

PERFORMANCE AND TISSUE RESIDUE OF TILAPIAS FED DIETARY AFLATOXIN

BY

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INTRODUCTION

Sarotherodon niloticus is a native African fish and has been reared in ponds, since ancient Egyptian times. Murals and engravings dating from 1400 B.C. show the fish held in ornamental ponds (Hepher and Pruginn, 1981). The tilapia species have become increasingly important in fish culture especially in warm climates. Diets can negatively influence the healthy growth of fish either by nutrient deficiencies, imbalances, introducing infective agents or toxins to fish. The mould *Asperigillus flavus* grows well on many feedstuffs and under appropriate temperature and humidity produce a very potent carcinogen of aflatoxin (Friedman and Shibko, 1972). Although the effect of feeding on feeds contained aflatoxin to livestock are well documented (Allcrart, 1969), much less is known about these effects on fish. Rainbow trout is the most sensitive animal to aflatoxin known, showing a significant incidence of hepatoma at dietary level as low as

0.5 ppb (Ashley et al., 1965). Contaminated aflatoxin B₁ (AFB₁) was reported as the cause of trout liver cancer (Halver, 1967), also Halver (1969) found that, prolonged feeding of AFB₁ at concentration as low as 0.4µg/kg diet, caused liver neoplasma, necrosis of hepatocytes and degenerative changes in pancreatic tissues and kidney of rainbow trout. Concerning channel catfish, Ashley (1966) and Friedman and Shibko (1972) reported that, catfish fed crude aflatoxin in diets ranging from 0 to 100 mg/kg body weight showed a relatively low response, while Jantrotor and Loveil (1990) found in channel catfish fed 10, 000µg/kg feed for 10 weeks, had shown a significant decrease in growth rate haematocrite, hemoglobin concentration, erythrocyt count and necrotic hepatocytes but not when fish fed the concentration of 2154µg/kg or less. In carp fish, at low dietary level (2ppb) AFB₁, had no effect on body weight, health physiology (Svobodova and Piskac, 1980).

The principle target organ in aflatoxicosis is the liver. The broad range of biological effect of aflatoxin properly related to their reaction with cell nucleoprotein and nucleic acid and the ultimate effect of these reaction on protein synthesis and cellular integrity (Patterson, 1976). Evidence from several developing countries showing an increase in incidence in aflatoxin content of the diet points to aflatoxin as a cause of human liver cancer.

This study was planned to provide more information on the potential risk involved the feeding tilapia species on aflatoxin contaminated feed. The objectives were to evaluate the effect of feeding of 50, 100 and 200 ppb aflatoxin on tilapias performance throughout 10 weeks feeding experiment. Moreover it is worth to detect aflatoxin residue in fish tissues.

MATERIAL AND METHODS

Table (1): Composition of the experimental ingredients % %.

Ingredients	Moisture	CP	EE	Ash
SBM	11.0	44.0	1.3	6.0
Fish meal	8.0	65.0	6.7	16.2
Wheat bran	11.0	15.0	1.4	7.0
Yellow corn	10.5	8.8	3.0	13

Diet formulation: Fish meal, SBM, wheat bran and yellow corn was done were the ingredients used

for diet formulation. The actual chemical analysis of these ingredients (Table, 1) according to A.O.A.C. (1975) Diet ingredients were analyzed to ensure freedom of AFB₁ before they were thoroughly mixed, pelleted and used in feeding trials.

Experimental diets: Contaminated

Table (2): Experimental diets in different treatments.

Ingredients (%)	Control diet	Aflatoxin level (ppb)		
		50	100	200
SBM	28	28	28	28
Fish meal	20	20	20	20
Wheat bran	20	20	20	20
Yellow corn	28	27	25.08	24.2
Contaminated y.corn	--	1.0	2.92	3.8
Mineral mixture	2.0	2.0	2.0	2.0
Vitamin mixture	2.0	2.0	2.0	2.0
Total	100	100	100	100
CALCULATED ANALYSIS %				
CP	30.8	30.8	30.8	30.8
EE	2.8	2.8	2.8	2.8
Ash	6.7	6.7	6.7	6.7
CP	4.7	4.7	4.7	4.7
Ca	1.24	1.24	1.24	1.24
P	1.13	1.13	1.13	1.13
ME Kcal /Kg	3359	3359	3359	3359
Cal/protein ratio	109	109	109	109

