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Using the Dried Yeast (Saccharomyces cerevisiae) as a Growth Promoter in the Nile Tilapia (Oreochromis niloticus) Diets

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ABSTRACT

This study aimed to assess the impact of incorporating dried yeast (Saccharomyces cerevisiae) on growth performance, feed utilization, body composition, blood parameters, and economical evaluation. 120 fish postacclimatization were randomly assigned to 12 experimental aquariums. In this setup, three aquariams acted as replicates for each treatment, with an average initial weight of 28.55±0.82g. Saccharomyces cerevisiae (Sc) levels of 0, 0.4, 0.8, and 1.2% (equivalent to 0, 4, 8, and 12g for diets D₁, D₂, D₃, and D₄, respectively) were used. The feeding trial lasted 56 days. The results revealed that diets varied in crude protein (CP) from 30.15 to 30.80% and gross energy from 4543 to 4559kcal/ kg DM. Mortality rates were 6.67% in the control and zero in the other groups. Protein efficiency ratio (PER) increased with 12g Sc/kg⁻¹ diets. Serum proteins rose at this level, while ALT, AST, and uric acid peaked at 4g Sc/kg. Body composition changed, where moisture, crude protein, and ash increased, whereas ether extract and growth energy decreased. Energy retention (ER)% decreased, while protein productive value (PPV)% was enhanced. Net improvements of 6.80, 9.47, and 19.03% were evident in D2, D3, and D4, respectively, compared to controls. In conclusion, Saccharomyces cerevisiae acts as a growth promoter, especially at 12g/kg⁻¹ feed. These findings illuminate the potential benefits of incorporating Sc in fish diets for enhanced performance and economic gains.

INTRODUCTION

The dry yeast *Saccharomyces cerevisiae* is one of the most common probiotics incorporated into aqua feeds, where the probiotics may improve feed utilization and enhance fish production (**Ramos et al., 2017; Chowdhury & Roy, 2020**). Studies by **Smith et al. (2003)** and **Sørum (2006)** have noted the increased usage of traditional antibiotics in aquaculture feeding regimes over the last few decades. However, **Han et al.**









(2015) observed that probiotics can lead to an improvement in gut health. Additionally, Safari *et al.* (2016) mentioned improved immune responses. Dawood and Koshio (2016) and Dawood *et al.* (2016, 2020a) have reported that probiotics also enhance feed utilization.

In their studies, Abu-Elala *et al.* (2013) and Yang *et al.* (2020) concluded that *Saccharomyces cerevisiae* is a promising probiotic in aqua-farming. Furthermore, Alderman and Hastings (1998) and Teuber (2001) have mentioned that the increased use of antibiotics has led to the prevention and control of bacterial diseases, but has also increased the prevalence of antibiotic-resistant bacteria.

On the other hand, **Kesarcodi-Watson** *et al.* (2008) have documented that the use of probiotics improves performance and feed utilization. Moreover, **Pooramini** *et al.* (2009) noted that *Saccharomyces* enhances performance. Therefore, this study aimed to investigate the effects of *Saccharomyces cerevisiae* at levels of 0, 4, 8, and 12g/kg⁻¹ diet on the productive and economical evaluation of the Nile tilapia fish.

MATERIALS AND METHODS

This study was conducted at the Fish Laboratory of Animal Production Department of Biological and Agriculture Research Institute in collaboration with the Hydrobiology Department of the Veterinary Research Institute, National Research Centre.

1. Experimental unit

A total of 120 fish with an initial body weight of $28.55\pm0.82g$ were used. After acclimatization, the fish were randomly assigned to experimental aquariums. There were 12 aquariums in total, with 10 fish per aquarium, and among these, 3 aquariums served as replicates for each treatment. The fish were placed in aquariums measuring $80\times40\times30$ cm, with a capacity of 60 liters each.

2. Experimental diets

Saccharomyces cerevisiae (Sc) was incorporated in the diets at four levels: 0, 0.4, 0.8, and 1.2%, which corresponded to 0, 4, 8, and 12g of Sc for diets D_1 , D_2 , D_3 , and D_4 , respectively, as shown in Table (1). The experimental diets were administered continuously for 56 days. The fish were hand-fed the tested diets for 56 days from approximately mid-October to mid-December 2023.

3. Growth parameters

Body weight gain (BWG) = Final weight - Initial weight. Survival rate (SR%) = Number of fish at final/ Number of fish at start x100. Specific growth rate (SGR) = [In final weight (g) - In initial weight (g)]/ Experimental days *100

Ingredient	Experimental diet					
	Control	4 g Sc*/kg ⁻¹	8 g Sc*/kg ⁻¹	12 g Sc*/ kg -1	Price	
	zero Sc*	diet	diet	diet	of ton	
	D 1	D ₂	D ₃	D 4	LE	
Composition of tested diet						
Dried yeast	0.00	0.40	0.80	1.20	150000	
Concentration (56% CP)	17.00	17.00	17.00	17.00	25000	
Soybean meal (44% CP)	40.00	40.00	40.00	40.00	33000	
Ground yellow corn (8% CP)	28.00	28.00	28.00	28.00	12500	
Wheat bran (13% CP)	10.00	9.60	9.20	8.80	9800	
Vegetable oil	3.00	3.00	3.00	3.00	50000	
Vitamins and minerals mixture**	2.00	2.00	2.00	2.00	180000	
Price of ten fed (LE)	24692	25253	25814	26.375		

Table 1. Composition of the different experimental diets

Sc*: Dried yeast (Saccharomyces cerevisiae)

** Vit. A (E672) (IU) 876.19, Vit. D3 (IU) 1141.39, Vit. E 114.30, Vit. K3 7.55, Vit. B1 13.71, Vit. B2 11.44, Vit. B6 15.33, Vit. B12 0.03, Niacin 60.96, Calpan 30.48, Folic Acid 3.04, Biotin 0.37, Vit. C 11.44, Selenium 0.27, Manganese 19.04, Iron 9.15, Iodine 0.77, Zinc 76.19, Copper 3.04, Cobalt 0.37, Choline Chloride 457.14.

3.1 Calculation of feed conversion ratio (FCR)

FCR = total dry matter intake, (TDMI) g/ total body weight gain (TBWG), g.

3.2 Calculation of protein efficiency ratio (PER)

(PER) = total body weight gain (TBWG) g/ total crude protein intake (TCPI), g.

3.3 Feed efficiency

Feed efficiency (FE %) = [weight gain (g)/ feed intake (g)]

Protein productive value (PPV %) = $[PR_1 - PR_0 / PI]$ 100.

