



# protective effects of date extract against toxic effects of lead acetate in rats.

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#### Abstract

Lead (Pb) is a common industrial and environmental pollutant. Prolonged exposure of a sublethal dose to this toxicant is associated with oxidative stressand considered to be a risk factor for kidney, liver in addition to many disorders. This study was carried out to investigate the most toxic effects of lead with trial to diminish this toxicity by supplementation of date extract. Both the enzymatic (superoxide dismutase (SOD), glutathione reductase (GR), glutathione peroxidase (GPx), glutathione-S-transferase (GSH-T) and catalase (CAT)} and non-enzymatic (gutathione, GSH) antioxidants had been studied, among mature male albino rats which have been received lead acetate in drinking water for 3 months, it is widely accepted that even small quantities of Pb are harmful to rats which implicated in a broad range of physiological conditions.the study proved that the use of date extract, through its antioxidant protective effect and immune potentiating properties, can reduce Pb-induced hepatic damage and toxicity

#### Introduction

Lead is ubiquitous, and the most common environmental pollutant naturally present in the earth's crust in small concentrations, For centuries, it has been mined and disseminated throughout the environment from where it has gradually become incorporated into the structural tissue of plants, animals and humans (Pracheta et al., 2009). There are two sources of exposure, i.e. exposure to lead based, paint and exposure to lead related industries, which are either inevitable or reducible at a rate so slow that is still a threat to public health (Martin et al., 2001).

Lead pollution can also cause irreversible encephalopathy, seizure, coma and even death. Fatigue, memory loss, nephropathy, pressure, high blood gastrointestinal disturbances, weight loss are immuno-suppression common toxic effects of lead exposure in animals. Prenatal exposure to metal may also cause birth defects, miscarriage and underdeveloped babies (Ehle and Mckee, 1990 and Pracheta et al., 2009).

The date fruit is listed in folk remedies for the treatment of various infectious diseases and cancer (Duke, 1992). The aqueous extracts of dates have potent antioxidant and antimutagenic activity (Allaith, 2007; Biglari et al., 2008 and Saafi et al., 2009). This study was carried out to investigate the most toxic effects of lead with trial to diminish this toxicity by supplementation of date extract

### Materials and Methods

This study was carried out at the Unit for Laboratory Animals at Hygiene and Management Department, Faculty of Veterinary Medicine, Cairo University.

### 3.1 Animals:

conducted on study was The Immature Wistar male rats (N=120) weighing approximately 120-140 g were obtained from the Unit for Laboratory Animals at Faculty of Veterinary

The University. Cairo Medicine, temperature in the laboratory animal house unit ranged from 25-30°C. The relative humidity ranged from 50-70%. Natural system of lighting was used throughout the study .Cages were cleaned twice weekly in summer and once per week in winter, Animals care as well as experimental protocols were in compliance with guidelines of ethical standards released by Cairo University policy on animal care and use. All efforts were made to minimize the numbers of animals and their suffering in this study through following the Guidelines on Laboratory Animal Care and Use.

### 3.2 Preparation of date palm extracts:

Fresh ripened Wahat date variety collected from the Wahat was extracted two times with distilled water (1/10, w/v) by grinding with a mortar and pestle. It was centrifuged at 41C for 20 min at 4000 g and the supernatant was collected.

#### 3.3 Experimental design

Immature male rats (120) were divided at random into 4 groups of 30 animals, each as follows:

Group 1: Control group: animals received a daily distilled water as drinking water for 12 weeks.

Group 2: High lead acetate group: animals received a daily lead acetate at a concentration of 0.5 mg / liter dissolved in distilled water and administered to the rats in drinking water for 12 weeks.

Group 3: Low lead acetate group: animals received a daily lead acetate at a concentration of 0.2 mg / liter dissolved in distilled water and administered to the rats in drinking water for 12 weeks.

Group 4: low lead acetate with date extract: animals received a daily oral of lead acetate at a concentration of 0.2 mg / liter dissolved in distilled water with date extract (4 ml / kg) for 12 weeks.