* Mineral mix. was prepared by mixing 50% dicalcium phosphate (25% Ca., 8% P) + 25% flamecal trace element premix + 25% NaCl. Each Kg. contains Ca 25 g., P 90 g., Fe 25000mg., Cu 2000 mg., Mn 6000 mg., Se 100 mg., Zn 40000 mg., NaCl 5000 mg.
* Each Kg contains: Vit.A 4000000 IU, Vit.D 8000 IU, Vit.E 10000mg, Vit.B₁ 2000 mg., Vit.B₂ 10000 mg., Vit.B₆ 1000 mg., Vit.B₁₂ 3 mg., Vit.C 10 mg., Folic acid 500 mg., Pantothenic acid 500 mg., Vit.K 1000mg and nicotine amide 5000 mg.

contaminated yellow corn was used in formulating the experimental diet contained 5200µg/aflatoxin/kg was prepared as described by El-Banna (1987). This corn used to formulate 3 experimental diets contained 50,100 and 200 ppb aflatoxin (Table, 2).

Laboratory conditions: Twelve 120-L glass aquaria equipped with an air pump, maintained at 28±2°C by using electric heaters 70 watts whilst PH throughout the experimental period was 7.5 ± 0.4 which are the optimal recommended

Tilapia species were used. Tap water was used and treated by anti-chlor reagent according to Boyd (1979).

Fish: Tilapia fingerlings were obtained from El-Abassa Fish Hatchery. They were divided into 4 groups each of 3 replicate with 10 fingerlings. The average initial body weight was 5.9 ± 0.22 g, while the initial standard length was 6.04 ± 0.08 cm.

Feeding regime: Fish were fed twice daily at a rate of 3% of body weight (Marek, 1975), which was adjusted in accordance to body weight change during 10 weeks experimental period. Each aquarium was drained and thoroughly cleaned weekly.

Performance Parameters: All fish of each group were weighed weekly, also standard length was measured, while weight gain and feed/gain were calculated. At the end of experimental period all alive fish in each group were killed, whole body was dried, ground and prepared to chemical analysis according to A.O.A.C. (1975). Three fish of each group were killed and examined internally for signs of aflatoxicosis. Histological examination was done by taking specimens from gills and liver from 3 fish of each group and fixed in 10% formaline solution, dehydrated, cleaned and embedded in paraffine wax blocks then sectioned at 5

micron, stained by haematoxylin-eosin stain (Carlton et al., 1967). Detection of aflatoxin residue: After 7 weeks feeding trials, 2 fish from each treatment also 3 fish from each treatment at the end of 10 weeks experimental period were taken to detect the presence of aflatoxin residue according to the method of A.O.A.C. (1975).

RESULTS AND DISCUSSION

Aflatoxins are potent hepatotoxins and also potent carcinogenic. In general, young animals of any species are more susceptible to acute toxic effect of aflatoxin than older animals of the same species (Edd, 1973).

Aflatoxicosis and tilapia performance:

Effect of AFB₁ on tilapia performance found in table (3). Concerning weight gain, fish fed the control diet was the best (5.46g) throughout the 10 weeks experimental period. Fish fed either 50 or 200 ppb AFB₁ showed nearly similar body weight gain (5.16 and 5.18g respectively). Diet contained 100 ppb AFB₁ showed the lowest weight gain (4.64g). About fish length, fish fed the control diet showed the highest increase in length (0.75 cm/fish). A similar increase in length was found in

both groups fed 50 or 200 ppb AFB₁ (0.54 and 0.51 cm/fish respectively). The least increase in fish length was in group fed 100 ppb of AFB₁. Growth rate (GR) was similar (0.9) in all groups except that fed 100 ppb of AFB₁ it was only 0.8. Regarding the mortality, only fish fed the highest level of AFB₁ (200ppb) showed

Table (3): Effect of aflatoxin on tilapia performance.

	Control diet	Aflatoxin level ppb		
		50	100	200
Initial weight (g/fish)	5.96 1.36	5.6 1.08	6.09 0.98	6.03 1.16
Final weight (g/fish)	11.42 1.49	10.77 0.89	10.70 0.61	11.2 2.3
Weight gain (g/fish)	5.64 ^a	5.16 ^a	4.64 ^b	5.18 ^a
Initial length (cm/fish)	6.04 0.61	5.93 0.9	6.13 0.72	6.06 0.05
Final length (cm/fish)	6.79 0.99	6.47 1.02	6.56 0.77	6.57 0.81
Increase in length (cm/fish)	0.75 ^a	0.54 ^a	0.43 ^b	0.51 ^a
Mortality %	0.0	0.0	0.0	16.7
Growth rate	0.89	0.9	0.79	0.86

Each mean represents 10 fish/treatment. a,b means with different superscripts are significantly different at (P<0.05)

16.7% mortality. There is no available literature dealing with the effect of aflatoxin on tilapia performance. The adverse effect that have been shown in fish fed the diet contained AFB₁ at level of 100 ppb regarding tilapia performance that was not clear in fish fed the level of 200 ppb of AFB₁, may be related to the high mortality rate of 16.7% in the later group, so fish of higher body weight may overcome the adverse effect of AFB₁ than fish of smaller body weight.

