



## Tomato Pomace with Exogenous Enzymes and/or Amino Acids Enhanced Growth Performance, Histological Parameters and Gene Expression for Growth in Nile tilapia Fish (*Oreochromis niloticus*)

Azza A. Hafez<sup>1</sup>, Eldsokey El-Nassef<sup>1</sup>, Abdelnasser A. Bakr<sup>1</sup>, Eman M. Moustafa<sup>2</sup>,  
Walied S. Abdo<sup>3</sup> and Elsayed M. Hegazi<sup>1</sup>

<sup>1</sup>Nutrition and Clinical Nutrition Department, Faculty of Veterinary Medicine, Kafrelsheikh University, Egypt.

<sup>2</sup>Fish Diseases and Management Department, Faculty of Veterinary Medicine, Kafrelsheikh University, Egypt.

<sup>3</sup>Pathology Department, Faculty of Veterinary Medicine, Kafrelsheikh University, Egypt.

### Abstract

**T**HIS study evaluated the impact of feeding tomato pomace with or without enzymes and amino acids to Nile tilapia for 8 weeks on growth performance, digestive enzymes, histological parameters and some liver gene expression. Five experimental diets were formulated with similar protein content (28%): (1) a control diet without tomato pomace (TP), (2) TP 15%, (3) TP 15% with lysine (1.79%) and methionine (0.94%), (4) TP 15% with gallizyme (0.2%) and (5) TP 15% with lysine, methionine and gallizyme. The results revealed significant improvements in growth performance, especially body gain by 31.05% and 33.03% and FCR by 21.83% and 23.35% in the 4<sup>th</sup> and 5<sup>th</sup> groups, respectively, as compared to the 2<sup>nd</sup> group. Enzymes alone or enzymes with amino acids significantly increased Villi length and goblet cells number in the intestine of tilapia. There was no significant difference in the activities of lipase and amylase and levels of total protein, albumin and globulin in all groups. The relative expression levels of IGF-1 and GH in the liver of tilapia were significantly increased ( $p < 0.05$ ) in the last three groups. Finally, addition of enzymes with or without amino acids to the Nile tilapia diets improved the growth performance parameters and gene expression.

**Keywords:** TP, gallizyme, lysine, methionine, GH, IGF-1.

### Introduction

For a very long time, the aquaculture industry has provided humans with delicious and safe animal protein [1]. However, more seafood protein is needed due to the ongoing increase in human consumption [2]. Since feed accounts for more than 60% of production costs, the aqua-feed business is essential to the growth of aquaculture [3]. High-quality protein sources are used by the aqua-feed industry to give aquatic animals the energy and protein they need for physiological and metabolic processes [4, 5]. Traditionally, nutritionally balanced feed formulations have been made using fish meal (FM) and plant components (such as soybean meal,

maize gluten, wheat middling, sunflower meal, etc.) [6]. Thus, in order to lower feed costs, it is essential to find a new supplier of unconventional feed ingredients [7-13]. Therefore, it is necessary for fish farmers worldwide to investigate the use of alternative, readily available, and reasonably priced plant-based feed ingredients. [14].

One of the more promising substitute sources of protein and energy for the monogastric animal feed business is tomato pomace. [15, 16]. Tomato pomace is a material consisting of the skin, pulp, and broken seeds that are left over after tomatoes are processed to make juice, paste, or ketchup. Protein (17–22%), fat (10–12%), crude fiber (33–57%), NDF

\*Corresponding author: Azza. A. Hafez, E-mail: [azza\\_ahmed@vet.kfs.edu.eg](mailto:azza_ahmed@vet.kfs.edu.eg), Tel.: 002 01067581222

(Received 20 April 2024, accepted 17 July 2024)

DOI: 10.21608/EJVS.2024.284053.2023

©2025 National Information and Documentation Center (NIDOC)

(50–72%), which is mostly made up of ADF (39–60%), and lignin concentration (20–30%) are all relatively high in tomato pomace [17]. Lycopene levels in tomato pomace, particularly the skin, are notable about five times higher than in pulp (on a moist basis) [18, 19]. Tomato pomace is a rich source of many bioactive substances, including tocopherols, polyphenols, terpenes, and sterols, in addition to carotenoids [20].

Non-starch polysaccharides (NSP) are challenging to digest and have a range of antinutritive effects because of modifications to the gut's morphology, physiology, digesta viscosity, and microbial composition [21]. Recirculating aquaculture systems (RAS) have a higher system waste load due to lower nutrient use, which also causes eutrophication when waste is dumped into nearby water bodies [22]. By adding NSP-degrading enzymes (such as xylanase,  $\beta$ -glucanase, and cellulase) and phytase to diets, it is possible to partially offset the detrimental effects of NSP and phytate on digestion, which in turn enhances growth of fish [23, 24]. The digestion and absorption of feed nutrients as well as the development performance of aquatic animals are markedly inhibited by dietary concentrations of NSPs above 8% [25, 26].

Thus, the purpose of this study was to determine how tomato pomace supplemented with exogenous enzymes and/or amino acids affected growth performance, digestive enzymes, histological parameters, and the expression of genes related to growth.

## Material and Methods

### Tomato Pomace (TP)

We obtained tomato pomace (TP) from a nearby supplier and let it dry in the sun for 4 hours per day for five days. After drying, it was ground and analyzed for chemical composition before diet preparation as shown in Table (1). Tomato pomace used in this study by inclusion rate of 15% according to previous studies.