Where: PR_1 = is the total fish body protein at the end of the experiment.

 PR_0 = is the total fish body protein at the start of the experiment.

PI = Protein intake.

3.4 Energy retention percentages (ER %)

The energy retention percentage was calculated according to the following equation:

Energy retention (ER %) = $E-E_0/E_F \times 100$

Where: E= the energy in fish carcass (kcal) at the end of the experiment.

 E_0 = the energy in fish carcass (kcal) at the start of the experiment.

 E_F = the energy (kcal) in feed intake.

4. Blood measurements

Six fish from each group were chosen randomly, then the blood was isolated using a 3ml syringe after anesthetizing the fish with clove oil (0.5ml L^{-1}). The samples were then centrifuged at 3000rpm for 15 minutes, and the isolated serum was stored at -20°C.

5. Body composition

The evaluation of fish body composition was carried out both before and after the feeding experiment. Initially, 8 fish were used at the start of the trial, followed by the inclusion of 6 fish from each treatment group later on. This assessment was conducted to determine values pertaining to both energy and protein retention.

5.1. Analytical procedures

The diets were analyzed, and the fish body composition was assessed using the methods of **AOAC** (2016). Total protein and albumin levels were determined colorimetrically following the procedures outlined by **Cannon** *et al.* (1974) and **Tietz** (1990). Globulin levels were calculated as the difference between total protein and albumin. Liver function parameters, including alanine aminotransferase and aspartate aminotransferase, were determined using the method outlined by **Reitman and Frankel** (1957). Kidney function parameters, such as uric acid and creatinine, were assessed colorimetrically using standard commercial kits (Bio-diagnostics, Giza, Egypt), following the procedure described by **Tietz** (1990).

6. Calculated data

Gross energy of experimental diets and fish body composition were calculated according to the method of **Blaxter (1968)** and **MacRae and Lobley (2003)**. While, metabolizable energy and protein energy ratio were calculated according to the guidelines of **NRC (2011)**.

7. Statistical analysis

The collected data were subjected to statistical analysis using one-way analysis of variance (ANOVA) as per **SPSS** (2020), with means separated using Duncan's multiple range test (Duncan, 1955).

RESULTS

1. Chemical analysis of the experimental diets

The crude protein percentages ranged from 30.15 to 30.80% among the four tested diets. Gross energy ranged from 4543 to 4559, and metabolizable energy (ME) ranged from 351.37 to 353.94. Additionally, the protein energy ratio varied from 85.18 to 87.66mg CP/Kcal ME among the four tested diets. These values are considered adequate to meet the requirements of the Nile tilapia fish (Table 2).

Ingredient	Tested diet							
-	Control zero Sc*	4 g Sc*/kg ⁻¹ diet	8 g Sc*/kg ⁻¹ diet	12 g Sc*/kg ⁻¹ diet				
	D 1	\mathbf{D}_2	D ₃	D4				
Moisture	8.15	9.74	9.13	9.48				
Dry matter	91.85	90.26	90.87	90.52				
Chemical analysis on DM basis								
OM	93.66	93.23	93.44	93.14				
CP	30.15	30.41	30.65	30.80				
CF	6.55	6.60	6.24	6.85				
EE	4.18	4.15	4.11	4.10				
NFE	52.78	52.07	52.44	51.39				
Ash	6.34	6.77	6.56	6.86				
Gross energy kcal/ kg DM	4559	4543	4553	4543				
Metabolizable energy kcal/ kg DM	353.94	352.39	354.37	351.37				
Protein energy ratio (mg CP/ Kcal ME)	85.18	86.30	86.49	87.66				
So*: Dried youst (Sacharomyous carevisi	(a) OM:	Organia mattar	CD: Cruda p	rotain				

Table 2. Chemical analysis of the different experimental diets

Sc*: Dried yeast (Saccharomyces cerevisiae)OM: Organic matterCP: Crude proteinCF: Crude fiberEE: Ether extractNFE: Nitrogen free extract.

2. Growth and survival ratio

As presented in Table (3), the results indicate that dietary treatments increased the FW, TBWG, ADG, SGR, and SR in the group of fish fed diets with 4, 8, and 12g of Sc/kg⁻¹ diet (D₂, D₃, and D₄, respectively) compared to the control group (D₁). Furthermore, the mortality rate was recorded at 6.67% in the control group (D₁) but was zero in the other three groups of fish (D₂, D₃, and D₄). Generally, with an increasing level of inclusion of *Saccharomyces cerevisiae* in the diet, the mentioned parameters' values tend to show significant improvements.

Table 3. Growth, specific growth rate and survival ratio of different experimental groups

Item						
	Control	4 g Sc*/kg ⁻¹	8 g Sc*/kg ⁻¹	12 g Sc*/kg ⁻¹		Sign.
	zero Sc*	diet	diet	diet	SEM	<i>P</i> <0.05
	\mathbf{D}_1	\mathbf{D}_2	D_3	\mathbf{D}_4		
Number of fish	30	30	30	30	-	-
Initial weight, g (IW)	287	283	288	284	0.82	NS
Final weight, g (FW)	548°	558°	580 ^b	614 ^a	7.90	*
Total body weight gain, g (TBWG)	261 ^d	275°	292 ^b	330 ^a	7.94	*
Duration experimental period			56 days			
Average daily gain, g (ADG)	4.66 ^d	4.91°	5.21 ^b	5.89ª	0.14	*
Specific growth rate (SGR)	0.51 ^d	0.53°	0.55 ^b	0.60 ^a	0.01	*
Number of fish at the starter	30	30	30	30	-	-
Number of fish at the end	28	30	30	30	-	-
Survival ratio (SR)	93.33 ^b	100 ^a	100 ^a	100 ^a	1.12	*
Number of dead fish	2	Zero	Zero	Zero	-	-
Mortality rate percentages	6.67	Zero	Zero	Zero	-	-

a, b, c and d: Means in the same row having different superscripts differ significantly (P < 0.05).

SEM: Standard error of mean NS: Not significant *: Significant at (P < 0.05). Sc*: Dried yeast (*Saccharomyces cerevisiae*).

3. Feed utilization of the different experimental groups

Results of feed utilization (Table 4) show that the values of FI, FCR, CPI, and PER increased when *Saccharomyces cerevisiae* was incorporated into the diets of the Nile tilapia fish. Values of FCR and PER increased with the addition of 4 to 12g per kg⁻¹ of *Saccharomyces cerevisiae* in the diet.