# 3.4 Determination of oxidative stress and lipid peroxidation:

Tissues (liver, spleen, brain and kidney) and blood were homogenized in ice-cold 1.15% KCl to make 10% (W/V) homogenate with Glas-Col motor driven homogenizer (USA) and the homogenate

was used for determination of malondialdehyde (MDA), reduced glutathione (GSH) levels as well as superoxide dismutase (SOD), catalase (CAT), Glutathione transferase (GST), and Glutathione Reductase (GSR) activities as recommended by (Buege and Aust, (1978), Burtes and Ashwood (1999) and Kakkar et al. (1984); respectively.

## 3.5 Determination of lipid peroxidation (MDA) concentration:

MDA concentration was assayed using the commercial MDA Assay Kit was purchased from (Bio Diagnostic Co., Egypt). The MDA adduct can be easily quantified colorimetrically at  $\lambda = 532 \text{ nm}$ .

### 3.6 Determination of reduced glutathione (GSH) (Burtis and Ashwood,1999):

GSH concentration was assayed using the commercial GSH Assay Kit was purchased from (Bio Diagnostic Co., Egypt).

### 3.7 Determination of superoxide dismutase (SOD) Activity:

SOD concentration was assayed using the commercial SOD Assay Kit was purchased from (Bio Diagnostic Co., Egypt).

### 3.8 Determination of catalase (CAT) Activity:

CAT activity was assayed using the commercial CAT Assay Kit was purchased from (Bio Diagnostic Co., Egypt).

# 3.9 Determination of glutathione-S-transferase activity:

GST activity was assayed using the GST commercial Colorimetric Activity Assay Kit was purchased from (Bio Diagnostic Co., Egypt) and detected by spectrophotometry at 340 nm. One unit of GST activity is defined as the amount of enzyme producing 1 /mol of GS-DNB conjugate/min under the conditions of the assay (Tounget al., 1990).

# 3.10 Determination of Glutathione Reductase Activity:

GR concentration was assayed using the commercial GR Assay Kit was

purchased from (Bio Diagnostic Co., Egypt).

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### Results and Discussion:

a metal with many Lead is recognized adverse health side effects (Sui et al., 2015). It is a ubiquitous environmental and industrial pollutant that induces a broad range of toxic manifestations within biological systems. Exposure to lead induces overproduction of reactive oxygen species and depletes the cellular antioxidant capacity. An imbalance of pro-oxidant/antioxidant ratio in tissue and cellular components is known to cause damage to membranes, DNA, or proteins, and finally destroy the tissues or systems (Hsu and Guo, 2002).

Lead causes oxidative stress by inducing the generation of reactive oxygen species (ROS), reducing the antioxidant defense system of cells via depleting interfering with glutathione, essential metal, inhibiting sulfhydryl antioxidant dependent enzymes or increasing activities or enzymes susceptibility of cells to oxidative attack by altering membrane integrity and fatty acid composition (Sharma et al., 2011).

In the present study, both the enzymatic SOD, GR, GPx, GSHT and CAT and non-enzymatic (glutathione GSH) antioxidants had been studied.

Quantification of lipid peroxidation is essential to assess oxidative stress in Lipid pathophysiological processes. Malondialdehyde peroxidation forms (MDA) and 4-hydroxynonenal (4-HNE), as natural bi-products. Measuring the end products of lipid peroxidation is one of the most widely accepted assays for oxidative damage. Pb is known to alter the activity of lipid metabolizing enzymes in liver (Kojima et al., 2005), which can limit the biosynthesis of bile acids. Bile acid plays elimination important role in cholesterol from the body (Mudipalli, 2007; Newairy and Abdou, 2009) in addition to increasing lipid peroxidation (Kamalakkanan and Prince, 2004).

Pb is known to produce oxidative damage by elevating peroxidation of membrane lipids. Generation of peroxyl

radicals after Pb-intoxication stimulates lipid peroxidation via a cyclisation reaction to endoperoxides (Marnett, 1999 and Sharma et al., 2010).

