	49
Contombor	Fifth week	24	43	52
September	sixth week	23	40	51
Overall means		25	43	50

Table (3): Average weekly room temperature and % (RH).

¹ The minimum and maximum room temperatures were recorded using digital thermometer recording minimum and maximum temperatures during the specified time (24-h period).

 2 The average values were calculated from minimum and maximum room temperature values recorded throughout the week.

Carcass and blood samples:

Five birds from each treatment were chosen at random, weighed, and killed for carcass dressing when they were six weeks old. The percentage of the live weight was used to express the weights of the carcass, liver, gizzard, heart, spleen, and intestinal organs. The American Public Health Association (A.P.H.A, 1985) was followed in the execution of the total intestinal anaerobic count, total coliform count, and aerobic count (APC). At the time of slaughter, ten blood samples from each experimental group were taken and split into two sections. To get serum, the first half was collected in heparin-containing tubes, and the second part was taken in non-heparin-containing tubes. Hematological parameters (red blood cells (RBCs), hemoglobin (Hb), packed cell volume (PCV), and white blood cells (WBCs)) were measured using fresh blood aliquots.

Measurements:

In accordance with the principles and procedures of Armstrong and Carr, 1964, blood biochemical parameters such as plasma total protein (g/dl) were determined using specialized kits supplied by sentinel CH Milano, Italy using a spectrophotometer (Beckman DU-530, Germany). Using specialized kits sent from sentinel CH Milano, Italy, plasma albumin (g/dl) was measured using the protocol of (Doumas et al., 1971). Since fibrinogen often consists of a small fraction, the plasma globulin level (g/dl) was determined by calculating the difference between total protein and albumin (Sturkie, 1986). Following the suggestion of Frings et al. (1972), serum total lipid concentrations (mg/dl) in blood serum were measured using spectrophotometer and specific kits supplied by CAL-TECH Diagnostics, Inc., Chino, CA, USA. Serum total cholesterol (mg/dl) was determined on individual bases using the specific kits according to the recommendation of (Bogin and Keller, 1987). Serum samples were analyzed for low-density lipoprotein (LDL) and high-density lipoprotein (HDL) using the colorimetric method by commercial kits obtained from Reactivos GPL, Barcelona, Spain according to the recommendation of (Warnick *et al.*, 1983). Plasma glucose concentration was measured by the method of (Trinder, 1969). The transaminase enzymes activities of serum aspartate aminotransferase (AST) and serum alanine aminotransferase (ALT), as U/L, were determined by the calorimetric method (Reitman and Frankel,

Egyptian J. Nutrition and Feeds (2023)

1957). Serum creatinine level was estimated according to (Husdan and Rapoport, 1968), while, serum uric acid level was measured according to the method explained by Patton and Crouch (1977). Plasma tri-iodothyronine (T3) and thyroxin (T4) concentrations were measured using by radioimmunoassay technique (RIA) according to (Darras et al., 1991). Serum immunoglobulin (Ig) types IgG and IgM fractions were determined using the Elisa technique according to (Micini et al., 1965). Total antioxidant capacity (TAC) is determined by the reaction of antioxidants in the sample with a defined amount of exogenously provided hydrogen peroxide (H2O2). The residual H2O2 is determined colorimetrically by an enzymatic reaction which involves the conversion of 3, 5, dichloro -2- hydroxyl, (Koracevic et al., 2001).

Statistical analysis:

The general linear model process of the Statistical Analysis System (SAS, 2002) was used to examine the statistical analysis data. One-way ANOVA was used in this model: $Yik=\mu + Ti + eik$. Where, Y is the dependent variable; μ is the general mean; T is the effect of experimental treatments; and e is the experimental random error. Significant differences between means were defined at P<0.05 compared using Duncan's multiple range test (Duncan, 1955).

RESULTS AND DISCUSSION

Growth performance:

Table (4) illustrates the impact of feeding Japanese quails diets with varying MOL and Vit. C concentrations on BW, BWG, FI, and FCR in subtropical climates. For every treatment, the quail chicks' initial BW was the same. Under subtropical climate circumstances, supplementing with vitamin C and Moringa oleifera leaves improved FCR and significantly (P<0.01) raised BW, BWG, and FI in comparison to the control group. When diet treatments T1, T2, T3, T4, and T5 were added, the BW, BWG, and FI of the control group increased by 13.13, 18.84, 14.11, 19.58, and 27.8%, 25.1, 23.17, 17.61, 24.23, and 34.25%, and 5.56, 8.48, 5.86, 9.45, and 12.69%, respectively. Furthermore, in comparison to the control group, quails fed the basal diet supplemented with 6g MOL/kg diet + Vit. C 200 ppm(T5), 3g MOL/kg diet + Vit. C 200 ppm (T4), 6g MOL/kg diet (T2), then Vit. C 200 ppm and 3g MOL/kg diet had significantly higher BW, BWG, and FI.

When compared the Vit. C 200 ppm group (T3) and the control group with 6g MOL/kg food (T2), the results indicated that the birds fed a diet supplemented with 6g MOL/kg food (T2) had the highest body weight, FI, BWG, and the best FCR. However, when compared to the other dietary treatments and the control group, the results indicated that the birds fed a diet supplemented with 6g MOL/kg food + Vit. C 200 ppm (T5) had the highest FI, BW, BWG, and the best FCR. It is well recognized that several plants contain naturally occurring antioxidants such vitamin C, flavonoids, tocopherol, and other phenolic substances. Rich in vitamins A and C, Moringa oleifera leaves are used externally for wound healing, believed to have purgative qualities, aid in digestion, and be beneficial in catarrhal ailments (The Wealth of India, 1962, A Dictionary of Indian Raw Materials and Industrial Products).

