



Evaluation the expression of SP1 and PAR3 in urinary bladder carcinoma

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Submit Date 04-05-2024

Revise Date 08-07-2024

Accept Date 03-06-2024



ABSTRACT

Endometrial cancer (EC) is a complex disease with various subtypes. Although it is considered a treatable disease due to early diagnosis and symptoms, the advanced stage, which represents approximately 8% of cases, is associated with poor outcomes, reflecting the absence of efficient systemic therapy. There are various molecular classifications available and many pathways involved in EC pathogenesis. Advances in understanding its biology have led to the development of a classification system to help tailor treatment strategies depending on both patients and disease features. Currently, immunotherapy and targeted therapies are key treatments for EC, whether recurrent or advanced. In this review, we aim to summarize the molecular classification of EC briefly and its impact on treatment strategies, as well as the proposed targeted therapy. The data collection is based on searches on scientific websites, clinical trials (ClinicalTrials.gov), and international guidelines from Europe's leading medical oncology society (ESMO) and the National Comprehensive Cancer Network (NCCN).

Keywords: Endometrial Cancer, Molecular Classifications, Target therapy.

INTRODUCTION

Urinary bladder cancer which is the second most common malignant genitourinary tumors globally, is responsible for over 199,000 fatalities annually, representing a significant burden of morbidity and mortality [1].

Despite improvement in treatment protocol with chemotherapeutic, radiation, and radical cystectomy approaches, prognosis for patients with urinary bladder cancer remain unsatisfactory due to aggressive behavior. Patients with high-risk non-muscle invasive bladder carcinoma (NMIBC) who do not exhibit a response to adjuvant therapy involving bacille Calmette-Guérin (BCG) immunotherapy present a formidable case for management. A considerable proportion of

individuals diagnosed with urothelial carcinoma develop distant metastasis, which reduces the 5-year survival rate to 8% [2].

Identification of the significant agent responsible for recurrence and metastasis required to improve the survival rates and/or response to therapy in patient with urinary bladder carcinoma.

Specificity protein1 (Sp1) is transcription factor that activates multiple genes that have important roles in cell cycle regulation, differentiation and apoptosis [3]. Sp1 plays key roles in tumorigenesis and progression of majority of malignancies such as lung, pancreatic, colon, glioma, and gastric cancer [4], [5].

Loss cell polarity and cell junction of epithelium are fundamental for tumorigenesis

and metastasis of carcinomas [6],[7]. Cell polarity is regulated by PAR, Crumbs, and Scribble. PAR polarity complex, including of Par3, Par6, and atypical PKCs, is essential at the apical-basal membrane border and the tight junction in epithelial cells [8]. Loss of Par3 is associated with metastasis and invasion of breast and lung tumors via regulation of aPKC activity and Stat3 activity [9].

The primary objective of the present study is to evaluate the expression of Sp1 and Par3 in urinary bladder carcinoma and to investigate their correlation with clinicopathological characteristics and patient survival.

MATERIAL AND METHODS:

This prospective cohort study included fifty cases of urothelial carcinoma. Pathology, Urology and Clinical Oncology departments at Zagazig University Hospitals conducted the current research from January 2018 to December 2023. For confirming the diagnosis, all the hematoxylin and eosin stained slides were examined. Tumor grades and stages were classified according to the TNM staging scheme (AJCC, eighth edition) [10]. Only cases of urothelial carcinomas with presented muscle layer in the examined sections were included in this research. All urinary bladder carcinoma underwent trans-urethral resection, partial and radical cystectomy. Clinical and survival data were obtained from electronic medical records from the diagnosis to last follow-up appointment or until death. Patient follow up for 36 months. The overall survival (OS) and disease free survival (DFS) were defined.

The adjuvant treatment

Treatment depends on the stage of the bladder cancer. Non muscle invasive bladder cancer patients (Stage Ta-T1; with or without CIS) were treated with Surgical resection of trans-urethral resection of bladder tumor (TURBT) that was done in the Urology department. The intermediate and high-risk patients were treated by TURBT, with standard regimen of BCG according to American urological association (AUA). Patients with locally advanced Non-metastatic muscle invasive

bladder cancer (stage II-III) were treated with maximal TURBT and concurrent chemoradiotherapy.

Radiation therapy was delivered using 3-dimensional conformal technique (3DCRT). Patients were instructed to be with empty bladder during simulation and during the course of treatment, multi-slice CT was done every 0.3-0.5cm in supine position. GTV = pre-TURBT tumor volume (as assessed on cystoscopy, CT, PET, or MRI). CTV = GTV + entire bladder + proximal urethra + prostate and prostatic urethra (in men) or proximal 2 cm of the female urethra + regional lymphatics (internal iliac, external iliac, and obturators). PTV = CTV + margin (1.5 to 2 cm). CTV boost = GTV + 0.5 cm, If GTV not well defined, entire bladder (empty) is CTV. PTVboost = CTVboost + 1 to 2 cm margin. A total dose of 64 Gy was given in 32 fractions. Concurrent chemotherapy that administered was weekly cisplatin in the dose of 40 mg/m² or weekly gemzar 75 mg/m² for patients were cisplatin intolerant. Metastatic bladder cancer patients were treated with systemic chemotherapy or concurrent chemoradiotherapy.

