PREVALENCE OF GIARDIA CYSTS IN PETS, RODENTS, WITH POSSIBLE IMPLICATION IN INFANTILE DIARRHEA IN KALIOBIA GOVERNORATE

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SUMMARY

This study was carried out in Kaliobia governorate from April to December 1997 to investigate the occurrence of Giardia cysts in dogs, cats, rodents and possible implication in infantile diarrhea.

A total of 400 faecal samples were collected from stray dogs and cats, 300 from rats and mice as well as 400 from children affected with diarrhea at age group 1-10 years. The prevalence of Giardia cysts in faecal samples amounted to, 24% dogs, 11% cats, 19.66% rats and mice; 21.48% (Rattus rattus), 25.55% (Rattus norvegicus) and 9.33% (Mus musculus). Giardia lamblia cysts were found in 21% in children affected with diarrhea.

Experimental infection of different species of rats and mice (Rattus rattus, Rattus norvegicus and Mus musculus) with Giardia cysts obtained from diseased children revealed no difference in morphological character of infective stage and emphasized the interspecies transmission concept of Giardia and its public health importance.

INTRODUCTION

Giardia is a flagellate protozoan showing worldwide ubiquity. On the basis of the difference in shape of the median bodies there appears to be at least three species: G. lamblia (duodenalis, intestinalis) in man and several other mammalian species, G.muris in mice and G. ranae in frogs (Bowman and Lynn, 1995).

Giardia lamblia is the commonest enteric pathogen causing gastroenteritis in children (Melloni et al. 1993) and causes outbreaks of diarrhea in children in day care centers (Pickering et al., 1984).

There is no evidence for host specific genotype of Giardia isolated from different host species

such as sheep, calves, dog and man (Ey et al. 1996).

Therefore, the present study has been carried out to throw light on the occurrence of Giardia in dogs, cats, rodents and children in Kaliobia governorate and the possibility of interspecies transmission of Giardia between children and rodents.

MATERIALS AND METHODS

This investigation was carried out in Kaliobia Governorate duing the period from April to December 1997.

Sampling:

Faecal samples were collected from 400 stray dogs and cats. 300 rats and mice were trapped alive by ordinary wire cage spring trap from different localities and transferred to laboratory where they were anaesthesized and identified according to Meehan (1984). Faecal samples were collected for microscopic examination. Moreover 400 faecal samples were collected from children at age group 1-10 years who were complaining from diarrhea and presented to the pediatric division, governmental hospitals where clinical data were collected.

Procedure:

The collected faecal samples were examined by direct wet mount method (Bowman and Lynn, 1995), flotation technique with 33% zinc sulphate (Soulsby, Formol-ether concentration 1982). technique (WHO, 1983), as well as iron hematoxylin staining method using Schaudinn's

solution fixative (Markell et al., 1992) were adopted for microscopic examination.

Experimental Infection:

inoculation of samples were Preparation and carried out according to Hewlett et al. (1982).

The inoculated samples were confirmed by observation of wet preparation of stools from infected children, the specimen was dispersed in 0.95% NaCl and filtered through four layers of gauze. The number of cysts in the resultant suspension was determined and the cysts were used within 24 hrs of collection.

Experimental infection was carried out after exclusion of infective rats and mice with Giardia cyst where each 7 of the same species were kept in separate cage and supplied with feed and water. Each one of the same group of the same species was infected orally with 0.2 ml of suspension of Giardia cysts obtained from infected children. 5 inoculated and 2 left without infection as control.

Daily examination of faecal materials of experimentally infected Rattus rattus, Rattus norvegicus and Mus musculus were containued until clearance of cyst excretion for a minimum of 30 days. Cysts excreted by experimentaly infected rats and mice were examined morphologically and were indistinguishable from the cysts used for inoculation. Inoculated rats and mice that did not become infected were killed and their duodenal contents were examined directly for Giardia.

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RESULTS AND DISCUSSION

Table (1) shows the prevalence of Giardia cyst in faecal samples of dogs and cats which amounted to (24%) and (11%) respectively.

The results achieved were higher than those recorded by Suobodova et al., (1995), while lies between those reported in Perth, Western Australia (Swan and Thompson, 1986) and in NewJersy, USA (Acha and Szyfres, 1992).

