



Microbial deterioration of a 13 AH-century manuscript housed in Al-Azhar library in Egypt: A case study

Noura Sh. A. Hagaggi^a & Taha Ayman Salah^{*b}

^a Botany Department, Faculty of Science, Aswan University, Aswan 81528, Egypt

^{b*} Conservation Department, Faculty of Archaeology, Aswan University, Aswan 81528, Egypt

* Corresponding author: E-mail address: aymansalahtaha82@yahoo.com

Abstract

Four saprophytic bacterial and fungal species attacked the manuscript that dated back to 1251 AH at Al-Azhar library in Cairo, Egypt were isolated. Based on the partial 16S rRNA gene sequence, the isolated bacteria were *Acinetobacter indicus* and *Exiguobacterium aurantiacum*. The fungal species were *Stachybotrys chartarum* and *Aspergillus flavus*. *Acacia nilotica* fruit extraction exhibited considerable antimicrobial activity at different concentrations against the isolated bacteria and fungi.

Key words: Saprophytic, manuscript, *Acacia nilotica*, antimicrobial.

Received; 4 May. 2016, In Revised form; 1 Jun. 2016, Accepted; 1 Jun. 2016

1. Introduction

Libraries keep all sources of human knowledge and cultural heritage [1], [15], [40]. Bio deterioration of ancient materials stored in libraries is a great problem over a world [8], [33], [64]. Industrial materials of paper including wood, cellulose, proteins and chemical additives represent a nutrition sources for saprophytic bacteria and fungi [7], [21], [28], [31], [38], [39], [52], [74]. In addition to ecological factors, bacteria and fungi cause degradation and discoloration of books and manuscripts stored in libraries [34], [52], [76]. Various proteolytic bacteria e.g. *Bacillus sp.*, *Staphylococcus sp.*, *Pseudomonas sp.*, *Virgibacillus sp.* and *Micromonospora sp.* were isolated from manuscripts and books [36]. Discoloration and purple spots of paper related to infection by alkaliphilic bacteria and various species of *Actinobacteria* [56], [58], [69]. Fungi represent important causative agents of deterioration of library materials [12], [37], [46], [53], [57], [62], [65], [67]. Chemical

disinfectants, ultra violet and gamma radiation used for controlling microbial contamination in libraries [19], [27], [52]. The disadvantages of these methods are paper ageing, discoloration and cancer [10], [27], [30], [53], [59], [60]. Therefore, the present study was designed to identify bacteria and fungi invading the historical 13 AH-century manuscripts and searching for simple, cheap, non-toxic and eco-friendly control against causative microorganisms. This study will introduce an effective strategy for the protection of books and manuscripts in libraries from microbial attack.

2. Material and methods

2.1. History of the manuscript

The manuscript "Resala fe alfaraid" dated back to 1251 AH at Al-Azhar library in Cairo, Egypt. Its Public number 85193 and special number 924. Its length is 16 cm² and width 11 cm² (Fig. 1).



Fig.1. the manuscript under investigation

2.2. Collection of samples

Sterile cotton swabs were wiped on the damaged area. Small fragments of paper (2-3 mm² width) were also collected from the margins of pages during restoration process to minimize invasive action on the manuscript [57].

2.3. Microbial analysis

The present work was performed in aseptic area under laminar-flow cabinet. Cotton swabs were inoculated directly on nutrient agar, Sabouraud-glucose agar and Czapek's agar plates. Multi-point inoculum method was also used to reduce the air-borne contaminants [24]. In this method small fragments of paper were washed in sterile water and divided into 25 sub-samples that were inoculated directly on nutritive agar plates. Plates were incubated at 37 °C for 5 days for bacteria and at 28 °C for 7 days for fungi. The distinct colonies were picked up and purified using streaking method. Isolates were preserved at 4 °C for identification and further studies.

2.4. Identification of isolates

The bacterial isolates were identified based on Gram's staining, morphological characteristics as well as the partial 16S rRNA gene sequence. The partial 16S rRNA was amplified using the universal primers 785 F and 907 R [73]. The reaction mixture of PCR contained 0.2 µM of each primer, 0.2 mM of dNTPs, 1X PCR buffer and 0.1 U of Taq polymerase. DNA was denatured for 5 min at 94 °C, primer annealing was at 55 °C for 1 min and strand extension was at 72 °C for 2 min. PCR products were separated using 1.5 % agarose gel. After that, the PCR products were sent to Macrogen, Inc., Korea (<http://www.macrogen.com/eng/>) for sequencing. The obtained 16S rRNA sequences were compared and aligned with the sequences deposited in the NCBI GenBank database using the BLAST tool (Blastn) that opened from the URL (www.ncbi.nlm.nih.gov).

Fungal isolates were identified microscopically on the basis of their morphologically characteristics according to the keys of Barnett and Hunter [11] and Moubasher [48].