Ethical approval:

The Institutional Review Board (IRB) of the Faculty of Medicine of Zagazig University, granted permission to collect data and samples (ZU-IRB# 300/21). The research followed the Declaration of Helsinki of the WMA. All patients or their legal representatives have signed informed consent forms

Immunohistochemical (IHC) assay

Using streptavidin-biotin labeling technique and the LSAB kit (Dako, Glostrup, Denmark) with appropriate positive and negative controls, immunohistochemistry was done. Samples embedded in paraffin were sectioned at a thickness of 5 um and subsequently put on polylysine for ten minutes, the sections were immersed in 0.3% hydrogen peroxide in order to inhibit endogenous peroxidase activity. In order to conduct antigen retrieval, sections were subjected to high-pressure heating (700 W microwave oven) for one

minute in a 10 mM citrate antigen retrieval solution (0.1 mM sodium citrate, 0.1 mM citric acid; pH=6.0). The sections were incubated at 4 °C overnight with a monoclonal antibody targeting Sp1 (Santa Cruz Biotechnology Inc., USA, 1:100) and Par3 (Sigma–Aldrich (USA) 1:50) after being washed with PBSTX (0.05 M PBS, 0.1% PBS, 0.1% Triton-X 100). Subsequently, the sections were incubated at 37 °C for 30 minutes with an HRP-conjugated secondary antibody (Concentrations 1:100) (sc-2005, Santa Cruz Biotechnology Inc., USA). In order to observe the sections, they were submerged in a solution of diaminobenzidine (10HDAB, TBS, 30% H₂O₂).

Evaluation of immunohistochemical staining

The nuclear and/or cytoplasmic staining for Sp1 was considered IHC-positive and for scoring cases divided according to the percentage of positive cells, into 0 for ≤5%, 1 for 6–30%, 2 for 31–80%, and 3 for >80%. Sp1 expression was evaluated by staining intensity and immunoreactive score ranging from 0 to 9 and considered as low expression (IHC score 0–2), and high expression (IHC score 3–9)[11].

Par3 immunostaining was evaluated as membranous and/or cytoplasmic. An isolated membranous immunostaining of Par3 was considered as low Par3 expression while both a membranous and cytoplasmic localization was considered as high Par3 expression [12].

Statistical analysis

The studied continuous parameters were expressed as mean ± SD & median, but the studied categorical variables were expressed as percentages that compared by Pearson's Chi-square test or Fisher's exact test. The stratification of DFS and OS was organized according to the studied immunohistochemical biomarkers. The time-to-event distributions were valued by Kaplan-Meier plot and compared by two-sided exact log-rank test. All the used tests were two sided. All our statistics were finished using SPSS 22.0 for windows and MedCalc

windows (MedCalc Software bvba 13,, Belgium

RESULTS

1. Clinicopathological data:

Resume of clinicopathological features of 50 studied cases were summarized in table 1.

2-Immunohistochemical expression of Sp1 and Par3:

In the current study, high Sp1 expression was found in 20 /50 (40%) of cases. There is a statistically significant correlation between high expression of Sp1 and tumor grade (51.5% of high grade tumors compared to 17.6 % in low grade tumors) (P=0.02), lymph nodes metastasis (p<0.007), and tumor stage whereas there was attend toward increased sp1 expression in higher stage (p=0.03). As regards to muscle invasion, 66.7 % of muscle invasive cases exhibited high Sp1 expression with statistically significant association (p=0.01). These result indicated that high Sp1 expression is associated with high grade, high stage and muscle invasive bladder carcinoma (As shown in table 2, figure 1).

In our study, high Par3 expression was found in 29 /50 (58% %) of cases. There is a statistically significant correlation between high expression of Par3 and low tumor grade (88.2% of low grade tumors compared to 42% high grade cases) (p=0.002), negatively lymph nodes metastasis (p=0.01), and tumor stage whereas there was attend toward low Par3 expression in higher stage (p=0.008).

Regarding the presence of muscle invasion in urothelial carcinoma cases, 88.5% of non muscle invasive cases showed par3 positivity, while 25% of muscle invasive cases showed Par3 positive expression (p=0.045). These results indicated that reduction in Par3 expression is associated with high grade, advanced tumor stage, lymph nodes metastasis, tumor multiplicity and muscle invasive bladder carcinoma (As shown in table 3, figure 2).

There is a significant negative correlation between Sp1 and Par3 expression (p<0.001)

which were significantly associated with worse overall survival. Overexpression of Sp1 inhibited the tumor suppressor effect of Par3.

3-Treatment outcome, survival, and progression analysis:

High Sp1 expression was correlated strongly with poor therapy response while patients with low Sp1 expression had achieved a complete response (p<0.001). as well as patients with high par3 expression had shown better treatment response rather than low Par3 expressed patients(p<0.001)..

