

AGE-RELATED CHANGES AND SEASONAL VARIATIONS IN THE DAILY SPERM PRODUCTION OF THE STALLION

BY

MOSTAFA M. ABOU-AHMED

Department of Theriogenology, Faculty of Veterinary Medicine, Cairo University,
Giza 12211, Egypt.

(Received: 5.10.1992).

INTRODUCTION

Although a variety of methods are available for quantifying rates of spermatozoal production, each of which entails certain assumptions and/or limitations (Berndtson, 1977; Berndtson et al., 1983). However, estimates of sperm production rates by quantitative histological analysis are imperative for clear and full evaluation of the effects of environments, drugs or other agents upon the gametogenic function of the testis (Amann, 1970).

In stallions, distinct effects of age and season on many aspects of reproduction have been documented (See Johnson and Neaves 1981). However, the corresponding influences of age and season on the quantitative expression of spermatogenesis have received limited attention. Such information may be useful in predicting a stallion's reproductive capacity and in establishing managerial practices which maximize reproductive efficiency in stallions.

Therefore, the present investigation was intended to: 1-determine the stallion's daily sperm production by a quantitative histological technique; and, 2-evaluate the extent of the changes due to age and season on daily sperm production of Arab and native horses in Egypt.

MATERIAL AND METHODS

Animals and thier assignment to groups:

The current study was carried out on 56 testes obtained from 28 Arab and native stallions along a complete annual cycle. No information was available on their reproductive history or breeding activity before castration. These animals were surgically castrated at the Department of Surgery, Faculty of Veterinary Medicine, Giza and the Veterinary Hospital of the Armed Forces, Cairo. The stallions were clinically sound and their testes were normal as determined by morphological and microscopi-

cal examinations. The age of each stallion was estimated by examination of the teeth and ranged from 3 to 18 years.

Age-related data were assigned to four groups: group I (3-5 yrs, n = 4), group II (6-8 yrs, n = 8), group III (9-13 yrs, n = 9) and group IV (14-18 yrs, n = 7 stallions). Concerning the influence of season, two seasonal periods were designated: breeding season (December-May, n=15) and non-breeding season (June-November, n=13 stallions).

Immediately after castration, the testes from each stallion were weighed. The tunica albuginea was removed and weighed, and the parenchyma weight was determined as the difference. Small pieces of testicular tissue were fixed in Bouin's solution and subsequently used for histological evaluations.

Histological analysis:

The fixed testicular tissues were processed by the usual histological technique. Paraffin sections of 5-7 μ thickness were made, stained with Periodic acid-Schiff's reagent and counterstained with hematoxylin.

Determination of the daily sperm production from quantitative histological data required the use of a technique proposed by Amann and Almquist (1962). The amount

of shrinkage associated with histological processing was determined using Archimedes, principle. The density of testicular tissue resulted from dividing the testis weight by the testis volume. The percentage of seminiferous tubules in the testis was determined by Chalkley's technique (1943). Stage I of the seminiferous epithelium cycle was characterized and determined in tubules from the complete disappearance of luminal spermatozoa to the onset of elongation of spermatid nuclei (Swierstra et al., 1974). The spermatid nuclei present in essentially round stage I seminiferous tubule cross-section was enumerated in 50 tubules per stallion. The resulting crude counts were corrected to true counts, where true count = crude count X (section thickness) \div section thickness + nuclear diameter (Berndtson, 1977). The area of stage I seminiferous tubule cross-section was calculated from the diameter of tubule cross-section evaluated using an ocular micrometer. The duration (12.2 days) of one cycle of the seminiferous epithelium of the stallion (Swierstra et al., 1974) has been used for calculation of DSP in the present study. The results were averaged to provide an estimate for each stallion.

Statistical analysis:

The effect of consecutive ages were tested by the one-way analysis of variance. The Student's "t"

test was used for statistical interpretation of the seasonal data. The interactions amongst the studied parameters were estimated by correlation coefficients. All statistical methods were done according to Snedecor and Cochran (1976).