### Exogenous enzymes

Exogenous enzymes used in the experiment is Gallizyme, it was purchased from (Tex bioscience company, India). The composition of these enzymes is amylase (1000000 U/kg), cellulase (10000000 U/kg), xylanase (1500000 U/kg), phytase (500000 U/kg), lipase (10000 U/kg), pectinase (30000 U/kg), protease (400000 U/kg), alpha-Galactosidase (10000 U/kg), beta- Glucosidase ((10000 U/kg), arabinase (7000 U/kg), and wheat bran up to 1 kg. Enzyme mixtures were added in this study by 0.2% according to [27]

### Fish and experimental diets

A 225 monosexual Nile tilapia fingerlings was used in this study. Prior to the beginning of the trial,

the fish were housed in glass tanks for two weeks for acclimation. The experimental fish with initial weight ( $21.2 \pm 0.08$ g/fish) were purchased from a private farm. Fish basal diet was developed in accordance with NRC (2011) [28], presented in Table (2). Fish were randomly distributed into 5 groups with 45 fish per aquarium with triplicate of 15 fish per replicate. G1 a control diet without TP, G2 a TP 15% (TP), G3 TP 15% with lysine (1.79%) and methionine (0.94%) supplementation (TPA), G4 TP 15% with exogenous enzymes (0.2% of the diet) (TPE) and G5 TP 15% with lysine (1.79%), methionine (0.94%) and (0.2%) exogenous enzymes (TPAE). After the trial started and for the next eight weeks, fish in each group were weighed every two weeks. Fish were fed 2 times per day (8 am and 4 pm) to apparent satiation. Every two weeks, body weight gain (BWG) was recorded. Then, the total BWG was determined. In a similar manner, FCR was calculated. After that, the total FCR was determined.

The following formulae were used to compute growth indicators:

weight gain (WG, g) = Final body weight (FBW, g) – Initial body weight (IBW, g).

Feed intake (FI, g fish<sup>-1</sup>) = Consumed feed of each pond / No. of fish.

Feed conversion ratio (FCR) = Feed given (g)/Weight gain (g).

Fish wastes and excreta were removed by siphoning and every day, nearly half of each tank's water level was refilled with dechlorinated fresh water. Temperature between (24°C–27°C), dissolved oxygen  $6.5 \pm 0.5$ mg l<sup>-1</sup>, pH  $7.1 \pm 0.8$ , EC  $219 \pm 2$   $\mu$  mho/cm and ammonia adjusted to the typical acceptable limits (<0.1 mg total ammonia) were observed during the study. Water quality assessments were conducted in accordance with Rice, E.W [29].

### Sampling

Blood samples were obtained from five fish per group after finishing the experiment, and caudal vein was used to collect the blood.[30]

### Biochemical parameters

Colorimetric analysis was used to determine the total proteins in serum (Spectrum: Egyptian Biotechnology Company, Egypt) at a wavelength of 546 nm [31]. At 630 nm in wavelength, albumins (Bio-diagnostic co. Egypt) were measured colorimetrically [32]. By mathematically deducting albumin from total protein, the globulin content was determined.

### Digestive enzymes

The colorimetric determination of lipase (Spectrum, Egyptian firm for Biotechnology, Egypt)

carried out at a wave length of 580 nm [33]. At a wave length of 660 nm, amylase (Bio-diagnostic co. Egypt) was measured colorimetrically [34].

#### *Intestine histological examination*

After eight weeks, the samples (five fish per replicate) from each group were taken. Freshly deceased fish were used to obtain intestinal tissue samples, which were then kept for at least 24 hours in 10% neutral buffered formalin. The fixed tissue samples were embedded using the conventional paraffin embedding technique. Hematoxylin and eosin (H&E) was then used to cut and stain 5- $\mu$ m-thick paraffin blocks for histopathological examination [35].

#### *Gene expression analysis*

The liver tissue was subjected to RNA extraction using the Qiagen RNeasy plus Mini kit. At 260 and 260/280 nm ratios, the Spectrophotometer Nano Drop ND-1000 (Nano Drop Technologies, Wilmington, Delaware, USA) was used to determine the concentration and purity of each RNA sample. In order to create cDNA, Reverse transcription of the RNA samples was performed with a High-Capacity RNA-to-cDNA Master Mix kit.

PCR primers special to each gene and Thermo Scientific Maxima® SYBR Green/ROXqPCR Master Mix (2 $\times$ ) by Rotor-Gene Q (Qiagen, USA) were used for the amplification process. As indicated in Table (3), G were created using Primer Express 3.0 (Applied Biosystems, USA) and blasted on NCBI/Blast to verify necessary gene's specificity. We analyzed the PCR products' melting curves to show the identity and specificity of each gene. The resulting Threshold Cycle (Ct) value allowed for the determination of the  $2^{-\Delta\Delta C_t}$  of the various genes' mRNA expression.

#### *Ethics statement*

The Medical Research Committee of the Faculty of Veterinary Medicine at Kafrelsheikh University in Egypt provided an approval for this study based on ethical guidelines (The approval number is KFS-IACUC/123/2023).

#### *Statistical analysis*

The acquired data were statistically analyzed using One-way analysis of variance (ANOVA).

For intergroup comparisons, Tukey's test was applied at ( $p \leq 0.05$ ).

## **Results**

#### *Growth performance*

Table (4) showed the impact of tomato pomace, with or without enzymes and/or amino acids, on Nile tilapia performance. When comparing G4 (TPE) and G5 (TPAE) to G1 (Control), G2 (TP), and G3 (TPA), the final body weight and body weight

gain are significantly higher ( $P < 0.05$ ). Between G4 and G5, there was no significance ( $P = 0.9$ ). In G1 and G2, A significant variation in total feed intake ( $P = 0.03$ ) which presented in Table (4).