Large concentrations of vitamin C, E, A, and B, carotene, polyphenols, calcium, iron, phosphorus, and protein can all be found in moringa leaves (Murro et al., 2003). The present study suggests that the observed improvement could be attributed to a decrease in the amount of bacteria causing intestinal disease and an enhancement in the intestinal lumen's condition, leading to better nutritional absorption and utilization. Rich in nutrients, moringa helps hens reach their full genetic potential by enhancing their ability to digest other foods and suppressing the growth of mildew and harmful bacteria (Gaia, 2005). Polyphenols including catechol, methyl gallate, ellagic acid, kaempferol, quercetin, and gallate are found in MOL, according to Atawodi (2010). The oxidative stability of chicken meat was successfully increased by adding supplements to the broiler diet that contained moringa (Qwele et al., 2013). The findings concur with those of Donkor et al. (2013) and Teteh et al. (2013), who reported that broilers given MOL had a considerably higher BWG than broilers fed a diet without MOL. Furthermore, the increase in FI of chickens fed MOL diets agrees with the findings of Melesse et al. (2011), who noted a greater FI for broiler chickens fed diets.

Additionally, BWG and FCR were highest (P<0.05) in birds supplemented with MOL (0.1-2.5%), according to Nkukwana et al. (2014). This suggests that birds fed meals based on MOL had improved capability for using nutrients, presumably as a result of becoming larger as inclusion levels increased (Onunkwo and George, 2015). El Tazi (2014) discovered that adding 5% MOL to broiler feed

considerably (P<0.05) increased the rates of BWG, FI, and FCR when compared to other experimental diets. However, it's possible that the MOL groups' improved performance was brought about by the vitamin C concentration of MOL, which might mitigate the negative effects of heat stress and promote positive reactions. According to El-Moniary et al. (2010), broiler feed supplemented with vitamin C during summer stress conditions can enhance product performance. When added to poultry feed, moringa enhances immunological response and growth performance (Abou Sekken, 2015). According to Hassan et al. (2016), FI followed the same trend as BWG, increasing considerably (P<0.05) as the MOL level rose. As MOL levels rose, the FCR was reported at better values. Additionally, the addition of MOL lessens the negative effects of heat stress, allowing birds to consume more feed and gain weight. It also demonstrated that the improvement in BWG and FCR could be attributed to either improved CP digestibility or nutrient utilization as a result of flavonoids acting as antioxidants and antibacterials, or to the positive effects of Moringa on the gut microbial environment, which could improve nutrient utilization through better absorption and digestion. According to research by Fouad and El-Rayes (2019), adding MOL to the diet enhanced productive performance.

Table (4):	Effect of	dietary	Moringa	oleifera	leaves	(MOL)	and	Vit.C o	on growth	performanc	e of:
	Japanese	quail un	der subtr	opical cl	limatic	conditio	ons.				

Treatments/ Traits	Control	T1	T2	Т3	T4	Т5	SEM	P- Value
IBW (g)	35.13	34.95	35.05	34.71	34.83	34.94	0.196	0.73
BW (g)	189.67 ^f	214.59 ^e	225.41°	216.45 ^d	226.82 ^b	242.40 ^a	0.191	0.01
BWG (g)	154.54 ^e	179.64 ^d	190.36 ^b	181.75 ^c	191.99 ^b	207.47 ^a	0.447	0.01
FI (g)	553.00 ^d	583.80°	599.90 ^b	585.43°	605.27 ^b	623.23ª	2.003	0.01
FCR (g feed/g gain)	3.62 ^a	3.23 ^b	3.14 ^c	3.21 ^b	3.15 ^c	3.00 ^d	0.015	0.01

^{A, b, c} Means with the different letters in the same row are significantly different ($P \le 0.05$). TI = (3g MOL/kg diet), T2 = (6g MOL/kg diet), T3 = (Vit.C 200 ppm), T4 = (3g MOL/kg diet + Vit. C 200 ppm), T5 = (6g MOL/kg diet + Vit.C 200 ppm), IBW = Initial body weight, BW = Body weight, BWG = Body weight gain, FI = Feed intake, FCR = Feed conversion ratio (g feed/g gain)

Carcass characteristics:

Under subtropical climate circumstances, the data given in Table (5) demonstrated the impact of dietary sources of MOL and vitamin C supplementation on the relative weights of dressing, liver, gizzard, heart, spleen, and intestinal. The internal organ relative weight of the treated groups of Japanese quail grew significantly (P<0.01) increased with vitamin C and an increase in MOL level, and was greater than that of the control group, according to the results. Furthermore, when compared to other dietary treatments and control groups, the results indicated that birds fed a diet supplemented with 6g MOL/kg food + Vit. C 200 ppm (T5) had the best dressing percentage. The rise in body weight at slaughter may be related to the improvement in dressings brought about by including MOL supplement in the meal. The improvement in dressings as a result of adding MOL supplements in the diet could be attributed to the increase in body weight at slaughter as it has been suggested that higher value of live body weight is attracted and could be related to the physiological condition of high value of carcass weight (Ojewole et al., 2000)

Table (5): Effect of dietary *Moringa oleifera* leaves (MOL) and Vit.C on some relative carcass characteristics of Japanese quail under subtropical climatic conditions.

Traits, %	Control	T1	T2	Т3	T4	Т5	SEM	P- Value
Dressing	64.46 ^c	65.94 ^{bc}	76.26 ^a	67.48 ^b	76.72ª	77.519 ^a	0.520	0.01
Liver	1.74 ^e	1.80 ^d	2.02 ^c	1.81 ^d	2.05 ^b	2.16 ^a	0.008	0.01
Gizzard	1.79 ^d	1.82 ^c	2.49 ^b	1.84 ^c	2.50 ^b	2.54 ^a	0.005	0.01
Heart	0.69 ^e	0.72 ^d	0.83 ^b	0.76 ^c	0.85 ^a	0.86^{a}	0.004	0.01
Spleen	0.040 ^c	0.045 ^{bc}	0.054 ^a	0.046 ^b	0.056 ^a	0.055 ^a	0.001	0.01
Intestinal	2.84 ^d	2.86 ^{cd}	2.95 ^b	2.88 ^c	2.98b	3.04 ^a	0.008	0.01

^{*a, b, c*} Means with the different letters in the same row are significantly different ($P \leq 0.05$).