RESULT

As no significant differences were found between the parameters studied for the right and left testes or between Arab and native horses, the results were tabulated irrespective of testis side and breed.

The pattern of changes in testicular and parenchymal weight due to age are shown in Table (1). Age variations in testicular and parenchymal weights were significant ($P < 0.05$). On the other hand, age and each of testis weight and parenchymal weight were positively correlated ($P < 0.01$) up to 8 years old stallions, but a reverse correlations were noted with the advancement of age (Table 2).

The mean density of testicular tissues was 1.040 (range 1.034 to 1.048). Neither age nor season significantly influence the densities of tissues either within stallions or among the testes of stallions within groups (Tables 1 & 3).

Tunica albuginea weight and percent value showed a consistent and gradual increase over the

whole age range studied. Although, tunica albuginea weight was not significantly influenced by age, the percent value was significantly ($P < 0.05$) affected. Moreover, age and tunica albuginea percent were correlated (Table 2).

The amount of shrinkage due to histological processing was 42% (range 39 to 45%) with no significant age and/or seasonal variations in the amount of shrinkage within stallions or among the testes of stallions within groups.

A gradual increase in the percent of the seminiferous tubules in the testis was reported up to 13 years old, followed by a slight drop thereafter (Table 1). Age variations in the percentage of seminiferous tubules did not reach the level of statistical significance. Also, correlation coefficients between age and seminiferous tubules percent was below the level of significance (Table 2).

The corrected number of spermatids per stage I tubule cross section was 50.03 at 3 to 5 years, attained a maximum of 71.40 in group III and declined (54.69) thereafter. Such variations being highly ($P < 0.01$) significant. Moreover, age and number of spermatids per stage I tubule were highly correlated up to 13 years old stallions (Table 2).

Diameter of seminiferous tubules

Sperm production

Table (1) : Age-related changes in daily sperm production of Arab and native horses (\pm SD) .

| Parameter | Group I (3-5 yrs.) | Group II (6-8 yrs.) | Group III (9-13 yrs.) | Group IV (14-18 yrs.) |
|----------------------------------------------------------------------|-----------------------|------------------------|--------------------------|--------------------------|
| Number of stallions | 4 | 8 | 9 | 7 |
| Testicular weight (g) | 166.65 \pm 19.00 | 185.55 \pm 20.74 | 176.47 \pm 16.05 | 151.34* \pm 19.04 |
| Testicular density | 1.041 \pm 0.004 | 1.044 \pm 0.002 | 1.039 \pm 0.003 | 1.036 \pm 0.003 |
| Parenchymal weight (g) | 149.50 \pm 17.57 | 166.81 \pm 19.47 | 157.10 \pm 13.69 | 131.66 \pm 17.26 |
| Tunica albuginea weight (g) | 17.15 \pm 2.49 | 18.74 \pm 2.18 | 19.40 \pm 2.62 | 19.89 \pm 2.81 |
| Tunica albuginea (%) | 10.34 \pm 1.21 | 10.16 \pm 1.06 | 10.97 \pm 0.64 | 13.16* \pm 1.10 |
| Seminiferous tubules (%) | 61.83 \pm 7.91 | 63.01 \pm 5.23 | 64.39 \pm 5.59 | 58.98 \pm 3.41 |
| Corrected number of spermatids / stage I tubule cross-section | 50.03 \pm 3.43 | 56.61 \pm 4.54 | 71.40 \pm 4.95 | 54.69** \pm 5.44 |
| Diameter of seminiferous tubules/ stage I (μ m) | 204.33 \pm 6.43 | 209.29 \pm 11.27 | 221.26 \pm 10.50 | 200.87* \pm 9.28 |
| Daily sperm production (DSP): DSP/testis ($\times 10^9$) | 3.75 \pm 0.80 | 4.56 \pm 0.86 | 5.30 \pm 0.67 | 3.38** \pm 0.81 |
| DSP/stallion ($\times 10^9$) | 7.50 \pm 1.60 | 9.11 \pm 1.72 | 10.60 \pm 1.35 | 6.76** \pm 1.63 |
| DSP/g parenchyma ($\times 10^6$) | 24.74 \pm 3.40 | 27.14 \pm 2.52 | 33.67 \pm 2.30 | 25.38** \pm 3.61 |

** Significant at 1% level & * Significant at 5% level

per stage I tubule displayed a consistent pattern of increase from group I through group III (Table 1). Age influenced ($P < 0.05$) and partially correlated with the diameter of seminiferous tubules (Tables

1 & 2).