2.5. Microbial cellulolytic activity

Cellulase activity of the bacterial isolates was detected according to Dingle et al. [20]. Colonies were streaked on carboxymethyl cellulose (CMC) agar plates composed of carboxymethyl cellulose, 10 g; NaNO₃, 3 g; K₂HPO₄, 5 g; MgSO₄.7H₂O, 5 g, Agar, 15 g and distilled water 1000 ml. pH was adjusted to 7.2. After incubation for 72 h, plates were flooded with Schulze's solution (Chlor-zinc-iodide) which consists of 1 % iodide and 3 % zinc chloride. Clear zones appeared around bacterial growth indicating CMC hydrolysis.

Cellulases production by fungal isolates was also determined following the method of Teather and Wood [71]. In the center of the CMC assay plates, 7 mm discs of 6 days old cultures of the tested fungi were inoculated. Plates were incubated at 25 °C for 7 days. After incubation, plates were flooded with 1 % Congo red and allowed to stand for 15 min at room temperature, then plates were counterstained using 1 M Na Cl. Clear zones

were appeared around fungal growth indicating cellulolytic activity.

2.6. Antimicrobial activities of the crude extract of *Acacia nilotica* fruits

2.6.1. Plant material and extraction procedure

Acacia nilotica also known as gum Arabic, Egyptian thorn in Egypt, scanted tree in English, thorn in mimosa in Australia and commonly known as “*Bagaruwa*” among the Hausas, is a species of vachellia and a genus of Acacia. It is a thorny Acacia found in different parts of the world [66]. Fruits of *Acacia nilotica* were collected from Aswan city, Egypt. Fruits were identified at the Department of Botany, Faculty of Science, Aswan University, Egypt. The fruits were washed under running tap water and air dried for 24 h. Methanolic extract was prepared from the fruits according to Alanis et al. [3]. Fruits were grounded into a fine powder by an electric blender. 50 grams of the powder was extracted with 150 ml of 99 % methanol in a conical flask, flask was shaken intermittently for 72 h at room temperature. It was then filtered through Whatman filter paper No. 1 and the solvent was evaporated at room temperature. Then methanol extract was stored at 4 °C in airtight bottles until further use.

2.6.2. Preparation of test samples

Samples used for antimicrobial assay were prepared according to NCCLS [5]. For antibacterial assay 2 g of the solid extract was dissolved in 2 ml of 99 % methanol to obtain a stock solution of 1000 mg/ml. Concentrations of 500, 250, 125, and 62.5 mg/ml were prepared using serial doubling dilution. For antifungal assay, the stock solution and diluting concentrations were prepared by the same method, but Dimethyl Sulphoxide (DMSO) was used instead of methanol.

2.6.3. Preparation of standard inoculums

The bacterial inoculum was prepared according to Mc Farland [42] and NCCLS [49]. The turbidity of 24 h old broth cultures of the bacterial isolates were adjusted to 10⁸ CFU/ml. using 0.5 Mc Farland standards.

The fungal inoculum was prepared following the method described by NCCLS [50]. Suspensions of 3 day old cultures were adjusted with UV-Visible spectrophotometer at 530 nm (OD 0.09- 0.11) to obtain standardized preparations containing approximately 10⁶ spores/ml.

2.6.4. Antibacterial activity test

Antibacterial activity of methanolic extract of *Acacia nilotica* fruits was detected using the disc diffusion method [9], [17]. Discs of Whatman filter paper No. 1 (6 mm) were prepared and sterilized by autoclaving. 10 µl of each extract concentration was loaded on discs to obtain the final concentrations 5, 2.5, 1.25 and 0.6 mg/disc respectively. 100 µl of the standardized bacterial suspension (containing 10⁸ CFU/ml) was spread uniformly on the surface of Mueller Hinton agar (Oxoid) using sterile cotton swabs. The discs were dispensed onto the surface of the inoculated agar plates. The positive control was chloramphenicol (30µg/disc) and the negative control was discs impregnated with methanol. Plates were incubated at 37 °C for 24 h. Duplicates were made. The antibacterial

activity was expressed as the mean diameter of inhibition zones in millimeter.

2.6.5. Antifungal activity test

Agar well diffusion method was used to detect the antifungal activity of the extract of *Acacia nilotica* fruits [17]. Plates containing 20 ml of Czapek's agar were uniformly inoculated with the test fungus by using sterile cotton swab. Wells of 7 mm diameter were made in the medium using sterile cork borer. 50 µl of each extract concentration was dropped into each labeled well. 50 µl of 1mg/ml fluconazole was used as positive control and 50 µl of DMSO was used as negative control. Plates were kept in the refrigerator for 1 h to allow the extracts to diffuse into the agar. Plates were then incubated at 28 °C for 72 h.

Antifungal activity was determined by measuring the diameter of inhibition zones in millimeter.

3. Results and discussion

3.1. Microbial analysis

Based on colony morphology and Gram's staining, two different isolates were selected and then sub-cultured onto a fresh nutrient agar medium for 24 h at 37 °C by the streaking method.