Item						
	Control	4 g Sc*/kg ¹	8 g Sc*/kg ⁻¹	12 g Sc*/kg ⁻¹		Sign.
	zero Sc*	diet	diet	diet	SEM	<i>P</i> <0.05
	D ₁	\mathbf{D}_2	D ₃	\mathbf{D}_4		
TBWG, g	261 ^d	275°	292 ^b	330 ^a	7.94	*
Feed intake (FI), g	864.51 ^{bc}	851.06 ^c	879.96 ^b	898.76^{a}	5.88	*
Feed conversion ratio (FCR)	3.31 ^d	3.09°	3.01 ^b	2.72 ^a	0.06	*
Crude protein %	30.15	30.41	30.65	30.80	-	-
Crude protein intake (CPI) g	260.65 ^c	258.81°	269.71 ^b	276.82 ^a	2.31	*
Protein efficiency ratio (PER)	1.00 ^c	1.06 ^b	1.08 ^b	1.19 ^a	0.02	*

Table 4. Feed	utilization	of the	different	experimental	groups
	will Lation	01 0110	GILLOLOLIU	onpointentent	SICOPD

a, b, c and d: Means in the same row having different superscripts differ significantly (P < 0.05).

FCR: Expressed as g of DM intake/g gain PER: Expressed as g of g gain/g CP intake.

Sc*: Dried yeast (Saccharomyces cerevisiae)

TBWG: Total body weight gain.

4. Biochemical parameters of the different experimental groups

Data from Table (5) indicate that the serum levels of total protein, albumin, and globulin increased in the group of fish that received a diet containing 12g Sc/kg⁻¹ (D₄) compared to the control group. Additionally, dietary treatments decreased the values of the albumin: globulin ratio compared to the control. Concerning the values of liver function, including ALT and AST, the highest values were recorded in the group of fish fed a diet containing 4g Sc/kg⁻¹ diet (D₂). Moreover, uric acid recorded the highest value in the group of fish that received the same diet (D₂). Furthermore, the group of fish that received the highest value of creatinine. Meanwhile, the lowest values of uric acid and creatinine were observed in the group of fish that received D₃, which contained 8g Sc/kg⁻¹ diet.

5. Fish body composition of different experimental groups

Feeding on diets containing *Saccharomyces cerevisiae* resulted in a significant (P < 0.05) increase in fish body composition, including moisture, crude protein, and ash% contents. Meanwhile, values of dry matter, organic matter, ether extract, and growth energy decreased (Table 6).

Parameter		_				
	Control zero Sc*	4 g Sc*/kg ⁻¹ diet	8 g Sc*/kg ⁻¹ diet	12 g Sc*/kg ⁻¹ diet	_	Sign.
	D 1	\mathbf{D}_2	D ₃	D 4	SEM	P<0.05
Total protein (g/dl)	3.15 ^b	3.26 ^b	2.64 ^c	4.17 ^a	0.17	*
Albumin (g/dl)	1.12 ^b	1.07 ^b	0.73 ^c	1.19 ^a	005	*
Globulin (g/dl)	2.03 ^c	2.19 ^b	1.91 ^d	2.98 ^a	0.13	*
Albumin: Globulin ratio	0.55 ^a	0.49 ^b	0.38 ^c	0.40^{c}	0.02	*
Liver function						
AST (Unit/l)	114.6 ^b	127.60 ^a	114.60 ^b	110.10 ^b	2.11	*
ALT (Unit/l)	85.35 ^b	90.06 ^a	88.71ª	84.36 ^b	0.74	*
Kidneys function						
Uric acid (mg/l)	3.80 ^c	6.37 ^a	1.21 ^d	5.47 ^b	0.59	*
Creatinine (mg/l)	0.84 ^b	0.89 ^{ab}	0.10 ^c	0.94 ^a	0.10	*

Ta	able 5	. Blood	parameters	of differen	t experimental	groups
_						

a, b, c and d: Means in the same row having different superscripts differ significantly (P < 0.05).

AST: Aspartate aminotransferase. ALT: Alanine aminotransferase

Sc*: Dried yeast (Saccharomyces cerevisiae).

Table (6. Fish	body	composition	of different	experimental	groups

Item	Initial	Tested diet					
	body composition	Control zero Sc*	4 g Sc*/kg ⁻¹ diet	8 g Sc*/kg ⁻¹ diet	12 g Sc*/kg ⁻¹ diet	_	Sign.
		\mathbf{D}_1	\mathbf{D}_2	D_3	D_4	SEM	P<0.05
Moisture	71.00	71.59°	71.65 ^c	72.94 ^b	73.80 ^a	0.28	*
Dry matter (DM)	29.00	28.41 ^a	28.35 ^a	27.06 ^b	26.20 ^c	0.28	*
Chemical analysis on DM bas	is						
Organic matter (OM)	82.25	85.23ª	84.03 ^b	82.10 ^c	81.16 ^d	0.48	*
Crude protein (CP)	53.60	56.15 ^d	59.78°	62.33 ^b	64.66 ^a	0.95	*
Ether extract (EE)	28.65	29.08 ^a	24.25 ^b	19.77°	16.50 ^d	1.43	*
Ash	17.75	14.77 ^d	15.97°	17.90 ^b	18.84 ^a	0.48	*
Gross energy kcal/100g	572.15	590.60 ^a	565.71 ^b	538.00 ^c	520.43 ^d	8.07	*
Gross energy cal/g DM	5.7215	5.9060 ^a	5.6571 ^b	5.3800°	5.2043 ^d	0.08	*

a, b, c and d: Means in the same row having different superscripts differ significantly (*P*<0.05). Sc*: Dried yeast (*Saccharomyces cerevisiae*).

6. Energy retention and protein productive value percentages

Data from Table (7) demonstrate that the incorporation of *Saccharomyces cerevisiae* in the Nile tilapia fish resulted in a significant (P < 0.05) decrease in their energy retention (ER)% values, while protein productive value (PPV)% was significantly (P < 0.05) increased. It can also be mentioned that protein ER% decreased by 1.66, 9.17, and 4.94% for fish that received D₂, D₃, and D4, respectively, compared to the control (D₁). Meanwhile, values of PPV% were improved by 119.03, 130.53, and 149.81% for fish that were fed D₂, D₃, and D4, respectively, compared to the control.