The difference between groups in FCR is presented Table (4). In G4 and G5 the results of total FCR are decreased than other groups ( $p < 0.002$ ).

#### *Digestive enzymes*

Table (5) presented the impact of tomato pomace with or without enzymes and /or amino acids on digestive enzymes of Nile tilapia. There is no significant difference in the activity of amylase enzymes in all groups except in G3 and G4 ( $P = 0.02$ )

lipase enzymes activity in all groups showed no notable variation ( $P = 0.98$ )

#### *Serum biochemical parameters*

Table (6) presented the impact of TP with or without enzymes and /or amino acids on the levels of total protein, albumin and globulin in Nile tilapia.

Serum total protein levels in the five groups showed no significant difference ( $p = 0.69$ ).

Also, the levels of serum albumin in the different groups showed no difference ( $p = 0.7$ ).

There is no significance between groups ( $p = 0.58$ ) in serum globulin levels

#### *Histological parameters*

The effects of tomato pomace combined with enzymes and/or amino acids on the morphology of the various intestinal sections in Nile tilapia are shown in Table (7).

It was observed that G5 had significantly more villi length than the G2 in the anterior, middle, and posterior sections of the intestine ( $p < 0.0001$ ). Additionally, G5 had a considerably higher number of goblet cells in the anterior and posterior regions than G2 did ( $p < 0.0001$ ).

#### *Growth - related genes expression*

Figure (1) showed the impact of tomato pomace, with or without enzymes and/or amino acids, on the mRNA expression of Nile tilapia genes associated to growth

#### *Growth hormone gene expression*

GH mRNA levels were substantially higher in G5 than in G2 ( $p < 0.0001$ ). Additionally, G1, G3, and G4 showed a significant difference ( $p < 0.01$ ). There was no discernible difference between G3 and G4 ( $P = 0.06$ ).

#### *Expression of IGF1 gene*

IGF1 mRNA levels in G5 were considerably higher than in the other groups ( $p < 0.0001$ ). Between G1 and G2, there was no significant difference ( $P =$

0.17). Moreover, no obvious difference ( $P= 0.08$ ) in G3 and G4. Otherwise, G3 and G4 had significantly higher *IGF1* mRNA levels than G1 and G2 ( $P<0.05$ ).

### Discussion

Tomato pomace is one of the very healthy by-products that can be utilized as a natural source of lipids (5–10%), protein (10–20%), and fibers (60–70%). Additionally, TP has anti-oxidant properties due to the bioactive substances like  $\beta$ -carotene and lycopene that are present in it [36]. But, the high fiber diet in monogastric animals can have adverse effect on the performance and health [37, 38]. According to published research, exogenous enzymes in Nile tilapia aquafeed reduce the negative effects of NSPs [24, 39]. As a result, these exogenous enzymes improve the digestion of nutrients, especially crude protein and crude lipids, in fish diets as demonstrated by *Dalsgaard* [40]. The present study revealed an elevation in the BWG and FBW and a decrease in FCR in the groups that contain tomato pomace with exogenous enzymes and amino acids (G4 and G5) as compared with tomato pomace 15% group with no significant difference in the feed intake and these findings are similar to *Adeoye*, [41-43] who highlighted the improved final body weight, specific growth rate, feed conversion ratio, and protein efficiency ratio of tilapia fed a diet supplemented with enzymes. Additionally, numerous studies have demonstrated that exogenous enzymes greatly increase fish development and decrease feed coefficient [44-46]. It was observed that tilapia's growth performance may be enhanced and its protein digestibility and digestive enzyme activity considerably increased when fed diets with multienzyme Natuzyme50® (Bioproton) containing protease, phytase, xylanase,  $\beta$ -glucanase,  $\alpha$ -amylase, and cellulase. But, These findings conflict with those of *Amirkolaie*, [47] who found that adding 10% tomato pomace considerably raised the common carp's final weight and specific growth rate. These findings differ from those of *Kavitha* [48] who showed that broiler performance increased when TP was supplemented with enzymes, while nitrogen utilization declined as tomato pomace levels increased. Additionally, *Sahin* [49] found that adding tomato powder at 2.5 and 5% inclusion rates increased the Japanese quail's growth and feed conversion ratio. *Rahmatnejad* [50] demonstrated that dried TP may be added to broiler chicken diets up to 8% without having a negative impact on performance. supplementation of enzymes with dried TP enhances body weight and feed utilization. The variation could be caused by how tomato pomace is composed, which varies depending on farming and processing methods as well as how much moisture is removed and dried [51], which affect in the % of skin ,pulp and seeds in the tomato and so influences on the available energy of diet with high fiber content as clarified by *Squires* [37].

Fish's ability to assimilate nutrients is significantly influenced by digestive enzymes [52]. The amounts of the enzymes lipase and amylase did not significantly differ between the groups. The cause may be inclusion of amino acids only or with exogenous enzymes with TP increase the digestive enzymes and these findings are similar with *Liaqat* [53] who clarified that supplementation of amino acids in stripped catfish increase digestive enzymatic activity and *Maas* [39] who claimed that supplementing tilapia diet with phytase and xylanase increased the activity of digestive enzymes.