Similar to the results of testis weight, percentage of seminiferous tubules, corrected number of spermatids and diameter of seminifer-

Table (2) : Correlation coefficients amongst the studied parameters .

| Parameters | Testis wt. | Parench. wt. | T.alb. (%) | SNT (%) | No.of spermatid | Diameter of SNT | DSP/ testis | DSP/ g. |
|-------------------------|------------|--------------|------------|---------|-----------------|-----------------|-------------|---------|
| Age | | | | | | | | |
| ≤8 yr. ^a | 0.434* | 0.427* | -0.472** | 0.099 | 0.433* | 0.229 | 0.400* | 0.344 |
| ≤13 yr. ^b | 0.091 | 0.054 | 0.134 | 0.228 | 0.839** | 0.554** | 0.614** | 0.747** |
| overall ^c | -0.412* | -0.469** | 0.643** | 0.234 | -0.032 | -0.106 | -0.259* | -0.097 |
| DSP/g | | | | | | | | |
| 8 yr. | 0.800** | 0.804** | -0.380* | 0.580** | 0.815** | 0.091 | 0.958* | |
| 13 yr. | 0.305* | 0.277 | 0.193 | 0.524** | 0.916** | 0.438** | 0.912** | |
| overall | 0.475** | 0.473** | -0.281* | 0.602** | 0.932** | 0.532** | 0.926** | |
| DSP/testis | | | | | | | | |
| 8 yr. | 0.950** | 0.933** | -0.415* | 0.649** | 0.800** | 0.285 | | |
| 13 yr. | 0.661** | 0.639** | 0.071 | 0.692** | 0.775** | 0.536** | | |
| overall | 0.797** | 0.795** | -0.441** | 0.764** | 0.819** | 0.630** | | |
| Diameter of SNT | | | | | | | | |
| 8 yr. | 0.479** | 0.468** | -0.061 | 0.245 | 0.360 | | | |
| 13 yr. | 0.438** | 0.402** | 0.241 | 0.505** | 0.522** | | | |
| overall | 0.559** | 0.545** | -0.209 | 0.562** | 0.604** | | | |
| No. of spermatid | | | | | | | | |
| 8 yr. | 0.657** | 0.678** | -0.493** | 0.134 | | | | |
| 13 yr. | 0.144 | 0.119 | 0.194 | 0.227 | | | | |
| overall | 0.344** | 0.348** | -0.270* | 0.338** | | | | |
| SNT(%) | | | | | | | | |
| 8 yr. | 0.727** | 0.695** | -0.026 | | | | | |
| 13 yr. | 0.662** | 0.629** | 0.161 | | | | | |
| overall | 0.797** | 0.701** | -0.230 | | | | | |
| T.alb.(%) | | | | | | | | |
| 8 yr. | -0.368 | -0.446* | | | | | | |
| 13 yr. | -0.169 | -0.267 | | | | | | |
| overall | -0.507** | -0.539** | | | | | | |
| DSP/g | | | | | | | | |
| 8 yr. | 0.997** | | | | | | | |
| 13 yr. | 0.996** | | | | | | | |
| overall | 0.994** | | | | | | | |

** Significant at 1% level. & * Significant at 5% level
^a_n = 12 ^b_n = 21 ^c_n = 28

Sperm production

Table (3) : Seasonal variations in daily sperm production of Arab and native horses (\pm SD) .

