

Impact of Phosphorus Sources on Biomass and Pigments Content of *Spirulina platensis*

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ABSTRACT

Spirulina sp. is a photosynthetic blue-green microalga with high-quality bioactive compounds including pigments. This study was designed to investigate various phosphorus sources, such as Na₂HPO₄ and KH₂PO₄, addressing their impact on the biomass and pigments of *Spirulina platensis* using different concentrations. The highest biomass obtained was 2.75 and 1.98g at the concentration of 0.5g/ l for each source, respectively, which is lower compared to the control obtained from Zarrouk Medium. While, the results of chlorophyll a, total chlorophyll, and carotenoids were 25.47, 38.14, and 12.04, respectively, with 0.5g/ l KH₂PO₄ compared with the other source, Na₂HPO₄. Notably, the pigments tested in Zarrouk medium were the best compared to the other sources of phosphorus. It was concluded that carotenoids were not significantly affected by the change in phosphorus concentration. Interestingly, in this respect, it appeared that increasing the production of chlorophyll a resulted in an enhancement in biomass and total chlorophyll production in *Spirulina*. Chl a and total Chl concentrations were significantly different ($P < 0.05$).

INTRODUCTION

Spirulina, a non-toxic blue-green algae found in marine, freshwater, and alkaline lakes, was approved by the Food and Drug Administration in 1981 as a safe food (Verne`s *et al.*, 2019). *Spirulina*, a highly cultivated microalga is well known for its valuable products (Ghaeni *et al.*, 2015, Matos *et al.*, 2017; Shao *et al.*, 2019). It has nutraceutical benefits owing to its richness in dietary compounds including highly valuable proteins, carbohydrates, vitamins, minerals, polyphenolics, polyunsaturated fats, and bio-pigments as chlorophyll group, carotenoids, and allo-phycoyanin (Shao *et al.*, 2019; Mohy El-Din, 2020). In addition, it includes vitamins, nutritional minerals, and vital amino acids (Belay, 2008; Sharoba, 2014; Gutiérrez-Salmeán *et al.*, 2015). These bioactive compounds can be used in potential applications. (Chandi & Gill, 2011; Dubini & Antal, 2015; Rizzo *et al.*, 2015). It can be used in various industries, such as cosmetics, pharmaceuticals, poultry, plant biological stimulants, foods, animal feed, and

fertilizers (Priyadarshani & Rath, 2012; El-Sheekh *et al.*, 2014; Suganya *et al.*, 2016; Soni *et al.*, 2019). *Spirulina* has a balanced composition of all essential components, thus represents a complete food containing high quality protein content ranges from 50 to 70 percent of its dry biomass (Falquet, 1997; Hosseini *et al.* 2013). Besides, it contains antioxidants which are essential for protecting the body from the free radicals, such as polyphenolics, flavonoids, and tocopherols (Kumar *et al.*, 2005; El-Baky *et al.*, 2008, Chu *et al.*, 2010; Michael *et al.*, 2018).

Carotenoids are responsible for the red and yellow colors visible in nature (Vonshak *et al.*, 1996; Habib *et al.*, 2008; Theodore & Georgios, 2013). Phosphorus stress impacts the efficiency of chloroplasts and decreases the process of photosynthesis (Rolead *et al.*, 2013). Phosphorus is a crucial element in algal growth and cell division, as it is a key component of DNA and RNA-phosphorylated sugar (Roopnarain *et al.*, 2014). Phosphorus is involved in algal growth, signal transduction, photosynthesis, and energy transfer through ATP and NADP (Roopnarain *et al.*, 2014; Yaakob *et al.*, 2021). In microalgae, the fast reprocessing of phosphorus in the natural environments has led to obvious limitations in these organisms (Alipanah *et al.*, 2018). Microalgae vary in their phosphorus requirements and can uptake orthophosphate and polyphosphate as macronutrients (Roopnarain *et al.*, 2014; Yaakob *et al.*, 2021).

MATERIALS AND METHODS

1. Microorganisms

Spirulina paltensis strain was collected from the Hydrobiology Lab, National Institute of Oceanography and Fisheries, Alqanater Elkhairya, Egypt, and cultivated in Zarrouk's medium.

2. Media

Spirulina paltensis was grown in Zarrouk medium, containing various elements and trace elements in distilled water. The pH was adjusted to 8.2 using a NaOH solution, as per Zarrouk's medium method (Zarrouk, 1966). Two different phosphorus sources (KH_2PO_4 and Na_2HPO_4) with three levels (0.5, 1.0, and 1.5) g/l were used instead of K_2HPO_4 (the standard source of phosphorus) and tested on Zarrouk medium. *Spirulina paltensis* was maintained in 500-ml Erlenmeyer flasks containing 90% medium and 10% subculture. Conical flasks were cleaned and sterilized using a steam autoclave at 121°C for 20min (Abou-El-Souod *et al.*, 2016).

3. Growth measurement

a. Biomass yield (g/l)

Wet biomasses of *Spirulina paltensis* were taken at 3-day intervals for an 18-day incubation period. Wet biomasses were dried at 105°C for two hours, and their dry weights were estimated (AOAC., 2000).