Our results revealed no significant difference in total protein, albumin and globulin between groups and these outcomes matched with *Elazab*, [54, 55] who found that adding 15% dried tomato pomace to growing rabbit diets raised the amounts of total protein, albumin, and globulins. Fish digestion and absorption is affected by intestinal villi length and goblet cells number [56]. they also affect the absorption area capacity so they considered a good healthy intestine indicators [57]. Goblet cells produce mucous in the different parts of intestine to make a protection of the intestinal mucosal layer [58]. The current study's findings showed that some intestinal villi parts were noticeably longer and had more goblet cells in G5 than in G2 and these results were similar to *Kim*, [59] who reported that comparing dietary supplementary phytase and basal diets, the exogenous multi-enzyme supplement that was added to the diet significantly improved the villus height, crypt depth, and the ratio of villus height to crypt depth.

*Mun* [60] stated that *GH* and *IGF1* play important function in the growth and health of fish. Few research has been done on how tomato pomace affects the genes linked to growth in Nile tilapia. There are few studies demonstrated impact of TP in tilapia growth- related genes. Our findings present\ed that addition of enzymes and amino acids to TP in tilapia diet has significantly upregulated the mRNA expression of *GH* and *IGF1* and these findings are similar to that of *Hassaan*, [61, 62] who mentioned that tilapia's levels of *IGF1* were raised by adding probiotics and/or exogenous enzymes (protease) to their plant protein diet. *A El-ashry*, [63] indicated that dietary addition of xylanase to plant based diet in tilapia increasing the *GH* levels digestive enzymes and growth performance. *Del Vesco*, [64] shown that adding DL-methionine to tilapia diet raised the amounts of *GH* and *IGF1* in the fish's liver, enhancing performance by promoting the synthesis and release of growth factors and these findings are similar to our results which levels of *GH* and *IGF1* increased in G3, G4 and G5. We can use tomato pomace in fish diet by 15% to improve performance as mentioned by *Abedalhammed* [65]. It was found that including tomato pomace in tilapia diet is lower in cost (9 LE/ kg) than SBM (27 LE

/kg). So, replacing part of SBM by TP in tilapia diet is economic. This work is the first to study inclusion of tomato pomace with lysine and methionine in tilapia fish. Also. Studies on growth related genes with tomato pomace in tilapia fish are very rare

### **Conclusion**

Based on the aforementioned findings, it is possible to conclude that Nile tilapia could make better use of TP up to 15% of the diet when combined with enzymes and/or amino acids. However, adding TP alone to a tilapia's diet negatively impacts both health and performance

### *Acknowledgment*

We would like to express our gratitude to everyone who contributed to this study

### *Conflicted interest*

There are no conflicts of interest to declare.

### *Author's contributions*

Every author made a contribution to this work.

### *Funding statement*

Self-supporting.

**TABLE 1. Chemical composition of dried tomato pomace**

| Nutrient      | %     |
|---------------|-------|
| Dry matter    | 93.0  |
| Crude protein | 17.64 |
| Crude fat     | 8.07  |
| Crude ash     | 9.5   |
| NDF           | 47.4  |
| ADF           | 42.7  |
| Lignin        | 18.8  |

**TABLE 2. Composition of feed ingredients and nutrients in Nile tilapia fish**

| Feedstuff              | G1     | G2    | G3     | G4     | G5     |
|------------------------|--------|-------|--------|--------|--------|
| Corn gluten meal       | 10%    | 10%   | 10%    | 10%    | 10%    |
| Soybean meal           | 35.92% | 32.29 | 32.29% | 32.29% | 32.29% |
| Corn grains            | 27.58% | 18.81 | 18.81% | 18.81% | 18.81% |
| Rice polishing         | 10%    | 8     | 8%     | 8%     | 8%     |
| Wheat middling         | 12%    | 12    | 12%    | 12%    | 12%    |
| Tomato pomace          | -      | 15    | 15     | 15%    | 15%    |
| Enzyme mixtures        | -      | -     | -      | 0.2%   | 0.2%   |
| Premix <sup>1</sup>    | 0.2%   | 0.2   | 0.2%   | 0.2%   | 0.2%   |
| Lysine Hcl             | 0.43%  |       | 0.46%  | -      | 0.46%  |
| Synthetic methionine   | 0.22%  | 0.09  | 0.28%  | 0.09%  | 0.28%  |
| Mono calcium phosphate | 1.32%  | 1.35  | 1.35%  | 1.35%  | 1.35%  |
| Limestone              | 0.59%  | 0.6   | 0.6%   | 0.6%   | 0.6%   |
| CMC binder             | 0.5    | 0.5   | 0.5    | 0.5    | 0.5    |
| Crude protein %        | 28%    | 28%   | 28%    | 28%    | 28%    |
| Starch%                | 27.04% | 21.25 | 21.25  | 21.25% | 21.25% |
| Lipid%                 | 6.25%  | 6.76  | 6.76%  | 6.76%  | 6.76%  |
| DE kcal/kg diet        | 3033   | 2957  | 2957   | 2957   | 2957   |
| Lysine %               | 1.43   | 1.43  | 1.79   | 1.43   | 1.79   |
| Methionine %           | 0.75   | 0.75  | 0.94%  | 0.75   | 0.94   |
| Calcium %              | 0.7    | 0.7   | 0.7    | 0.7    | 0.7    |
| Available phosphorus   | 0.45   | 0.45  | 0.45   | 0.45   | 0.45   |

<sup>1</sup> premix; Each kilogram contained 1000 mg of copper, 1 mg of cobalt, 1000 mg of iodine, 100 mg of selenium, 100,000 mg of iron, 10,000 mg of manganese, 30,000 mg of zinc, 200,000 IU of vitamin A, 10,000 mg of vitamin E, and 2000 IU of vitamin D3.