| Parameter | Non-breeding season | Breeding season | Overall mean |
|------------------------------------------------------------|-----------------------|-------------------------|-----------------------|
| Number of stallions | 13 | 15 | 28 |
| Age (Years) | 9.46 \pm 4.48 | 10.13 \pm 3.31 | 9.80 \pm 4.23 |
| Testicular weight (g) | 158.85 \pm 19.57 | 182.24** \pm 17.45 | 171.38 \pm 22.00 |
| Testicular density | 1.041 \pm 0.003 | 1.039 \pm 0.004 | 1.040 \pm 0.004 |
| Parenchymal weight (g) | 141.05 \pm 18.83 | 162.29** \pm 16.39 | 152.42 \pm 20.87 |
| Tunica albuginea weight (g) | 18.34 \pm 1.52 | 19.70 \pm 2.75 | 19.01 \pm 2.55 |
| Tunica albuginea (%) | 11.54 \pm 1.60 | 10.82 \pm 1.37 | 11.20 \pm 1.51 |
| Seminiferous tubules (%) | 60.00 \pm 5.26 | 64.25 \pm 7.05 | 62.35 \pm 6.26 |
| Corrected number of spermatids/stage I tubule crosssection | 54.98 \pm 7.32 | 64.51** \pm 8.14 | 60.10 \pm 9.28 |
| Diameter of seminiferous tubules/stage I (μ m) | 201.31 \pm 6.69 | 218.10** \pm 11.14 | 210.32 \pm 12.77 |
| <u>Daily sperm production (DSP):</u> | | | |
| DSP/testis ($\times 10^9$) | 3.66 \pm 0.85 | 5.01** \pm 0.89 | 4.39 \pm 1.12 |
| DSP/Stallion ($\times 10^9$) | 7.33 \pm 1.70 | 10.03** \pm 1.78 | 8.77 \pm 2.25 |
| DSP/g parenchyma ($\times 10^6$) | 25.45 \pm 4.78 | 30.84* \pm 4.25 | 28.45 \pm 4.78 |

** Significant at 1% level. & * Significant at 5% level

ous tubules per stage I, daily sperm production (DSP) per testis, DSP per stallion and DSP per gram parenchyma increased from 3.75×10^9 , 7.50×10^9 and 24.74×10^6 (group I) to 4.56×10^9 , 9.11×10^9 and 27.14×10^6 , respectively

(group II). Nevertheless, in age group III, inspite of the slight drop in testis weight, DSP per testis, DSP per stallion and DSP per gram continued to increase where maximum values of 5.30×10^9 , $10.60 \times$

and 33.67×10^6 were achieved. In age group IV, the decline in rates corresponds to a similar pattern in the other criteria studied (Table 1). Age exerted a highly ($P < 0.01$) significant influence on DSP per testis, DSP per gram. Moreover, age and each of DSP per testis and DSP per gram were highly correlated (Table 2).

Testicular weight, parenchyma weight, number of spermatids per stage I tubule cross-section and DSP per gram accounted for 64%, 67% and 86% of the variation in the DSP per testis, respectively. Also, the number of spermatids per stage I tubule accounted for 87% of the variation in DSP per gram parenchyma. The coefficient correlations were high enough to allow accurate prediction of the latter from the former.

As regard seasonal influence, testis and parenchyma weights showed a substantial differences between the breeding and non-breeding season (182.24 vs 158.85 ; $P < 0.01$) and (162.29 vs 141.05 ; $P < 0.01$), respectively.

Although a slight increase in tunica albuginea weight was noted in the breeding season, differences in either weight or percent contribution were not significantly influenced by season (Table 3).

The percentage of seminiferous

tubules in the testis of stallions castrated in the breeding season was slightly higher than (64.25 vs 60%) those in the non-breeding season. Seasonal variation in the percentage of seminiferous tubules was not significant. On the other hand, season exerted a highly significant effect on the diameter of seminiferous tubules (201.31 vs $218.10 \mu\text{m}$; $P < 0.01$) and the corrected number of spermatids (54.98 vs 64.51 ; $P < 0.01$) per stage I, where about 8% increase for the former and 15% for the latter criterion were recorded in the breeding season (Table 3).