Lead may disturb the antioxidant barrier via inhibition of the functional SH groups present also in free radical-scavenging enzymes such asGR, GPx,GST,SOD, CAT and d-aminolevulinic acid dehydratase (D-ALAD) (Moniuszko-Jakoniuk et al., 2007; Olaleye et al., 2007). These enzymes are believed to be the major antioxidant agents in the mammalian body that protect against ROS toxicity (Ashry et al., 2010).

The observed increase in the MDA was accompanied by a 70% decrease in GSH levels in the PbAc-group, which may be due to its utilization either in lead detoxification or in scavenging lead-generated free radicals (Othman and El-Missiry, 1998 and Olaleye et al., 2007). Under oxidative stress, the GSH-related enzymes merely catalyze reactions to detoxify peroxides in the water phase by reacting them with GSH (Moniuszko-Jakoniuk et al., 2007).

Thus, the enhanced concentration of MDA and severe depletion in GSH activity suggests that the increased peroxidation is a consequence of depleted GSH stores and diminished GSH-related enzymes, which are otherwise capable of degree of the moderating peroxidation (Ashry et al., 2010). It is lipid increased that hypothesized peroxidation as a result of reduced endogenous antioxidant capacity may be the initial event in the mechanism of Pbinduced hepatic damage (Ashry et al., 2010).

In this study, Pb intoxication caused significant enhancement of thio barbituric acid reactive substances (TBARSs) levels in blood and organs (liver, spleen, brain and kidney). Date extract significantly lowered TBARSs levels in blood and other organs in a dose dependant manner. The protective effect may be due to the radical scavenging activity of phenolics present in the extract. Ubiquinones (co-enzymes Q)

function as important cellular electron carriers (Ernster and Dallner, 1995)

The result of SOD and CAT activity suggest that date extract contains a free radical scavenging activity, which could against action beneficial a exert pathological alteration caused by the presence of free radicals. This action could involve mechanisms related to scavenging activity.

We selected an aqueous extract antioxidant the of most because components in dates are extracted in water Al-Farsi et al., (Vayalil, 2002 and 2005b). During the experience, the aqueous date fruit extract (Wahat date) was daily prepared and administrated to rats.

The aqueous extracts of dates have potent antioxidant and anti-mutagenic activity, the antioxidant activity is activity, attributed to its content of the wide range of phenolic compounds including p coumaric, ferulic, and sinapicacids, flavonoids and procyanidins (Farsi et al., 2005; Mansouri et al., 2005 and Hong et al., 2006) and also to the presence of vitamin C (Allaith, 2007 and Mrabet et al., 2008). Flavonoids can act in the initiation stage of peroxidation interfering with the metabolism oxidative agent either by scavenging the free radicals or by impairing the microsomal enzymatic system needed for this metabolism (Singh and Handa, 1995). It is clear that date extract showed an antioxidant protective effect

Table (1): Mean  $\pm$  SD of oxidative stress and lipid peroxidation parameters among rats which received lead acetate.

	Control group	High lead acetate treated group	Low lead acetate treated group	Date extract with low lead acetate treated group
G.S.H	9.80 ± 1.92 a	4.00 ± 2.00 c	6.80 ± 2.39 b	$8.40 \pm 1.52$ ab
G.R	12.40 ± 2.07 c	45.80 ± 8.04 a	33.60 ± 4.72 b	18.00 ± 5.43 c
G.Px	16.00 ± 4.06 c	46.40 ± 8.08 a	37.20 ± 5.17 b	23.00 ± 5.24 c
MDA	14.00 ± 4.06 c	17.60 ± 3.91 bc	21.40 ± 4.39 b	29.80 ± 4.32 a
CAT	56.00 ± 2.65 c	65.40 ± 5.03 b	$78.00 \pm 8.37$ a	48.80 ± 5.63 c
SOD	610.40 ± 17.84 c	698.80 ± 16.75 b	839.60 ± 32.54 a	579.60 ± 6.43 d

Means with different letters (a, b, c, d) within the same row are significantly different at P value ≤ 0.05 G.S.H: Glutathione G.S.H.R: Glutathione Reductase G.S.H.Px: Glutathione peroxidase Malondialdehyde concentration CAT: Catalase Activity SOD: Superoxide Dismutase Activity.