Effect of aflatoxin on Feed/gain:

Calculated feed/gain in different

Table (4): Effect of aflatoxin on gain in body mass, feed consumption and feed conversion in experimental groups (average of 3 replications with 10 fish each)

	Control diet	Aflatoxin level ppb		
		50	100	200
Gain in body mass (g/group)	163.8	154.8	139.2	99.35
Feed consumption (g/group)	447.0	412.2	426.5	356.95
Feed/gain (g/g)	2.73	2.66	3.05	3.59

groups was found in table (4). Data showed that fish fed the control diet and that fed the lowest AFB₁ level had a similar values (2.75 and 2.66 respectively) while feed/gain was increased with increase the dose of AFB₁ (3.05 and 3.59 respectively) in fish fed 100 and

Table (5): Effect of aflatoxin on whole body consumption of tilapia fish.

%	Control diet	Aflatoxin level ppb			initial
		50	100	200	
Dry matter	25.54 1.05	25.73 0.56	25.51 2.46	25.18 0.23	20.78 0.9
CP	61.37 1.42	62.83 0.74	61.83 0.1	63.53 0.18	62.94 0.51
EE	26.1 0.13	21.18 0.62	26.4 0.19	25.85 0.81	18.0 1.2
Ash	13.5 0.4	15.6 0.9	15.5 0.7	16.9 1.0	12.2 0.8

200ppb AFB₁. At the end of 10 weeks experimental period, the weight of fish fed the highest level of AFB₁ was 20% less than the fish fed the control diet. In carp, Svabodova et al. (1982) reported that neither concentration of AFB₁ (20 or 200µg/kg feed) affected feed intake of increase the mortality.

Effect of AFB₁ on fish body composition:

The data concerning fish body composition is found in table (5). All values concerning moisture, CP, EE and ash% were within the normal values of tilapia body composition.

Aflatoxin residue in fish body:

The results in table (6) showed that the amount of AFB₁ residues in fish body were directly related to both the level of AFB₁ and the duration of feeding period. The greatest amount of AFB₁ residue (31.5 ng/g) was detected in fish fed 200 ppb AFB₁ for 10 weeks. It can be observed that, with as minimum as 50 ppb AFB₁ in tilapia diet, the tissue residue was high (13.4ng/g), therefore the calculated feed/tissue ratio for this level was 3.7 which is considerably very narrow when compared with that reported for laying hens. For example, Stoloff (1977) who reported that aflatoxin have been detected in the eggs of

Table (6): Aflatoxin residue and feed/tissue in whole fish body (g/g).

	AFB ₁ residue after		Feed/tissue after	
	7 weeks	10 weeks	7 weeks	10 weeks
Control diet	Nil	Nil	Nil	Nil
50 ppb AFB ₁	6.53	13.4	7.67	3.7
100 ppb AFB ₁	13.10	26.71	7.63	3.7
200 ppb AFB ₁	17.82	31.50	11.22	6.34

laying hen receiving in their rations 2200ug/kg of AFB₁ in ration will show up to 1ug/kg of AFB₁ in eggs. An interesting point to recognized is the identical effect that

produced by two levels of AFB₁ (50 and 100ppb) on the feed/tissue ratio (3.7). Regarding duration of AFB₁ feeding, it is noticed that up to 7 weeks of feeding, a positive relationship between the level of AFB₁ and the residue found in fish tissue, However, AFB₁ feeding for 10 weeks demonstrated duplication of the amount or residue that found in the same group after 7 weeks feeding period. This effect empha-

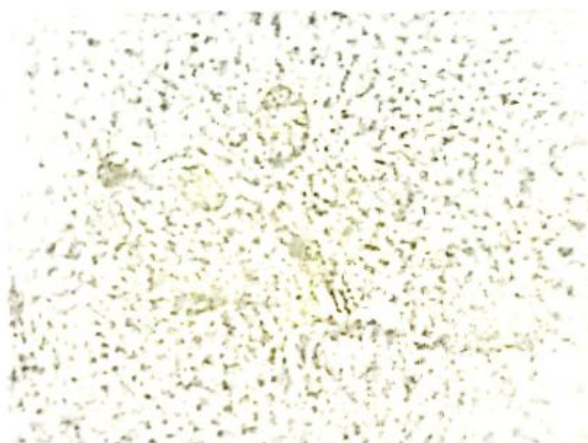


Fig.(1) : Liver of fish fed 50 ppb AFB₁ showing vacuolation of hepatocytes with necrotic debris with the bile canaliculi

sised the cumulative effect or AFB₁ in tissue which was supported by feed tissue ratio.