The production of sperm (per testis/day or stallion/day) was significantly lowest (3.66×10^9 vs 5.01×10^9 and 7.33×10^9 vs 10.03×10^9 ; $P < 0.01$) in the non-breeding season, averaging only 73% of the DSP in the breeding season. Whereas, the seasonal influence on the DSP per gram parenchyma was at a lower level ($P < 0.05$) of significance.

DISCUSSION

Estimates of testicular density, tunica albuginea weight and percent, and amount of shrinkage due to histological processing are crucial, when histological methods are practiced for determination of daily spermatozoal production. In the present study, the mean density of

testicular tissues reported for Arab and native horses was similar to the 1.042 (Hemeida et al., 1980) for the same breeds, 1.041 (Gebauer, et al., 1974) for the same species and 1.038 to 1.041 reported for the testes of the bull, boar and rabbit (Swierstra, 1966, 1986; Amann, 1970). The mean weight and percent contribution of tunica albuginea (19.01 g and 11.20%) were very close to 19.18 g and 12.01% (Hemeida et al., 1980) and 20 g and 12.3% (Swierstra et al. 1974). The increase in tunica albuginea weight and percent value with advancement of age could be related to the progressive testicular fibrosis, which is a histological feature of aging (Humphrey and Ladds, 1975; Hemeida et al., 1980). In agreement with Gebauer et al. (1974), the amount of shrinkage due to histological processing was more consistent among the stallion testes than that (41 to 58%) reported for bull (Swierstra, 1966) and 45 to 55% for boar testes (Swierstra, 1968).

Testicular weight of Arab and native horses (171.38g) was close to 170 g (Amann et al., 1979), 167g (Hemeida et al., 1980), 163g (Swierstra et al., 1974); higher than 125 to 149 g (Berndtson et al. 1983) and lower than 213 and 216 g (Gebauer et al., 1974). Large inherent variability in testicular size (Berndtson et al., 1983), breed, rearing system and varied ages

could account for such discrepancy. The results of age-related changes in testicular weight are in partial agreement with that reported by Johnson and Neaves (1981), Berndtson et al. (1983), and Johnson and Thompson (1983) who reported that testicular weight increased with age.

The present findings confirm the results of Swierstra et al. (1974) that the seminiferous tubules made up to 61.3 % of the stallion testis ; 58 to 61 .6 % reported by Berndtson et al., (1983) with no significant differences between the age groups studied. In corroboration with Johnson and Neaves (1981), the diameter of the seminiferous tubules was significantly higher in mature than young horses. Slightly higher values (212 to 242 μm) (Johnson and Neaves, 1981) and lower values (156 μm) (Swierstra et al., 1974) and 115 to 158 μm (Berndtson et al., 1983) were reported.

The corrected number of spermatozoa per stage I tubule displayed a similar pattern with the advancement of age as did the DSP/testis, DSP/stallion and DSP/g. The overall mean DSP/testis and DSP/stallion determined in the current study, coincided with the 4.0×10^9 testis (range 2.5 to 5.4×10^9) and mean of 8.0×10^9 /stallion estimated by quantitative histological analysis (Gebauer et al., 1974), 3.9×10^9

6×10^9 / testis estimated by homogenate method (El-Wishy et al., 1980) and higher than 1.27 to 1.18×10^9 / testis reported by Johnson and Neaves (1981). The mean value for DSP/g parenchyma of Arab and native stallions was close to the 26.72×10^6 (Hemeida et al., 1980) using a divisor time of 5.1 days and higher than those (21.0 to 11.5×10^6) reported by Gebauer et al. (1974) for Quarter and Thoroughbred horses which have larger testes (213 and 216 g). The lower figure given by Amann et al. (1979) was due to the use of a longer divisor time of 6.0 days. Moreover, difference due to breed, selection, rearing system, age and nutritional regimen may be contributing factors.