Blast analysis of the sequences of the present isolates with 16S rRNA gene sequences in the NCBI GenBank database, showed similarity with *Acinetobacter indicus* (96 %) and *Exiguobacterium aurantiacum* (99 %). The partial 16S rRNA gene sequences of isolates were deposited to NCBI Gen Bank under the accession numbers KX998197 and KX998198 respectively. (Fig.2-3)

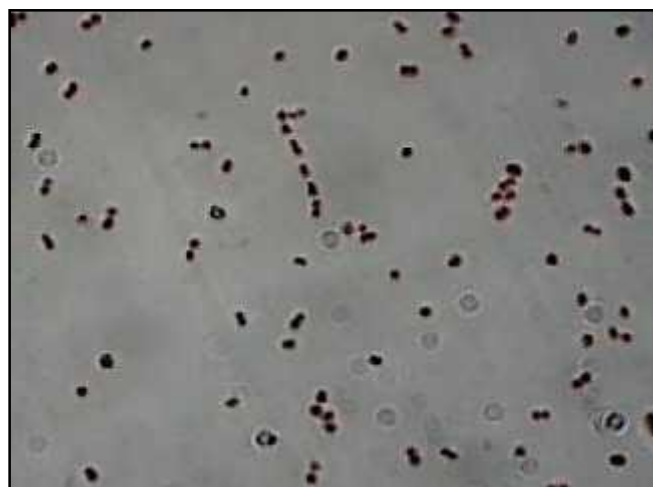


Fig.2. *Acinetobacter indicus* (1000 X)

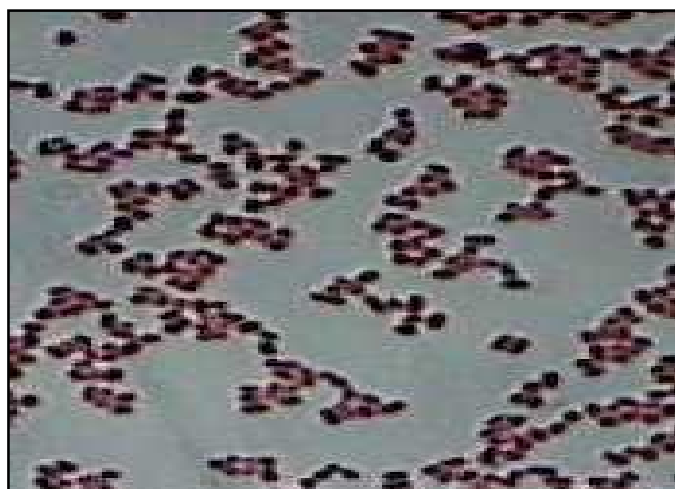


Fig.3. *Exiguobacterium aurantiacum* (1000 X)

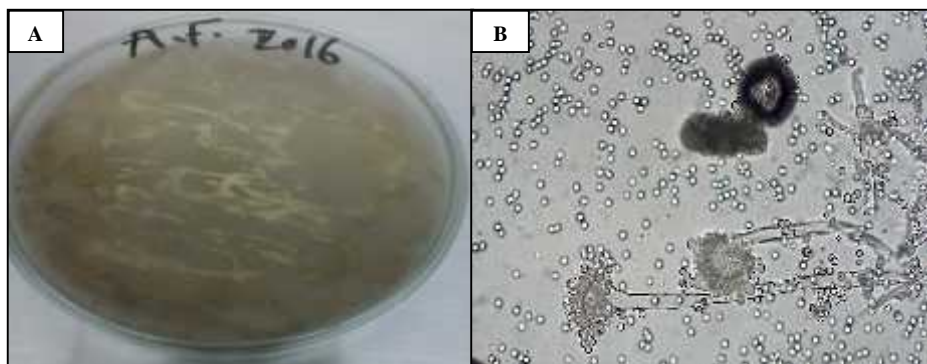
Several species of bacteria belong to *Pseudomonas*, *Cellulomonas*, *Cellvibrio*, *Myxobacteria*, *Actinomycetes*, *Streptomyces*, *Nocardia*, *Massilia timonae*, *Lysobacter dokdonensis*, *Bacillus sp.*, *Microbacterium sp.*, *Curtobacterium sp.*, *Virgibacillus* and *Brevibacterium*

pityocampae were isolated from paper and documents [4], [25], [29], [32], [36], [55], [72].

