

Veterinary Medical Journal-Giza (ISSN 1110-1423) Faculty of Veterinary Medicine, Cairo University

Accredited from national authority for Quality Assurance and Accreditation

Giza, 12211, Egypt



Antimicrobial susceptibility of coagulase negative staphylococci Isolated from bovine mastitis

Karam Alawer¹, Azzam Alkurdi¹, Ebrahim Rifai¹ and Mostafa A. Shalaby²

¹ Dept. of Microbiol., ² Dept. of Pharmacol.. Faculty of Veterinary Medicine, Cairo University, Egypt

Abstract

The present study aimed to determine the in vitro susceptibility of coagulase negative staphylococci (CNS) isolated from milk samples in cases of clinical and subclinical bovine mastitis to some antimicrobial drugs. The antibacterial sensitivity was tested using disk diffusion method and performed according to the National Committee for Clinical Laboratory Standards (NCCLS) guidelines using Mueller-Hinton agar. A total of 180 milk samples from apparently healthy cows and 130 milk samples from cows suffering from bovine mastitis were collected. Subclinical mastitis was detected using California test and bacterial isolation and identification of CNS isolates were carried out depending on the morphology of colony and biochemical tests. The results showed that a total of 368 (45.77%) isolates of CNS were obtained from clinical and subclinical mastitis. Coagulase negative staphylococci were dominant in 366 (%54.3) isolates from subclinical mastitis and only 2 isolates from clinical mastitis. The highest number of CNS isolates was susceptible to chloramphenicol (84.78%), followed by tetracycline (80.43%) and kanamycin (74.46%). Whereas, 40.76 % of CNS isolates was resistant to penicillin. Before starting the treatment of mastitis, the antibiogram for different antibacterial agents is necessary to attain the maximum efficacy of used antibiotic and to prevent emergence of bacterial resistance.

Keywords: Antimicrobial susceptibility, Coagulase negative *staphylococci*, Bovine mastitis **Corespondind Author**: E *Karam Alawer* mail. karamawer@yahoo.com

Introduction

Bovine mastitis is an inflammation of the mammary glands of dairy cows physical, chemical, accompanied by pathological and bacteriological changes in milk and the glandular tissue. It is the most common infectious diseases of dairy cows all over the world which adversely affects dairy industry (Halasa et al., 2009 and Pachauri, et al., 2013). Mastitis may be caused by a wide variety of microorganisms including bacteria, fungi, yeast and mycoplasma. However, bacteria are the most frequent pathogens causing this disease (Halasa et al., 2007). Mastitis is continuously the most expensive frequent and disease of dairy cows (Halasa et al., ²⁰⁰⁹). It is one of the

most prevalent and costly diseases in the dairy industry with losses attributable to reduced milk production, discarded milk, early culling, veterinary services, and costs (Thompson-Crispi labor al.,2014). In most cases, mastitis results migrating into bacteria from mammary gland through the teat canal. The invaded bacteria interact with the host tissue and activate the cow's immune system, promoting neutrophil migration from the blood into the milk, so leading to increase in somatic cell count (SCC) in milk (Veerle, 2011 and Archer et al., 2014). The increase in milk SCC of an individual cow or quarter above threshold is indicative for the presence of an intramammary infection (IMI) and is inversely correlated with milk production and quality (Archer et al., 2014). The cases of mastitis vary from severely clinical with visible signs of inflammation to subclinical without signs, but with an elevation of SCC. Not only clinical mastitis, but also subclinical form can result in reduction of milk production (Halasa et al., 2009 and Fuenzalida, et al., 2015).

staphylococci Coagulase-negative (CNS) have been traditionally considered to be the minor mastitis pathogens, but now it is the most bacteria commonly isolated from mastitis (Pitkälä et al., 2004; Roberson et al., 2006 and Tenhagen et al., 2006). Mastitis caused by CNS in most cases remains subclinical, or the clinical signs are mild (Taponen, 2008). One of the most important reasons for failure of treatment of mastitis is attributed to the indiscriminate use of antibiotics without the in vitro sensitivity testing causative bacterial strains. The practice at one hand increases economic losses and on the other results in the development of bacterial resistance to commonly used antimicrobial drugs (Tremblay et al., 2014). For suitable and successful antibiotic therapy, the bacterial isolation and antibiotic sensitivity tests are always essentials. Antimicrobial susceptibility tests help to guide the veterinarians in selecting the most appropriate and effective antimicrobial agent for treatment of bovine mastitis (Rüegsegger et al, 2014). Therefore, the present investigation was carried out to determine the in vitro susceptibility of CNS isolated from bovine mastitis to some antimicrobial

Material and Method

Milk samples for bacteriological examination were collected aseptically from 180 apparently healthy cows and 130 clinical from and suffering cows Samples from subclinical mastitis. subclinical mastitis were detected by California test. From each animal, 15ml of milk samples were collected, cooled and immediately transported to the laboratory on ice. The bacteriological examination was performed according to Devriese et al., (1994)

Antimicrobial sensitivity of CNS isolated from milk samples was tested method and using disk diffusion performed according to guidelines of NCCLS (2013). The plates were prepared and checked for sterility by incubating the plates overnight at 37°C. The antibiotics discs were kept at room temperature for one hour then the agar plates were with different inoculums overlaid coagulase negative staphylococcus isolates that showed turbidity equivalent to that of 0.5 McFarland turbidity tube as a standard.