It was found that the higher Sp1 expression was significantly associated with recurrence after treatment (p=0.01), disease free survival (p<0.001).and overall survival(p=0.004). High Sp1 expression was predictor of recurrence, short disease free survival and overall free survival

Also, we found that low Par3 expression was associated with shorter disease free survival (p<0.001).and lower overall survival (p<0.001). Low Par3 expression was related with poor survival progression in bladder carcinoma (As shown in figure 3)

Table (1): Demographic and clinical data of the tested cases

Variable		(n=50)	
Age: (years)	Mean ± Sd Range	61.28±8.57 45-84	
Variable		No	%
Sex:	Female	22	44
	Male	28	56
Grade:	Low	17	34
	High	33	66
Size: (cm ²)	Mean ± Sd Range	2.82±0.97 1-5	
Number:	Unifocal	40	80
	Multifocal	10	20
Stage:	T1	20	40
	T2	20	40
	T3	8	16
	T4	2	4
Muscle invasion:	Non-invasive	20	40
	Invasive	30	60
Vascular invasion:	No	45	90
	Yes	5	10
LN metastasis:	No	45	90
	Yes	5	10
Distant metastasis:	No	42	84
	Yes	8	16
Relapse:	No	41	82
	Yes	9	18
Survival	Lived	44	88
	Dead	6	12

SD: Standard deviation

Table (2): Relation between demographic and clinical data of the tested cases and Sp1 expression

Variable		Low (n=30)(60%)		High (n=20)(40%)		P
Age: (years)	Mean ± Sd Range	60.53±9.29 45-84		62.4±7.44 52-80		0.46 NS ^s
Variable		No	%	No	%	P
Sex:	Female	16	72.7	6	27.3	0.10
	Male	14	50	14	50	NS [#]
Grade:	Low	14	82.4	3	17.6	0.02**
	High	16	48.5	17	51.5	
Size: (cm ²)	Mean ± Sd Range	2.67±0.79 1-4.5		3.06±1.16 1-5		0.17 NS ^s
Number:	Unifocal	27	67.5	13	32.5	0.03**
	Multifocal	3	30	7	70	
Stage:	T1	14	70	6	30	0.03**
	T2	14	70	6	30	
	T3	1	12.5	7	87.5	
	T4	1	50	1	50	
Muscle invasion:	Non-invasive Invasive	22 8	84.6 33.3	4 16	15.4 66.7	0.01**
Vascular invasion:	No Yes	29 1	64.4 20	16 4	35.6 80	0.14 NS [#]
LN metastasis:	No Yes	30 0	66.7 0	15 5	33.3 100	0.007* #
Distant metastasis:	No Yes	27 3	64.3 37.5	15 5	35.7 62.5	0.16 NS [#]
Relapse:	No Yes	27 3	69.2 27.3	12 8	30.8 72.7	0.01**
Survival	Lived	28	66.7	14	33.3	0.03**
	Dead	2	25	6	75	
Par3 expression:	Low	2	9.5	19	90.5	<0.001 ***
	High	28	96.6	1	3.4	

SD: Standard deviation, \$:Independent t test, #: Chi square test with Fischer exact correction when needed, NS: Nonsignificant (P>0.05), *:Significant (P<0.05), **:Extremely significant (P<0.001)

Table (3): Relation between demographic and clinical data of the studied tested and Par3 expression

Variable		Low (n=21)		High (n=29)		P
Age: (years)	Mean ± Sd Range	61.62±7.84 48-80		61.03±9.19 45-84		0.82 NS ^s
Variable		No	%	No	%	P
Sex:	Female	7	31.8	15	68.2	0.20
	Male	14	50	14	50	NS [#]
Grade:	Low	2	11.8	15	88.2	0.002**
	High	19	57.6	14	42.4	
Size: (cm ²)	Mean ± Sd Range	3.11±1.15 1-5		2.62±0.76 1.5-4.5		0.08 NS ^s

Number:	<i>Unifocal</i>	13	32.5	27	67.5	0.01**
	<i>Multifocal</i>	8	80	2	20	
Stage:	<i>T1</i>	6	30	14	70	0.008**
	<i>T2</i>	6	30	14	70	
	<i>T3</i>	7	87.5	1	12.5	
	<i>T4</i>	2	100	0	0	
Muscle invasion:	<i>Non-invasive</i>	3	11.5	23	88.5	0.003**
	<i>Invasive</i>	18	75	6	25	
Vascular invasion:	<i>No</i>	16	35.6	29	64.4	0.01**
	<i>Yes</i>	5	100	0	0	
LN metastasis:	<i>No</i>	16	35.6	29	64.4	0.01**
	<i>Yes</i>	5	100	0	0	
Distant metastasis:	<i>No</i>	15	35.7	27	64.3	0.05**
	<i>Yes</i>	6	75	2	25	
Relapse:	<i>No</i>	13	33.3	26	66.7	0.02**
	<i>Yes</i>	8	72.7	3	27.3	
Survival	<i>Lived</i>	14	33.3	28	66.7	0.004**
	<i>Dead</i>	7	87.5	1	12.5	