*b. Estimation of pigments***Chlorophyll group (mg/g dw)**

Chlorophyll a and total Chlorophyll concentrations measured by spectrophotometry and calculated by using equations assessed in the study of Seely *et al.* (1972).

Total carotenoids (mg/g dw)

Total carotenoids were determined by the spectrophotometric method at 450nm. A standard compound was used for preparing the calibration curve according to the AQAS (1990).

RESULTS AND DISCUSSION

Table 1. Dry biomass yield (g/l) and pigments content ($\mu\text{g/g}$ dry weight) of *Spirulina platensis* under different phosphorus sources

Biochemical compound	KH ₂ PO ₄ concentration			Na ₂ HPO ₄ concentration			Zarrouk medium (control)
	0.5 g L ⁻¹	1.0 g L ⁻¹	1.5 g L ⁻¹	0.5 g L ⁻¹	1.0 g L ⁻¹	1.5 g L ⁻¹	
Dry biomass	2.75	2.52	0.46	1.94	1.89	1.05	2.82
Chl a	25.47	16.87	11.77	25.3	14.71	14.28	26.54
Total Chl	37.04	27.38	19.64	36.26	19.03	13.39	37.23
Carotenoids	12.04	12.54	8.48	8.82	7.19	3.54	12.67

Biomass yield

The dry biomass yield (g/l) of *Spirulina platensis* under different phosphorus sources was compared to the control (Zarrouk medium), as shown in Table (1) and Figs. (1, 2). The results revealed that by using 0.5 g/l KH₂PO₄, the highest biomass yield was 2.75g compared to 2.82 g obtained from Zarrouk's medium. Upon using Na₂HPO₄ at levels of 0.5g/ l, the biomass yield was 1.94g compared to the control (2.82g/ l). Therefore, the Zarrouk medium is superior to the two phosphorus sources; however, when considering costs, it is better to use KH₂PO₄ rather than K₂HPO₄ in the Zarrouk medium. Additionally, 0.5g of the KH₂PO₄ compound should be used, while 2.5g is used in Zarrouk. Phosphorus concentration doesn't affect dry biomass, but phosphate limitation is linked to decreased productivity, cell division, and chlorophyll synthesis (Khozin, 2006; Rocha, 2018). Researchers have confirmed that a phosphate level of 0.5g/ l is optimal for improving the biomass yield by *Spirulina platensis* due to a favorable N:P weight ratio. (Costa *et al.*, 2002; Radmann *et al.*, 2007). Actually, Bulgakov and Levich (1999) discovered that the

N:P ratio of 5 to 10 is commonly found in cyanophyta-controlled communities, with optimal values varying between various algae such as *S. platensis* and *Scenedemus obliquus* (Çelekli *et al.*, 2008). The present study found that the blue-green alga produced a higher biomass than that recorded in prior research on *Spirulina* (Lodi *et al.*, 2005; Soletto *et al.*, 2005; Colla *et al.*, 2007). The Tukey HSD comparison test revealed significant differences in biomass production at various phosphate concentrations upon increasing the concentrations, causing a decrease in biomass yield (Figs. 1, 2). The maximum ($P < 0.05$) biomass production was recorded at 2.75 & 0.5g/ l phosphorus (Fig. 1). Raof *et al.* (2006) elucidated that, increasing phosphorus levels of 1.0–1.5g/ l in Zarrouk medium reduced the biomass yield of *S. platensis*.

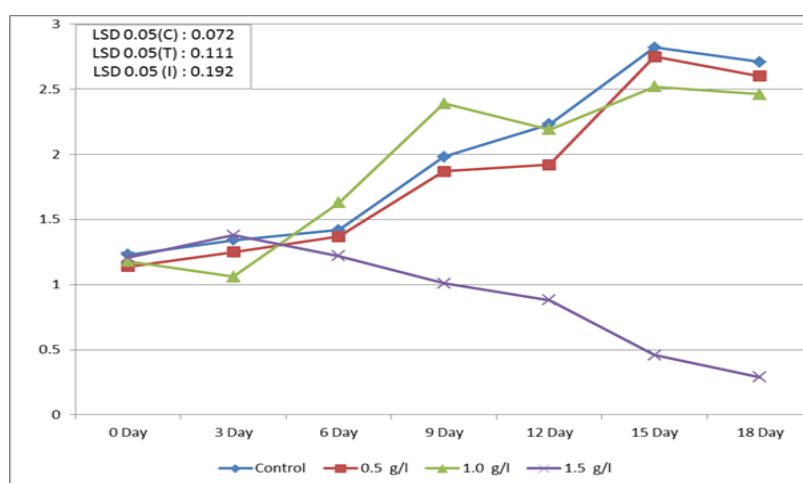


Fig. 1. Growth curves of *Spirulina platensis* dry weight grown on different levels of KH_2PO_4