T1 = (3g MOL/kg diet), T2 = (6g MOL/kg diet), T3 = (Vit.C 200 ppm), T4 = (3g MOL/kg diet + Vit.C 200 ppm), T5 = (6g MOL/kg diet + Vit.C 200 ppm).

Several studies have revealed that the optimum response in terms of growth performance, feed efficiency, FCR, carcass quality, and survival rate in broilers under heat stress appeared with average supplements of 200-250 mg/kg Vit. C. (Abidin et al., 2013, Sahin et al., 2002 and Seven et al., 2010). This important production of immune cells may also be due to the antioxidant activity of some components of MOL such as Vit. C and E (Rocha et al., 2010), phenols especially flavonoids (Diallo et al., 2009), and the plant's capacity of polysaccharides to modulate the immune system (Dong et al., 2007). This result was also confirmed by Olugbemi et al. (2010) who reported improving broiler gut health with MOL. Karthivashan et al. (2015) reported that broiler feed 0, 0.5, 1.0, or 1.5% of MOL extract supplementation had a significantly (P < 0.05) higher dressing rate compared to the control group. while 1.0% MOL showed the highest dressing rate. Kout Elkloub et al. (2015) reported that MOL significantly reduced abdominal fat in Japanese quail, and indicated that 0.2, 0.4, or 0.6% MOL improved spleen ratio without significant differences compared to the control group. Fouad and El-Rayes (2019) reported that adding moringa leaves to the diet improved the relative weight of chicks from the treated groups compared to the control group. In addition, birds treated with 7g MOL/kg had the best carcass weight compared to the control group and the other treatments. While Juniar et al. (2008) reported that the inclusion of MOL up to 10% did not produce significant effects (P<0.05) on carcass weight. Also, Ayssiwede et al. (2011) found that up to 24% of MOL had no effect on the organs and carcass characteristics of native Senegalese chickens. Moreover, Zanu et al. (2012) reported that no effect of MOL was found on all carcass traits.

Blood constituents:

WBC(10³/mm3)

Heterophils (%)

Eosinophils (%)

Monocytes (%)

IgG (mg/100 ml)

Igm (mg/100 ml)

TAC(mg/dl)

Lymphocytes (%)

116.73

53.10

35.17

2.63

8.53

411.21^d

828.53^d

228.09^d

Tables 6 and 7 report the results of the hematological parameters of Japanese quails fed different doses of MOL and vitamin C supplementation in subtropical climate conditions. The findings showed that quail metrics RBCs, Hb, and -PCV-hematological characteristics were considerably (P<0.01) elevated by MOL and vitamin C supplementation. On WBCs, however, no discernible changes were found as compared to the control. Furthermore, there was a substantial (P<0.01) increase in antioxidant indices like TAC and immunological indices like IgG and IgM of quail parameters with vitamin C, and the MOL level increased more than that of the control group.

a C	onditions.	mulces a	na minun	le muices	of Japane	se quan	under subt	ropical c	matic
Traits		Control	T1	T2	Т3	T4	T5	SEM	Р-
									value
RBC (10 ⁶ /m)	m3) 2.	.78 ^d	2.89°	3.63 ^b	2.92°	3.66 ^b	3.76 ^a	0.009	0.01
HB(g/dl)	14	4.16 ^d	16.03 ^c	17.17 ^{bc}	17.80 ^{ab}	18.90 ^a	19.10 ^a	0.313	0.01
PCV%	34	4.83 ^e	38.27 ^d	41.00 ^c	40.23°	43.47 ^b	44.97 ^a	0.438	0.01

118.06

53.36

34.96

2.70

8.97

421.07^b

832.16^c

239.10 °

118.13

54.30

33.97

2.83

8.90

422.01^b

845.73^b

247.97^b

118.36

54.10

34.17

2.90

8.83

429.23^a

854.97^a

257.13 a

0.513

0.484

0.543

0.079

0.486

0.842

0.859

1.008

0.266

0.278

0.691

0.243

0.842

0.01

0.01

0.01

117.70

53.80

34.80

2.86

9.10

422.75^b

842.90^b

246.37 ^b

Table (6): Effect of dietary Moringa oleifera leaves (MOL) and Vit.C on some hematological blood, ntiovident indices and immune indices of Io ail undan auhtuani

^{*a, b, c*} Means with the different letters in the same row are significantly different ($P \leq 0.05$).

117.46

52.83

34.86

2.80

9.50

418.25^c

831.33^{cd}

237.30^c

TI= (3g MOL/ kg diet), T2= (6g MOL/ kg diet), T3= (Vit.C 200 ppm), T4= (3g MOL/ kg diet+ Vit. C 200 ppm), T5= (6g MOL/kg diet+ Vit.C 200 ppm), RBC=red blood cell; PCV=packed cell volume; WBC=white blood cell; HB= Hemoglobin TAC = total antioxidant capacity; Immunoglobulin G (IgG), IgM = Immunoglobulin M.

Furthermore, the findings demonstrated that, in comparison to the other treatments and the control group, birds fed a diet supplemented with 6g MOL/kg food + Vit. C 200 ppm (T5) had the best hematological features, antioxidant indices, and immunological indices. The blood biochemical parameters of quail, including total protein, albumin, globulin, HDL, T4, and T3 hormone, significantly improved (P<0.01) when Moringa oleifera leaves and vitamin C supplementation were given.

Conversely, total serum lipids, cholesterol, glucose concentration, LDL, ALT, AST, creatinine, and urea significantly decreased (P<0.01) when compared to the control group. Higher numbers suggest better health and greater capacity for these functions since red blood cells are important for producing hemoglobin and transferring oxygen and carbon dioxide in the blood (Olugbemi et al., 2010).

Table (7): Effect of dietary Moringa oleifera leaves (MOL) and Vit.C on some blood constituents of
Japanese quail under subtropical climatic conditions.