From the available literature dogs were implicated as an important reservoir of human giardiasis in Costa Rica (Aguilar et al., 1988), London (Sykes and Fox, 1989), Brisbane, Australia (Collyer et al. 1992), and Konya, Turkiye (Guclu and Aydenizoz, 1995). In Egypt several researches detected Giardia in stray dogs, in Ismailia city (Abou-Eisha and Abdel-Aal, 1995), Sharkia (Magda-Amin, 1992) and in carnivora in zoological gardens in Giza (Siam et al. 1994).

Table (2) displayed the occurrence of Giardia cysts (19.66%) in trapped rats and mice; 21.48, 25.55 and 9.33% in Rattus rattus, Rattus norvegicus and Mus musculus respectively. A finding substantiates what had been reported by Meywer and Jarrol (1982).

Faecal samples were collected from children affected with diarrhea at age group 1-10 years.

Table (3) shows a prevalence of 21% of Giardia lamblia cysts in children with diarrhea. The results obtained support what had been conducted

in Jamaica (Christie et al., 1988).

Giardia lamblia was detected in (22%) in preschool children complaining from diarrhea at age group 1-5 years. A finding which is almost to similar those obtained in Houston day care centers (Pickering et al. 1984) and in Lesotho preschool children (Esrey et al. 1989), where as it was higher than those reported in San Vincente University Paul Hospital, Colombia (Hermandez et al. 1987) and in Ghana (Annan, 1985), and lower than those found in Turkiye (Kocman et al., 1982), Bangladesh (Islam et al. 1983) and in Bilbies, Egypt (Dupont and Sullivan 1986).

In the present study Giardia lambia is implicated as a cause of diarrhea in children at age group 5-10 years with a prevalence rate of 20%. A result obtained was lower than that recorded in children in public school in Chile (Cobrera et al. 1981).

Regarding experimental infection of rats and mice with Giardia lamblia cysts procured from children (Table 4), the prepatent and patent periods were nearly similar in Rattus rattus, Rattus norvegicus and Mus musculus after infection. Rats and mice inoculated with Giardia cysts and did not respond to a infection were killed and no Giardia cysts could be detected in their dvodenal contents.

From the available literature, several studies reported the isolation of Giardia lamblia cysts from human and succeeded in experimental infection of animals. Mongorel dogs were experimentally infected with G.lamblia cysts

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Table (1): Prevalence of Giardia cysts among examined pets in Kaliobia Governorate

Species of animal	Examined number	Positive samples		
		No.	%	
Dogs Cats	200 200	48 22	24 22	
Total	400	70	17.5	

Table (2): Prevalence of Giardia cysts among examined rodents in Kaliobia Governorate

Species of	Examined	Positive samples		
rodents	number	No.	%	
Rattus rattus	135	29	21.48	
Rattus norvegicus	90	23	25.55	
Mus musculus	75	7	9.33	
Total	300	59	19.66	

Table (3): Prevalence of Giardia cysts among examined children affected with diarrhea in in Kaliobia Governorate

Age group/year	Examined number	Positive samples	
group/year		No.	%
1-5 5-10	200 200	44 40	22 20
Total	400	84	21

Table (4): Results of experimental infection of rodents with Giardia cysts recovered from children.

Species of rodents	No. of infective rats & mice/ control	No. of + ve/infective	Prepatent period/d.	Patent period/d.
Rattus rattus	5/2	3/5	7-12	19-23
Rattus norvegicus	5/2	4/5	6-10	18-30
Mus musculus	5/2	2/5	8-14	16-19

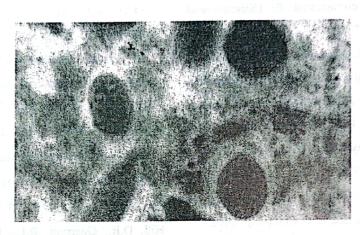


Fig (1): Giardia lamblia cyst from human (X1250)

from human stool and trophozites from axenic culture (Hewlett et al. 1982). Mongolian gerbils (Belosevic et al. 1983) and Swiss mice (Hill et al. 1983) responded to infection with Giardia lamblia cysts recovered from human. Moreover Siam et al. (1994) succeeded in experimental infection of Swiss mice with Giardia cysts obtained from zoo carnivora and their attendants.

In addition, recent researches indicated no difference between genotype of Giardia isolates obtained from human and animals and concluded that man and animals do not seem to possess their own unique species (Bertram et al. 1983, Nash et al. 1985 and Ey et al. 1996).

From Zoonotic point of view, infection with Giardia lamblia is not restricted to humans and can be transmitted to animals and possibly back to humans. Dogs and cats as pets should be considered as reservoir of infection. Rodents play a role in spread of infection through contamination of food and water sources.

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