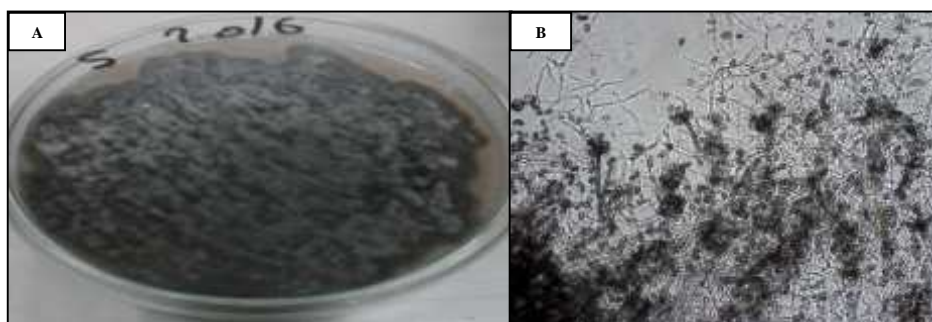
Two fungal species *Stachybotrys chartarum* and *Aspergillus flavus* were also isolated. Previously different species belong to *Penicillium*, *Mucor*, *Phoma*,

Cladosporium, *Aspergillus*, *Stachybotrys*, *Alternaria*, *Botrytis*, *Chaetomium*, *Chromelosporium*, *Epicoccum*, *Phlebiopsis* and *Toxicocladosporium* were isolated from

ancient paper and documents by many researches [36], [43], [44], [45], [47], [61], [68], [70], [76]. (Fig. 4 A & B and Fig. 5 A & B).



**Fig. 4: A: pure culture of *Aspergillus flavus* on Czapek's agar
B: *Aspergillus flavus* under light microscope (400 X)**



**Fig. 5: A: Pure culture of *Stachybotrys chartarum* on Czapek's agar
B: *Stachybotrys chartarum* under light microscope (400 X)**

3.2. Microbial cellulolytic activity

Cellulolytic activity of *Acinetobacter indicus* and *Exiguobacterium aurantiacum* was determined by streaking on CMC agar plates. It was found that the two tested bacteria exhibited strongly cellulolytic activities which indicated their effective role in the deterioration

process of the manuscript by decomposing cellulose in paper and binding textiles. Cellulases secretion was previously reported for various genera of bacteria which have been isolated from papers (Fig. 6 and 7) [2], [4], [25], [55], [63], [75].



Fig. 6 . Cellulase activity of *Acinetobacter indicus*



Fig.7. Cellulase activity of *Exiguobacterium aurantiacum*

Cellulase activity of the fungal isolates was qualitatively determined using CMC agar plates. Interestingly, both *Stachybotrys chartarum* and *Aspergillus flavus* exhibited strong ability to produce cellulases. Many cellulolytic fungi such as *Alternaria sp.*, *Aspergillus sp.*, *Fusarium sp.*, *Humicola grisea*, *Myrothecium verrucaria*, *Penicillium sp.*, *Stachybotrys sp.*, *Stemphylium sp.*, *Trichoderma sp.*, *Ulocladium sp.* and *Chaetomium sp.* were isolated from books and documents [22], [34], [35].

3.3. Antimicrobial activities of the crude extract of *Acacia nilotica* fruits

3.3.1. Antibacterial activity test

The methanolic crude extract from *Acacia nilotica* fruits was clearly inhibited the bacterial growth at all the tested concentrations (Table 1) and (Fig. 8 & 9). This correlates with the findings of previous researchers who reported that the extracts from *Acacia nilotica* fruits contain polyphenolic compounds and volatile oils which cause inhibition of a wide range of microorganisms [13], [16], [23].

Table 1

Antibacterial activity of *Acacia nilotica* fruits extract at different concentrations against the growth of *Acinetobacter indicus* and *Exiguobacterium aurantiacum*

Extraction concentration (mg/disc)	Inhibition zone (mm)	
	<i>Acinetobacter indicus</i>	<i>Exiguobacterium aurantiacum</i>
5	26	20
2.5	22	16
1.25	19	14
0.6	15	10
chloramphenicol (30µg/disc)	26	25



Fig.8. Effect of *A. nilotica* fruits extract on extract on *Exiguobacterium aurantiacum*



Fig.9. Effect of *A. nilotica* fruits *Acinetobacter indicus*

3.3.2. Antifungal activity test

Results in table (2), Fig. (10 and 11) showed that the *Acacia nilotica* fruits extract exhibited strong antifungal activity at various concentrations ranging from 500 to 62.5 mg/ml. Many researchers reported that *A. nilotica* fruits can be used as antifungal drugs due to its

high activity [41]. Chemical analysis of *A. nilotica* fruit extract exhibited that it contains highly effective compounds such as phenolic compounds, tannins, saponins and flavonoids which have a wide spectrum of antimicrobial activity [6], [18], [41].