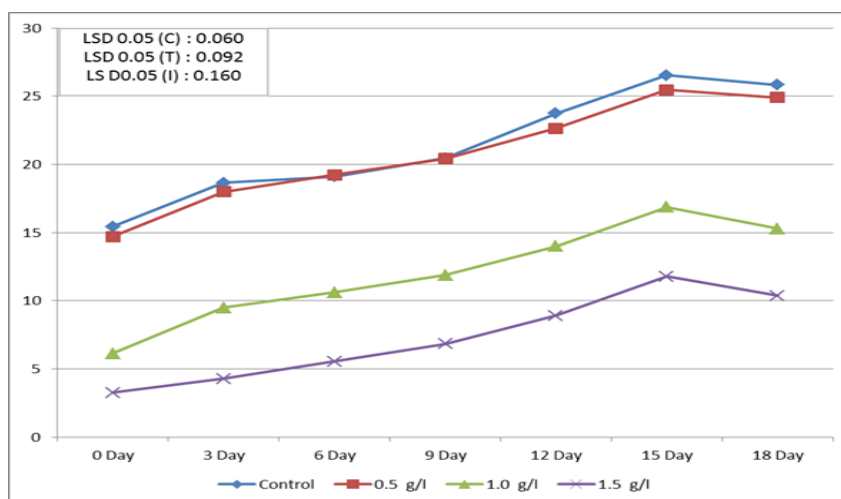


Fig. 2. Growth curves of *Spirulina platensis* dry weight grown on different levels of Na_2HPO_4

Pigments content

Table (1) shows the concentrations of Na₂HPO₄ addressed for the pigment production of the tested isolate. Chl a and total Chl were significantly higher when Na₂HPO₄ was used at a concentration of 0.5g/ L, while carotenoids were not significantly affected by the change in Na₂HPO₄ concentration. Similar findings were reported in the studies of **Pribyl et al. (2005)** and **Ontawong et al. (2013)**. In this respect, **Ikegaya et al. (2008)** reported that, upon increasing the production of Chl a, an increase was detected in the production of total Chl and biomass in *Spirulina* sp. The Chl a and total Chl concentrations were considerably ($P<0.05$) affected by altering the culture medium with the increase of Na₂HPO₄. Figs. (3- 10) show chlorophyll a, total chlorophylls, and carotenoids contents affected by phosphorus concentrations of 0.5, 1.0, and 1.5g/ l, respectively, during 18 days. The maximum production of Chl a, total Chl, and carotenoids was obtained by Zarrouk medium at 23.45, 35.08, and 12.01mg/ g, respectively, followed by the low levels of the two phosphorus sources. KH₂PO₄ as a source of phosphorus, recording the highest content of pigments at a level of 0.5g/ l, which followed the control treatment, where the levels of decrease were 25.3, 36.26, and 8.82mg/ g, giving lower values than the control, followed by Na₂HPO₄ at the same level for the previously mentioned pigments, respectively. The study found no significant difference in the pigment content at high phosphorus concentrations ($P<0.05$). Phosphorus concentration in *Azolla pinnata* blue-green alga reduced chlorophyll content while enhancing astaxanthin content in *Haematococcus pluvialis* green algae (**Brinda et al., 2004; He et al., 2007**). Previous studies also showed that phosphorus is effective for algae carotenoids and chlorophyll production (**Latas, 1994; Buapet et al., 2008**). Phosphorus starvation has been reported to stimulate carotenoids and chlorophylls (**Subudhi, 1979; Brinda et al., 2004; He et al., 2007**).

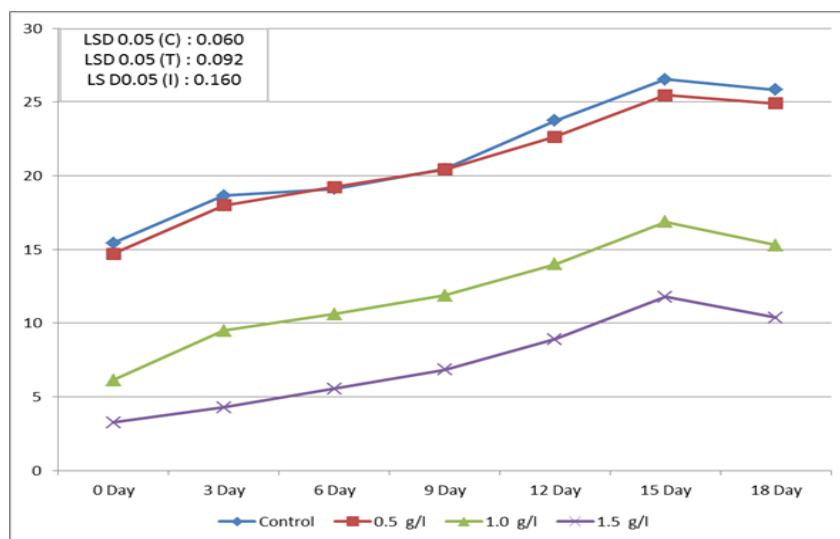


Fig. 3. Chl a content of *Spirulina platensis* in different concentration of KH₂PO₄

