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ANTIMICROBIAL ACTIVITY OF NIGELLA SATIVA AND ZINGBER OFFICINALE

BY

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SUMMARY

Aqueous and ethanolic extracts of Zingber officinale had antimicrobial activity against large number of pathogenic bacteria and fungi.

Aqueous extract of Nigella sativa possessed antibacterial activity in higher concentration. In addition all fractions of both Zingber officinale and Nigella sativa exhibited antimicrobial activities against various microorganisms.

Saponin and oil fractions of the test plants increased the anitbacterial acitivity of ampicillin against pyogenic microorganisms while resin fraction decreased that activity.

INTRODUCTION

Nigella sativa and Zingber officinale exhibit antimicrobial activities against large number of pathogenic microorganisms (Sinha et al 1977. saxena and Vyas 1986. Hassan et al., 1989 and Mascolo et al., 1989). Alkaloids, saponins and volatile oil (Essential oil) of the tested plants had antimicrobial activity (El-Fatatry et al. 1975. Rathee et al., 1982, Sauza et al., 1987 and Akgul 1989).

The present work was designed to investigate the antimicrobial activity of aqueous and ethanolic extracts of Nigella sativa and Zingber officinale as well as saponin, alkaloid and oil of Nigella sativa and saponin, resin and volatile oil of Zingber officinal. Moreover to study their therapeutic uses.

MATERIAL AND METHODS:

Preparation of the tested materials:

Aqueous and ehtanolic extracts were prepared by complete exhausion of Nigelia sativa seeds and Zingber officinale rhizomes with distilled water or ethanol. The solvents were evaporated under reduced pressure till complete dryness. The crude extracts were disolved in tween 80 and then diluted with distilled water to 10-200 mg/ml.

Saponin of the tested plants was isolated and purified according to Basu and Rastorgi (1967) and Sandermann (1962). Extraction of resin (Baily and Pridhan 1962), alkaloid (Seiber, 1970 and Mangold 1969). Essential oil (Nursten 1970) was also carried. out The latter was identified according to (Kaldwey, 1969). Each isolate was dissolved in tween 80 and diluted with distilled water to concentrations of 10-1000 ug/ml.

The prepared concentrations were poured (0.2 ml in each pore) in plates and incubated at optimal temperature for suitable time of each tested organism.

Bacterial strains:

1- Gram positive bacteria.

Staphylococcus aureus, Streptococcus pyogens and Corynebacterium pyogens.

2- Gram negative bacteria:

Salmonella typhimurium, Escherichia coli, Psendomonas aeruginosa and Klebsiella pneumoniae.

Fungi:

A- Moulds:

Aspergillus niger, aspergillus flavus and Penicillum spp.

B- Dermatophytes:

Microsporum canis, Microsporum gypseum.

c- Yeast:

Candida albicans all the tested microorganisms were obtained by personal contact from the Department of Microbiology faculty of et, Med. Cairo University. Antibacterial activity of the graded concentrations of both aqueous and ethanolic extracts (10-200 mg/ml) and isolates principale (Alkaloid, saponin, resin and oils 10-1000 ug/ml) of Nigella sativa and Zingber officinale was studied by the pore method as described by Cooper and Woodman (1964) using nutrient agar No I and borer No 8 with dianeter (8 mm± 0.1 mm). The plates were incubated at 37°C for 18 hours. Antifungal activity of the studied extracts and isolates were studied in vitro as described by Robell and Lamb (1953) using sabaroud agar medium and the plates were incubated at 25°C for 3 days for mould species and 21 days for dermatophytes.

Nutrient agar No 1		Sabaroud agar
Peptone	6.0	glucose 40
Pancreatic digest of	casine 4.0	peptone 10
Yeast extract	3.0	Agar 20
Beef extract	1.5	water to 1000
Dextrose	1.0	
Agar	15.0	
Distilled water to	1000.0	
PH 6.55±0.05		

Concomitant combiations of the studied fractions (1 mg/ml) with ampicillin (1 ug/ml) were tested against certain pathogenic bacteria (Streptococcus pyogens, Salmonella typhimurium and

Corynebacterium pyogens) using the samethod.