Table 2

Antifungal activity of *Acacia nilotica* fruits extract at different concentrations against the growth of *Stachybotrys chartarum* and *Aspergillus flavus*

Extraction concentration (mg/ml)	Inhibition zone (mm)	
	<i>Stachybotrys chartarum</i>	<i>Aspergillus flavus</i>
500	25	35
250	22	32
125	20	28
62.5	14	20
Fluconazole (1mg/ml)	25	19



Fig.10. Effect of *A. nilotica* fruits extract on *Stachybotrys chartarum*

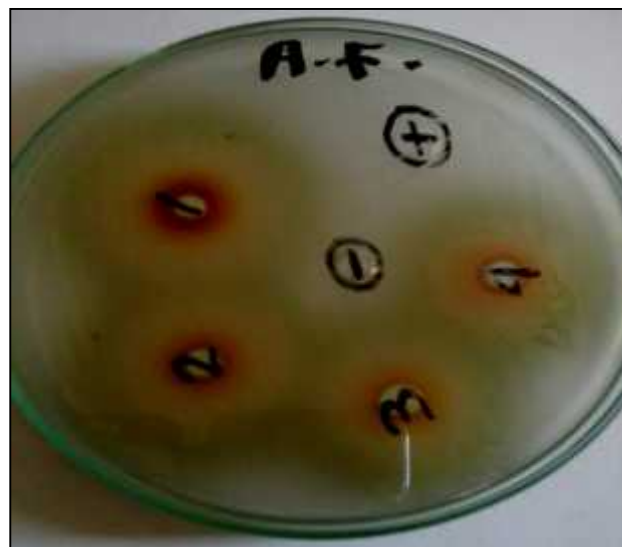


Fig.11. Effect of *A. nilotica* fruits extract on *Aspergillus flavus*

4. Conclusions

The microbial analysis of the infected manuscript "Resala fe alfaræid" which had kept in Al-Azhar library at Cairo, Egypt was carried out. Two bacterial and two fungal species were isolated which were identified as *Acinetobacter indicus*, *Exiguobacterium aurantiacum*, *Stachybotrys chartarum* and *Aspergillus flavus* respectively. Interestingly, the methanolic crude extract of *Acacia nilotica* fruits was used as non- toxic, cheap and eco- friendly control against the causative microbial isolates.

Conflicts of interest

The authors declare that there is no conflict of interest.

Acknowledgements:

Authors are introducing their sincere thanks and gratitude to the staff members of Al-Azhar library in Cairo for their kind assistance and cooperation for samples collection. Grateful thanks to Botany department, Faculty of science, Aswan University and Conservation department, Faculty of Archaeology, Aswan University for their supporting. We thank Dr. Fatma Fakhry Abdel-Motaal, lecture of mycology at Botany department, Faculty of science, Aswan University for her kind helping in the identification of fungal isolates. We thank Macrogen, Inc., Korea for helping in 16S rRNA gene sequencing.