Item	Tested diet					
	Control	4 g Sc*/kg ⁻¹ 8 g	Sc*/kg ⁻¹	12 g Sc*/kg ⁻¹	-	
	zero Sc*	diet	diet	diet	SEM	Sign.
	D 1	\mathbf{D}_2	D ₃	D 4	-	<i>P</i> <0.05
Initial weight (IW) g	287	283	288	284	0.82	NS
Final weight (FW) g	548 ^c	558°	580 ^b	614 ^a	7.90	*
Energy content in final body fish (cal/g)	5.9060 ^a	5.6571 ^b	5.3800 ^c	5.2043 ^d	0.08	*
Total energy at the end in body fish (E)	3236 ^a	3157°	3120 ^d	3195 ^a	13.07	*
Energy content in initial body fish (cal/g)			5.7215			
Total energy at the start in body fish (E ₀)	1642 ^a	1619 ^b	1648 ^a	1625 ^b	3.85	*
Energy retained in body fish $(E-E_0)$	1594 ^a	1538°	1472 ^d	1570 ^b	13.89	*
Energy of the feed intake (Cal/g feed)	4.559	4.543	4.553	4.543	-	-
Quantity of feed intake	864.51 ^{bc}	851.06°	879.96 ^b	898.76 ^a	5.88	*
Total energy of feed intake (EF)	3941°	3866 ^d	4006 ^b	4083 ^a	24.18	*
Energy retention (ER)%	40.45^{a}	39.78 ^b	36.74 ^d	38.45°	0.43	*
Crude protein% in final body fish	56.15 ^d	59.78°	62.33 ^b	64.66 ^a	0.95	*
Total protein at the end in body fish (PR ₁)	308 ^d	334 ^c	362 ^b	397 ^a	10.08	*
Crude protein % in initial body fish			53.60			
Total protein at the start in body fish (PR ₂)	154	152	154	152	1.29	NS
Protein Energy retained in body fish	154 ^d	182 ^c	208 ^b	245 ^a	10.26	*
$(\mathbf{PR}_3) = (\mathbf{PR}_1 - \mathbf{PR}_2)$						
Crude protein in feed intake (CP%)	30.15	30.41	30.65	30.80	-	-
Total Protein intake (PI), g	260.65 ^c	258.81°	269.71 ^b	276.82 ^a	2.31	*
Protein productive value (PPV)%	59.08 ^d	70.32 ^c	77.12 ^b	88.51ª	3.22	*

Table 7. Energy retention (ER) and protein productive value (PPV) percentages

a, b, c and d: Means in the same row having different superscripts differ significantly (P<0.05).

Sc*: Dried yeast (Saccharomyces cerevisiae).

7. Economical evaluation of different experimental groups

Values of economic evaluation showed that the incorporation of *Saccharomyces cerevisiae* in feed formulations increased the cost of feed formulation from 24.692LE in the control diet (D_1) to 25.253, 25.814, and 26.375LE per ton for the other diets (D_2 , D_3 , and D_4 , respectively). However, a net improvement was realized by 6.80, 9.47, and 19.03% for D_2 , D_3 , and D_4 , respectively, compared to the control that did not contain *Saccharomyces cerevisiae* in the diet (Table 8).

Item	Tested diet						
	Control zero Sc*	4 g Sc*/kg ⁻¹ diet	8 g Sc*/kg ⁻¹ diet	12 g Sc*/kg ⁻¹ diet			
	\mathbf{D}_1	\mathbf{D}_2	D ₃	\mathbf{D}_4			
Costing kg feed (LE)	24.692	25.253	25.814	26.375			
Relative to control (%)	100	102.27	104.54	106.81			
Feed conversion ratio (FCR)	3.31	3.09	3.01	2.72			
Feeding cost (LE) per (Kg weight gain)	81.73	78.03	77.70	71.74			
Relative to control (%)	100	95.47	95.07	87.78			
Net improving in feeding cost (%)	Zero	6.80	9.47	19.03			

Table 8. Economical evaluation of different experimental groups

LE .: Egyptian pound

Diet formulation calculated according to the local prices at year 2023, as presented in Table (1) Feed cost (L.E) FCR×FI. Cost per Kg diet

Sc*: Dried yeast (Saccharomyces cerevisiae).

DISCUSSION

The results of the growth performance and survival ratio of the different experimental groups showed that the inclusion of *Saccharomyces cerevisiae* in fish diets led to an increase in their FW, TBWG, ADG, specific growth rate (SGR), and survival ratio (SR). Previous studies by **Goda** *et al.* (2012) have shown that fish fed yeast exhibited higher growth than the control group, suggesting that *S. cerevisiae* enhances growth performance. Additionally, they noted that the SR of the Nile tilapia at 119 days was recorded at 100%.

The impact of different dietary supplements of yeast on growth has been observed in the rainbow trout, *Oncorhynchus mykiss* (Irianto & Austin, 2002), the Nile tilapia (Ng *et al.*, 2002; Wing-Keong & Chong, 2002; Medri *et al.*, 2005), and common carp, *Cyprinus carpio* (Singh *et al.*, 2011). On the other hand, Tewary and Patra (2011) noted an increase in weight gain and SGR with the addition of *S. cerevisiae* at 5%. Furthermore, Tolan (2006) showed an enhancement in the total gain of the Nile tilapia by increasing the level of dry yeast supplementation from 1 up to 3g/kg diet. Additionally, Diab *et al.* (2006) established that dietary dried yeast fed to the Nile tilapia from 1 up to 5% recorded higher average body weight. The same trend was observed by Tolan and Sherif (2007) for the Nile tilapia fed diets containing 4% *S. cerevisiae*. Goda *et al.* (2012) indicated that the addition of *S. cerevisiae* improved FCR, dietary protein, and energy utilization. Similar results were found by Lara-Flores *et al.* (2003) when *S. cerevisiae* was used in diets for the Nile tilapia. Furthermore, Pooramini *et al.* (2009) stated that the use of probiotics decreased the amount of feed necessary for growth, resulting in reduced production costs.

Values of feed intake FI, FCR, CPI, and PER increased when *Saccharomyces cerevisiae* is included in the diets of the Nile tilapia fish. **Goda** *et al.* (2012) mentioned that the exogenous application of enzymes improved the FCR.

Biochemical analyses are important tools for evaluating the health status of fish and their response to new food additives. Total protein, albumin, and globulin have important immunological and nutritional impacts (**Vali** *et al.*, **2020**). The significant increase in total protein and globulin in the present study, particularly in group D₄ which was fed the highest yeast concentration (12g *Saccharomyces cerevisiae*/kg⁻¹ diet) revealed an improved feed intake and an increase in the immunity status of fish. Similar to this, **Abdel-Tawwab** *et al.* (**2008**) estimated the growth-promoting influence of bakers' yeast on the Nile tilapia at a concentration of 1.0 to 5.0g yeast/kg diet.