In the present study, the increase in DSP efficiency with subsequent leveling-off up to 13 years old and the decrease in spermatogenic activity of the testes in older horses is consistent with age-related studies in stallions. The peak values for DSP rates reported herein coincided with a similar peak in the gonadal and epididymal sperm reserve (El-Wishy et al., 1980), the extragonadal sperm reserve (Amann et al., 1979) and the number of ejaculated spermatozoa (Pickett et al., 1979, El-Baghdady et al., 1990). With the approach of senescence, the testis of the stallion diminish their capacity to produce spermatozoa as indicated by the drastic drop in DSP rates reported

for older horses. It is possible that age-related changes in Leydig cell population (Johnson and Neaves, 1981) may be involved. Also, a progressive reduction in Sertoli cell functions and/or regression in their numbers (Jones and Berndtson, 1986) occurred in the stallion testes with senescence. The deleterious effect of the aged basement membrane and tunica propria of the seminiferous tubules (which separate the germinal epithelium from blood supply due to fibrosis and thickening) on the spermatogenic processes (Paniagua et al., 1987) might be implicated. Finally, testosterone is required for the completion of meiosis during spermatogenesis (Steinberger, 1971), the apparent reduction in its level (both peripherally and intratesticularly) at older ages (Gusmao et al., 1988; Berndtson and Jones, 1989) may increase the rate of cell loss during the postprophase division which ultimately result in reduced rate of daily sperm production (Johnson and Thompson, 1983).

Although Arab and native horses are capable of breeding all the year round, the testes in the breeding season (December-May) are more adapted for high levels of sperm production. However, testes from horses in the non-breeding season (June-November) did continue to produce spermatozoa albeit at a lower rate. In the current study, some significant seasonal

changes were detected and these may have implications for the breeding potential of stallions.

Testicular weight, parenchymal weight, diameter of seminiferous tubules, corrected number of spermatids per stage I tubule, DSP/testis and DSP/stallion increased by 13%, 14%, 8%, 15%, 27% and 27%, respectively in the breeding season. The current results confirms the observations of Berndtson et al. (1983) and Johnson and Thompson (1983) on the seasonality of equine spermatogenesis and the seasonal nature of spermatozoa production by a quantitative histological technique. Furthermore, seasonal changes in testicular weight and DSP rates reported in this study, might be driven seasonal fluctuations in concentrations of LH (Thompson et al., 1977) and testosterone (Berndtson et al., 1974) in the blood or intratesticular (Berndtson et al., 1983; Berndtson and Jones, 1989). These seasonal changes possibly could be induced by photoperiod, which drives seasonal changes in hormone (FSH, LH and testosterone) concentrations (Clay et al., 1988). Moreover, the increase in the testicular weight and DSP rates reported in the breeding season could be explained by an elevated population of A spermatogonium (Johnson, 1985; Johnson and Tatum, 1989), the number of Sertoli cell/testis (Johnson and Nguyen, 1986) and Leydig cells (Johnson

and Thompson, 1986). An elevated Sertoli cell population in the breeding season would accommodate a larger number of germinal cells than would normally be supported during the non-breeding season, when sperm output and sperm production continued at lower rate (Thompson et al., 1977; Johnson and Thompson 1983).

Based upon the present results the onset of breeding season in Arab and native horses was associated with a significant increase in testicular weight, parenchymal weight, diameter of seminiferous tubules, corrected number of spermatids per stage I tubule cross section and DSP rates. Therefore stallions are capable of producing significantly more spermatozoa (2.70×10^9 additional sperm/stallion/day) during the breeding season (December-May) and could supply sufficient numbers of sperm for insemination of more mares at this time. Presumably, it is reasonable that stallions of 6 to 13 year old are adapted for greater sperm production potential as well that could either younger or older horses.

ACKNOWLEDGEMENT

The author wish to express his appreciation to Dr. Y.M. Shaheen Professor of Histology, and Dr. N.A. Hemeida, Professor of Theri

ogenology, Faculty of Veterinary Medicine, Cairo University, for their help with the histological analysis.