Results

The sensitivity of coagulase negative *staphyloccoci* (CNS) isolated from milk samples to antimicrobial drugs are presented in Table (1) and illustrated in Figs (1) and (2).

The results of antibiotic sensitivity test of CNS isolated from mastitis are shown in Fig. (1). It clear that the prevalence of bovine mastitis was found to in 368 (45.77%) isolates of CNS which obtained from clinical and subclinical mastitis. The CNS was dominant in 366 isolates (%54.3) from subclinical mastitis, and 2 isolates from clinical mastitis. The CNS were belonging to

8 species as follows: S. chromogenes 80 (21.86%); S. xylosus 86 (23.5%); S. epidermidis 65 (17.76%); S.haemolyticus 64 (17.49%); S. sciuri 32 (8.74%); S. saprophyticus 19 (5.19%); S. hominis 18 (4.91%) and S. lentus 2 (0.55%).

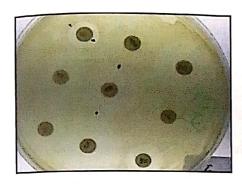
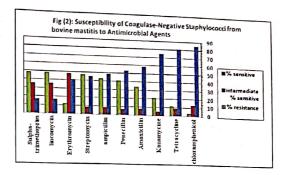


Fig.(1): Showing antibiotic sensitivity test of CNS isolated from mastitis.

As recorded in table (1) and shown in Fig. (2), the susceptibility of coagulase negative staphyloccoci (CNS) to the tested antimicrobial drugs on the isolates were in the following order: the highest numbers of CNS chloramphenicol susceptible to were (84.78%), followed by tetracycline (80.43%), kanamycin (74.46%), amoxicillin (58.15%), penicillin (52.99%), ampicillin (48.64%), erythromycin (44.84%),streptomycin (40.49%). The lowest susceptibility was shown to lincomycin (16.58%), and sulphatrimethoprim (16.58%).

Table (1): Susceptibility of CNS isolated from mastitis to some antimicrobial agents.

Antimicrobial	High sensitive		Intermediate sensitive		Resistant	
	No.	%	No.	%	No.	%
Chloramphenicol	312	84.78	46	12.5	10	2.72
Tetracycline	296	80.43	31	8.41	41	11.14
		74.46	17	4.62	77	20.92
Kanamyeln	274		23	6.25	150	40.76
Penicillin	195	52.99		8.42	158	42.93
Ampleillin	179	48.64	31	8.15	173	47.01
Streptomycin	165	14.84	30	8.15		
Lincomycin	61	16.58	130	35.33	177	48.10
Erythromycin	149	10.49	175	47.55	44	11.96
			20	7.88	125	33.97
Amoxicillin	214	58.15	29	1,		T
Sulpha- trimethoprim	61	16.58	130	35.33	177	48.10



Discussion

The present study was performed to evaluate the in *vitro* susceptibility of coagulase negative *staphylococci* (CNS) isolated from bovine mastitis to some antimicrobial drugs.

Previous studies have heen reported that the response rate of CNS to different antimicrobial drugs was qualified as poor when cure rate of mastitis less than or equal to 25% and as favorable when it attained 75% or above (Silley, et al., 2012; Moser, et al., 2012 and Tremblay et al , 2014). It has been also reported that the highest sensitivity of CNS isolated from bovine mastitis was to tetracycline chloramphenicol, kanamycin (Basappa et al., 2011). The result of our study that CNS was highly sensitive to chloramphenicol followed by tetracycline and kanamycin was closely similar to the results reported by Abdulla et al., (2011); Basappa et al., (2011); Rüegsegger, et al., (2014) and Tremblay, et al., (2014).