1000 mg folic acid, 1000 mg B1 and B2; 4000 mg B6 and 4 mg B12; 20,000 mg niacin; 20 mg biotin; 10,000 mg pantothenic acid; calcium carbonates up to 1000 gm

G1 (Control), G2 (Tomato pomace TP), G3 (Tomato pomace with amino acids TPA), G4 (Tomato pomace with enzymes TPE), G5 (Tomato pomace with amino acids and enzymes TPAE)

**TABLE 3. Primers used for real time PCR analysis**

| Primers        | (5'-3')   | Accession number |
|----------------|---|------------------|
| <b>GH</b>      | F: GTTGTGTGTTTGGGCGTCTC<br>R: CAGGTGCGTGACTCTGTTGA    | HM565014.1       |
| <b>IGF-1</b>   | F: GTTTGTCTGTGGAGAGCGAGG<br>R: GAAGCAGCACTC GTCCACG   | Y10830.1         |
| <b>β-actin</b> | F: CCACACAGTGCCCATCTACGA<br>R: CCACGCTCTGTCAGGATCTTCA | EU887951.1       |

**TABLE 4. Effect of tomato pomace with enzymes and /or amino acids supplementation on performance of Nile tilapia**

|                            | G1                      | G2                      | G3                       | G4                       | G5                       | P value |
|----------------------------|-------------------------|-------------------------|--------------------------|--------------------------|--------------------------|---------|
| <b>Initial BW (g/fish)</b> | 21.2±0.08 <sup>a</sup>  | 21.2±0.08 <sup>a</sup>  | 21.2±0.08 <sup>a</sup>   | 21.2±0.08 <sup>a</sup>   | 21.2±0.08 <sup>a</sup>   | 0.43    |
| <b>Final BW (g/fish)</b>   | 42±0.58 <sup>b</sup>    | 41.5±0.58 <sup>b</sup>  | 44.5±0.87 <sup>ab</sup>  | 47.8±0.73 <sup>a</sup>   | 48.2±0.72 <sup>a</sup>   | 0.004   |
| <b>BWG(g/fish)</b>         | 20.84±0.54 <sup>b</sup> | 20.26±0.6 <sup>b</sup>  | 23.3±0.83 <sup>ab</sup>  | 26.56±0.74 <sup>a</sup>  | 26.96±0.96 <sup>a</sup>  | 0.002   |
| <b>Feed intake(g/fish)</b> | 41.24±0.32 <sup>a</sup> | 39.71±0.11 <sup>a</sup> | 40.17±0.14 <sup>ab</sup> | 40.65±0.38 <sup>ab</sup> | 40.62±0.51 <sup>ab</sup> | 0.04    |
| <b>FCR</b>                 | 1.98±0.06 <sup>a</sup>  | 1.96±0.06 <sup>a</sup>  | 1.76±0.12 <sup>ab</sup>  | 1.53±0.05 <sup>b</sup>   | 1.51±0.05 <sup>b</sup>   | 0.002   |

G1 (Control), G2 (Tomato pomace TP), G3 (Tomato pomace with amino acids TPA), G4 (Tomato pomace with enzymes TPE), G5 (Tomato pomace with amino acids and enzymes TPAE)

**TABLE 5. Effect of tomato pomace with enzymes and /or amino acids supplementation on Digestive enzymes of Nile tilapia**

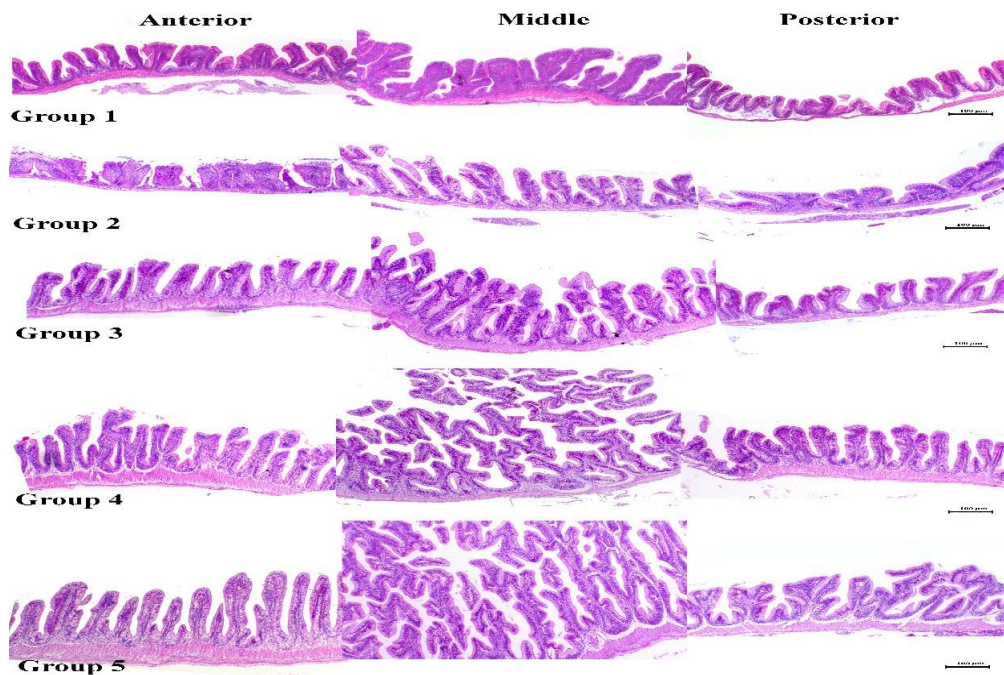
|                       | G1                       | G2                       | G3                      | G4                      | G5                      | P value |
|-----------------------|--------------------------|--------------------------|-------------------------|-------------------------|-------------------------|---------|
| <b>Amylase (U/ml)</b> | 41.93±2.37 <sup>ab</sup> | 38.74±1.64 <sup>ab</sup> | 32.13±2.59 <sup>b</sup> | 48.34±4.4 <sup>a</sup>  | 39.25±3 <sup>ab</sup>   | 0.04    |
| <b>Lipase (U/ml)</b>  | 32.22±4.01 <sup>a</sup>  | 34.76±3.17 <sup>a</sup>  | 30.56±6.45 <sup>a</sup> | 32.60±3.78 <sup>a</sup> | 20.03±0.76 <sup>a</sup> | 0.98    |