SD: Standard deviation, \$:Independent t test, #: Chi square test with Fischer exact correction when needed, NS: Nonsignificant (P>0.05), *:Significant (P<0.05), **:Extremely significant (P<0.001)

Table (4): Disease free survival and over-all survival among the studied cases according to Sp1 & Par3 expression

Variable	N	DFS			Overall survival			
		Median	95% CI	P	Median	95% CI	P	
Total	50	32.67	30.85-34.53	--	34.38	33.27-35.5	---	
SP1	<i>Low</i>	30	35.09	34.08-36.09	0.001	35.60	35.02-36.17	0.004*
	<i>High</i>	20	28.66	24.76-32.56		*	32.22	
Par3	<i>Low</i>	29	28.31	24.45-32.18	0.001*	31.70	29.15-34.25	<0.001
	<i>High</i>	21	35.13	34.15-36.09			35.88	

CI: Confidence interval, P:Log Rank test, **: extremely significant (P<0.001)

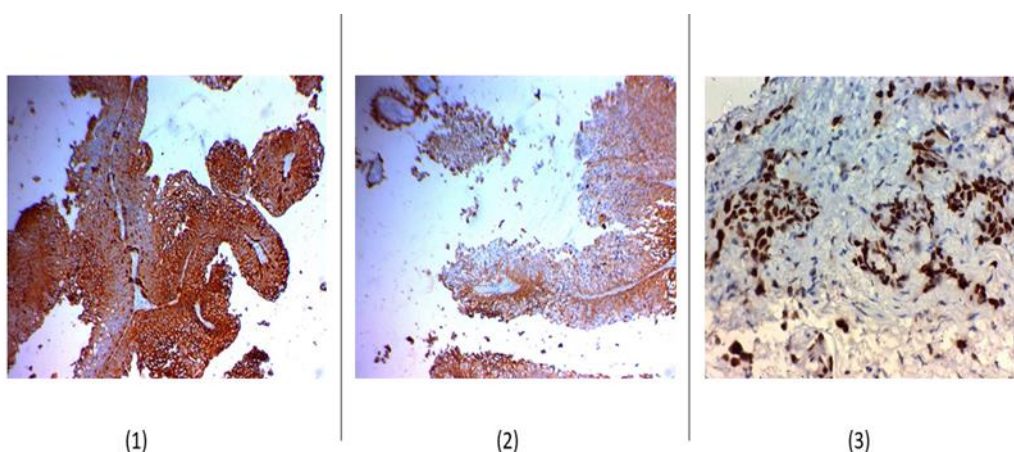


Figure 1. Representative images of Sp1 immunohistochemical staining expression in papillary urothelial carcinoma. (1) high SP1 expression in papillary urothelial carcinoma grade 2 immunohistochemical x 200. (2) low SP1 expression in low grade papillary urothelial carcinoma immunohistochemical x 200. (3) high SP1 expression in high grade urothelial carcinoma immunohistochemical x (400); mainly nuclear stain

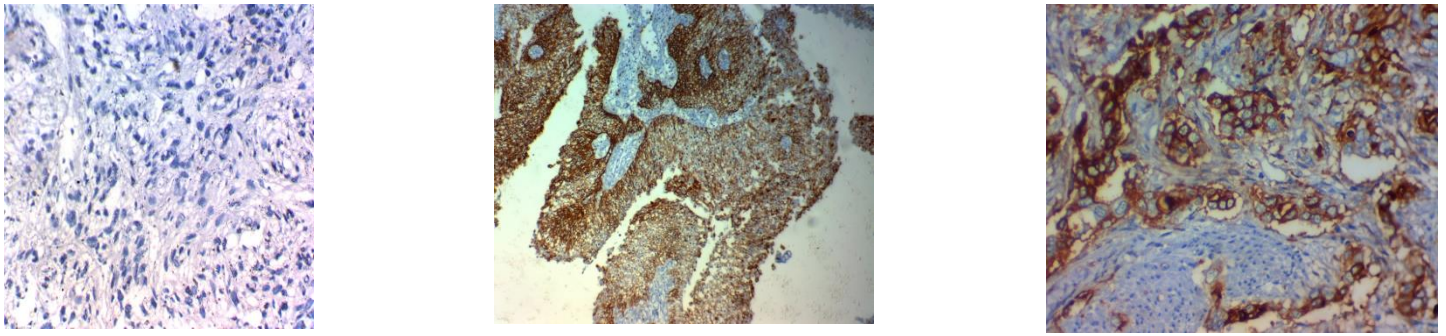


Figure 2. Representative images of Par3 immunohistochemical staining: expression in urothelial carcinoma. 1- negative Par3 expression in high grade urothelial carcinoma (immunohistochemical x 400). 2- high Par3 expression in low grade urothelial carcinoma (immunohistochemical x 200). 3- high Par3 expression in low grade invasive urothelial carcinoma (immunohistochemical x 400).