### References

AbuhFY, Wambebe, C., Rai, P.P., and Sokamba, E.N. (1990): Hypoglycaemic activity of Anthocleistavoglii(Planch) aqueous extract in rodents. Phytother Res 4: 20-24.

Abu Abeeleh, M., Ismail, Z., Alzaben, K., Abu-Halawah, Al-Essa, S., Abuabeeleh, J. and Alsmady, M. (2009): Induction of diabetes mellitus in rats using intraperitoneal streptozotocin: A comparison between 2 strain Rats. European Journal of scientific Research. 32:398-402.

Aebi, H., (1974): Catalase. In: Bergmeyer H.U, editor. Methods of Enzymatic Analysis. Weinheim: Verlag Chemie; pp. 673-8.

Afanas'ev, A., Dorozhko, A.I., Brodskii, A.V., Kostyk, V.A. and Potapovitch, A.I. (1989): Chelating and free radical

scavenging mechanism action of rutin and quercetin in lipid peroxidation

Agca, Y. and Critser, J. K. (2006): Assisted reproductive technologies and genetic modifications in rat. In: The Laboratory Rat. (2<sup>nd</sup> edition), by Suckow, M. A., Weisbroth, S. H., and Franklin, C. L. (eds). Elsevier Inc. p: 165-189,

Akbarzadeh, A. N. D., Mehrabi, M.R., Jamshidi, S., Farhangi, A., Allah, A., Mofidian, S., and B. L.R., (2007): Induction of diabetes by streptozotocin in

Alasalvar, C., Morris, A., Baron, M., and Shahidi, F. (2005a): Barous, (2005a):
Compositional and sensory characteristics of three native sun-dried date (Phoenix dactyliferal...) varieties grown in Oman. J. of Agricultural and Food Chemistry, 53, 7586-

All, B.H., (1997): The effect on plasma glucose, insulin and glucagon levels of treatment of diabetic rats with the medicinal plant Rhazyastricta and with glibenclamide, alone and in combination. J. Pharm,

Pharmacol. 49; 1003-1007

Allaith, A.A.A.(2007): Antioxidant activity of Bahraini date palm (Phoenix dactyliferal..) fruit of various cultivars. Int J Food Sci

Al-Jubori, A. (2009): A comparative study of the effect of insukin and alcoholic extract of Apium graveolens seed on the treatment of experimentally-induced diabetes mellitus in mature male rabbit. M Sc. thesis, College of Vet. Med., Al-Qadisiya Univ., Iraq.

Andrade Cetto, A., Wiedenfeld, H., Revilla, M.C., and Sergio, L. A., (2000): Hypoglycaemic effect of Equisetum myriochaetum aerial parts on streptozotocin diabetic rats. J. Ethnopharmacol. 72: 129-

Anuradha, C.V. and Selvam, R. (1993): Effect of oral methione on tissue lipid peroxidation and antioxidants in alloxan induced diabetes rats. J. Nutr. Biochem. 4: 212-217.

Arison, R. N., Ciaccio, E. I., Glitzer, M. S., Cassaro, J.A., and Pruss, M.P. (1967): Light and electron microscopy of lesions in rats rendered diabetic with streptozotocin. Diabetes 16:51-56. Biochem. Pharmacol. 38: 1763-1769.

Asensi, M.; Medina, I.; Ortega, A.; Carretero, J.; Bano, M.C.; Obrador, E., and Estrela J.M. (2002): Inhibition of cancer growth by resveratrol is related to its low bioavailability. Free Radic Biol Med.

Atkinson, M.A., and Maclaren, N. K. (1994): The pathogenesis of insulin-dependent diabetes mellitus N Engl J Med331 1428-36.

Augusti, K.T., Joseph, P., and Babu, T.D. principles (1995): Biologically active isolated from Salaciaoblongawall. Indian J Physiol Pharmacol 39: 415-417.

Babu, P.S., and Srinivasan K. (1997): Influence of dietary capsaicin and onion on the metabolic abnormalities associated with streptozotocin induced diabetes mellitus. Mol Cell Biochem 175: 49-57.