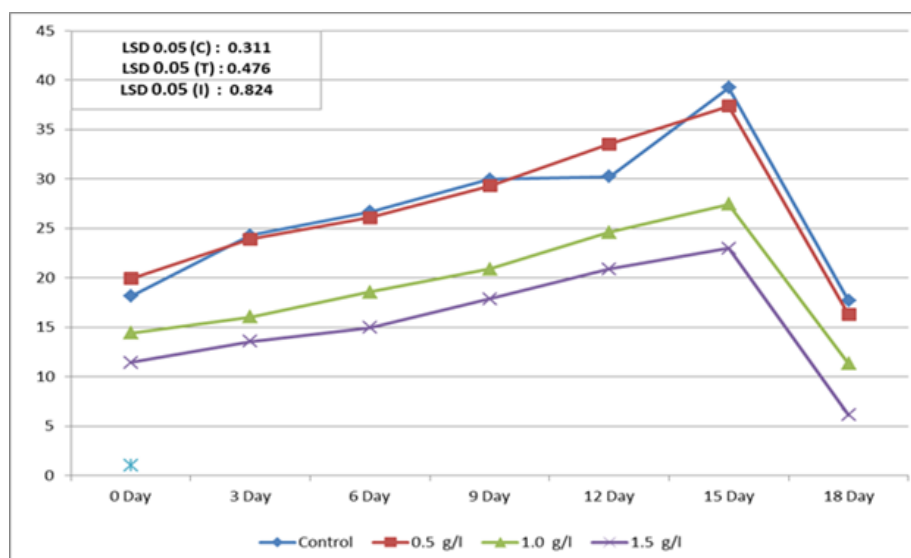


Fig.4. Total chlorophyll content of *Spirulina platensis* in different concentrations of KH_2PO_4

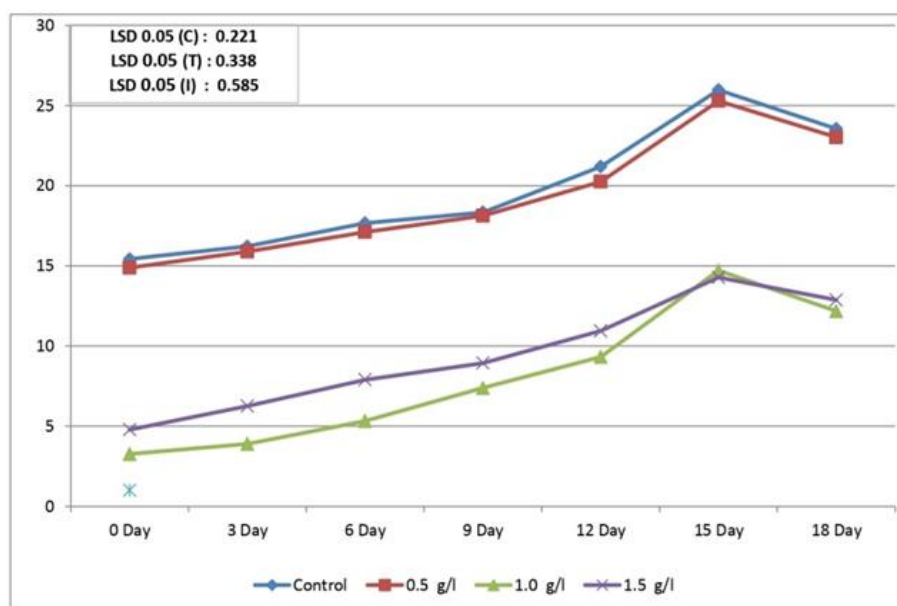


Fig.5. Chl a content of *Spirulina platensis* in different concentration of Na_2HPO_4 .

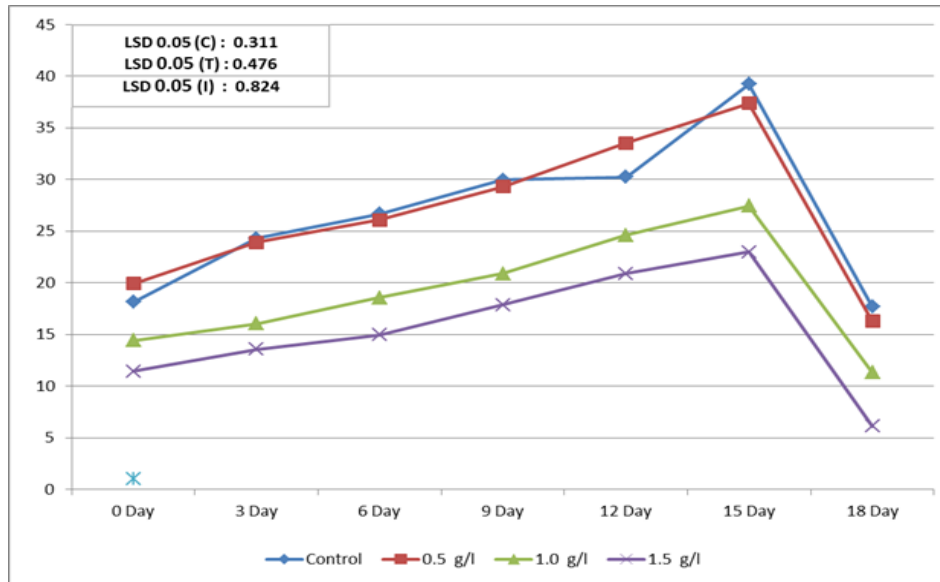


Fig. 6. Total chlorophyll content of *Spirulina platensis* in different concentrations of Na_2HPO_4