Traits	Control	Т1	ТĴ	Т2	Т4	т <i>5</i>	SEM	Р-
	Control	11	12	13	14	15	SEM	value
Total protein (g/dl)	3.33 ^e	3.61 ^d	4.00 ^b	3.64 ^c	4.01 ^b	4.31 ^a	0.007	0.01
Albumin (g/dl)	1.40 ^d	1.64 ^c	1.71 ^b	1.66 ^c	1.71 ^b	1.98ª	0.007	0.01
Globulin (g/dl)	1.93 ^d	1.97 ^{cd}	2.29 ^b	1.98°	2.30 ^{ab}	2.33ª	0.011	0.01
Glucose, (mg/dl)	270.67ª	263.00 ^b	220.33°	261.00 ^b	218.33 ^{cd}	216.33 ^d	0.651	0.01
Total lipids (mg/dl)	294.00 ^a	286.67 ^b	267.33°	285.67 ^b	267.00 ^c	266.00 ^c	0.788	0.01
Cholesterol(mg/dl)	265.33ª	226.00 ^b	203.66 ^d	221.00 ^c	201.00 ^d	197.00 ^e	1.011	0.01
HDL (mg/dl)	47.00 ^d	56.00 ^c	72.67 ^b	57.67°	73.00 ^b	88.33ª	0.764	0.01
LDL (mg/dl)	122.33ª	119.00 ^b	112.00 ^c	119.67 ^b	112.33°	98.00 ^d	0.602	0.01
AST (U/L)	58.00 ^a	51.00 ^b	46.00 ^c	50.00 ^b	45.00 ^c	42.67 ^d	0.628	0.01
ALT (U/L)	23.55ª	21.72b	21.20b	21.67 ^b	20.70 ^b	18.37 ^c	0.526	0.01
Creatinine (mg/dl)	0.197ª	0.180^{ab}	0.163 ^{bc}	0.173 ^{ab}	0.156 ^{bc}	0.143 ^d	0.006	0.01
Urea (mg/dl)	16.33ª	14.00 ^b	12.33 ^{bc}	14.33 ^{ab}	12.00 ^{bc}	11.00 ^c	0.638	0.01
T4 (ng/ml)	8.89 ^c	9.37 ^{bc}	10.77 ^{ab}	10.67 ^{ab}	10.90 ^a	12.10 ^a	0.342	0.01
T3 (ng/ml)	1.86 ^e	1.97°	2.17 ^b	2.06 ^c	2.18 ^b	2.33 ^a	0.014	0.01

^{*a, b, c*} Means with the different letters in the same row are significantly different ($P \leq 0.05$).

T1= (3g MOL/ kg diet), T2= (6g MOL/ kg diet), T3= (Vit.C 200 ppm), T4= (3g MOL/ kg diet+ Vit. C 200 ppm), T5= (6g MOL/ kg diet+ Vit.C 200 ppm), AST=aspartate amino transferase; ALT=alanine amino transferase; HDL=high-density lipoprotein; LDL=low-density lipoprotein; T3= triiodothyronine; T4=thyroxine;

Moringa oleifera leaves have a beneficial effect on immune responses (Olugbemi et al., 2010). These results were in agreement with Onu and Aniebo (2011), which found that PCV was significantly higher in birds fed 2.5 or 5% MOL compared to control birds. Sameh (2017) showed that chicks fed 5% MOL had significantly higher PCV than the control group and those supplemented with 8% MOL, while birds fed 3% MOL had average medium and no difference from others. Makanjuola et al. (2014) noted that the decrease in ALT activity observed in birds on the MOL diet could indicate that it has properties that can promote liver health. In addition, it has been used as a natural antioxidant due to its antioxidant activity and is rich in antioxidant compounds such as ascorbic acid, flavonoids, carotenoids, and phenols (Vongsak et al., 2014). Moringa oleifera extract (in both the mature and tender leaves have strong antioxidant activity against free radicals, provides significant protection against oxidative stress, and prevents oxidative damage to main biomolecules (Sreelatha and Padma, 2009). Kout Elkloub et al. (2015) reported that MOL significantly reduced plasma cholesterol, especially LDL, and improved the immune system and blood components in Japanese quail. Hassan et al. (2016) reported that MOL addition up to 0.3% improved physiological parameters and enhanced the ability to resist heat stress conditions for broilers where Hb increased with increasing MOL level, while hematocrit (Ht) values did not affect. Plasma total protein, globulin, and thyroid hormones (T3 and T4) increased significantly (P<0.05) with increasing MOL levels. Albumin was not affected. While the AST was significantly decreased (P<0.05). Found and El-Rayes (2019) reported that supplementation of Moringa leaves at levels of 3, 5, and 7g MOL/kg to the diet improved blood components, hormones, antioxidant indices, and immune parameters.

Bacteriological count:

The results of the intestinal microbial count of Japanese quail are presented in Table (8). All levels of MOL and Vit. C supplementation under subtropical climatic conditions led to a decrease in the total aerobic count, total coliform, and total anaerobic count, compared with the control group. In this study, the beneficial effect on the number of gut microbes may be due to the addition of MOL to bird feed because it contains flavonoid and phenol components, as well as its antimicrobial activity (Luqmans et al., 2012). Moreover, MOL is rich in nutrients that improve the digestion of other foods and reduce the activity of pathogenic bacteria and molds, helping chickens to display their natural genetic potential (Gaia, 2005). Atawodi (2010) noted that MOL contains polyphenols such as catechol, methyl gallate,

Egyptian J. Nutrition and Feeds (2023)

ellagic acid, kaempferol quercetin, and gallate. Supplementation to the MOL diet for broilers was effective in enhancing the oxidative stability of chicken meat (Qwele et al., 2013).

Table (8): Effect of dietary *Moringa oleifera* leaves (MOL) and Vit.C on total anaerobic, total coliform and aerobic count of bacteria in intestine of Japanese quail under subtropical climatic conditions.