References

- [1] Adams, J., Analysis of printing and writing papers by using direct analysis in real time mass spectrometry. *Int. J. Mass Spectrom* 301, (2011)109-126.
- [2] Aktuganov, G. E., Galimzyanova, N. F., Melent'ev, A. I., Kuz'mina, L. Y., "Extracellular hydrolases of strain *Bacillus* sp. 739 and their involvement in the lysis of micromycete cell walls," *Microbiology* 76 (4), (2007) 413–420.
- [3] Alanis, A.D., Glazada, F., Cervantes, J.A., Tarres, J. and Ceballas, G.M. Antibacterial properties of some plants used in mexican traditional medicine for treatment of gastrointestinal disorders. *Journal of Ethnopharmacology* 100 (1-2): (2005) 153-157.
- [4] Altibrandi, M.G., Il deterioramento di natura biologica. In: *Chimica e biologia applicate alla conservazione degli archivi. Pubblicazione degli Archivi di Stato, Roma, Saggi 74*, (2002) 367-379.
- [5] Altschul, S. F., Gish, W., Miller, W., Myers, E.W., Lipman, D.J., "Basic local alignment search tool." *J. Mol. Biol.* 215, (1990) 403-410.
- [6] Amjad, U. R., Abdul, S., Gul, Z., Abdul, S., Yasir, I., Abdulaziz, N., Comparative antimicrobial activity of *Acacia nilotica* L. Leaves extracts against pathogenic bacteria and fungi. *J Med Res.* 8(29), (2012) 975-82.
- [7] Area, M.C., Cheradame, H., Paper aging and degradation: Recent findings and research methods. *Bioresources* 6, (2011) 5307-5337.
- [8] Bankole, O. M., A review of biological deterioration of library materials and possible control strategies in the tropics. *Lib Rev.* 59(6), (2010) 414-29.
- [9] Bauer, A.W., Kirby, W.M., Sherris, J.C., Turck, M., Antibiotic susceptibility testing by a standardized single disc method. *Am. J. Clin. Pathol.*; 45 (4), (1966) 493-96.
- [10] Beebe, W.L., Carbo gasoline methods for disinfection of books. *J. Am. Public Health Assoc.* 1, (1911) 54-60.
- [11] Barnett, H.L., Hunter, B.B., *Illustrated Genera of Imperfect Fungi*, 3rd edition, Burgess Publishing Co, Minneapolis, (1987).
- [12] Blyskal, B., Fungi utilizing keratinous substrates. *Int Biodeterior Biodegrad* 63, (2009) 631-653.
- [13] Boakye-Yiadom, K., Fiagbe, N. I. Y., Ayim, J. S. K., Antimicrobial properties of some West African Medicinal Plants. IV. Antimicrobial activity of xylopic and other diterpenes from the fruits of *Xylopic aethiopic* (Annonaceae) *Lloydia* 40, (1977) 543–545.
- [14] Bwai, M.D., Uzama, D., Abubakar, S., Olajide, O.O., Ikokoh, P.P., Magu, J., Proximate, elemental, phytochemical and anti-fungal analysis of *Acacia nilotica* fruit. *Pharmaceutical and Biological Evaluations* 2 (3), (2015) 52-59.
- [15] Byers, B., A simple and practical fumigation system. *Abbey Newslett* 7, (1983) 1-4.
- [16] Cheesbrough, M., *Tropical Health Technology*, Cambridge University press U.K. (1984) 2:2-392.
- [17] Clinical and Laboratory Standard Institute, Performance standards for antimicrobial disk susceptibility tests. Approved standard M2–A10. Wayne, PA: Clinical and Laboratory Standards Institute. (2009)
- [18] Cowan, M. M., Analysis of phytochemicals constituents and anti-microbial activity of some medicinal plants in Tamilnadu, India. *Clin Microbiol Rev.*12(4), (1999) 564–82.
- [19] da Silva, M., Moraes, A.M.L., Nishikawa, M.M., Gatti, M.J.A., Vallim de Alencar, M.A., Brandão, L.E., Nóbrega, A., Inactivation of fungi from deteriorated paper materials by radiation, *International Biodeterioration & Biodegradation* 57(3), (2006)163-167.
- [20] Dingle, J., Reid, W. W., Solomons, G.L., The enzymic degradation of pectin and other polysaccharides. 11. Application of the 'cup-plate' assay to the estimation of enzymes. *Journal of the Science of Food and Agriculture* 4, (1953)149-155.
- [21] Doncea, S.M, Ion, R.M., Fierascui, R.C., Spectral methods for historical paper analysis: composition and age approximation. *Instrum. Sci. Tech.* 38, (2010) 96-106.
- [22] Doria, E., Altobelli, E., Girometta, C., Nielsen, E., Zhang, T., Savino, E., Evaluation of lignocellulolytic activities of ten fungal species able to degrade poplar wood. *International Biodeterioration & Biodegradation* 94, (2014)160-166.
- [23] El-Shanawny, M. A. A., Medicinal plants used in Saudi traditional Medicine, King Abdul- Aziz City for Science and Technology, Riyadh. (1996) 277.
- [24] Gallo, F., Biological factors in the deterioration of books, *Technical Notes. ICCROM, Rome.* (1985).
- [25] Gallo, F., Il biodeterioramento di libri e documenti. In: *Centro di Studi per la Conservazione della Carta. ICCROM, Roma.* (1992).
- [26] Guggenheim, S., Martin, R.T., Definition of clay and clay mineral: Journal report of the AIPEA nomenclature and CMS nomenclature committees. *Clay Miner* 43, (1995) 255-256.
- [27] Hanus, J., Gamma radiation for use in archives and libraries. *Abbey Newsletter* 9, (1985) 34–36.
- [28] Henniges, U., Schiehsser, S., Ahn, K., On the structure of the active compound in mass deacidification of paper. *Holzforchung: Int. J. Biol. Chem. Phys. Technol. Wood* 66, (2012) 447-450.
- [29] Heyndrickx, M., Lebbe, L., Kersters, K., De Vos, P., Forsyth, C., Logan, N.A., *Virgibacillus*: a new genus to accommodate *Bacillus pantothenicus* (Proom and Knight 1950). Emended description of *Virgibacillus pantothenicus*. *International Journal of Systematic Bacteriology* 48, (1998) 99-106.
- [30] Justa, P., Gamma radiation as an alternative means for disinfection of archives. *Proceeding of Second International Conference on Biodeterioration of Cultural Property, Yokohama, [Japan]*, (1992) 51-54.
- [31] Karbowska-Berent, J., Go'rný, R.L., Strzelczyk, A.B., and Wlazło, A., Airborne and dust borne microorganisms in selected Polish libraries and archives. *Building and Environment*, 46(10), (2011) 1872 -1879.
- [32] Kati, H., Ince, I.A., Demir, I., Demirbag, Z., *Brevibacterium pityocampae* sp. nov., isolated from

caterpillars of *Thaumetopoea pityocampa* (Lepidoptera, Thaumetopoeidae).

International Journal of Systematic and Evolutionary Microbiology 60, (2010) 312-316.

[33] Koestler, R. J., When bad things happen to good art. *International Biodeterioration & Biodegradation* 46 (4), (2000) 259-260.

[34] Kowalik, R., a. "Microbio deterioration of library materials. Part 1," *Restaurator*, 4, (1980) 99-114.