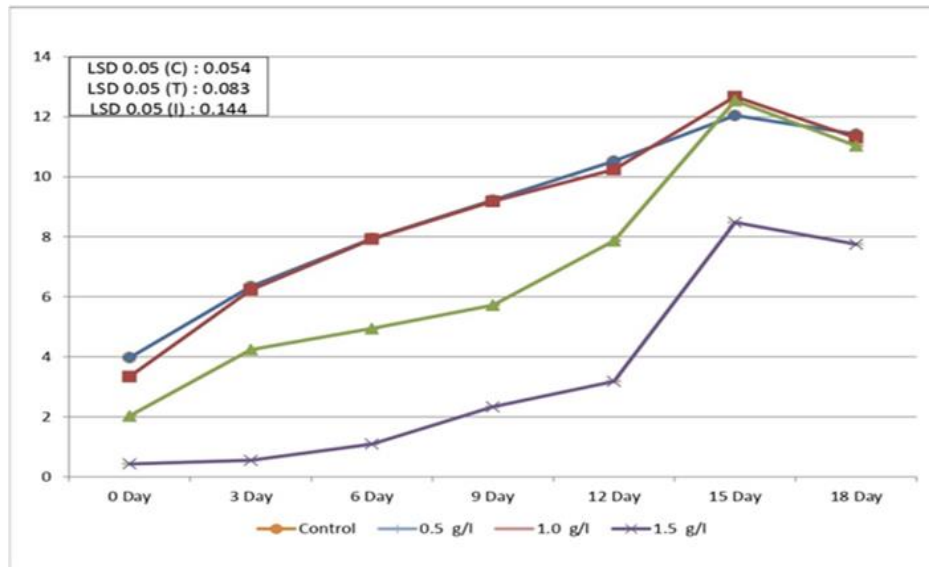


Fig. 7. Carotenoids content of *Spirulina platensis* at three levels of KH_2PO_4 .

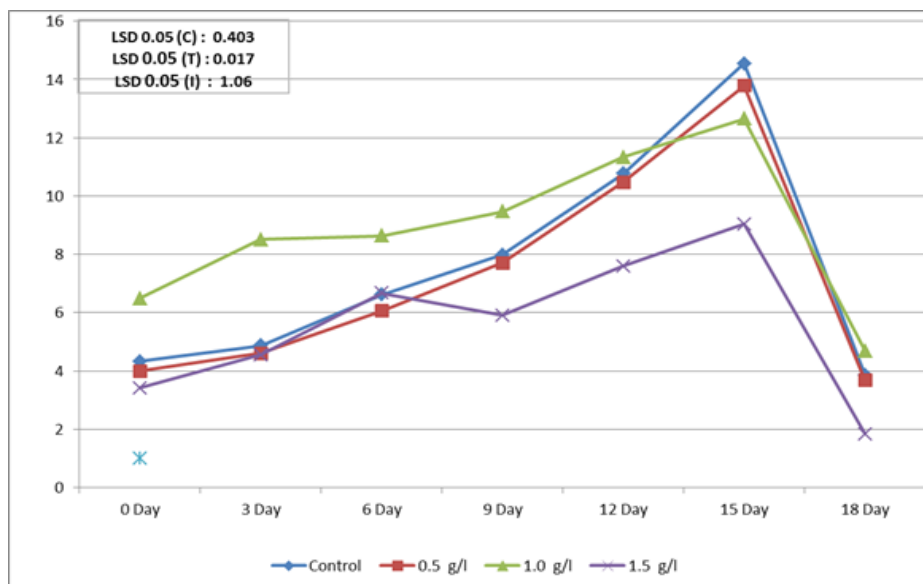


Fig. 8. Carotenoids content of *Spirulina platensis* at three levels of Na_2HPO_4

CONCLUSION

The study examined the impact of various phosphorus sources, cultivating modified Zarrouk medium, and the incubation period of 18 days on *S. platensis* biomass and pigment production. According to the study's results, 0.5g/ l of phosphate generated the greatest amount of biomass and pigments. The statistical analysis revealed that the cultivation period and phosphate concentrations were significant factors in the growth process. The phosphorus concentration (1.0–1.5 g/L) is considered sufficient to decrease total chlorophyll, carotenoids, and biomass yield by *S. platensis*. The results showed that *S. platensis* has the potential to produce biomass and pigments, depending on the optimum phosphorus concentration.

REFERENCES

- Abou-El-Souod, G.W.; Morsy, E.M. and Hassan, L.H.S. (2016).** Comparison of different media formulations and the optimal growing conditions on growth, morphology, and chlorophyll content of green alga, *Chlorella vulgaris*. *Journal of American Science*, 12(6): 86–95.
- Alipanah, L.; Winge, P.; Rohloff, J.; Najafi, J.; Brembu, T. and Bones, A.M. (2018).** Molecular adaptations to phosphorus deprivation and comparison with nitrogen deprivation responses in the diatom *Phaeodactylum tricornutum*. *PLoS One* 13:e0193335.

AOAC, (1990). Official Methods of Analysis of the Association of Official Analytical Chemists. 15th Edition, Association of Official Analytical Chemists, Washington,

AOAC, (2000). Analysis of the Association of Official Analytical Chemists. (Ed. William, H.), 17th ed., Gaithersburg, MD, USA, pp: 141–144.