G1 (Control), G2 (Tomato pomace TP), G3 (Tomato pomace with amino acids TPA), G4 (Tomato pomace with enzymes TPE), G5 (Tomato pomace with amino acids and enzymes TPAE)

**TABLE 6. Effect of tomato pomace with enzymes and /or amino acids supplementation on total protein, albumin and globulin of Nile tilapia**

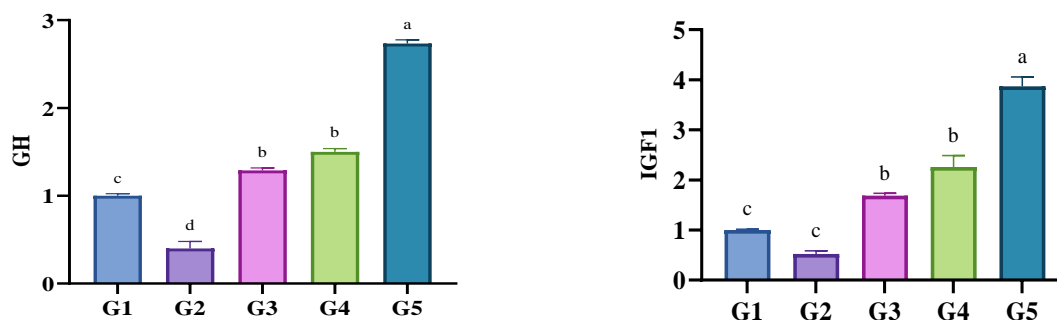
|                             | G1                     | G2                     | G3                     | G4                     | G5                     | P value |
|-----------------------------|------------------------|------------------------|------------------------|------------------------|------------------------|---------|
| <b>Total protein (g/dl)</b> | 4.59±0.16 <sup>a</sup> | 5.15±0.64 <sup>a</sup> | 5.16±0.29 <sup>a</sup> | 5.17±0.56 <sup>a</sup> | 5.48±0.28 <sup>a</sup> | 0.69    |
| <b>albumin (g/dl)</b>       | 1.63±0.03              | 1.63±0.1               | 1.64±0.07              | 1.56±0.04              | 1.53±0.06              | 0.7     |
| <b>Globulin (g/dl)</b>      | 2.96±0.17              | 3.51±0.5               | 3.51±0.54              | 3.6±0.24               | 3.94±0.31              | 0.58    |

G1 (Control), G2 (Tomato pomace TP), G3 (Tomato pomace with amino acids TPA), G4 (Tomato pomace with enzymes TPE), G5 (Tomato pomace with amino acids and enzymes TPAE)



**Fig.1. The effect of tomato pomace with enzymes and or amino acids in histological parameters in Nile tilapia**

G1 (Control), G2 (Tomato pomace TP), G3 (Tomato pomace with amino acids TPA), G4 (Tomato pomace with enzymes TPE), G5 (Tomato pomace with amino acids and enzymes TPAE)



**Fig.2. Effect of tomato pomace with enzymes and/or amino acids supplementation on GH and IGF1 of Nile tilapia**

G1 (Control), G2 (Tomato pomace TP), G3 (Tomato pomace with amino acids TPA), G4 (Tomato pomace with enzymes TPE), G5 (Tomato pomace with amino acids and enzymes TPAE)

## References

- Bartley, D.M., Beveridge, M.C., Phillips, M.J., Tacon, A.G. and Verdegem, M., Enhancing the nutritional values of farmed fish production systems. *Reviews in Aquaculture*, **15**(1), 3-6 (2023).
- Action, S. World fisheries and aquaculture. *Food and Agriculture Organization*, **2020**, 1-244 (2020).
- Colombo, S.M., Roy, K., Mraz, J., Wan, A.H., Davies, S.J., Tibbetts, S.M., Øverland, M., Francis, D.S., Rocker, M.M. and Gasco, L., Towards achieving circularity and sustainability in feeds for farmed blue foods. *Reviews in Aquaculture*, **15**(3), 1115-1141 (2023).
- Mugwanya, M., Dawood, M.A., Kimera, F. and Sewilam, H., Replacement of fish meal with fermented plant proteins in the aquafeed industry: A systematic review and meta-analysis. *Reviews in Aquaculture*, **15**(1), 62-88 (2023).
- Somdare, P.O., Hamid, N.K.A. and Kari, Z.A., Effect of papaya leaf extract inclusion on growth performance and haematological parameters of red hybrid tilapia, *Oreochromis mossambicus* × *Oreochromis niloticus* fed diets formulated with *Hermetia* meal and *Azolla*. *Agriculture Reports*, **2**(1), 29-45 (2023).