The results of this present study indicated the existence of CNS resistance for penicillin at a high rate (40.76%). The most significant factors affecting the cure rates from coagulase-negative staphylococci (CNS) causing bovine mastitis were the ability of CNS to produce β-lactamase (Pyorala et al., 2000 and Veerle, 2011) which inactivate

Karam Alawer

penicillin. Other mechanism of bacterial resistance of CNS to penicillin might be due to its slime-producing ability i.e. biofilm formation (Tremblay et al., 2014). The emergence of bacterial resistance to antimicrobial drugs among pathogens that affects animal health is of growing veterinary in concern pathogens Antimicrobial-resistant animals have also been considered as a potential health risk for humans (Basappa et al., 2011). However, the effectiveness of antibacterial therapy against udder pathogens depends not only on its into administration exogenous intramammary gland tissues, but it is also related to the various indigenous inhibitors in milk such as immunoglobulin (Ig), complement, lactoferrin, lactoperoxidase, etc. Many of these are activated by the Interactions inflammatory process. between antibiotics and immunological factors could lead either to augmenting the antibacterial effect on the target tissue or diminishing it (Saran and Leitner, 2000 and Basappa et al., 2011). Therefore, antimicrobials for mastitis therapy should be selected not only on bacterial sensitivity, but also for their positive interactions with the innate immune response of the mammary gland (Mazzilli and Zecconi, 2010 and Leitner et al., 2013),

Conclusion

Before starting the treatment of mastitis, the antibiogram for different antibacterial agents is necessary to attain the maximum efficacy of used antibiotic and to prevent emergence of bacterial resistance. Appropriate elimination of bacteria in mastitis requires both drug and well functioning immune defense system of the animal.

References

Abdulla, B.A., Al-Jammaly, M.M. and Sadoon, A.S. (2011): Isolation and identification of some bacteria causing subclinical mastitis in cows. Iraq. Mag. Vet. Med.; 63:67-72.

Archer, S.C., Mc Coy, F., Wapenaar, W. and Green, M.J. (2014): Association between somatic cell count during the first lactation and the cumulative milk yield of cows in Irish dairy herds. J. Dairy Sci.; 97(4):2135-2144.

Basappa, B. K., Sadashiv, S. O., Mahantesh, M. K. and Rajeshwari, D. S. (2011): Prevalence and antimicrobial susceptibility of coagulase negative *staphylococci* isolated from bovine mastitis. Veterinary World; 4(4):158-161.

Devriese, L.A., Lawvens H. F., Haesebrouck. F. and Hommez, J. (1994): A simple identification scheme for coagulase negative *staphylococci* from bovine mastitis. Res. Vet. Sci.; 57: 240-244.

Fuenzalida, M.J., Fricke, P.M. and Ruegg, P.L. (2015): The association between occurrence and severity of subclinical and clinical mastitis on pregnancies per artificial insemination at first service of Holstein cows. J. Dairy Sci.; 98(6):3791-3805.

Halasa, T. K., Huijps, O., Osteras, M. and Hogeveen, H. (2007): Economic effects of bovine mastitis and mastitis management: A review. Vet. Quart.; 29:18-31.

Halasa, T. K., Nielsen, M. A., De Roos, R., Van Hoorne, G., de Jong, T., Lam, J. G., van Werven, G.T. and Hogeveen, H. (2009): Production loss due to new subclinical mastitis in Dutch dairy cows estimated with a test-day model. J. Dairy Sci.; 92:599-606.

Leitner, G., Pinchasov, Y., Morag, E., Spanier, Y., Jacoby, S., Eliau, D., Pitcovski, J. (2013): Immunotherapy of mastitis. Vet. Immunol. Immunopathol.; 153(3): 209-216.

Mazzilli, M. and Zecconi A. (2010):

Assessment of epithelial cells' immune and

- inflammatory response to Staphylococcus aureus when exposed to a macrolide. J. Dairy Sci.; 77(4): 404-410.
- Moser, A., Stephan, R., Ziegler, D. and Johler, S. (2013): Species distribution and resistance profiles of coagulase-negative staphylococci isolated from bovine mastitis in Switzerland. Schweiz. Arch. Tierheilkd.; 155(6): 333-338.
- NCCLS (2013): National Committee for Clinical Laboratory Standards, Antimicrobial Susceptibility Testing; Twentieth Informational Supplement. Approved Standard M100-S23, Clinical and Laboratory Standards Institute, Wayne, Pennsylvania, USA.
- Pachauri, S., Varshney P., Dash, S.K. and Gupta, M.K. (2013): Involvement of fungal species in bovine mastitis in and around Mathura, India. Vet. World; 6(7):393-395. Doi:10.5455/vetworld.2013.
- Pitkälä, A., Haveri, M., Pyörälä, S., Myllys, V. and Honkanen-Buzalski, T. (2004): Bovine mastitis in Finland: Prevalence, distribution of bacteria and antimicrobial resistance. J. Dairy Sci.; 87: 2433-2441.
- Pyorala, S., Taponen, S., Jantmen, A. and Pyorala, E. (2000): Efficacy of targeted 5-day parenteral and intramammary treatment of clinical Staphylococcus aureus mastitis caused by penicillin-susceptible or penicillin-resistant bacteria strain. Proceeding of the International Symposium, Immunology of Ruminant Mammary Gland, Stresa, Italy, pp. 382-384.
- Roberson, J., Mixon, J., Oliver, S., Rohrbach, B. and Holland, R. (2006): Etiologic high SCC dairy herds. The 24th World Buiatrics Congress, Nice, France, pp. 1-19.
- Rüegsegger, F., Ruf, J., Tschuor, A., Sigrist, Y., Rosskopf, M. and Hässig, M. (2014):