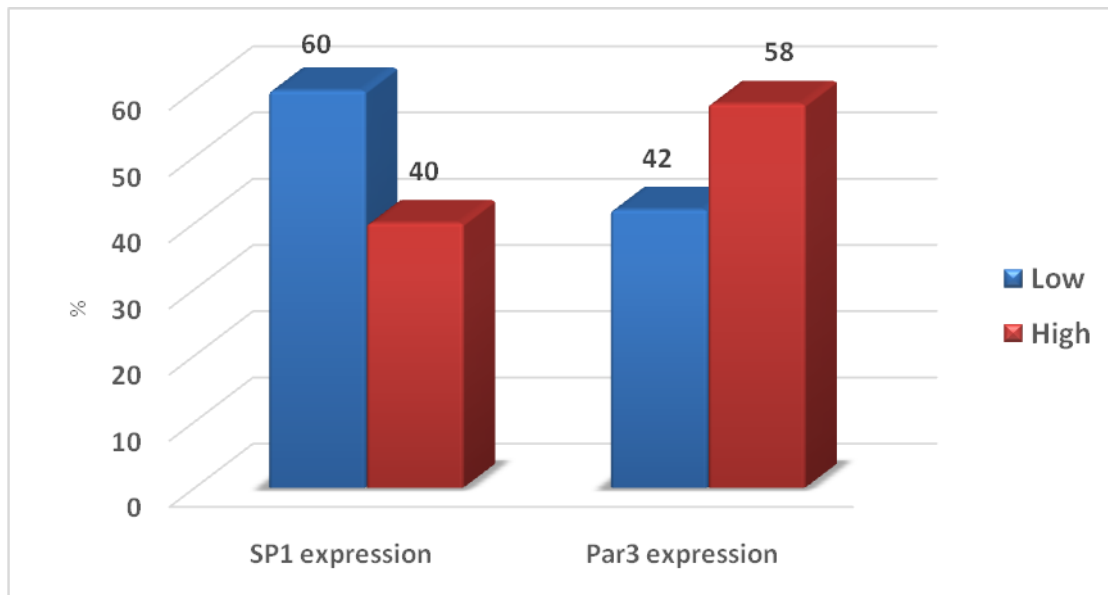
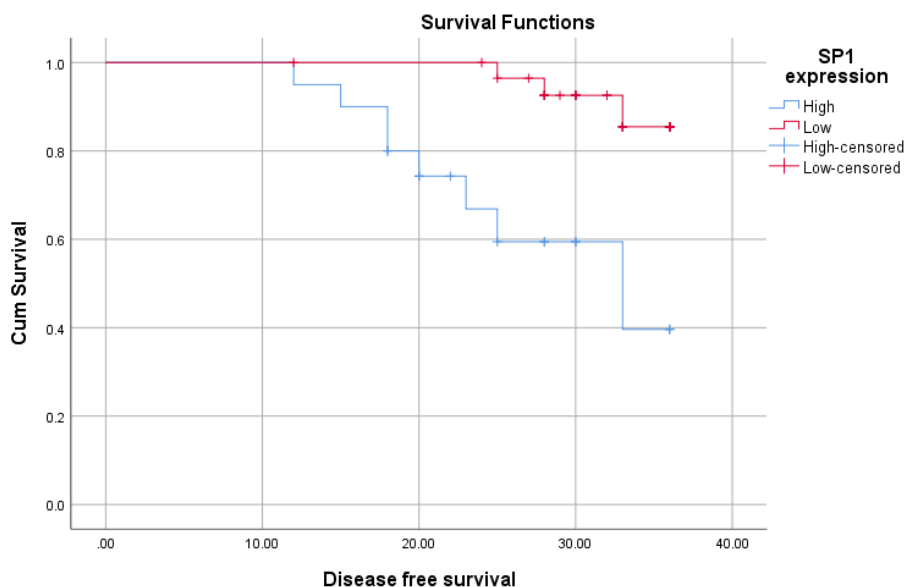


Figure 3. Frequency distribution of Sp1 & Par3 expression among the studied cases