Control	T1	T2	Т3	T4	T5	SEM	P-
							value
0.97ª	0.80^{ab}	0.40 ^c	0.77 ^b	0.37°	0.30 ^c	0.056	0.01
11.00 ^a	9.00 ^b	6.00 ^c	9.33 ^{ab}	5.50 ^c	4.80 ^c	0.593	0.01
7.80 ^a	5.53 ^b	4.13 ^{cd}	5.30 ^{bc}	3.87 ^d	1.67 ^e	0.393	0.01
	Control 0.97 ^a 11.00 ^a 7.80 ^a	Control T1 0.97 ^a 0.80 ^{ab} 11.00 ^a 9.00 ^b 7.80 ^a 5.53 ^b	$\begin{array}{c c} \textbf{Control} & \textbf{T1} & \textbf{T2} \\ \hline 0.97^a & 0.80^{ab} & 0.40^c \\ 11.00^a & 9.00^b & 6.00^c \\ 7.80^a & 5.53^b & 4.13^{cd} \end{array}$	$\begin{array}{c c c c c c c c c c c c c c c c c c c $	$\begin{array}{c c c c c c c c c c c c c c c c c c c $	ControlT1T2T3T4T5 0.97^{a} 0.80^{ab} 0.40^{c} 0.77^{b} 0.37^{c} 0.30^{c} 11.00^{a} 9.00^{b} 6.00^{c} 9.33^{ab} 5.50^{c} 4.80^{c} 7.80^{a} 5.53^{b} 4.13^{cd} 5.30^{bc} 3.87^{d} 1.67^{e}	ControlT1T2T3T4T5SEM 0.97^{a} 0.80^{ab} 0.40^{c} 0.77^{b} 0.37^{c} 0.30^{c} 0.056 11.00^{a} 9.00^{b} 6.00^{c} 9.33^{ab} 5.50^{c} 4.80^{c} 0.593 7.80^{a} 5.53^{b} 4.13^{cd} 5.30^{bc} 3.87^{d} 1.67^{e} 0.393

^{A, b, c} Means with the different letters in the same row are significantly different ($P \leq 0.05$).

T1 = (3g MOL/kg diet), T2 = (6g MOL/kg diet), T3 = (Vit.C 200 ppm), T4 = (3g MOL/kg diet + Vit.C 200 ppm), T5 = (6g MOL/kg diet + Vit.C 200 ppm).

CONCLUSION

It can be concluded that MOL supplementation in Japanese quail diets is a good natural source and can be used as a safe alternative to synthetic Vit.C under subtropical climatic conditions.

REFERENCES

- A. P. H. A. American Public Health Association (1985). Standard Methods for the Examination of Water and Wastewater, 16th Ed. APHA, Washington, D.C.
- Abidin, Z. And A. Khatoon (2013). Heat stress in poultry and the beneficial effects of ascorbic acid (vitamin C) supplementation during periods of heat stress. World's Poult. Sci. J., 69: 135–152.
- Abou Sekken, M.S.M. (2015). Performance, immune response and carcass quality of broilers fed low protein diets contained either Moringa Oleifera leaves meal or its extract. J. Am. Sci. 11(6): 153-164.
- Altan, O., A. Pabuccuoglu, A. Altan, S. Konyalioglu and H. Bayraktar (2003). Effect of heat stress on oxidative stress, lipid peroxidation and some stress parameters in broilers. Br. Poult. Sci., 44: 545-550.
- Armstrong, W.D. and C.W. Carr (1964). Physiological chemistry laboratory direction, 3rd edition. Burges publishing, Minneololis, Minnesota, USA.
- Atawodi, S.E. (2010). Nigerian foodstuffs with prostate cancer chemopreventive polyphenols. Infect Agent Cancer Sep.23:6 Suppl.,2:S9.
- Ayssiwede, S.B., A. Dieng, H. Bello, C.A.M. Chrysostome, M.B. Hane, A. Mankor, M. Dahouda, M.R. Houinato, J.L. Hornick and A. Missohou (2011). Effects of Moringa oleifera (Lam.) leaves meal incorporation in diets on growth performances, carcass characteristics and economics results of growing indigenous Senegal chickens. Pakist. J. Nut. 10 (12): 1132-1145.
- Bogin, E., and P. Keller (1987). Application of clinical biochemistry to medically relevant animal models and standardization and quality control in animal biochemistry. J. Clin. Chem. Clin. Biochem, 25, 873-878.
- Darras, V. M., A. Vanderpooten, L. M. Huybrechts, L. R. Berghman, E. Dewil, E. Decuypere and E. R. Kühn (1991). Food intake after hatching inhibits the growth hormone induced stimulation of the thyroxine to triiodothyronine conversion in the chicken. Hormone and metabolic research, 23(10), 469-472.
- Diallo,A., K. Eklu-Gadegkeku, T. Mobio, S. Moukha, A. Agbonon, K. Aklikokou, E.E. Creppy and M.Gbeasso (2009). Protective effect of Moringa oleifera Lam. and Lannea kerstingii extracts against cadmium and ethanol-induced lipid peroxidation J. of Pharm. and Toxico., 4. issue: 4 p: 160-166.
- Doba, T., G.W. Burton and K.U. Ingold (1985). Antioxidant and co-antioxidant activity of vitamin C. The effect of vitamin C, either alone or in the presence of vitamin E or a water-soluble vitamin E

analogue, upon the peroxidation of aqueous multilamellar phospholipid liposomes. Biochim. Biophys. Acta, 835, 298–303.