Liver enzymes (ALT and AST) represent important biomarkers that indicate any disturbance in liver function (McGill, 2016). According to our observations, the lowest liver enzymes (ALT and AST) were recorded in D₄ (12g Sc*/kg⁻¹ diet), which is the best treatment compared to the control and other treatments. As noted by **Decie and Lewis** (1991) and Goda *et al.* (2012), blood is a pathophysiological reflector of the whole body therefore, blood parameters are important in diagnosing the status of fish health. On the

other hand, Goda *et al.* (2012) found that the albumin: globulin ratio decreased when fish received a diet containing *Saccharomyces cerevisiae* compared to the control diet. A similar observation was reported by Siwicki *et al.* (1994) for rainbow trout, and by Esteban *et al.* (2000), Ortuno *et al.* (2002) and Cuesta *et al.* (2003) for Gilthead Sea bream. Sahoo and Mukherjee (2001) and Abozaid *et al.* (2023) mentioned that the albumin: globulin ratio might be due to the increase in globulin level, which signifies increased protective mechanisms for fish.

Feeding fish on diets containing *Saccharomyces cerevisiae* revealed a significant (P < 0.05) increase in their body composition, including moisture, crude protein, and ash% contents, while organic matter and growth energy decreased compared to the control. **Goda** *et al.* (2012) mentioned that the incorporation of *Saccharomyces cerevisiae* in the Nile tilapia fish showed no differences among treatments for body moisture content (%). On the other hand, Ali and El-Feky (2019) and Abo-State *et al.* (2021) noted that no differences were recorded in whole-body moisture, ether extracts, and ash when prebiotics were used in commercial diets of the Nile tilapia fingerlings. Furthermore, El-Dakar *et al.* (2023) and Abozaid *et al.* (2024) showed significant differences between crude protein, lipids, ash, and gross energy in fish body composition; however, they observed no differences between treatments (P > 0.05).

The inclusion of *Saccharomyces cerevisiae* in the diet of the Nile tilapia fish resulted in a significant (P < 0.05) decrease in their values of energy retention (ER)%, however significantly (P < 0.05) increased the protein productive value (PPV)%. These results are in line with those found by **Abo-State** *et al.* (2021), who revealed differences (P < 0.05) in PPV and ER% between treatments. Additionally, they noted that no differences (P > 0.05) were recorded among various levels of MOS and β -glucan on PPV and ER.

Net revenue showed an improvement of 6.80, 9.47, and 19.03% when *Saccharomyces cerevisiae* was incorporated at 4, 8, and 12g Sc/kg⁻¹ diet for D₂, D₃, and D₄, respectively, compared to the control that did not contain *Saccharomyces cerevisiae* in the diet. **Goda** *et al.* (2012) mentioned that the increasing price of feed is considered one of the most important factors limiting profitability in fish culture. They also showed that the diet containing 1g *Saccharomyces cerevisiae* per 100g diet⁻¹ was the cheapest and recommended for culturing the Nile tilapia fingerlings. Economic efficiency includes both technical efficiency and productive efficiency, along with price efficiency. Thus, economic efficiency is the result of multiplying productive efficiency by price efficiency (Azevedo *et al.*, 2015).

CONCLUSION

The present results indicate that the inclusion of dried yeast (*Saccharomyces cerevisiae*) improves the feed utilization efficiency in the Nile tilapia fingerlings fed diets containing 4, 8, and 12g *Saccharomyces cerevisiae* per kg-1 diet for 56 days.

REFERENCES

- Abareethan, M. and Amsath, A. (2015). Characterization and evaluation of probiotic fish feed. Int. J. Pure Appl. Zool., 3: 148-153.
- Abdel-Aziz, M.F.; Abdel-Tawwab, Y.A.; Sadek, M.F. and Yones, A.M. (2021). Evaluation of use effective microorganisms (EM) with different feeding strategies on growth performance, body chemical composition and economic efficiency of mono sex Nile tilapia *Oreochromis niloticus* juveniles. Aquat. Living Resour., 34: 21.
- Abdel-Tawwab, M.; Abdel-Rahman, A.M. and Ismael, N.E. (2008). Evaluation of commercial live bakers' yeast, *Saccharomyces cerevisiae* as a growth and immunity promoter for Fry Nile tilapia, *Oreochromis niloticus* (L.) challenged *in situ* with Aeromonas hydrophila. Aquaculture, 280(1-4): 185-189.
- **Abo-State, H.A.; El-Monairy, M.M.; Hammouda, Y.A. and Hassan, H.M.A. (2021).** Effect of dietary supplementation of manna oligosaccharide and β-glucan on the performance and feed utilization of Nile tilapia fingerlings. Curr. Sci. Int., 10(1): 226-233.
- Abozaid, H.; Elnady, A.S.M.; Aboelhassan, D.M.; Mansour, H.; Abedo, A.E.;
 Abdelrahman, H.; Ghaly, I.S.; Radwan, H.A.; Abbas, W.T. and Farag, I.M. (2023). Impact of Spirulina platensis as a Dietary Supplement on Growth Performance, Blood Biochemical Parameters, and Expression of Growth-Related Genes in Nile Tilapia (*Oreochromis niloticus*). Egypt. J. Vet. Sci., 54(6): 1256-1277.
- Abozaid, H.; Elnadi, A.S.M.; Mansour, H.; Aboelhassan, D.M.; Awad, E.; El-Nomeary, Y.A.A.; Omar, H.A.A.; Abbas, W.T. and Farag, I.M. (2024). Nutritional Effect of Using a Bioactive Mixture (Lemon, Onion and Garlic) on Growth Performances, Feed Utilization, Immune Status and Gene Expression of Nile tilapia (*Oreochromis niloticus*). Egypt. J. Vet. Sci., 55(6): 1669-1684.
- Abu-Elala, N.; Marzouk, M. and Moustafa, M. (2013). Use of different Saccharomyces cerevisiae biotic forms as immune-modulator and growth promoter for Oreochromis niloticus challenged with some fish pathogens. Int. J. Vet. Sci. Med., 1: 21-29.
- Akhter, N.; Wu, B.; Memon, A.M. and Mohsin, M. (2015). Probiotics and prebiotics associated with aquaculture: a review. Fish Shellfish Immunol., 45(2): 733-741.
- Alderman, D.J. and Hastings, T.S. (1998). Antibiotic use in aquaculture: development of antibiotic resistance-potential for consumer health risk. Int. J. Food Sci. Technol., 33: 139-155.