SUMMARY

Fifty-six testes were collected from 28 Arab and native stallions (aged 3 to 18 years) along a complete annual cycle to determine the influence of age and season on the daily sperm production (DSP) by a quantitative histological technique. Overall mean values for testicular measurements, percentage of seminiferous tubules (SNT%), corrected number of spermatids and diameter of SNT/stage I cross-section were given in the text. Estimates of 4.39×10^9 sperm/testis/day, 8.77×10^9 sperm/stallion/day and 28.45×10^6 sperm/g parenchyma were noted. highly significant ($P < 0.01$) increase with age in these traits was evident up to 13 years old stallions, where peak values of 5.30×10^9 , 10.60×10^9 and 33.67×10^6 for the foregoing parameters were achieved. On the other hand, testicular weight, parenchymal weight, number of spermatids per stage I tubule cross-section and DSP/g accounted for 64%, 63%, 67% and 86% of the variation in the DSP/testis, respectively. Age-related changes in the above mentioned parameters were also scrutinized.

The onset of breeding season (December-May) in Arab and native horses was associated with a significant increase in testicular weight (13%), parenchymal weight (14%), diameter of SNT/stage I cross-section (8%), corrected number of spermatids/stage I tubule cross-section (15.%), DSP/testis (27%) and DSP/stallion (27%). The production of sperm (per testis/day or stallion/day) was lowest (3.66×10^9 vs 5.01×10^9 and 7.33×10^9 vs 10.03×10^9 ; $P < 0.01$) in the non-

breeding season, averaging only 73% of the DSP in the breeding season. Therefore, stallions are capable of producing significantly more spermatozoa (2.70×10^9 additional sperm/stallion/day) in the breeding season and could supply sufficient numbers of sperm for insemination of more mares at this time.

REFERENCES

- Amann, R.P. (1970): Sperm production rates. In the testis. Vol. 1, pp. 433-382. Eds A.D. Johnson, W.R. Gomes and N.L. VanDemark. Academic Press, New York and London.
- Amann, R.P. and Almquist, J.O. (1962): Reproductive capacity of dairy bulls. VIII-Direct and indirect measurements of testicular sperm production. *J. Dairy Sci.* 45: 774-781.
- Amann, R.P., Tompson, D.L., Jr, Squires, E.L. and Pickett, B.W. (1979): Effect of age and frequency of ejaculation on sperm production and extragonadal sperm reserves in stallions. *J. Reprod. Fert., Suppl.* 27: 1-6.
- Berndtson, W.E. (1977): Methods for quantifying mammalian spermatogenesis. A review. *J. Anim. Sci.* 44: 818-833.
- Berndtson, W.E. and Jones, L.S. (1989): Relationship of intratesticular testosterone content of stallions to age, spermatogenesis, Sertoli cell distribution and germ cell-Sertoli cell ratio. *J. Reprod. Fert.* 85: 511-519.
- Berndtson, W.E., Pickett, B.W. and Nett, T.M. (1974): Reproductive physiology of the stallion. IV. Seasonal changes in the testosterone concentration of peripheral plasma. *J. Reprod. Fert.* 39: 115-118.
- Berndtson, W.E., Squires, E.L. and Tompson, D.L., Jr. (1983): Spermatogenesis, testicular composition and the concentration of testosterone in the equine testis as influenced by season. *Theriogenol.* 20: 449-457.