- Dong, X.F., W.W. Gao, J.M. Tong, H.Q. Jia, R.N. Sa and Q. Zhang (2007). Effect of polysavone (alfalfa extract) on abdominal fat deposition and immunity in broiler chickens. Poult. Sci., 86: 1955-1959.
- Donkor, A.M., R.L.K. Glover, D. Addae and K.A. Kubi (2013). Estimating the nutritional value of the leaves of Moringa oleifera on poultry. Food Nut. Sci., 4: 1077-1083.
- Doumas, B. T., W. A. Watson and H. G. Biggs (1971). Albumin standards and the measurement of serum albumin with bromcresol green. Clinica chimica acta, 31(1), 87-96.
- Duncan, D. B., (1955). Multiple range and multiple F test. Biometrics. 11: 1-42.
- Ebenebe, C.I., C.O. Umegechi, Aniebo and B.O. Nweze (2012). Comparison of haematological parameters and weight changes of broiler chicks fed different levels of Moringa oleifera diet. Int. J. Agric. Biosci., 1: 23-25.
- El Tazi, S.M.A. (2014). Effect of feeding different levels of Moringa oleifera leaf meal on the performance and carcass quality of broiler chicks. Int. J. Sci. Res., 3: 147-151.
- El-Moniary, M.M.A., A.A. Hemid, I. El-Wardany, A.E. Gehad and A. Gouda (2010). The effect of early age heat conditioning and some feeding programs for heat-stressed broiler chicks on: 1-Productive performance. World J. Agric. Sci., 6: 689-695
- Faisal, B.A. (2008). Immunocompetence, hepatic heat shock protein 70 and physiological responses to feed restriction and heat stress in two body weight lines of Japanese quail. Int. J. of Poult. Sci., 7 (2): 174-183.
- Fouad W. And T. K. El-Rayes (2019). Effect of using Moringa Oleifera leaves on Productive performance and some physiological parameters of Japanese quail. Egypt. Poult. Sci. Vol. (39)(I): (193-205).
- Franchini, A., F. Sirri, N. Tallarico, G. Minelli, N. Iaffaldano and A. Meluzzi (2002). Oxidative stability and sensory and functional properties of eggs from laying hens fed supranutritional doses of vitamins E and C. Poult. Sci., 81:1744–1750.
- Frings, C.S., T.W. Fendly, R.T. Dunn and C.A. Queen, (1972). Improved determination of total serum lipids by the Sulfo-phospho-vanlin reaction. Clin. Chem., 18:673-674.
- Gaia, S. (2005). Wonder tree 100 facts moringa fact 04 exceptional animal feed moringa as livestock feed & pet food. Moringa Mission Trust. Available at: http:// gaiathelivingplanet.blogspot.com/200 5/06/wondertree-100-facts-moringafact-04.html (Accessed 31 October 2013).
- Hassan, H.M.A., M.M. El-Moniary, Y. Hamouda, Eman F. El-Daly, Amani W. Youssef and Nafisa A. Abd El-Azeem (2016). Effect of different levels of Moringa oleifera leaves meal on productive performance, carcass characteristics and some blood parameters of broiler chicks reared under heat stress conditions. Asian J. of Anim. and Vet. Advances, 11: 60-66.
- Husdan, H., and A. Rapoport (1968). Estimation of creatinine by the Jaffe reaction: A comparison of three methods. Clin Chem Mar;14(3):222-38.
- Juniar, I.; Widodo, E. and Sjofjan, O. 2008. Effect of Moringa oleiferaleaf meal in feed on broiler production performance. J. Ilmuil. Petern. Brawij. 18: 238–242. Karthivashan, G.; Arulselvan, P.; Alimon, A.; Ismail, I.S.; and Sharida Fakurazi, S. (2015). Competing Role of Bioactive Constituents in Moringa oleifera Extract and Conventional Nutrition Feed on the Performance of Cobb 500 Broilers. BioMed Research International. Article ID 970398, 13 pages.
- Koracevic, D., G. Koracevic, V. Djordjevic, S. Andrejevic and V. Cosic (2001). Method for the measurement of antioxidant activity in human fluids. Journal of clinical pathology, 54(5), 356-361.
- Kout Elkloub, M. EL. Moustafa.; Riry, F.H. Shata.; Mousa, M.A.M.; Hanan, A.H.Alghonimy and Youssef, S.F. 2015. Effect of using moringa oleifera leaf meal on performance of Japanese quail. Egypt. Poult. Sci. Vol. (35): 1095- 1108.
- Luqmans, S., S. Srivastava, R. Kumar, A.K. Maurya and D. Chanda (2012). Experimental assessment of Moringa oleifera leaf and fruit for its antistress, antioxidant and scavenging potential using in vitro and in vivo assays. Evidence-Based Complement. Altern. Med. 10.1155/2012/519084
- Makanjuola, B. A., O. O. Obi, T. O. Olorungbohunmi, O. A. Morakinyo, M. O. Oladele-Bukola and B. A. Boladuro (2014). Effect of Moringa oleifera leaf meal as a substitute for antibiotics on the performance and blood parameters of broiler chickens. Livestock Research for Rural Development 26 (8).