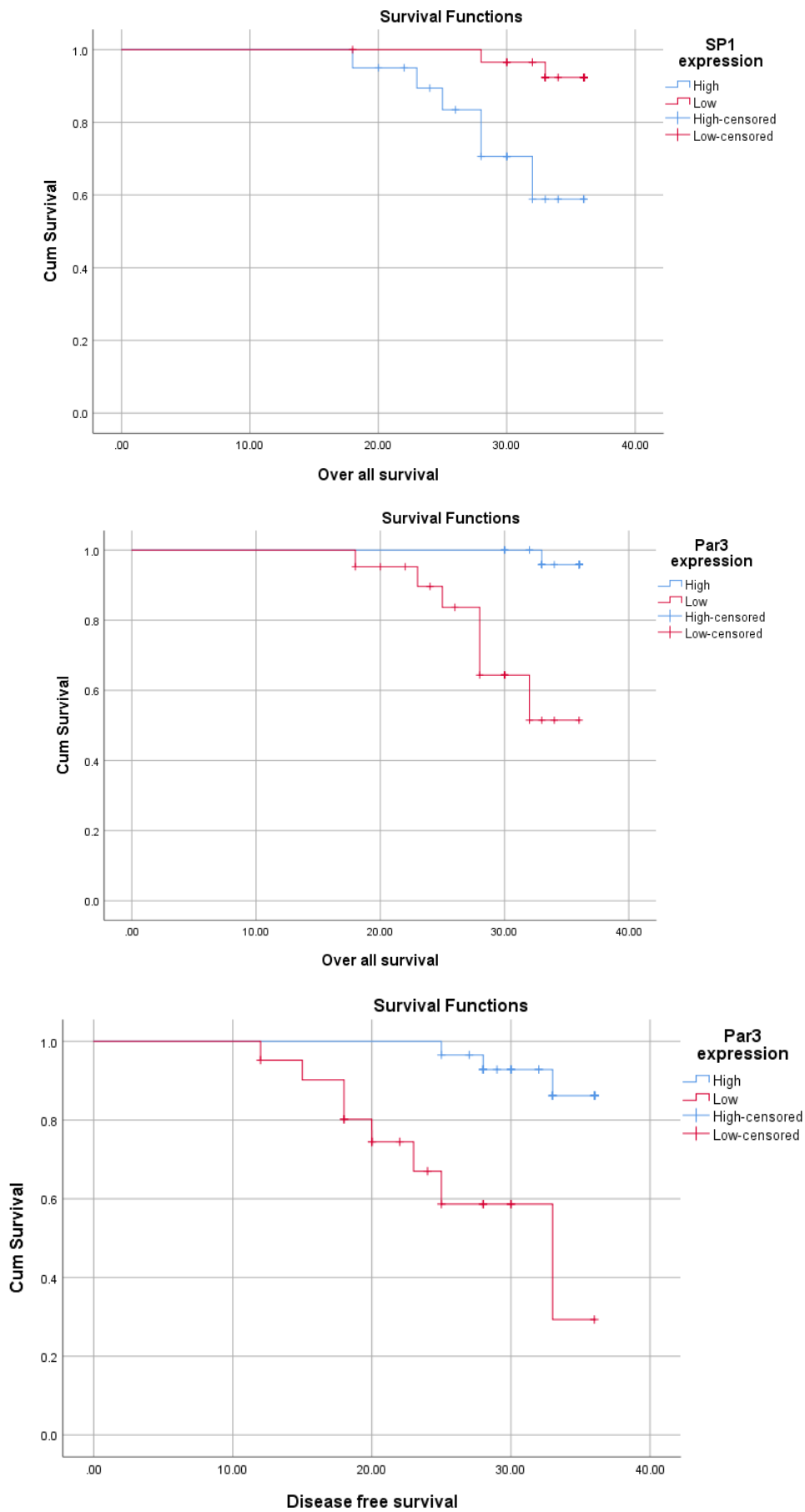


Figure 4. Survival analysis of Sp1 and Par3 expression by Kaplan Meier method

DISCUSSION

Urothelial carcinoma is classified as non-muscle invasive urothelial carcinoma (Tis, Ta, T1) and muscle-invasive urothelial carcinoma (T2, T4) [10]. High risk non-muscle invasive cases have a high relapse rate up to 80% within 5 years and 30% progress to muscle invasion following primary therapy [13]. The identification markers to predict the invasion of the tumor is important for management.

In the present study, Sp1 was over expressed in urothelial carcinoma. High Sp1 expression was significantly associated with high grade, lymph node metastasis and poor survival progression. This findings are consistent with **Chen** [11] stated that Sp1 was overexpressed in urothelial carcinomas and knockdown of Sp1 could accelerate apoptosis and suppress invasion of bladder urothelial tumor cells by enhancing PTEN expression and down regulation of AKT/mTOR pathway related protein in vitro and vivo studies.

Our results revealed significant difference in the Sp1 expression in MIBC when compared to NMIBC, which were consistent with previous study by **Zhu et al** [14] reported statistically significant difference between the invasive and non-invasive bladder tumors concerning IHC staining of Sp1. Sp1 may be used to identify cases with aggressive outcomes following surgery and design more effective therapeutic protocol.

Up-regulation of Sp1 expression may accelerate cell invasion in lung cancer [15] and glioma [16] by enhancing matrix metalloproteinase-2 (MMP-2) and matrix metalloproteinase-9 (MMP-9) expression. Besides, SP1 regulates vascular endothelial growth factor (VEGF) expression to promote angiogenesis in gastric carcinoma [17] Sp1 can also facilitate metastasis and invasion in colorectal cancer [18] and ovarian cancer [19] via the Fyn-SP1-SATB1 axis and Sp1-CD147 positive feedback loop, respectively.