Bajpai, M.B., Asthana, R.K. and Sharma, N.K. (1991): Hypoglycaemic effect of Swerchirin from the hexane fraction Swertia chirayita. Planta. Med. 57: 102-104

Barnett, A. H., Eff, C., Leslie, R. D. and Pyke, D. A. (1981): Diabetes in identical twins. A study of 200 pairs. Diabetologia 20 87-93.

Basnet, P., Kadota, S. and Terashima, S. (1993):Two new 2arylbenzofuran derivatives from hypoglycaemic activitybearing fractions of Morusinsignis. Chem Pharm Bull (Tokyo) 41: 1238-1243.

Basnet, P., Kadota, S., Shimizu, M. and Namba, T. (1994): Bellidifolin: a potent hypoglycaemic agent in streptozotocin (STZ)- induced diabetic rats from Swertia japonica. Planta Med 60: 507-511.

Baynes, J.W. (1991): Role of oxidative stress in development of complications in diabetes. Diabetes; 40: 405-12.

Bedoya, F. J.; Solano F. and Lucas M. N-monomethyl-arginine (1996): nicotinamide prevent streptozotocin-induced double strand DNA break formation in pancreatic rat islets. Experientia 52 344-7.

Bendayan, M. and Ito, S. (1979): The J. of Histochemistry and Cytochemistry. 27, 1029-1034.

Bennett P. (1990): Epidemiology of diabetes mellitus, in Ellenberg and Rifkin's Diabete Mellitus Elsevier: New York 363.

Biglari, F., AlKarkhi A.F.M., Easa, A.M.(2008): Antioxidant activity and phenolic content of various date palm (Phoenix dactylifera) fruits from Iran. Food Chem; 107:1636-41.

Bonoli, M., Verardo, V., Marconi, E. and Caboni, M.F. (2000): Antioxidant phenols in barley (HordeumVulgareL.) spectrophotometric Comparative among extraction methods.

Brentjens, R. and Saltz, L. (2001): Islet cell tumors of the pancreas: the medical oncologist's perspective. Surg Clin North Am 81 (3): 527-42 (1978):

Buege, J.A. and Aust, S.D. Microsomal lipid peroxidation. Enzemol. 51:302-310.

Burtis, C. and Ashwood, E.R. (1999):Tetize Fundamental of clinical biochemistry. 4th Edition. W.B. Saunders Company chap 22.

Bustin S A, and Nolan, T. (2004): Analysis of mRNA expression by real-time PCR.In Real-Time PCR: An Essential Guide (K. Edwards, J. Logan, and N. Saunders, eds.) pp. 125-184. Horizon Bioscience, Norfolk,

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- Handbook phytochemicals of GRAS herbs and other Duke, economic plants. CRC Press: Boca Raton,
- Ernster, L., Dallner, G. (1995): Biochemical, physiological and medical aspects of ubiquinone function. Biochim. Biophy. Acta
- Haleagrahara, N., Jackie, T., Chakravarthi, S. and Rao, M. (2010): Protective effects of Etlingeraelatior extract on lead acetateinduced changesin oxidative biomarkers in bone marrow of rats.
- Hong, Y.J., Tomas-Barberan, F.A., Kader, A.A. and Mitchel, A.E. (2006): The flavonoid glycosides and procyanidin composition of Deglet Noor dates (Phoenix dactyllifera). J. Agric. Food Chem. 54 (6), 2405-2411.
- Hsu, P.C. and Guo, Y.L.(2002): Antioxidant nutrients and lead toxicity. Toxicology 180,33-44
- Kakkar, P., Das, B. and Visvanathan, P.N. (1984): A modified spectrophotometric assay of superoxide dismutase. Ind. J. Biochem. Biophy. 2, 130-132
- Kamalakkanan, N., Prince, P.S.M.( 2004): Antidiabetic and anti-oxidant activity ofAegle marmelosextract in streptozotocininduced diabetic rats. Pharm. Biol. 42,125-130.
- Kojima, M., Sekikawa, K., Nemoto, K., Degawa, M. (2005): Tumor necrosis factoraindependent down regulation of hepatic cholesterol 7a-hydroxylase gene in mice treated with lead nitrate. Toxicol. Sci. 87, 537-542
- Marnett, L.J. (1999): Lipid peroxidation-DNA by malondialdehyde. Mut. damage Res.:Fund. Mol. Mech. Mut. 424, 83-95.
- Mudipalli, A. (2007): Lead hepatotoxicity and potential health effects. Ind. J. Med.Res. 126, 518–527.
- Newairy, A.S.A., Abdou, H.M.(2009): Protective role of flax lignans against lead