RESULTS

The obtained data showed that aqueous a ethanolic extracts of Zingber officinale concentrations ranged from 10 to 200 mg/exhibited a powerful antibacterial and antifugactivity against, the tested pathogenic a nonpathogenic bacteria and fungi (Table 1-2) addition, aqueous extract of Nigella san possessed antibacterial activity in concentration over 50 mg/ml especially against Staphylocoa aureus, Streptococcus pyogens, E. coli a Pseudomonas. aeruginosa, while the ethanone had no effect. Both extracts of Nigella san at the tested concentrations had no antifugactivity in vitro.

Both antibacterial and antifungal activities of studied fractions of Zingber officinale and Nigsativa are recorded in tables (3-6). It is clearly volatile oil of Zingber officinale is the most activation against the tested bacterial and functions while saponin and resin fractions are active than volatile oil (Table 3).

Saponin, alkaloid and fixed oil fraction of Nissativa in concentrations over 50 ug/ml exhibit potent antibacterial activity especially agu Streptococcus pyogens. E. coli and Salmond typhimurium (Table 4) Furthermore sapos alkaloid and oil fractions of nigella sativic concentrations over lmg/ml had the ability inhibit the growth of Candida albical Aspergillus niger. penicillium special Microsporum gypseum and Microsporum can recorded in table (6).

Our findings showed that saponin and oil fraction of Nigella sativa and Zingber officinale increase the antibacterial activity of ampicillin in against Streptococcus pyogens, Salme typhimurium and Corynebacterium pyogens the other hand resin fraction of the tested precessed the antibacterial activity of ampicilling

DISCUSSION

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Table 1:In vitro antibacterial activity of ethanolic and aqueous extracts of Zingber officinale.

Microorganism	Concentration	Diameter of the ihibition Zone in (mm) ±S		
	mg/ml	Ethanolic	Aqueous	
Staphylococcus	10			
aureus	25	12.0 ± 0.50		
unitina	50	13.5 ± 0.67		
	100	15.0 ± 0.50	13.30 ± 0.67	
	200	17.67 ± 0.33	17.67 ± 0.88	
	200	17.07 ± 0.55	17.07 ± 0.86	
Streptococcus	10			
pyogens	25	12.5 ± 0.29	·	
and the second second	50	14.0 ± 0.29		
	100	17.0 ± 0.58		
	200	20.67 ± 0.67	15.67 ± 0.67	
E.Coli	10			
E.Con	25	1 - 1 - 1 1 - 1 - 1 - 1 - 1 - 1 - 1 - 1		
	50			
	100		12.33 ± 0.33	
	200		15.63 ± 0.57	
S-1	10	3-9-10.		
Salmonella	25			
Typhimurium	50	11.33 ± 0.33		
		13.0 ± 0.00	10.00 ± 0.00	
	100		14.67 ± 0.33	
	200	14.67 ± 0.67	14.07 ± 0.33	
Klebsiella	10		-	
pneumoniae	25	12.67 ± 0.17		
	50	14.83 ± 0.17		
	100	16.67 ± 0.58	12.33 ± 0.33	
	200	21.20 ± 0.49	18.32 ± 0.88	
Pseudomonus	10			
gerueinosa	25			
MEL HETTI AND	50			
	100		12.33 ± 0.33	
	200		15.67 ± 0.67	
Corynebacterium	10	12.02 . 0.44		
pyogens	25	13.83 ± 0.44		
	50	15.50 ± 0.50	**	
	100	17.0 ± 0.58		
	200	20.67 ± 0.67		

Table (2): In vitro anitfungal activity ethanolic and aqueous extracts of Zingber officinale.