Ayehunie, S.; Belay, A.; Baba, T.W. and Ruprecht, R.M. (1998). Inhibition of HIV-1 replication by an aqueous extract of *Spirulina platensis* (*Arthrospira platensis*). JAIDS Journal of Acquired Immune Deficiency Syndromes, 18(1):7–12.

Belay, A. (2008). *Spirulina* (*Arthrospira*): production and quality assurance. Gershwin ME, Belay A (eds.) *Spirulina in Human Nutrition and Health*. CRC Press, London, Boca Raton, pp. 1–26 Bligh EG, Dyer WJ (1959) A rapid method of total lipid extraction and purification. *Can J Biochem Physiol* 37:911–917.

Brinda, B. R.; Sarada, R.; Kamath, B. S. and Ravishankar, G. A. (2004). Accumulation of astaxanthin in flagellated cells of *Haematococcus pluvialis*: cultural and regulatory aspects," *Curr. Sci.*, 87:1290–1295.

Buapet, P.; Hiranpan, R.; Ritchie, R. J. and Prathep, A. (2008). Effect of nutrient inputs on growth, chlorophyll, and tissue nutrient concentration of *Ulva reticulata* from a tropical habitat", *ScienceAsia*, 34:245-252.

Bulgakov, N.G. and Levich, A.P. (1999). The nitrogen-phosphorus ratio is a factor regulating phytoplankton community structure. *Arch. Hydrobiol.* 146:3–22.

Çelekli, A.; Balcı, M. and Bozkurt, H. (2008). Modelling of *Scenedesmus obliquus*; function of nutrients with a modified Gompertz model. *Bioresour. Technol.* 99: 8742–8747.

Chandi, G.K. and Gill, B.S. (2011). Production and characterization of microbial carotenoids as an alternative to synthetic colours: A review. *International Journal of Food Properties*, 14: 503–513.

Chauhan, K. and Pathak, N. (2010). Effect of different conditions on the production of chlorophyll by *Spirulina platensis*. *Biomass Utiln.*, 1 (4):89–99.

Chu W.L.; Lim Y.W.; Radhakrishnan, A.K. and Lim, P.E. (2010). Protective effect of aqueous extract from *Spirulina platensis* against cell death induced by free radicals. *BMC Complement Altern Med* 10:53. <https://doi.org/10.1186/1472-6882-10-53>.

Colla, L.M.; Reinehr, C.O.; Reichert, C. and Costa, J.A.V. (2007). Production of biomass and nutraceutical compounds by *Spirulina platensis* under different temperature and nitrogen regimes. *Bioresour. Technol.* 98: 1489–1493.

COSTA, J.; COLLA, L.; FILHO, P.; KABKE, K. and WEBER, A. (2002). Modelling *Spirulina platensis* growth in fresh water using response surface methodology. *World J Microbiol Biotechnol*, 18(7): 603-607.

Desikachary, V. (1959). Cyanophyta. Indian Council of Agricultural Research, New Delhi, pp. 187–198.

Dubini, A. and Antal, T.K. (2015). Generation of high-value products by photosynthetic microorganisms: From sunlight to biofuels. *Photosynthesis Research*, 125: 355–356.

El-Baky HHA; Baz FKE. and El-Baroty, GS (2008). characterised nutraceutical compounds in blue-green alga *Spirulina maxima*. *J Med Plant Res* 2:292–300.

El-Sheekh, M.; Daboor, S.; Swelim, M.A. and Nohamed, S. (2014). Production and characterization of an antimicrobial active substance from *Spirulina platensis*. *Iranian Journal of Microbiology*, 6(2): 112-119.

Falquet, J. (1997). The nutritional aspects of spirulina. Antenna Foundation Available online at: https://www.antennach/wp-content/uploads/2017/03/AspectNut_UK.pdf. Accessed April 15, 2019 (25).

Ghaeni, M.; Roomiani, L. and Moradi, Y. (2015). Evaluation of carotenoids and chlorophyll as natural resources for food in *Spirulina* microalgae. *Applied Food Biotechnology*, 2(1): 39–44. Shao, W., Ebaid, R., El-Sheekh, M., Abomohra, A.; and Eladel, H. (2019). Pharmaceutical applications and consequent environmental impacts of *Spirulina* (*Arthrospira*): An overview. *Grasas y Aceites*, 70(1): 292-304.

Grobbelaar, J.U. (2013). Inorganic algal nutrition. In: Richmond A, Hu Q (eds), *Handbook of Microalgal Culture: Applied Phycology and Biotechnology*, 2nd edn., John Wiley & Sons, NY, pp. 123–133.

Gutiérrez-Salmeán G.; Fabila-Castillo L. and Chamorro-Cevallos G (2015). Nutritional and toxicological aspects of *Spirulina* (*Arthrospira*). *Nutr Hosp* 32:34–40.

Habib, B.; Parvin, M.; Huntington, C. and Hasan, R. (2008). Review of the Culture, Production, and Use of *Spirulina* as Food for Humans and Feeds for Domestic Animals and Fish. *FAO Fisheries and Aquaculture Circular No. 1034*: 33pp.