- acetate induced oxidative damage and acetate hyperlipidemia in rats. Food Chem, Toxicol, 47, 813-818
- Olaleye, S.B., Adaramoy, O.A. and Erigbali, O.S. (2007): Lead exposure increases oxidative stress in the gastric mucosa of World J. HCl/ethanol-exposed Gastroenterol. 13, 5121-5126
- Othman, A.I. and El-Missiry, M.A. (1998): Role of selenium against lead toxicity in male rats. J. Biochem. Mol. Toxicol. 12, 345-349.
- Pracheta M. and Singh L. (2009): Effect of lead nitrate (Pb(NO3)2 on plant nutrition. physical and chemical as well as parameters on Lobia (Vigna unguiculata Linn. Walp.). J. Plant Develop. Sci., 1(1 and 2): 49-56
- Saafi, E.B., El Arem .A., Issaoui .M., Hammami .M. and Achour. L. (2009): Phenolic content and antioxidant activity of four date palm (Phoenix dactyliferaL.) fruit varieties grown in Tunisia. Int J Food Sci Technol.; 44:2314-9
- Sharma, V., Sharma, A. and Kansal, L. (2010): The effect of oral administration ofAllium sativum extracts on lead nitrate induced toxicity in male mice. Food Chem. Toxicol. 48, 928-936
- Sharma, S., Sharma, V., Paliwal, P. and Pracheta (2011): Lead toxicity, oxidative damage and health implications
- Sui, L., Zhang, R.H., Zhang, P., Yun, K.L., Zhang, H.C., Liu, L. and Hu, M.X. (2015): Lead toxicity induces autophagy to protect against cell death through mTORC1 pathway in cardiofibroblasts. Biosci. Rep. 35(2):art:e00186.doi:10.1042/BSR20140164
- Vayalil, P. K. (2002): Antioxidant and antimutagenic properties of aqueous extract date fruit (Phoenix dactyliferal. Arecaceae). J. of Agricultural and Food Chemistry, 50, 610-617

### الملخص العربي

الرصاص هو من الملوثات الصناعية والبيلية المشتركةويرتبط التعرض لفترات طويلة لجرعات اقل من المميتة ذو تاثير علي الكلي والكبد ويؤدي إلى العديد من الاضطرابات. وقد أجريت هذه الدراسة للتحقيق في الآثارالأكثر سمية للرصاص مع محاولة تقليل هذه السمية عن طريق إضافة مستخلص ثمار البلح.

ويتم في هذه الدراسة تأثير الرصاص على بعض الزيمات الأكسدة في الكبد والكلي بين ذكور الفنران الناضجة حيث تتلقي ذكور الفنران عي حده الدراسة دراسة تاثير الرصاص على بعض الريعات المصلح في بين المسلم من الرصاص ضارة للفلران واتضح أيضا تأثير الناضجة خلات الرصاص في مياه الشرب لمدة 3 شهور. وفي الختام اتضح أنه حتى كميات صغيرة من الرصاص ضارة للفلران واتضح مستخلص البلح المضاد للأكسدة في الظروف الفسيولوجية من خلال تأثير وقاني وبالتالي يؤدي الي رفع الخصائص المناعبة ويؤدي إلى تقليل تأثير والتالي يؤدي المضاد للأكسدة في الظروف الفسيولوجية من خلال تأثير وقاني وبالتالي يؤدي الي رفع الخصائص المناعبة ويؤدي إلى تقليل تأثير المناد المناعبة المناعبة ويؤدي المناعبة ويؤ تأثير الرصاص على الكبد والكلي.