Pathological effect of AFB₁:

Liver of fish that fed 50ppb AFB₁ after 7 weeks feeding period revealed vacular degeneration of hepatocytes, slight activation of epithelial lining the bile duct with the presence of necrotic debris

within the bile canaliculi (Fig. 1) together with slight congestion of central vein. At end of low weeks feeding period we found the same picture, moreover there were focal coagulative necrosis and areas of intravascular haemolysis. Gills showed telangectasia or the fine blood capillaries of the secondary lamellae together with proliferation or the epithelial lining of the secondary lamellae.

Liver of fish fed 100ppb AFB₁; 7 weeks feeding period, showed proliferative changes of the epithelial lining the bile duct, leucocytic infiltration in the portal areas, focal areas of congestion and activation of melanine carrying cells. After 10 weeks feeding period found the same picture was noticed in addition to dissolution of cytoplasm of hepatocytes together with marked appearance of vascular channels (sinusoids) Channels takes rays appearance. Gills showed slight proliferative changes in the epithelial lining of the lamellae with severe congestion or the main branchial blood vessels (Fig. 2).

Fish fed the highest level of AFB₁ 200 ppb after 7 weeks feeding period, their liver showed focal areas of vacuolar degeneration leucocytic infiltration in the portal areas, the vaculation of hepatocytes was more pronounced and bile duct proliferation was very clear. After

10 weeks feeding, the same picture was noticed in addition to intravascular haemolysis associated

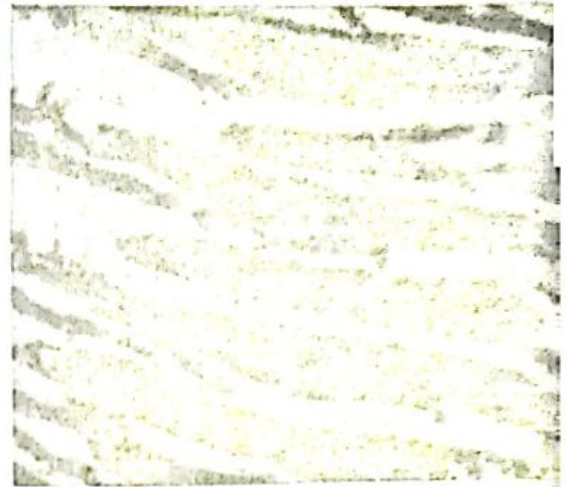


Fig.(2): Gills showing congestion of the main branchial blood vessels

with focal areas of necrosis. Gills showed proliferative changes in the epithelial lining with severe congestion of the main branchial blood vessels. These findings indicate that the gills showed no specific reduction while in liver there is a negative effect which is related to both AFB₁ level in the diet and feeding period which agree with the findings dealing with performance and residue found in fish tissue.

The most important aim of the food control organization in any country is the protection of the consumers from health hazards posed by feeds containing pathogenic microorganisms or their toxins, in amount sufficient to cause disease or toxicosis. From this work, it can be concluded that, tilapia fed as low as 50ppb of AFB₁,

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were normal in appearance and performance although they contain high amount of AFB₁ residue in their body that hazard human health. further studies, on levels below 50ppb AFB₁, are needed to detect the critical level that produce a residue in fish body.

SUMMARY

In 10 weeks feeding period, tilapia fingerlings were fed on diets contained 0.50, 100 and 100ppb of aflatoxin B₁ (AFB₁). Fish fed on diet contained 50ppb of AFB₁ showed little or no effect on fish performance, but the negative effect was increased by both level of AFB₁ and the duration of feeding period. These levels of AFB₁ showed no effect on fish body composition. Histopathological examination revealed an adverse effect on liver, not in gills, that related to both AFB₁ level and duration of feeding. Even with as low as 50ppb of AFB₁, its residue was detected in fish body in higher amount than that reported in other species. AFB₁ residue showed a cumulative effect related to level of AFB₁ and feeding period.

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