- Badiaa A.A. and El-Feky A. (2019). Enhancing growth performance and feed utilization using prebiotics in commercial diets of Nile Tilapia (*Oreochromis niloticus*) fingerlings. Egypt. J. Nutr. Feeds, 22 (1): 219-225.
- Ali, H.M.; Ghazalah A.A.; Gehad E.A.; Hammouda Y.A. and Abo-State H.A. (2010). Practical aspects and immune response of probiotics preparations supplemented to Nile Tilapia (*Oreochromis niloticus*) diets, Int. J. Nature Sci., 8(5): 39-45.
- Andani, H.R.R.; Tukmechi, A.; Meshkini, S. and Sheikhzadeh, N. (2012). Antagonistic activity of two potential probiotic bacteria from fish intestines and investigation of their effects on growth performance and immune response in rainbow trout (*Oncorhynchus mykiss*). J. Appl. Ichthyol., 28: 728-734.
- **AOAC.** (2016). Official Methods of Analysis, 18th ed. Association of Official Analytical Chemists, Washington, DC, USA (2005). All content following this page was uploaded by Hayder N. Al-Mentafji on 02 February 2016.
- Azevedo, R.V.D.; Fosse Filho, J.C.; Cardoso, L.D.; Mattos, D.D.C.; Vidal Júnior, M.V. and Andrade, D.R.D. (2015). Economic evaluation of prebiotics, probiotics and symbiotics in juvenile Nile tilapia. Rev. Ciência Agron., 46: 72-79.
- **Blaxter, K.L.** (1968). The energy metabolism of ruminants. 2nd ed. Charles Thomas Publisher. Spring field. Illinois, USA.
- **Blazer, V.S. and Wolke, R.E. (1984).** The effect of ά-tocopherol on the immune response and non-specific resistance factors of Rainbow trout (*Salmo gairdneri* Richardson). Aquaculture, 37: 1-9.
- Cannon, D.C.; Olitzky, I. and Inkpen, J.A. (1974). Proteins. In: Clinical Chemistry, principles and techniques, 2nd ed., R.J. Henery, D.C. Cannon, J.W. Winkelman (eds.), Harper and Row New York, pp. 407-421.
- Chowdhury, M.A. and Roy, N.C. (2020). Probiotic supplementation for enhanced growth of striped catfish (*Pangasianodon hypophthalmus*) in cages. Aquac. Rep., 18: 100504.
- Cuesta, R.A.; Ortun, J.; Esteban, M.A. and Meseguer, J. (2003). Immunostimulant properties of a cell wall-modified whole *Saccharomyces cerevisiae* strain administered by diet to seabream (*Sparus aurata* L.). Vet. Immunol. Immunopathol., 96: 183-192.
- **Dawood, M.A.O. and Koshio, S. (2019).** Application of fermentation strategy in aqua feed for sustainable aquaculture. Rev. Aquac., 12(2): 987-1002.
- Dawood, M.A.O.; Eweedah, N.M.; Khalafalla, M.M.; Khalid, A.; El Asely A.; Fadl S.E.; Amin A.A.; Paray B.A. and Ahamed, H.A. (2020a). *Saccharomyces*

cerevisiae increases the acceptability of Nile tilapia (*Oreochromis niloticus*) to date palm seed meal. Aquac. Rep., 17: 100314.

- **Dawood, M.A.O.** (2016). Effect of Various Feed Additives on the Performance of Aquatic Animals. PhD. Diss., Kagoshima University.
- **Dawood, M.A.O. and Koshio, S. (2016).** Recent advances in the role of probiotics and prebiotics in carp aquaculture: a review. Aquaculture, 454: 243-251.
- Dawood, M.A.O.; Koshio, S.; Ishikawa, M.; El-Sabagh, M.; Esteban, M.A. and Zaineldin, A.I. (2016). Probiotics as an environment-friendly approach to enhance red sea bream, *Pagrus major* growth, immune response and oxidative status. Fish Shellfish Immunol., 57: 170-178.
- **Dawood, M.A.O.; Zommara, M.; Eweedah, N.M. and Helal, A.I. (2020b).** The evaluation of growth performance, blood health, oxidative status and immune-related gene expression in Nile tilapia (*Oreochromis niloticus*) fed dietary nanoselenium spheres produced by lactic acid bacteria. Aquaculture, 515: 734571.
- Debnath, D.; Sahu, N.P.; Pal, A.K; Jain, K.K.; Yengkokpam, S. and Mukherjee, S.C. (2005). Mineral status of *Pangasius pangasius* (Hamilton) fingerlings in relation to supplemental phytase: Absorption, whole body and bone mineral content. Aquac. Res., 36: (326-335.
- **Decie, S.I.V. and Lewis, S.M. (1991).** Practical Haematology (VII Ed), J and A Chuechill Ltd., Livingston, London, Melbourne and NewYork.
- Diab, A.S.; Abdel-Hadi, Y.M.; Ahmed, M.H.; Saber, S.F. and Aboel-Atta, M.E. (2006). Out 255 door study on the use of Echinacea (*Echinacea purpurea*), Marjoram (*Origanum dictamnus*) and yeast (*Saccharomyces cerevisiae*) as feed additives for *Oreochromis niloticus*. Egypt. J. Agric. Res., 84(18): 537-551.
- Dossou, S.; Koshio, S.; Ishikawa, M.; Yokoyama, S.; Dawood, M.A.O.; El Basuini, M.F.; El- Hais, A.M. and Olivier, A. (2018a). Effect of partial replacement of fish meal by fermented rapeseed meal on growth, immune response and oxidative condition of red sea bream juvenile, Pagrus major. Aquaculture, 490: 228-235.
- **Dossou, S.; Koshio, S.; Ishikawa, M.; Yokoyama, S.; Dawood, M.A.O.; El Basuini, M.F.; Olivier, A. and Zaineldin, A.I. (2018b).** Growth performance, blood health, antioxidant status and immune response in red sea bream (Pagrus major) fed Aspergillus oryzae fermented rapeseed meal (RM-Koji). Fish Shellfish Immunol., 75: 253-262.
- El-Dakar, A.; Mouhmed, A.; Elgamal, A.; Amer, M.; Shalaby, S. and Abdel-Aziz, M.F.A. (2023). Effect of re-feeding regime under different stocking density of Nile tilapia, *Oeochromis niloticus* on growth performance, nutritional efficiency and fish body composition. Mediterr. Aquac. J., 10(1): 1-9.