Microorganism	Concentration	Diameter of the in ihibition Zone in (mm) 2 S.E.		
	mg/ml	Ethanolic	Aqueous	
Candida albicans	10		Por Labor.	
	25		-	
	50	11.66 ± 0.33		
	100	14.00 ± 0.58	11.00 ± 0.58	
	200	15.67 ± 0.67	17.33 ± 0.67	
Asperigillus	10			
niger	25			
	50			
40 (100)	100	11.67 ± 0.88		
	200	16.00 ±0.58		
Asperigillus	10	••		
falvous	25			
	50	11.33 ± 0.67		
100	100	14.00 ± 0.58		
	200	17.00 ± 0.58		
Pencillium Spp	10		des pro-	
	25			
	50		13.00 ± 0.33	
	100	11.33 ±0.33	18.00 ± 0.58	
	200	14.00 ± 0.58	23.33 ± 0.88	
Microsporum	10		-	
gypseum	25			
	50	••		
	100		15.33 ± 0.33	
	200		18.67 ± 0.67	
Microsporum	10			
canis	25	••		
	50	••	12.67± 0.33	
	100		18.67 ± 0.67	
	200		22.33 ± 1.20	

Table(3): In vitro antibacterial activity of Zingber officinale isolated fractions.

Microorganism	Concentration	Diameter of the ihibition Zone in (mm) ±S.		
	mg/ml	Saponin	Resin	Volatile oil
Staphylococcus	10			
aureus	50	7	••	••
	100			$12.0 \neq 0.57$
	1000		35° •• 17	14.5 ± 0.47
	5000		ad in the	15.0 ± 0.00
Streptococcus	10	70- Table 1	8" ++	
pyogens	50		••	11.7 ± 0.16
	100	12.2 ± 0.33	10.5 ± 0.16	13.0 ± 0.00
	1000	15.5 ± 0.57	13.2 ± 0.33	15.5 ± 0.28
	5000	17.0 ± 0.33	15.0 ± 0.00	19.0 ± 0.50
E.Coli	10			11.0 ± 0.00
1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1	50		••	13.0 ± 0.16
	100	11.5 ± 0.47	11.0 ± 0.00	14.5 ± 0.28
	1000	13.5 ± 0.57	13.5 ± 0.47	17.2 ± 0.33
	5000	15.2 ± 0.88	17.0 ± 0.00	20.0 ± 0.57
Salmonella	10	-		man .
Typhimurium	50			11.2 ± 0.13
	100	4 15.	10.5 ± 0.16	13.0 ± 0.57
	1000	10.5 ± 0.16	13.3 ± 0.33	16.3 ± 0.33
	5000	13.0 ± 0.57	15.0 ± 0.57	18.0 ± 0.00
Kiebsiella	10			
pneumoniae	50		••	••
	100	11.0 ± 0.10		••
	1000	14.5 ± 0.47	••	12.0 ± 0.47
	5000	16.2 ± 0.33	•	14.2 ± 0.50
Pseudomonus	10			10.4
aeruginosa	50		••	
	100			11.5 ± 0.15
	1000	••	10.5 ± 0.15	13.0 ± 0.28
	5000	•	12.0 ± 0.00	14.2 ± 0.33
Corynebacterium	10			10.3 ± 0.15
prozens	50			12.2 ± 0.33
	100			13.2 ± 0.33
	1000	11.5 ± 0.28		15.7 ± 0.47
	5000	13.0 ± 0.15		17.0 ± 0.00