- Maurice, D., S. Lightsey and J.Toler (2004). Ascorbic acid biosynthesis in hens producing strong and weak eggshells. Br. Poult. Sci., 45, 404–408.
- Melesse, A., S. Maak, R. Schmidt, R. Schmidtc and G. von Lengerken (2011). Effect of long-term heat stress on some performance traits and plasma enzyme activities in Naked-neck chickens and their F1 crosses with commercial layer breeds. Livestock Sci., 141: 227-231.
- Melesse, A., Y. Getye, K. Berihun and S. Banerjee (2013). Effect of feeding graded levels of Moringa stenopetala leaf meal on growth performance, carcass traits and some serum biochemical parameters of Koekoek chickens. Livestock Sci., 157: 498-505.
- Micini, G., A. O. Garbonara and H. Harmans (1965). Immunochemical quantitation of antigen by single radial immunodiffusion. Immunochemistry 2, 235–241.
- Mujahid, A., Y. Akiba- and M. Toyomizu (2009). Olive oil-supplemented diet alleviates acute heat stressinduced mitochondrial ROS production in chicken skeletal muscle. Am. J. Physiol. Regul. Integr. Comp. Physiol. 297:R690–R698.
- Murro, J.K., V.R.M. Muhikambele and S.V. Sarwatt (2003). Moringa oleifera leaf meal can replace cotton seed cake in the concentrate mix fed with Rhodes grass (Chloris gayana) hay for growing sheep. Livestock Res. Rural. Develop. 15(11).
- Nkukwana, T.T., V. Muchenje, E. Pieterse, P.J. Masika, T.P. Mabusela, L.C. Hoffman and K. Dzama (2014). Effect of Moringa oleifera leaf meal on growth performance, apparent digestibility, digestive organ size and carcass yield in broiler chickens. Livestock Sci., 161: 139-146.
- NRC (1994). Nutrient Requirement of Domestic Animal, Nutrient Requirement of Poultry . Second Edition National Academy of Science Washington D.C., USA.
- Ojewole, G.S., S.N. Uka and F.Onyenucheya (2000). Comparative carcass characteristics of indigenous poultry feds different agro-industrial by-product, Tropical J.I of Anim. Sci., 3(2): 159 -161.
- Olugbemi T. S., S. K. Mutayoba and F. P. Lekule (2010). Effect of Moringa (Moringa oleifera) inclusion in cassava-based diets fed to broiler chickens. Int. J. of Poult. Sci., 9:363–367. http://www.pjbs.org/ijps/fin1681.pdf
- Onu, P.N. and A.O. Aniebo (2011). Influence of Moringa oleifera leaf meal on the performance and blood chemistry of starter broilers, Nigeria. Int. J. Food Agric. Vet. Sci., 1 (1): 38-44.
- Onunkwo, D.N. and O.S. George (2015). Effects of Moringa oleifera leaf meal on the growth performance and carcass characteristics of broiler birds. IOSR J. Agric. Vet. Sci., 8: 63-66.
- Patton, C. J., and S. R. Crouch (1977). Spectrophotometric and kinetics investigation of the Berthelot reaction for the determination of ammonia. Analytical chemistry, 49(3), 464-469.
- Puthpongsiriporn, U., S.E. Scheideler, J.L. Sell and M.M. Beck (2001). Effects of vitamin E and C supplementation on performance, in vitro lymphocyte proliferation, and antioxidant status of laying hens during heat stress. Poult. Sci., 80:1190–1200.
- Qwele, K., V. Muchenje, S.O. B. Oyedemi, B. Moyo and P.J. Masika (2013). Effect of dietary mixtures of moringa (Moringa oleifera) leaves, broiler finisher and crushed maize on anti-oxidative potential and physicochemical characteristics of breast meat from broilers African J. of Biotec., 12(3): 290-298.
- Reitman, S., and S Frankel (1957). A colorimetric method for the determination of serum glutamic oxalacetic and glutamic pyruvic transaminases. American journal of clinical pathology, 28(1), 56-63.
- Rocha, J.S.R., L.J.C. Lara, N.C. Baiao, R.J.C. Vasconcelos, V.M. Barbosa, M.A. Pompeu and M.N.S. Fernandes (2010). Antioxidant properties of vitamins in nutrition of broiler breeders and laying hens. World's Poult. Sci. J., 66: 261-270.
- Sahin, K., N. Sahin and S. Yaralioglu (2002a). Effects of vitamin C and vitamin E on lipid peroxidation, blood serum metabolites and mineral concentrations of laying hens reared at high ambient temperature. Biol. Trace Elem. Res., 85: 35-45.
- Sahin, K., N. Sahin, O. Kucuk, A. Hayirli and A.S. Prasad (2009). Role of dietary zinc in heat-stressed poultry: A review. Poult. Sci. 88:2176–2183.
- Sahin, K., N. Sahin, S. Yaralioglu and M. Onderci (2002). Protective role of supplemental vitamin E and selenium on lipid peroxidation, vitamin E, vitamin A, and some mineral concentrations of Japanese quails reared under heat stress. Biol. Trace Elem. Res., 85:59–70.
- Sahin, K., O. Kucuk, N. Sahin and M. Sari (2002b). Effects of vitamin C and vitamin E on lipid peroxidation status, serum hormone, metabolite, and mineral concentrations of Japanese quails reared under heat stress (34°C). Int. J. Vitam. Nutr. Res., 72, 91–100

- Sahin, N., F. Akdemir, C. Orhan, O. Kucuk, A. Hayirli and K.Sahin (2008). Lycopene enriched quail egg as functional food for humans. Food Res. Int. 41:295–300
- Sameh, G.A. (2017). Impact of supplementation of Moringa Oleifera in diet of broiler chicks on their behavior, welfare, performance and immune responses. Alex. J. of Vet. Sci., Vol. 55 (2): 50-59. www.alexjvs.com AJVS.

SAS Institute (2002). SAS/STAT User's guide statistics. SAS institute INC., Cary. NC, USA.

- Seven, I., T. Aksu and P.T. Seven (2010). The effects of propolis on biochemical parameters and activity of antioxidant enzymes in broilers exposed to lead-induced oxidative stress. Asian-Australas J. Anim. Sci., 23: 1482–1489.
- Siddhuraju, P. and K. Becker (2003). Antioxidant properties of various solvent extracts of total phenolic constituents from three different agroclimatic origins of drumstick tree (Moringa oleifera Lam.) leaves. J. Agric. Food Chem., 51:2144-2155.
- Sreelatha, S. and P.R. Padma (2009). Antioxidant activity and total phenolic content of Moringa oleifera leaves in two stages of maturity. Plant Foods Hum. Nut. 64(4): 303- 311.
- Sturkie, P. D.(1986). Heart: contraction, conduction, and electrocardiography. In Avian physiology (pp. 167-190). Springer, New York, NY.
- Teteh, A., E. Lawson, K. Tona, E. Decuypere and M. Gbeassor (2013). Moringa oleifera leave: Hydroalcoholic extract and effects on growth performance of broilers. Int. J. Poult. Sci., 12: 401-405.
- The Wealth of India (A Dictionary of Indian Raw Materials and Industrial Products); Raw Materials, Vol. VI, L-M; Council of Scientific and Industrial Research: New Delhi, India, 1962; pp 425-429.
- Trinder, P.(1969). Determination of glucose in blood using glucose oxidase with an alternative oxygen acceptor. Annals of clinical Biochemistry, 6(1), 24-27.
- Tuzcu, M., N. Sahin, M. Karatepe, G. Cikim, U.;Kilinc, and K. Sahin (2008). Epigallocatechin-3-gallate supplementation can improve antioxidant status in stressed quail. Br. Poult. Sci., 49:643–648.
- Vongsak, B., P. Sithisam and W. Gritsanapan (2014). Simultaneous HPLC quantitative analysis of active compounds in leaves of Moringa oleifera Lam. J. Chromatogr. Sci., 52(7): 641-645.
- Wang, R.R. X.J. Pan and Z.Q. Peng (2009). Effects of heat exposure on muscle oxidation and protein functionalities of pectoralis majors in broiler. Poult Sci., 88:1078–1084.
- Warnick, G. R., V. Benderson and N. Albers (1983). Selected methods. Clin. Chem., 10: 91-99.
- Zanu, H.K., P. Asiedu, M. Tampuori, M. Asada and I. Asante (2012). Possibilities of using Moringa (Moringa oleifera) leaf meal as a partial substitute for fishmeal in broiler chickens diet. J. of Anim. Feed Resour. 2 (1): 70-75.