- Antimicrobial susceptibility of mastitis pathogens of dairy cows in Switzerland. Schweiz. Arch. Tierheilkd.; 156(10): 483-488.
- Saran, A. and Leitner, G. (2000):
 Interactions between bacteria, immunity
 and therapy in mammary gland.
 Proceeding of International Symposium on
 Immunology of Ruminant Mammary
 Gland. Stresa, Italy, pp. 290-297.
- Silley, P., Goby, L. and Pillar, C.M. (2012): Susceptibility of coagulase negative *staphylococci* to a kanamycin and cefalexin combination. J. Dairy Sci.; 95(6):3448-3453.
- Taponen, S. (2008): Bovine mastitis caused by coagulase negative *staphylococci*. University of Helsinki, Finland.
- Tenhagen, B.A., Köster, G., Wallman, J. and Heuwieser, W. (2006): Prevalence of mastitis pathogens and their resistance against antimicrobial agents in dairy cows in Germany. J. Dairy Sci.; 89: 2542-2551.
- Thompson-Crispi, K., Atalla, H., Miglior, F. and Mallard, B.A. (2014): Bovine mastitis: frontiers in immunogenetics. Front. Imminol.; 5:493-499.
- Tremblay, Y.D., Caron, V., Blondeau, A., Messier, S. and Jacques, M. (2014): Biofilm formation by coagulase-negative staphylococci: Impact on the efficacy of antimicrobials and disinfectants commonly used in dairy farms. Vet. Microbiol.; 172(3-4):511-518.
- Veerle, P. (2011): Epidemiology and characterization of coagulase negative staphylococcus species from dairy farms. Discussion in Faculty of Veterinary Medicine, Ghent University, Germany.

حساسية العنقوديات السالبة لاتزيم التختر المعزولة من التهاب ضرع الابقار للمضادات الميكروبية \mathbf{z} كرم العوير \mathbf{z} ، عزام الكردي \mathbf{z} ، ابراهيم الرفاعي \mathbf{z} ومصطفى عباس شلبي \mathbf{z}

أقسم الأحياء النقيقة - كلية الطب البيطري - جامعة حماه - سورية قسم الأدوية - كلية الطب البيطري - جامعة القاهرة

الملخص

استهدفت هذه الدراسة قباس حساسية العقوديات السائبة لانزيم التخثر المعزولة من التهاب الضرع السريرى وتحت السريرى في الإبقار لبعض المضادات الميكروبية بطريقة الإقراص على بيئة مولر هنتون. ولقد تم تجمع عينات الحليب من ١٣٠ بقرة مصابة بالتهاب الضرع السريري ومن ١٨٠ بقرة سليمة ظاهرياً. وقد تم اجراء اختبار كاليقورنيا لتحديد التهاب الضرع تحت السريرى. وتم التعرف على المعزولات اعتماداً على الفحص الميكروسوبي والاختبارات الكيميةية. وتم زرع عينات الحليب على الأوساط الجرثومية للفحص البكتريولوجي.. وأظهرت النتاتية أن العقوديات السلبة لانزيم التغثر كانت موجود في ٣٦٨ معزولة من مجمل حالات التهاب الضرع بنسبة ٧٧,٥١%, وكانت منها ٣٦٦ معزولة من مجمل حالات التهاب الضرع بنسبة ٧٧,٥١%, وكانت منها ٣٦٦ معزولة من حالات التهاب الضرع السريري. واثبتت النتائيج أن حساسية العقوديات السلبة لانزيم التغثر للمضاد الحيوى كلورامفينيكول كانت بنسة ٤٧,١٨ % ثم التتراسيكالين بنسبة ٤٦،٨ % ثم الكالمليسين بنسبة لانزيم التغثر للمضاد الحيوى كلورامفينيكول كانت بنسبة ٤٧,١٨ % ثم التتراسيكالين بنسبة المتعروبية المنادات الميكروبية ضرورى للحصول على أعلى كفاءة في علاج التهاب الضرع ولتجنب ظهور مقاومة البكتيريا للمضاد الميكروبي.