Table(4): In vitro antibacterial activity of Nigella sativa isolated fractions

Microorganism	Concentration	Diameter of the ihibition Zone in (mm) ±S.		
•	mg/ml	Saponin	Alkaloid	Volatile oil
Staphylococcus	10			
gureus	50			
MILL THE	100	12.0 ± 0.00		11.5 ± 0.28
	1000	15.0 ± 0.28	$10.5 \neq 0.16$	14.5 ± 0.33
	5000	17.3 ± 0.17	12.0 ± 0.16	17.0 ≠ 0.00
Streptococcus	10			
procens	50	11.3 ± 0.16		10.3 ± 1.60
	100	13.0 ± 0.16		12.5 ±0.00
	1000	16.3 ± 0.33	12.7 ± 0.78	14.0 ± 0.16
	5000	19.5 ± 0.57	14.5 ± 0.16	19.0 ± 0.28
E.Coli	10			
	50	11.3 ± 0.33	11.5 ± 0.00	••
	100	13.0 ± 0.10	12.0 ± 0.16	12.0 ± 0.00
	1000	16.6 ± 0.28	14.5 ± 0.28	15.5 ± 0.57
	5000	18.0 ± 0.16	17.2 ± 0.16	18.7 ± 0.68
Salmonella	10			
Typhimurium	50			
	100		10.5 ± 0.00	11.0 ± 0.00
a figure in the same	1000	12.3 ± 0.33	13.3 ± 0.16	13.2 ± 0.16
	5000	14.0 ± 0.16	16.0 ± 0.28	15.5 ± 0.47
Klebsiella	10			
pneumoniae	50			
	100			12.2 ± 0.16
	1000		10.5 ± 0.16	14.5 ± 0.47
	5000	12.0 ± 0.00	12.7 = 0.60	17.2 ± 0.33
Pseudomonus	10			
aerusinosa	50			11.5 ± 0.28
	100	10.0 ± 0.47	••	13.0 ± 0.16
	1000	13.0 ± 0.00	12.3 ± 0.16	16.5 ± 0.97
	5000	14.5 ± 0.28	13.0 ± 0.00	20.0 ± 0.57
Correctacterium	10			
STOREGIE	50			
	160	11.0 ± 0.00	12.0 ± 0.16	12.0 ± 0.28
	1600	13.0 ± 0.16	14.0 ± 0.57	14.5 ± 0.16
	5000	14.3 ± 0.33	15.3 ± 0.33	17.0 ± 0.33

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Table(5): In vitro antibacterial activity of Zingber officinale isolated fractions.

Microorganism	Concentration	Diameter of the in ihibition Zone in (mm): S.E. ±S.E.		
	mg/ml	Saponin	Resin	Volatile oil
Candida	1	11.0 ± 0.00	_	12.2 = 0.33
albicans	5	14.7 = 0.57	11.9 ± 0.73	17.4 = 0.41
	10	18.2 ± 0.33	14.2 ± 0.33	21.0 = 1.50
1-40	25	20.0 ± 0.28	17.2 ± 0.33	25.9 = 0.57
	50	23.7 ± 0.57	20.0 = 0.00	29.2 ± 0.33
Asperieillus	1		_	-
niger	5	- 1	-	-
meer	10		-	10.5 = 0.10
and the state of	25	- 1	12.2 ± 0.33	13.2 ± 0.33
	50		15.0 ± 0.00	17.5 = 0.20
Asperieillus	1	_	_	_
falvous	5	-		
Jan Villa	10	- 1	-	11.9 = 0.57
	25	10.0 ± 0.00	11.2 ± 0.30	14.3 ± 0.70
1	50	12.5 ± 0.10	13.5 ± 0.27	17.0 ± 0.00
Pencillium Spp	1			-
	5	10.0 ± 0.00		11.2 = 0.27
	10	12.5 ± 0.57		13.2 ± 0.33
The second of	25	14.2 ± 0.33	11.5 ± 0.50	17.7 ± 0.57
	50	173 ± 0.39	13.1 = 0.43	22.0 ± 1.50
Microsporum	1		-	-
gypseum	5	10.5 ± 0.30	-	121 = 0.30
	10	12.0 ± 0.00	10.5 = 0.01	13.0 ± 0.50
THE PERSON NAMED IN	25	14.2 ± 0.33	12.0 = 0.30	16.8 = 0.44
The second	50	16.8 ± 0.73	14.5 = 0.00	18.7 = 0.61
Microsporum	1			11.0 ± 0.57
canis	5	11.0 ± 0.00	••	14.0 ± 0.00
	10	13.2 ± 0.33	11.5 ± 0.57	18.2 = 0.33
	25	17.9 = 0.50	143 = 0.44	25.7 = 0.33
1.400	50	23.2 ± 0.33	17.0 ± 0.00	29.6 ± 0.88