6. Midhun, S.J. and Arun, D., *Alternative feed technology in aquaculture*, in *Recent Advances in Aquaculture Microbial Technology* Elsevier., 291-306 (2023).
7. Naylor, R.L., Goldburg, R.J., Primavera, J.H., Kautsky, N., Beveridge, M.C., Clay, J., Folke, C., Lubchenco, J., Mooney, H. and Troell, M., Effect of aquaculture on world fish supplies. *Nature*, **405** (6790), 1017-1024 (2000).
8. Goddard, S., Al-Shagaa, G. and Ali, A., Fisheries by-catch and processing waste meals as ingredients in diets for Nile tilapia, *Oreochromis niloticus*. *Aquaculture Research*., **39**(5), 518-525 (2008).
9. MUTHUI, J.N., Matofari, J.W., Kingori, A., and Hülsebusch, C., Beneficial effects of non-conventional feedstuffs on carcass characteristics and ileal mucosal morphology of finisher pigs. *Egyptian Journal of Veterinary Sciences*, **49**(2), 167-177 (2018).
10. Ghany, A.-E., El-Gebali, M. and Abuelhamd, M., Nutritional Evaluation of Fermented Potato (*Solanumtuberosum*) and Green Bean (*Phaseolus vulgaris*) Vines in Growing Rabbit Diets. *Egyptian Journal of Veterinary Sciences*, **54**(7), 57-66 (2023).
11. EL Dakroury, M., Evaluation of Moringa Oleifera Leaves Powder on Some Blood Parameters and Performances of Broilers. *Egyptian Journal of Veterinary Sciences*. **50**, 81-88 (2019). The 8th International Conference of Veterinary Research Division (NRC) Cairo, Egypt, 3rd-5th December, 2019.
12. Azzam, E.R. and Radif, W.T., Synergistic Effectiveness of Phoenix dactylifera Seed Extract with Some Antibiotics on Staphylococcus aureus Bacteria In-vitro. *Egyptian Journal of Veterinary Sciences*, **55**(7), 1949-1954 (2024).
13. Abozahra, N.A., Albalawi, A., Althobaiti, N., Mohamed, R. and Diab, A., Effects of Pomegranate (*Punica granatum*) Peel Methanolic Extract Dietary Supplementation on *Oreochromis Niloticus* Performance, Blood Health, Intestine Morphometry and Immunity. *Egyptian Journal of Veterinary Sciences*, **54**(7), 221-236 (2023).
14. Bhosale, S.V., Bhilave, M. and Nadaf, S., Formulation of fish feed using ingredients from plant sources. *Research Journal of Agricultural Sciences*, **1**(3), 284-287 (2010).
15. Mansoori, B., Modirsanei, M., Radfar, M., Kiaei, M., Farkhoy, M. and Honar zad, J., Digestibility and metabolisable energy values of dried tomato pomace for laying and meat type cockerels. *Animal Feed Science and Technology*, **141**(3-4), 384-390 (2008).
16. Peiretti, P., Gai, F., Rotolo, L., Brugiapaglia, A. and Gasco, L., Effects of tomato pomace supplementation on carcass characteristics and meat quality of fattening rabbits. *Meat Science*, **95** (2), 345-351 (2013).
17. Travieso, M.D.C., de Evan, T., Marcos, C.N. and Molina-Alcaide, E., Tomato by-products as animal feed. *Tomato Processing by-Products*, 33-76 (2022).
18. Choudhari, S.M. and Ananthanarayan, L., Enzyme aided extraction of lycopene from tomato tissues. *Food Chemistry*, **102**(1), 77-81 (2007).
19. Papaioannou, E.H. and Karabelas, A.J., Lycopene recovery from tomato peel under mild conditions assisted by enzymatic pre-treatment and non-ionic surfactants. *Acta Biochimica Polonica*, **59**(1), 71-74 (2012).
20. Kalogeropoulos, N., Chiou, A., Pyriochou, V., Peristeraki, A. and Karathanos, V.T., Bioactive phytochemicals in industrial tomatoes and their processing byproducts. *LWT-Food Science and Technology*, **49**(2), 213-216 (2012).
21. Sinha, A.K., Kumar, V., Makkar, H.P., De Boeck, G. and Becker, K., Non-starch polysaccharides and their role in fish nutrition—A review. *Food Chemistry*, **127**(4), 1409-1426 (2011).
22. Amirkolaie, A.K., Reduction in the environmental impact of waste discharged by fish farms through feed and feeding. *Reviews in Aquaculture*, **3**(1), 19-26 (2011).
23. Castillo, S. and Gatlin III, D.M., Dietary supplementation of exogenous carbohydrase enzymes in fish nutrition: A review. *Aquaculture*., **435**, 286-292 (2015).
24. Maas, R.M., Verdegem, M.C., Dersjant-Li, Y., and Schrama, J.W., The effect of phytase, xylanase and their combination on growth performance and nutrient utilization in Nile tilapia. *Aquaculture*, **487**, 7-14 (2018).
25. Amirkolaie, A.K., Leenhouwers, J.I., Verreth, J.A. and Schrama, J.W., Type of dietary fibre (soluble versus insoluble) influences digestion, faeces characteristics and faecal waste production in Nile tilapia (*Oreochromis niloticus* L.). *Aquaculture Research*, **36**(12), 1157-1166 (2005).
26. Glencross, B., Rutherford, N. and Bourne, N., The influence of various starch and non-starch polysaccharides on the digestibility of diets fed to rainbow trout (*Oncorhynchus mykiss*). *Aquaculture*, **356**, 141-146 (2012).
27. Dalólio, F.S., Moreira, J., Vaz, D.P., Albino, L.F.T., Valadares, L.R., Pires, A.V. and Pinheiro, S.R.F., Exogenous enzymes in diets for broilers. *Revista Brasileira de Saúde e Produção Animal*, **17**, 149-161 (2016).
28. Jobling, M., National Research Council (NRC): Nutrient requirements of fish and shrimp: The National Academies Press, Washington, DC, 2011, 376+ XVI pp, £ 128 (Hardback), ISBN: 978-0-309 (2012).
29. Rice, E.W., Bridgewater, L. and Association, A.P.H., *Standard methods for the examination of water and wastewater*.: American public health association Washington, DC. Vol. **10** (2012).
30. Jain, N.C., *Schalm's veterinary hematology*. Lea & Febiger (1986).
31. RJ, H., Clinical chemistry: principles and technics. *New-York: Hoeber Medical Division Harper and Row* (1964).
32. Young, D.S., Effects of drugs on clinical laboratory tests.. 4th Edition, AACC Press, Washington (1995).