- **El-Haroun, E.R.; Goda, A.S. and Kabir Chowdhury M.A. (2006).** Effect of dietary probiotic biogen supplementation as a growth promoter on growth performance and feed utilization of Nile tilapia *Oreochromis niloticus* (L.). Aquac. Res., 37(14): 1473-1480.
- Esteban, M.A.; Mulero, V.; Cuesta, A.; Ortun, J. and Meseguer, J. (2000). Effects of injecting chitin particles on the innate immune responses of gilthead seabream (*Sparus aurata* L.). Fish Shellfish Immunol., 10: 543-554.
- **Fuller, R. (1992).** History and development of probiotics. In: Fuller, R. (Ed.), Probiotics: the Scientific Basis, vol. 232. Chapman & Hall, London, pp. 1-18.
- Gatesoupe, F.J. (1999). The use of probiotics in aquaculture. Aquaculture, 180: 147-165.
- Giri, S.S.; Sahoo, S.K.; Sahu, A.K. and Meher, P.K. (2003). Effect of dietary protein level on growth, survival, feed utilization and body composition of hybrid Clarias catfish (*Clarias batrachus × Clarias gariepinus*). Anim. Feed Sci. Technol., 104 (1-4): 169-178.
- **Go'ngora, C.M. (1998).** Mecanismos de resistencia bacteriana ante la medicina actual McGraw-Hill, Barcelona, 456 pp.
- Goda, A.M.A.; Mabrouk, H.A.H.; Wafa, M.A and El-Afifi TM. (2012). Effect of using Baker's yeast and exogenous digestive enzymes as growth promoters on growth, feed utilization and hematological indices of Nile tilapia, *Oreochromis niloticus* fingerlings. J. Agric. Sci. Technol., B 2(1B): 15-28.
- **Hai, N.V. (2015).** Research findings from the use of probiotics in tilapia aquaculture: a review. Fish Shellfish Immunol., 45: 592-597.
- Han, B.; Long, W.Q.; He, J.Y.; Liu, Y.J.; Si, Y.Q. and Tian, L.X. (2015). Effects of dietary Bacillus licheniformis on growth performance, immunological parameters, intestinal morphology and resistance of juvenile Nile tilapia (*Oreochromis niloticus*) to challenge infections. Fish Shellfish Immunol., 46: 225-231.
- Helmy, A.M.; Badawi, H.K. and El Bishry, A. (1974). Seasonal variations in the protein composition of blood serum of *Anguilla vulgaris* and *Mugill cephalic*, Egypt. J. Aquat. Res., 4: 369-375.
- **Ibrahem, M.D. (2015).** Evolution of probiotics in aquatic world: potential effects, the current status in Egypt and recent prospectives. J. Adv. Res., 6: 765-791.
- **Irianto, A. and Austin, B. (2002).** Use of probiotics to control furunculosis in rainbow trout, Oncorhynchus mykiss (Walbaum). J. Fish Dis., 25: 333-342.
- Jackson, L.S.; Li, M.H. and Robinson, E.H. (1996). Use of microbial phytase in channel catfish *Ictalurus punctatus* diets to improve utilization of phytate phosphorus. J. World Aqua. Soc., 27(3): 309-313.

- Kesarcodi-Watson, A.; Kaspar, H.; Josie Lategan, M. and Gibson, L. (2008). Probiotics in aquaculture: the need, principles and mechanisms of action and screening processes. Aquaculture, 274: 1-8.
- Klaenhammer, T.D. and Sullen, M.J. (1999). Selection and design of probiotics. Int. J. Food Microbiol., 50: 45-57.
- Lara-Flores, M.; Olvera-Novoa, M.A.; Guzman-Méndez, B.E. and Lopez-Madrid, W. (2003). Use of the bacteria *Streptococcus faecium* and *Lactobacillus acidophilus* and the yeast *Saccharomyces cerevisiae* as growth promoters in Nile tilapia (*Oreochromis niloticus*). Aquaculture, 216: 193-201.
- Lazado, C.C. and Caipang, C.M.A. (2014). Mucosal immunity and probiotics in fish. Fish Shellfish Immunol., 39: 78-89.
- Li, P. and Gatlin, D.M. (2004). Dietary brewers yeast and the prebiotic GroBiotick[™] AE influence growth performance, immune responses and resistance of hybrid striped bass (*Morone chrysops* × *M. saxatilis*) to Streptococcus iniae infection. Aquaculture, 231: 445-456
- Lin, S.; Mai, K. and Tan, B. (2007). Effects of exogenous enzyme supplementation in diets on growth and feed utilization in tilapia, *Oreochromis niloticus* x *O. aureus*. Aquac. Res., 38: 1645-1653.
- MacRae, J. and Lobley, G.E. (2003). Some factors which influence thermal energy losses during the metabolism of ruminants. Livest. Prod. Sci., 9(4): 447-456.
- **Majed, S.H. (2008).** Biomass production of *Saccharomyces cerevisiae* (Baker's Yeast) using the Cactus Cladodes extract as a Culturemedi, M. Sc Thesis, Islamic University of Gaza Faculty of Science, Microbiology Department, pp. 73.
- McGill, M. R. (2016). The past and present of serum aminotransferases and the future of liver injury biomarkers. EXCLI J., 15: 817.
- Medri, V.; Medri, W. and Filho, M.C. (2005). Growth of Nile tilapia (*Oreochromis niloticus* L.) fed diets with different levels of proteins of yeast in tank-net. Acta. Sci. Anim. Sci., 27: 221-227.
- Moss, A.S.; Koshio, S.; Ishikawa, M.; Yokoyama, S.; Nhu, T.H.; Dawood, M.A.O. and Wang, W. (2018). Replacement of squid and krill meal by snail meal (*Buccinum* striatissimum) in practical diets for juvenile of kuruma shrimp (*Marsupenaeus* japonicus). Aquac. Res., 49: 3097-3106.
- Nayak, S.K. (2010). Probiotics and immunity a fish perspective. Fish Shellfish Immunol., 29: 2-14.