He, P.; Duncan, J. and Barber, J., "Astaxanthin accumulation in the green alga *Haematococcus pluvialis*: Effects of cultivation parameters," **J. Integr. Plant Biol.** (2007), 49: 447–451.

Hindak, F. (1988). Studies on chlorococcal algae (Chlorophyceae). Publishing hours of the Slovak Academy of Science, VEDA, Bratislava, 4:599–624.

Hindak, F. (1990). Studies on chlorococcal algae (Chlorophyceae). Publishing hours of the Slovak Academy of Science, VEDA, Bratislava, 5:647–671.

Hosseini SM; Khosravi K. and Mozafari M. (2013). Nutritional and medical applications of *Spirulina* microalgae. *Mini-Rev Med Chem* 13: 1231–1237. <https://doi.org/10.2174/1389557511313080009>.

Ikegaya, H; Hayashi, T.; Kaku, T.; Iwata, K.; Sonobe, S. & Shimmen, T. (2008). Presence of xyloglucan-like polysaccharide in *Spirogyra* and possible involvement in cell-cell attachment. *Phycological Research*, 56 (3): 216-222.

Khozin-Golberg, I. and Cohen, Z. (2006). The effect of phosphate starvation and the lipid-fatty acid composition of the freshwater eustigmatophyte *Monodus subterraneus*. *Phytochemistry*, 67(7): 696–701.

Kumar M.; Sharma M.K. and Kumar A. (2005). *Spirulina fusiformis*: a food supplement against mercury-induced hepatic toxicity. *J Health Sci* 51:424–430.

Latasa M. and E. Berdalet, (1994). "Effect of nitrogen or phosphorus starvation on the pigment composition of cultured *Heterocapsa* sp." **J. Plankton Res.**, 16: 83–94.

Leonardos, N. and Lucas, I. (2000). The nutritional values of algae grown under different culture conditions for *Mytilus edulis* L. larvae. *Aquaculture*, 182 (3–4): 301-315. [http://dx.doi.org/10.1016/S0044-8486\(99\)00269-0](http://dx.doi.org/10.1016/S0044-8486(99)00269-0).

Lodi, A.; Binaghi, L.; Faveri, D.D.; Carvalho, J.C.M. and Converti, A. (2005). Fed-batch mixotrophic cultivation of *Arthrospira* (*Spirulina*) *platensis* (Cyanophyceae) with carbon source pulse feeding. *Ann. Microbiol.* 55 (3): 181–185.

Mani, U.V.; Iyer; U.M.; Dhruv, S.A.; Mani, I.U. and Sharma, K.S. (2007). Therapeutic utility of *Spirulina*. *Spirulina*. In: "Human Nutrition and Health," Chapter 4, pp: 71–99.

Matos, J.; Cardoso, C.N.; Bandarra, M. and Afonso, C. (2017). Microalgae as healthy ingredients for functional food: A review. *Food and Function*, 8: 2672–2685.

Michael A.; Kyewalyanga MS; Mtolera MS.and Lugomela CV (2018). Antioxidant activity of the cyanobacterium *Arthrospira* (*Spirulina*) *fusiformis* cultivated in a low-cost medium. *Afr J Food Sci* 12:188–195. <https://doi.org/10.5897/AJFS2018.1688>

Mohy EL-Din, S.M. (2020): Evaluation of different biological activities of *Spirulina platensis* extracts. *Egyptian Journal of Botany*, 60(1): 249–259.

Ontawong, A.; Saowakon, N.; Vivithanaporn, P.; Pongchaidecha, A.; Lailerd, N.; Amornlerdpison, D.; Lungkaphin, A. and Srimaroeng, C. (2013). Antioxidant and renoprotective effects of *Spirogyra neglecta* (Hassall) Kützing extract in experimental type 2 diabetic rats. *BioMed Research International*.

Prescott, A. (1978). *How to Know the Fresh Water Algae*. (Third Edition), WMC Brown Company Publishers, UK, 293 pp.

Pribyl, P.; Cepák, V. and Zachleder, V. (2005). Cytoskeletal alterations in interphase cells of the green alga *Spirogyra decimina* in response to heavy metal exposure: I. The effect of cadmium. *Protoplasma*, 226 (3): 231-240.

Priyadarshani, I. and Rath, B. (2012). Commercial and industrial applications of microalgae: A review. *Journal of Algal Biomass Utilisation*, 3: 89–100.

Radmann, E.M.; Reinehr, C.O. and Costa, J.A.V. (2007). Optimisation of the repeated batch cultivation of microalga *Spirulina platensis* in open raceway ponds. *Aquaculture* 265 (1–4): 118–126.

Raof, B.; Kaushik, B.D. and Prasanna, R. (2006). Formulation of a low-cost medium for mass production of *Spirulina*. *Biomass Bioenergy*, 30: 537–542.

Rizzo, R.F.; Nascimento, B.; Dos-Santos, C.; Castro, G.F.P.S.de, Passos, T.S.; Nascimento, M.A.; Guerra, H.D.; da Silva, C.G.; Dias, D.d.S.; Domingues, J.R. and de Lima-Araújo, K.G. (2015). Production of phycobiliproteins by *Arthrospira platensis* under different light conditions for application in food products. *Food Science and Technology Campinas*, 35: 247–252.