Table(6): In vitre antifungal activity of Nigella sativa isolated frations.

Microorganism	Concentration	Diameter of the inhibition Zone in (mm) ± S.E. ±S.E.		
	mg/ml	Saponin	Alkaloid	Fixed oil
Candida	1	12.5 ± 0.17	11.5 ± 0.90	13.0 ± 0.16
albicans	5	15.2 ± 0.33	13.5 ± 0.44	17.2 ± 0.33
	10	18.7 ±0.57	15.2 ± 0.33	21.7 ± 0.57
	25	21.0 = 0.67	17.9 ± 0.47	30.1 ± 0.88
	50	25.9 ± 0.83	21.2 ± 0.5	39.5 ± 1.87
Asperigillus	1			12.5 ± 0.10
niger	5			14.6 ± 0.16
	10	10.7 ± 0.13	11.5 ± 0.88	19.8 ± 0.33
	25	12.5 ± 0.30	13.2 ± 0.43	25.2 ± 0.88
	50	15.0 ± 0.50	11.5 ± 0.27	33.7 ± 0.67
Asperigillus	1			11.5 ± 0.33
falvous	5	10.7 = 0.30	$12.0 \neq 0.10$	13.5 ± 0.44
	10	12.7 ± 0.44	14.2 ± 0.33	16.5 ± 0.51
	25	13.7 ± 0.80	17.9 = 0.57	19.7 ± 0.56
	50	15.2 ± 0.33	23.7 ± 0.88	25.0 ± 0.67
Pencillium Spp	1			
	5	10.5 ± 0.30	12.0 ± 0.10	12.5 ± 0.16
	10	12.4 ± 0.44	13.2 ± 0.33	15.4 ± 0.28
	25	13.5 ± 0.33	15.9 ± 0.88	19.5 = 0.33
	50	17.0 ± 0.57	19.5 ± 0.70	22.7 ± 0.88
Microsporum	1	11.5 ± 0.17	12.5 ± 0.70	14.0 ±16
Expseum	5	13.1 ± 0.37	14.2 ± 0.33	18.5 ± 0.33
	10	16.1 ± 0.27	18.6 ± 0.90	20.0 ± 0.28
	25	21.9 ± 0.44	22.7 ± 0.64	27.2 ± 0.57
100	50	27.0 ± 0.78	29.2 ± 0.88	35.3 ± 0.58
Microsporum	1			11.0 ± 0.00
canis	5	12.1 ± 0.17	13.5 ± 0.37	13.2 ± 0.33
	10	14.0 ± 1.00	15.7 ± 0.80	163 ± 0.16
	25	16.0 ± 0.64	18.2 ± 0.33	21.0 = 0.88
	50	20.6 ± 0.84	22.7 ± 0.88	32.0 ± 0.67

Nigella sativa & Zingber officinale

Table(7): In vitro study on the activity of ampicillin (1 ug/ml), isolated tested plant fractions (1 mg/ml) and their combination against three bacterial strains.