- Ng, W.K.; Lim, H.A.; Lim, S.L. and Ibrahim, C.O. (2002). Nutritive value of palme kernel meal pretreated with enzyme or fermented with *Trichoderma koningii* as an dietary ingredient for red hybrid tilapia *Oreochromis* sp. Aquac. Res., 33: 1199-1207.
- Noh, H.; Han, K.I.; Won, T.H. and Choi, Y.J. (1994). Effect of antibiotics, enzymes yeast culture and probiotics on the growth performance of Israeli carp. Korean J. Anim. Sci., 36: 480-486.
- NRC. (2011). National Research Council. Nutrient Requirement of Fish. National Academy Press, Washington, DC, USA.
- Ortuno, J.; Cuesta, A.; Rodriguez, A.; Esteban, M.A. and Meseguer, J. (2002). Oral administration of yeast, *Saccharomyces cerevisiae*, enhances the cellular innate immune response of gilthead seabream (*Sparus aurata* L.). Vet. Immunol. Immunopathol., 85: 41-50.
- Pooramini, M.; Kamali, A.; Hajimoradloo, A.; Alizadeh, A. and Ghorbani, R. (2009). Effect of using yeast (*Saccharomyces cerevisiae*) as probiotic on growth parameters, survival and carcass quality in rainbow trout, *Oncorhynchus* mykiss fry. Int. Aquat. Res., 1: 39-44.
- Ramos, M.A.; Batista, S.; Pires, M.A.; Silva, A.P.; Pereira, L.F.; Saavedra, M.J.; Oz'orio, R.O.A. and Rema, P. (2017). Dietary probiotic supplementation improves growth and the intestinal morphology of Nile tilapia. Animal, 11: 1259-1269.
- **Reitman, S. and Frankel, S. (1957).** Colorimetric determination of glutamic oxaloacetic and glutamic pyruvic transaminases. Am. J. Clin. Pathol., 28: 53-56.
- Safari, R.; Adel, M.; Lazado, C.C.; Caipang, C.M. and Dadar, M. (2016). Hostderived probiotics *Enterococcus casselifavus* improves resistance against *Streptococcus iniae* infection in rainbow trout (*Oncorhynchus mykiss*) via immunomodulation. Fish Shellfish Immunol., 52: 198-205.
- Sahoo, P.K. and Mukherjee, S.C. (2001). Immuonocompressive effect of aflatoxin B1 in Indian major carp (*Labeo rohita*). Comp. Immunol. Micro. Inf. Dis., 24(3): 143-149.
- Salnur, S.; Gultepe, N. and Hossu, B. (2009). Replacement of fish meal by yeast (*Saccharomyces cerevisiae*): effects on Gilthead Sea bream (*Sparus aurata*). J. Anim. Vet. Adv., 8(12): 2557-2561.
- Singh, P.; Maqsood, S.; Samoon, M.H.; Phulia, V.; Danish, M. and Chalal, M. (2011). Exogenous supplementation of papain as growth promoter in diet of fingerlings of *Cyprinus carpio*. Int. Aquat. Res., 3: 1-9.

- Siwicki, A.K.; Anderson, D.P. and Rumsey, G.L. (1994). Dietary intake of immuno stimulants by rainbow trout affects non-specific immunity and protection against furunculosis. Vet. Immunol. Immunopathol., 4: 125-139.
- Smith, V.J.; Brown, J.H. and Hauton, C. (2003). Immunostimulation in crustaceans: does it really protect against infection?. Fish Shellfish Immunol., 15: 71-90.
- Sørum, H. (2006). Antimicrobial drug resistance in fish pathogens. In: Aarestrup FM (Ed) Antimicrobial resistance in bacteria of animal origin. ASM Press, Washington DC, pp. 213-238
- **SPSS.** (2020). Statistical Package for Social Science (Software version: 22.0).
- Suzer, C.; Çoban, D.; Kamaci, H.O.; Saka, Ş.; Firat, K.; Otgucuoğlu, Ö. and Küçüksari, H. (2008). *Lactobacillus spp.* bacteria as probiotics in Gilthead Sea bream (*Sparus aurata*, L.) larvae: effects on growth performance and digestive enzyme activities. Aquaculture, 280: 140-145.
- Talpur, A.D.; Memon, A.J.; Khan, M.I.; Ikhwanuddin, M.; Danish, M.M. and Daniel, AbolMunafi, A.B. (2012). Inhibition of pathogens by lactic acid bacteria and application as water additive multi isolates in early stages larviculture of P. pelagicus (Linnaeus, 1758). J. Anim. Plant Sci., 22: 54-64.
- **Teuber, M. (2001).** Veterinary use and antibiotic resistance. Curr. Opin. Microbiol., 4: 493-499.
- Arup Tewary, A.T. and Patra B.C. (2011). Oral administration of baker's yeast (Saccharomyces cerevisiae) acts as a growth promoter and immunomodulator in *Labeo rohita* (Ham.). J. Aquac. Res. Dev., 2(1).
- **Tietz, N.W. (1990).** Clinical guide to laboratory tests. 2^{*nd*} ed. Philadelphia, WB Saunders. pp. 566.
- **Tizard, I. (1992)**. Veterinary Immunology: An Introduction, 4th ed., W.B. Saunders Co., Philadelphia, London and Toronto PA, USA, 1992, pp. 11-113.
- **Tolan, A.E. (2006).** Evaluation of some feed additives at different levels in diets of Nile tilapia. Egypt. J. Agric. Res., 84(18): 385-402.
- Tolan, A.E. and Sherif, A.H. (2007). Effect of some growth promoters on growth performance of Nile tilapia fingerlings. J. Arab. Aquacult. Sci., 2(1): 89-102.
- Vali, S.; Mohammadi, G.; Tavabe, K.R.; Moghadas, F. and Naserabad, S.S. (2020). The effects of silver nanoparticles (Ag-NPs) sublethal concentrations on common carp (*Cyprinus carpio*): Bioaccumulation, hematology, serum biochemistry and immunology, antioxidant enzymes, and skin mucosal responses. Ecotoxicol. Environ. Saf., 194: 110353.

- Wing-Keong, N.G. and Chong, K.K. (2002). The nutritive value of palm kernel meal and the effect of enzyme supplementation in practical diets for Red Hybrid Tilapia (*Oreochromis* sp.). Asian Fish. Sci., 15: 167-176.
- Wong, M.H.; Tang, L.Y. and Kwok, F.S.L. (1996). The use of enzyme digested soy bean residue for feeding common carp, J. Biol., 9(4): 418-442.
- Yang, X.; Chi, S.; Tan, B.; Nie, Q.; Hu, J.; Dong, X.; Yang, Q.; Liu, H. and Zhang,
 S. (2020). Yeast hydrolysate helping the complex plant proteins to improve the growth performance and feed utilization of Litopenaeus vannamei. Aquac. Rep., 17: 100375.

Zaki, M.A.A.; Alabssawy, A.N.; Nour, A.E.-A.M.; El Basuini, M.F.; Dawood, M.A.O.; Alkahtani, S. and Abdel-Daim, M.M. (2020). The impact of stocking density and dietary carbon sources on the growth, oxidative status and stress markers of Nile tilapia (*Oreochromis niloticus*) reared under biofloc conditions.