[35] Kowalik, R., b. "Microbio decomposition of basic organic library materials. Microbio deterioration of library materials. Part 2," *Restaurator*, 4 (34), (1980) 135-219.

[36] Kraková, L., Chovanová, K., Selim, S. A., Šimonovi ová, A., Puškarová, A., Maková, A., Pangallo, D. A multiphasic approach for investigation of the microbial diversity and its biodegradative abilities in historical paper and parchment documents. *Int. Biodeterior. Biodegrad.* 70, (2012) 117-125.

[37] López-Miras, M., Piñar, G., Romero-Noguera, J., Bolivar-Galiano, F. C., Ettenauer, J., Sterflinger, K., Martin-Sanchez, I., Microbial communities adhering to the obverse and reverse sides of an oil painting on canvas: identification and evaluation of their biodegradative potential. *Aerobiol* 29, (2013) 301-314.

[38] Lugauskas, A., and Kriktaonis, A., Microscopic fungi found in the libraries of Vilnius and factors affecting their development. *Indoor and Built Environment*, 13(3), (2004) 169-182.

[39] Mabee, W., Roy, D.N., Modeling the role of paper mill sludge in the organic carbon cycle of paper products. *Environ. Rev.* 11, (2003) 1.

[40] Maggi, O., Persiani, A.M., Gallo, F., Valenti, P., Pasquariello, G. Sclocchi, M.C., Scorrano, M., Airborne fungal spores in dust present in archives: Proposal for a detection method, new for archival materials. *Aerobiologia*, 16(3-4), (2000) 429-434.

[41] Mahesh, B., Satish, S., Antimicrobial Activity of Some Important Medicinal Plant against Plant and Human Pathogens. *World J Agricultural Sci.* 4(5), (2008) 839-43.

[42] McFarland, J., Nephelometer: An Instrument for Estimating the Number of Bacteria in Suspension Used for Calculating the Opsonic Index for Vaccines. *Journ. American Med. Assoc.*; 14: (1907) 1176-1178.

[43] Mesquita, N., Portugal, A., Videira, S., Rodríguez-Echeverría, A., Bandeira, A.M.L., Santos, M.J.A., Freitas, H., Fungal diversity in ancient documents. A case study on the Archive of the University of Coimbra. *International Biodeterioration & Biodegradation* 63, (2009) 626–629.

[44] Meynell, G.G., Newsam, R.J., Foxing, a fungal infection of paper. *Nature* 274, (1978) 466–468.

[45] Michaelsen, A., Pinzari, F., Ripka, K., Lubitz, W., Piñar, G., Application of molecular techniques for identification of fungal communities colonizing paper material. *International Biodeterioration & Biodegradation* 58, (2006) 133–141.

[46] Micheluz, A., Manente, S., Tigini, V., Prigione, V., Pinzari, F., Ravagnan, G., Varese, G. C., The extreme environment of a library: Xerophilic fungi inhabiting indoor niches. *International Biodeterioration & Biodegradation* 99, (2015) 1-7.

[47] Montemarini-Corte, A., Ferroni, A., Salvo, V.S., Isolation of fungal species from test samples and maps damaged by foxing, and correlation between these species and the environment. *International Biodeterioration & Biodegradation* 51, (2003) 167–173.

[48] Moubasher, A. H., Soil fungi in Qatar and other Arab Countries. The Centre for Scientific and Applied Research, Doha, Qatar. 1993.

[49] National Committee for Clinical Laboratory Standards, Performance Standards for Antimicrobial Disk Susceptibility Tests. 4th ed. 10:7, (1990) 10.

[50] National Committee for Clinical Laboratory Standards, Reference Method for Broth Dilution Antifungal Susceptibility Testing of Conidium-forming Filamentous Fungi: Proposed Standard M38-P. NCCLS, Wayne, PA, USA. (1998).99

[51] National committee for clinical laboratory standards: Methods for dilution, antimicrobial susceptibility tests for bacteria that grow aerobically. 5th ed. Pp 30 (2000).

[52] Nittérus, M., Fungi in archives and libraries, a literary survey. *Restaurator* 21, (2000) 25 -40.

[53] Nunes, I., Mesquita, N., Verde, S. C., Bandeira, A. M. L., Carolino, M. M., Portugal, A., Botelho, M. L., Characterization of an airborne microbial community: A case study in the archive of the University of Coimbra, Portugal, *International Biodeterioration & Biodegradation* 79, (2013) 36-41.

[54] Pavon, F., Gamma radiation as fungicide and its effects on paper. *Bull. Am. Inst. Conserv. Hist. Art Works* 16, (1975) 15-44.

[55] Pasquariello, G., Valenti, P., Maggi, O., Persiani, A.M., Carta e i processi di biodeterioramento in relazione ai materiali dei beni culturali. Da Biologia vegetale per i Beni Culturali. In: *Biodeterioramento e Conservazione*, Vol. 1. Nardini Editore, Firenze, (2005) 107- 112.