			Diameter of inihibition Zone		
Plant	Isolate	Strain	Ampicillin	Fraction	Ampicillin + Fraction
	Saponin	Strept.	24.1 ± 0.3	28.7 ± 0.7	35.9 ± 0.3
		Sal.	12.5 ± 0.0	14.1 ± 0.2	14.1 ± 0.3
		Coryne.	47.7 ± 0.3	40.1 ± 0.9	50.5 ± 0.9
Zingber officinale	Volatile	Strept.	16.0 ± 0.0	18.0 ±0.1	22.3 ± 0.2
ojjie	oil	Sal.	13.3 ± 0.3	28.0±0.0	20.1 ± 0.3
		Coryne.	45.3 ± 0.7	40.0 ±0.3	46.7 ± 0.9
	Saponin	Strept.	25.7 ± 0.3	40.0 ± 0.3	40.0 ± 0.3
2 4 4 1	Suponin	Sal.	12.0 ± 0.0	14.0 = 0.1	16.5 ± 0.9
100		Coryne.	45.7 ± 0.6	44.0 ± 0.3	-47.5 ± 0.9
Nigella	Alkaloid	Strept.	26.0 ± 0.2	40.7 ± 0.3	45.2 ± 1.5
stiva	Aikaioia	Sal.	13.5 ± 0.7	18.2 ± 0.2	16.1 ± 0.1
Jura		Coryne.	44.3 ± 0.9	40.3 ± 0.5	40.3 ± 0.4
	Fixed oil	Strept.	26.7 ± 0.3	25.7 ± 0.3	25.6 ± 0.4
		Sal.	13.1 ± 0.2	14.0 ± 0.0	-
		Coryne.	44.7 ± 0.8	45.5 ± 0.7	45.6 ± 0.3

Streptococcus pyogens. Salmonella typhimurium. Corynebacterium pyogens.

The obtained data proved that aqueous and ethanolic extracts of Zingber officinale exhibit a potent antimicrobial activity in low concentrations. This effect may be attributed to the presence of saponin, resin and volatile oil as it has been previously recorded by Kaur and Sinha (1982) and Sauza et al., (1987). In addition, the antimicrobial activity of both extracts of Zingber officinale recorded here was similar to that finding recorded by Gugnani and Ezenwanze (1985), and Mascolo et al (1989).

Our findings showed that saponin, resin and volatile oil fractions of Zingber officinale exhibited a more potent antibacterial and antifungal activities against several pathogenic and nonpathogenic bacterial and fungal strains in vitro. The present antimicrobial activities of equeous, ethanolic extracts and fractions was in consistence with the findings recorded by El-Fatatry et al (1975). Rathee et al., (1982), Saxena and Vays (1986) and Hassan et al (1989).

The ability of aqueous and ethanolic extracts as well as the tested fractions of Zingber officinale to inhibit the growth and multiplication of various pathogenic fungi and bacteria, encouraged the authors to use these materials as a chemotherapeutic agents in treatment of different diseases as ring worm, wound infection and diahrriea in veterinary medicine but these trials still need further field application study to demonestrate the ability of these extracts and fractions for treatment of systemic infections and also to determine the proper form and dose for use on larg scale.

The obtained results demonstrated that aqueous extract of Nigella sative seeds has a potent intibacterial activity against pathogenic bacteria. In addition, alkaloids, saponin and fixed oil fractions of Nigella sativa seeds exhibit very strong antibacterial and antifungal activities so they can be used effectively in therapy in the form

of solution.

The present data showed that saponin and oil fractions of Nigella sativa and Zingber Officinale increased the antibacterial activity of ampicillin. This effect may be attributed to a tested synergestic action with amipicillin. On the other hand, resin fraction of the tested plants decreased the antibacterial effect of ampicillin against pathogenic bacteria. This may be referred to the formation of less active compound during their combination resulted in antagonestic effect, therfore this fraction must be used alone as antibacterial substance.

Nearly the same synerge-stic activity between Nigella sativa seeds extracts and streptomycin and gentamycin antibiotics was recorded by Hanafy and Hatem (1991).

Conclusively the extracts and isolates of Zingber Officinale and Nigella sativa are effective against pathogenic bacteria and fungi and they are safer than other antibacterials of synthetic origin.

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