These findings are consistent with previous report suggesting that Sp1 was downregulated in bladder urothelial carcinoma in response to betulinic acid, arsenic trioxide, curcumin, and celastrol, which have antitumor effects [20].

Par3 is an important polarity protein for maintaining polarity and tight junctions of

epithelium. Loss of polarity is a prerequisite for tumor progression [6]. The current study revealed that low Par3 expression was significantly associated with high grade, lymph node metastasis, advanced stage, and worse overall survival. Similarly, **Wang et al.** [21] reported that urothelial carcinoma with muscle invasion and bad prognosis is accompanied with considerable loss of Par3 expression.

Fomicheva et al. [22] also demonstrated the association of down regulation of Par3 with Epithelial-Mesenchymal Transition (EMT) and tumor aggressiveness. Transforming growth factor- β 1 (TGF- β 1), SNAIL, Wnt/ β catenin pathway that initialize EMT, could decrease expression of Par3 (encoded by PARD3 genes) and weaken cell-cell junction [23][24].

In addition to the critical role of par3 in polarity, it is complex independent functions to control cellular processes and exert its tumor suppressor function [23]. Low expression of Par3 facilitated TIAM1/Rac1 and JAK/STAT3 activation accelerated tumor growth and metastasis in breast cancer [25], [26], and lung carcinomas [27]. These findings are line with **Zhang et al** [28] who found that loss of PARD3 disturbed the Hippo pathway by dissociating the PARD3/merlin/LATS1 complex and increased YAP nuclear translocation in prostatic carcinoma.

We found a negative correlation between Sp1 and Par3 expression which was significantly associated with worse overall survival. Sp1 exerts its oncogenic activities such as EMT, angiogenesis, inflammatory signaling and immune escape via enhancing transcription cancer related genes. **Baba et al** [29] demonstrated that Sp1 directly activates to snail and TGF- β 1 genes. Sp1-mediated Snail expression represses par3 and E-cadherin expression [30][31]. TGF- β 1 also induced the nuclear expression of Snail in late stages of carcinoma resulting in tumor progression and metastasis [29]. Furthermore, **Wang et al** [21] have illustrated that Snail is major down-regulator for Par3 expression via its binding to PARD3 E3-box. Snail/Par3/ZO-1 axis is a key step in muscle invasion and metastasis of urothelial carcinoma which may

provide targets for treatment of bladder urothelial carcinoma.

Conclusion: In urothelial carcinoma, Sp1 was highly expressed and was related to more malignant clinicopathological phenotype and lower survival rates, while Par3 down-regulation was associated with a poor outcome. Furthermore, Sp1 and Par3 regulate cancer behaviors of urothelial carcinoma, including proliferation, invasion, and metastasis, making them prognostic indicators and novel target of urothelial carcinoma.