[56] Piñar, G., Pinzari, F., Sterflinger, K., Modern technologies as basis for the preservation of parchment. In: LópezMontes AM, Collado Montero F, Medina Flórez V, Espejo Arias T, García Bueno A (eds). 18th International meeting on heritage conservation. Edited by Univ. of Granada. GR 4206–2011 Granada; ISBN: 978-84-338-5339-4. (2011) 250-253.

[57] Pinzari, F., Cialei, V., Barbabietola, N., Measurement of the microaeroflora deteriorating potentialities in the indoor environments. *Preserv. Sci.* 7, (2010) 29-34.

[58] Pinzari, F., Cialei, V., Piñar, G., A case study of ancient parchment biodeterioration using variable pressure and high vacuum scanning electron microscopy. In: *Historical technology, Materials and conservation: SEM and microanalysis*. Meeks N, Cartwright C, MeekA, Mongiatti A (eds). Archetype Publications - International AcademicProjects, 1 Birdcage Walk, London, ISBN: 9781904982654. (2012).

[59] Rakotonirainy, M.S., Lavédrine, B., Screening for antifungal activity of essential oils and related compounds to control the biocontamination in libraries and archives

storage areas. *Int. Biodeter. Biodegr.* 55, (2005) 141-147.

[60] Rakotonirainy, M.S., Fohrer, F., Flieder, F., Research on fungicides for aerial disinfection by thermal fogging in libraries and archives. *Int. Biodeter. Biodegr.* 44, (1999) 133-139.

[61] Rakotonirainy, M.S., Heude, E., Lave´ rdine, B., Isolation and attempts of biomolecular characterization of fungal strains associated to foxing on a 19th century book. *Journal of Cultural Heritage* 8, (2007) 126–133.

[62] Santos, A., Cerrada, A., Garca, S., San Andres, M., Abrusci, C., Marquina, D., Application of molecular techniques to the elucidation of the microbial community structure of antique paintings. *Microb. Ecol.* 58, (2009) 692-702.

[63] Schabereiter-Gurtner, C., Pi˜nar, G., Lubitz, W., Rofleke, S., An advanced strategy to identify bacterial communities on art objects. *Journal of Microbiological Methods* 45, (2001) 77–87.

[64] Shamsian, A., Fata, A., Mohajeri, M., Ghazvini, K., Fungal contaminations in historical manuscripts at Astan Quds Museum library, Mashhad, Iran. *Int. J. Agri. Biol.* 8, (2006) 420-422. [65] Sterflinger, K., Fungi: their role in deterioration of cultural heritage. *Fungal Biol. Rev.* 24, (2010) 47-55.

[66] Soforowora LA., Medicinal plants and traditional medicine in Africa. Spectrum Books Ltd, Ibadan. Harbone, (1993) 55-71.

[67] Sterflinger, K., Pi˜nar, G. Microbial deterioration of cultural heritage and works of art -tilting at windmills? *Appl Microbiol Biotechnol* 97 (22): (2013) 9637–9646.

[68] Strzelczyk, A. B., Observations on aesthetic and structural changes induced in Polish historic objects by microorganisms. *International Biodeterioration & Biodegradation* 53 (3), (2004) 151-156.

[69] Strzelczyk, A. B., Karbowska, J., The role of Streptomycetes in the biodeterioration of historic parchment. Copernicus Univ Press, Torun, (2000) 158.

[70] Szczepanowska, H., Cavaliere, A.R., Fungal deterioration of 18th and 19th century documents: a case study of the Tilghman Family Collection, Wye House, Easton, Maryland. *International Biodeterioration & Biodegradation* 46, (2000) 245–249.

[71] Teather R.M., Wood P.J., Use of Congo red Polysaccharide interactions in enumeration and characterization of cellulolytic bacteria from the bovine rumen. *Applied and Environmental Microbiology* 43, (1982) 777-780.

[72] Tiano, P., Biodegradation of Cultural Heritage: Decay, Mechanisms and Control Methods. Seminar article. New University of Lisbon, Department of Conservation and Restoration. (2002).

[73] Weidner S, Arnold W, Puhler A "Diversity of uncultured microorganisms associated with the seagrass *Halophila stipulacea* estimated by restriction fragment length polymorphism analysis of PCR-amplified 16S rRNA genes". *Appl Environ Microbiol* 62 (3): (1996) 766–71.

[74] Xiaoyu, C., Xueren, Q., Xianhui, A., Using calcium carbonate whiskers as paper making filler. *Bioresource* 6, (2011) 2435-2447.

[75] Yin, L. J., Huang, P. S., Lin, H. H., Isolation of cellulase-producing bacteria and characterization of the cellulase from the isolated bacterium *Cellulomonas* Sp.YJ5. *J. Agric. Food Chem.* 58, (2010) 9833-9837.

[76] Zyska, B., Fungi isolated from library materials: a review of the literature. *International Biodeterioration and Biodegradation* 40, (1997) 43-51.