Rizwana, M.; Mujtabab, G.; Memonc, S.A.; Leed, K. and Rashid, N. (2018). Exploring the potential of microalgae for new biotechnology applications and beyond: A review. *Renewable and Sustainable Energy Reviews*, 92: 394–404.

Rocha, G. S.; Parrish, C. C.; Lombardi, A. T. & Melão, M. D. G. G. (2018). Biochemical and physiological responses of *Selenastrum gracile* (Chlorophyceae)

acclimated to different phosphorus concentrations. *Journal of Applied Phycology*, 30(4): 2167–2177.

Roleda M.Y.; Slocombe S.P.; Leakey R.J.G.; Day J.G.; Bell E.M. & Stanley, M.S. (2013). Effects of temperature and nutrient regimes on biomass and lipid production by six oleaginous microalgae in batch culture employing a two-phase cultivation strategy. *Bioresource Technology* 129: 439–449. DOI: 10.1016/j.biortech.2012.11.043.

Roopnarain A; Grey VM. and Sym SD (2014): Phosphorus limitation and starvation effects on cell growth and lipid accumulation in *Isochrysis galbana* U4 for biodiesel production. *Bioresour Technol.* 156:408–411.

Salunke, K.I.; Magar, S.A.; Joshi, R.R. and Adikar, M.S. (2016). A comparative study on the growth of *Spirulina platensis* on different cultural media. *Bioscience Discovery*, 7(1): 90–92.

Seely, R.; Duncan, J. and Vidaver, E. (1972). Preparative and analytical extraction of pigments from brown algae with dimethyl sulfoxide. *Mar. Biol.* 12: 184–188.

Shao, W.; Ebaid, R.; El-Sheekh, M.; Abomohra, A. and Eladel, H. (2019). Pharmaceutical applications and consequent environmental impacts of *Spirulina* (*Arthrospira*): An overview. *Grasas y Aceites*, 70(1): 292-304.

Sharoba, AM. (2014). Nutritional value of spirulina and its use in the preparation of some complementary baby food formulas. *J Food Dairy Sci, Mansoura Univ* 5:517–538.

Snedecor GW; Cochran WG (1994). *Statistical Methods*, 9th Ed., Iowa State University . Press, Ames, Iowa, USA.

Soletto, D.; Binaghi, L.; Lodi, A.; Carvalho, J.C.M. and Converti, A. (2005). Batch and fed-batch cultivations of *Spirulina platensis* use ammonium sulphate and urea as nitrogen sources. *Aquaculture*, 243: 217–224.

Soni, R.A.; Sudhakar, K. and Rana, R.S. (2019). A comparative study on the growth performance of *Spirulina platensis* on modifying culture media. *Energy Reports*, 5: 327–336.

Stein, J. (1973), "Handbook of Phycological Methods." Culture methods and growth measurements. Cambridge University Press. 448 pp.

Subudhi B.; P. R. and Singh, P. K., "Effect of phosphorus and nitrogen on growth, chlorophyll, amino nitrogen, soluble sugar contents, and algal heterocysts of the water fern *Azolla pinnata*," *Biol. Plantarum* (1979), 21: 401–40.

Suganya, T.; Varman, M.; Masjuki, H. and Renganathan, S. (2016). Macroalgae and microalgae as a potential source for commercial applications along with biofuel production: A biorefinery approach. *Renewable and Sustainable Energy Reviews*, 55: 909–941.

Theodore, S. and Georgios, S. (2013). Health aspects of the *Spirulina* (*Arthrospira*) microalga food supplement. *J. Serb. Chem. Soc.*, 78 (3): 395–405.

Verne`s, L.; Abert-Viana, M.; El Maâtaoui, M.; Tao, Y., Bornard, I. and Chemat, F. (2019). Application of ultrasound for green extraction of proteins from *Spirulina*. Mechanism, optimization, modelling, and industrial prospects. *Ultrasonics and Sonochemistry*, 54: 48–60.

Vonshak, A.; Chanawongse, L.; Bunnag, B. and Tanticharoen, M. (1996). Role of light and photosynthesis on the acclimation process of the cyanobacterium *Spirulina platensis* to salinity stress. *J. Appl. Phycol.*, 8:119–124.

Vonshak, A. (1997). *Spirulina platensis* (*Arthrospira*): Physiology, Cell Biology, and Biotechnology. Taylor and Francis, London.

Yaakob MA; Mohamed RMSR; Al-Gheethi A; Aswathnarayana Gokare R. and Ambati RR (2021), Influence of nitrogen and phosphorus on microalgal growth, biomass, lipid, and fatty acid production: an overview. *Cells* 10:393.

Zarrouk, C. (1966). Contribution à l'étude de l'unecyanophycée. Influence de divers' facteurs physiques et chimiques sur la croissance et la photosynthèse de *Spirulina maxima*. Ph.D. Thesis, Université de Paris, France, 84 pp.

Zhou, Y.; Zou, M.; Woods, S. A. and Wu, C.-H. (2019). The restorative effect of work after unemployment: An intraindividual analysis of subjective well-being recovery through reemployment. *Journal of Applied Psychology*, 104(9): 1195–1206. <https://doi.org/10.1037/ap:10000393>.