No conflict of interest

Financial none

REFERENCES

1. van Hoogstraten LMC, Vrieling A, van der Heijden AG, Kogevinas M, Richters A, et al. Global trends in the epidemiology of bladder cancer: challenges for public health and clinical practice. *Nat Rev Clin Oncol.*2023; 20: 287-304.
2. Lenis AT, Lec PM, Chamie K, Mshs MD. Bladder Cancer: A Review. *Jama.*2020; 324: 1980-1991.
3. Fosso E, Leo M, Muccillo L, Mandrone VM, Di Meo MC, et al. Quercetin's Dual Mode of Action to Counteract the Sp1-miR-27a Axis in Colorectal Cancer Cells. *Antioxidants* 2023;12(8): 1547
4. Safe S, Abbruzzese J, Abdelrahim M, Hedrick E. Specificity protein transcription factors and cancer: opportunities for drug development. *Cancer prevention research.*2018; 11: 371-382.
5. Safe S. Specificity Proteins (Sp) and Cancer. *International Journal of Molecular Sciences.*2023; 24: 5164.
6. Riga A, Castiglioni VG, Boxem M. New insights into apical-basal polarization in epithelia. *Curr Opin Cell Biol.*2020; 62: 1-8.
7. Williams ED, Gao D, Redfern A, Thompson EW. Controversies around epithelial-mesenchymal plasticity in cancer metastasis. *Nat Rev Cancer.* 2019; 19: 716-732.
8. Liu Z, Yang Y, Gu A, Xu J, Mao Y, et al. Par complex cluster formation mediated by phase separation. *Nat Commun.*2020; 11: 2266.
9. Jung HY, Fattet L, Tsai JH, Kajimoto T, Chang Q, et al. Apical-basal polarity inhibits epithelial-mesenchymal transition and tumour metastasis by PAR-complex-mediated SNAI1 degradation. *Nat Cell Biol.*2019; 21: 359-371
10. Wang G, McKenney JK. Urinary Bladder Pathology: World Health Organization Classification and American Joint Committee on Cancer Staging Update. *Arch Pathol Lab Med.* 2019;143(5):571-577
11. Chen Z. The Role of Specificity Protein 1 (SP1) in Bladder Cancer Progression through PTEN-Mediated AKT/mTOR Pathway. *Urol Int.* 2023; 107: 848-856.
12. Zhao Y, Yao D, Li Y, Zhang S, Tao Z, et al. Loss of polarity protein Par3 is mediated by transcription factor Sp1 in breast cancer. *Biochem Biophys Res Commun.*2021; 561: 172-179.
13. Van Hoogstraten L M, Vrieling A, van der Heijden A G, et al. Global trends in the epidemiology of bladder cancer: challenges for public health and clinical practice. *Nature Reviews Clinical Oncology*, 2023; 20(5),287-304.
14. Zhu J, Lu Z, Ke M. and Cai X. Sp1 is overexpressed and associated with progression and poor prognosis in bladder urothelial carcinoma patients. *International Urology and Nephrology*, 2022; 54(7), 505-1512.
15. Bi Q, Wang M, Zhao F, Wang M, et al. N-Butanol fraction of Wenxia formula extract inhibits the growth and invasion of non-small cell lung cancer by down-regulating Sp1-mediated MMP2 expression. *Frontiers in Pharmacology*, (2020); 11, 594744.
16. Guan H, Cai J, Zhang N, Wu J, et al. Sp1 is upregulated in human glioma, promotes MMP-2-mediated cell invasion and predicts poor clinical outcome. *International journal of cancer*, (2012); 130(3), 593-601.
17. Jiang W, Jin Z, Zhou F, Cui J, Wang L, et al. High co-expression of Sp1 and HER-2 is correlated with poor prognosis of gastric cancer patients. *Surg Oncol.* 2015; 24: 220-225.
18. Wang Q, Qian J, Wang F, Ma Z. Cellular prion protein accelerates colorectal cancer metastasis via the Fyn-SP1-SATB1 axis. *Oncol Rep* 2012; 28:2029-2034.
19. Zhao J, Ye W, Wu J, Liu L, Yang L, et al. Sp1-CD147 positive feedback loop promotes the invasion ability of ovarian cancer. *Oncol Rep.* 2015; 34: 67-76.
20. Safe S. Specificity Proteins (Sp) and Cancer. *Int J Mol Sci.* 2023; 24(6):5164..
21. Wang S, Cai J, Zhang S, Dong M, Zhang L, et al. Loss of polarity protein Par3, via transcription factor Snail, promotes bladder cancer metastasis. *Cancer Sci.*2021;112: 2625-2641.
22. Fomicheva M, Tross EM, Macara IG. Polarity proteins in oncogenesis. *Curr Opin Cell Biol.*2020; 62: 26-30.
23. Atashrazm F, and Ellis S. The polarity protein PARD3 and cancer. *Oncogene*, 2021; 40(25), 4245-4262.
24. Islam SS, Mokhtari RB, Noman AS, Uddin M, Rahman MZ, et al. Sonic hedgehog (Shh) signaling promotes tumorigenicity and stemness via activation of epithelial-to-mesenchymal transition (EMT) in bladder cancer. *Mol Carcinog.* 2016; 55: 537-551
25. Archibald A, Mihai C, Macara IG, McCaffrey L. Oncogenic suppression of apoptosis uncovers a Rac1/JNK prolifer Single- molecule trackation pathway activated by loss of Par3. *Oncogene.* 2015;34:3199-206.
26. McCaffrey LM, Montalbano J, Mihai C, Macara IG. Loss of the Par3 polarity protein promotes breast tumorigenesis and metastasis. *Cancer Cell.* 2012; 22: 601-614.
27. Song T, Tian X, Kai F, Ke J, Wei Z, et al. Loss

- of Par3 promotes lung adenocarcinoma metastasis through 14-3-3 ζ protein. *Oncotarget*. 2016;7:64260–73
28. **Zhang P, Wang S, Wang S, Qiao J, et al.** Dual function of partitioning-defective 3 in the regulation of YAP phosphorylation and activation. *Cell Discov* 2016;2:16021
29. **Baba A B, Rah B, Bhat G R, Mushtaq I, et al.** Transforming growth factor-beta (TGF- β) signaling in cancer-A betrayal within. *Frontiers in pharmacology*, 2022; 13, 791272
30. **Lee M S, Byun H J, Lee J, Jeoung D I, et al** Tetraspanin CD82 represses Sp1-mediated Snail expression and the resultant E-cadherin expression interrupts nuclear signaling of β -catenin by increasing its membrane localization. *Cellular Signalling*, 2018; 52, 83-94.
31. **Wu W S, You R I, Cheng C C, Lee M Cet al.** Snail collaborates with EGR-1 and SP-1 to directly activate transcription of MMP 9 and ZEB1. *Scientific reports*, 2017; 7(1), 17753.
- 32.

Citation:

Fouad, E., Gharieb, S., Hanna, A., Abdelaleem, M., abozaid, Ep., Abdullatif, A., Evaluation the expression of Sp1 and Par3 in urinary bladder carcinoma. *Zagazig University Medical Journal*, 2024; (2334-2344): -. doi: 10.21608/zumj.2024.284107.3347