- Chalkley, H.W. (1943):** Method for the quantitative morphologic analysis of tissues. *J. Nat. Cancer inst.* 4: 47-53.
- Clay, C.M., Squires, E.L., Amann, R.P. and Nett, T.M. (1988):** Influences of season and artificial photoperiod on stallions: Luteinizing hormone, follicle stimulating hormone and testosterone. *J. Anim. Sci.* 66: 1246-1255.
- El-Baghdady, Y.R.M., Hemeida, N.A., Abou-Ahmed, M.M., El-Belely, M.S. and Ismail, S.T. (1990):** Agr-related changes in seminal and behavioural characteristics of Arab horses. 4th Sci. Cong. Fac. Vet. Med., Assiut Univ., Egypt. Vol. II: 510-519.
- El-Wishy, A.B., Abou-Ahmed, M.M., Hemeida, N.A. and El-Sayed, M.A.I. (1980):** Sperm producing capacity of Arab and native horses in Egypt. *J. Reprod. Fert., Suppl.* 32: 27-30.
- Gebauer, M.E., Pickett, B.W. and Swierstra, E.E. (1974):** Reproductive physiology of the stallion. II. Daily production and output of sperm. *J. Anim. Sci.* 39: 732-736.
- Gusmao, A.L., Klug, El., Merkt, H., Hoogen, H. and Hoppen, H.O. (1988):** Hormonal profiles and hormone challenge test throughout the reproductive life of Hannoverian stallions. *Proc. 11th. Int. Cong. Anim. Reprod. & A.I. Dublin Univ.* Vol. 2, PP. 30, Dublin Ireland.
- Hemeida, N.A., Abou-Ahmed, M.M. and El-Wishy, A.B. (1980):** Reproductive capacity of horses in Egypt. II. Effect of breed and age on testes characteristics and sperm production. *Egypt. J. Vet.Sci.* 17: 149-169.
- Humphery, J.D. and Laddds, P.W. (1975):** A quantitative histological study of changes in the bovine testis and epididymis associated with age. *Res. Vet. Sci.* 19: 135-140.
- Johnson, L. (1985):** Increased daily sperm production in the breeding season of stallion is explained by an elevated population of spermatogonia. *Biol. Reprod.* 32: 1181-1190.
- Johnson, L. and Neaves, W.B. (1981):** Age-related changes in the Leydig cell population, seminiferous tubules and sperm production in stallions. *Biol. Reprod.* 24: 703-712.
- Johnson, L. and Nguyen, H.B. (1986):** A cycle of the Sertoli cell population in adult stallions. *J. Reprod. Fert.* 76: 316.
- Johnson, L. and Tatum, M.E. (1989):** Temporal appearance of seasonal changes in numbers of Sertoli cells, Leydig cells and germ cells in stallions. *Biol. Reprod.* 994-999.
- Johnson, L. and Thompson, D.L., Jr. (1985):** Age-related and seasonal variation in Sertoli cell population, daily sperm production and serum concentrations of follicle-stimulating hormone, luteinizing hormone and testosterone in stallions. *Biol. Reprod.* 29: 777-789.
- Johnson, L. and Thompson, D.L., Jr. (1986):** Seasonal variation in the total volume of Leydig cells in stallions is explained by variation in cell number rather than size. *Biol. Reprod.* 35: 971-979.
- Jones, L.S. and Berndtson, W.E. (1986):** A quantitative study of Sertoli cell and germ cell populations as related to sexual development and aging in the stallion. *Biol. Reprod.* 35: 138-148.
- Paniagua, R., Nistal, M., Amat, P. and Linares, A. (1987):** Seminiferous tubules and germ cell population in elderly men. *Biol. Reprod.* 36: 939-946.
- Pickett, B.W., Voss, J.L. and Squires, E.E. (1979):** Factors affecting sperm output of the stallion. *Aust. Adv. Vet. Sci.* pp. 23-24.
- Snedecor, G.W. and Cochran, W.G. (1980):** Statistical methods, 6th ed. Iowa State University Press, Ames.
- Steinberger, E. (1971):** Hormonal control of mammalian spermatogenesis. *Physiol. Rev.* 51: 1-22.
- Swierstra, E.E. (1966):** Structural comparison of shorthorn bull testes and daily spermatozoal production as determined by quantitative testicular histology. *Can. J. Anim. Sci.* 46:107-119.
- Swierstra, E.E. (1968):** A comparison of spermatozoa production and spermatozoal output of Yorkshire and Lacombe boars. *J. Reprod. Fert.* 17: 459-469.

ierstra, E.E., Gebauer, M.R. and Pickett, B.W. (1974): Reproductive physiology of the stallion. I. Spermatogenesis and testis composition. *J. Reprod. Fert.* 40: 113-123.

ompson, D.L., Jr., Pickett, Berndtson, W.B., W.E., Voss, J.L. and Nett, T.M. (1977): Reproductive physiology of the stallion. VIII. Artificial photoperiod, collection interval and seminal characteristics, sexual behaviour and concentration of LH and testosterone in serum. *J. Anim. Sci.* 44: 656-664.