تأثير اضافة أوراق المورينجا أوليفيرا وفيتامين ج على أداء النمو ومكونات الدم ومضادات الأكسدة والخصائص الهرمونية لطائر السمان الياباني تحت الظروف المناخية شبه الاستوائية

> وليد فؤاد احمد طه¹ ,وايمن يوسف كساب ² , ومحمود عبد الحميد امبادر ¹ ^{لقسم} انتاج الدواجن، كلية الزراعة، جامعة الوادي الجديد، مصر ^يقسم الانتاج الحيواني، كلية الزراعة، جامعة الوادي الجديد، مصر

أجريت هذه التجربة لتقييم تأثير أوراق المورينجا (MOL) كمصدر طبيعي آمن في علائق السمان كبدبل لفيتامين ج المخلق صناعياً تحت الظروف المناخية شبه الاستوائية على الأداء الإنتاجي ومكونات الدم والمناعة والخصائص المضادة للأكسدة والهرمونات وخصائص الذبيحة وعدد البكتيريا في الأمعاء. اشتملت هذه الدراسة على عدد 180 كتكوت سمان ياباني عمر سبعة أيام قسمت عشوائياً على 6 معاملات تجريبية بكل منها 3 مكررات بكل مكرر 10 كتاكيت. تمت تغذية كتاكيت المجموعة الأولى على العليقة الاساسية بدون أي إضافة (كنترول) ، بينما تمت غذيت المجموعات الأخرى الثانية (T1) ، والثالثة (C2) ، والرابعة (T3) ، والخامسة (T4) والسادسة (T5) على العليقة الاساسية مضاف إليها 3 جم من ورق المورينجا (MOL) / كجم علف ، 6 جم (MOL) / كجم علف ، ولندسة (T5) على العليقة الاساسية مضاف إليها 3 جم من ورق المورينجا (MOL) / كجم علف ، 6 جم (MOL) / كجم علف ، فيتامين C (200 جزء في المليون) ، 3 جم (MOL) / كجم علف + فيتامين C (200 جزء في المليون)، و 6 جم (MOL) / كجم علف + فيتامين C (200 جزء في المليون) على التوالي ، وتراوح متوسط درجة الحرارة بين 40 درجة مئوية مع رطوبة نسبية 50٪ خلال فترة التجربة حتى عمر 6 أسابيع.

أظهرت النتائج ان اضافة 6 جم (MOL)/ كجم علف + فيتامين C (200 جزء في المليون) ادى الى زيادة معنوية فى وزن الجسم الحي ومعدل الزيادة فى وزن الجسم والعلف المستهلك مع تحسن الكفاءة الغذائية ونسبة الذبيحة، بالضافى الى تحسن نسب الاعضاء الداخلية للسمان اليابانى تليها فى ذلك المعامله (CT)و (T4) ثم (T3) و (T1) مقارنة بمجموعه الكنترول. كان للمعاملة زيادة ملحوظة (CO-Q)في مكونات الدم والمناعة والخصائص المضادة للأكسدة، وتحسين صفات الدم الكيميائية مثل البروتين الكلي، والألبومين، والجلوبيولين، والبروتينات الدم والمناعة والخصائص المضادة للأكسدة، وتحسين صفات الدم الكيميائية مثل البروتين الكلي، والألبومين، والجلوبيولين، والبروتينات الدهنية عالية الكثافة، وهرمونات الغدة الدرقية بالمقارنة بالكنترول وانخفضت نسبة الدهون الكلية والكوليسترول وتركيز الجلوكوز والبروتينات الدهنية منخفضة الكثافة LDL وأنزيمات الكبد والكرياتينين واليوريا معنويا (P-0.01) مقارنة بمجموعة الكنترول وكذلك لم توجد أي فروق معنوية في خلايا الدم البيضاء بين المعاملات والكريرا المعاد نتجمع عند الكنترول. والكنترول. النوريا معنويا (MOL) مقارنة بالمقارنة بالكنترول وانخفض الكلية مقارنة بمجموعة الكنترول وكذلك لم توجد أي فروق معنوية في خلايا الدم البيضاء بين المعاملات والكنترول. انخفض عدد البكتيريا في الامعاء نتيجة إضافة المعاملات المختلفة مقارنة بمجموعة الكنترول. لذلك يمكن الاستنتاج أنه يمكن اضافة أوراق المورينجا (MOL) الامعاء نتيجة إضافة المعاملات المختلفة مقارنة بمجموعة الكنترول. لذلك يمكن الاستنتاج أنه يمكن المانخية شرول المورينجا المرونيا ميزيا في لعف السمان الياباني كمصدر طبيعي جيدة وبديل آمن بدلاً من فيتامين C

مفاتيح البحث: السمان، أوراق المورينجا، فيتامين ج، أداء النمو، الظروف المناخية شبه الاستوائية، الهرمون.