33. Moss, D. and Henderson, A., Digestive enzymes of pancreatic origin. *Tietz textbook of clinical chemistry*. 3rd ed. Philadelphia: WB Saunders Company, 689-708 (1999).
34. Caraway, W.T., A stable starch substrate for the determination of amylase in serum and other body fluids. *American Journal of Clinical Pathology*, **32**, 97-99 (1959).
35. Suvarna, K.S., Layton, C. and Bancroft, J.D., *Bancroft's theory and practice of histological techniques E-Book*. Elsevier health sciences (2018).
36. Szabo, K., Diaconeasa, Z., Cătoi, A.-F. and Vodnar, D.C., Screening of ten tomato varieties processing waste for bioactive components and their related antioxidant and antimicrobial activities. *Antioxidants*, **8**(8), 292 (2019).
37. Squires, M., Naber, E. and Toelle, V., The effects of heat, water, acid, and alkali treatment of tomato cannery wastes on growth, metabolizable energy value, and nitrogen utilization of broiler chicks. *Poultry Science*, **71**(3), 522-529 (1992).
38. Persia, M., Parsons, C., Schang, M. and Azcona, J., Nutritional evaluation of dried tomato seeds. *Poultry Science*, **82**(1), 141-146 (2003).
39. Maas, R.M., Verdegem, M.C., Stevens, T.L. and Schrama, J.W., Effect of exogenous enzymes (phytase and xylanase) supplementation on nutrient digestibility and growth performance of Nile tilapia (*Oreochromis niloticus*) fed different quality diets. *Aquaculture*, **529**, 723-735 (2020).
40. Dalsgaard, J., Verlhac, V., Hjermslev, N., Ekmann, K.S., Fischer, M., Klausen, M. and Pedersen, P.B., Effects of exogenous enzymes on apparent nutrient digestibility in rainbow trout (*Oncorhynchus mykiss*) fed diets with high inclusion of plant-based protein. *Animal Feed Science and Technology*, **171**(2-4), 181-191 (2012).
41. Adeoye, A.A., Yomla, R., Jaramillo-Torres, A., Rodiles, A., Merrifield, D.L. and Davies, S.J., Combined effects of exogenous enzymes and probiotic on Nile tilapia (*Oreochromis niloticus*) growth, intestinal morphology and microbiome. *Aquaculture*, **463**, 61-70 (2016).
42. Hassaan, M.S., Mohammady, E.Y., Soaudy, M.R. and Abdel Rahman, A.A., Exogenous xylanase improves growth, protein digestibility and digestive enzymes activities in Nile tilapia, *Oreochromis niloticus*, fed different ratios of fish meal to sunflower meal. *Aquaculture Nutrition*, **25** (4), 841-853 (2019).
43. Nascimento, A.A., de Macêdo, É.S., Gonçalves, G.S., da Cruz, T.P., Wernick, B., Furuya, V.R.B. and Furuya, W.M., Xylanase and  $\beta$ -glucanase in tandem improve performance, digestive enzymes activity and digestibility in juvenile Nile tilapia fed corn distillers' dried grains with solubles-added diet. *Animal Feed Science and Technology*, **306**, 115816 (2023).
44. Diógenes, A.F., Castro, C., Carvalho, M., Magalhães, R., Estevão-Rodrigues, T.T., Serra, C.R., Oliva-Teles, A. and Peres, H., Exogenous enzymes supplementation enhances diet digestibility and digestive function and affects intestinal microbiota of turbot (*Scophthalmus maximus*) juveniles fed distillers' dried grains with solubles (DDGS) based diets. *Aquaculture*, **486**, 42-50 (2018).
45. Ghomi, M.R., Shahriari, R., Langroudi, H.F., Nikoo, M. and von Elert, E., Effects of exogenous dietary enzyme on growth, body composition, and fatty acid profiles of cultured great sturgeon *Huso huso* fingerlings. *Aquaculture International*, **20**, 249-254 (2012).
46. Hlophe-Ginindza, S.N., Moyo, N.A., Ngambi, J.W. and Ncube, I., The effect of exogenous enzyme supplementation on growth performance and digestive enzyme activities in *O. reochromis mossambicus* fed kikuyu-based diets. *Aquaculture Research*, **47**(12), 3777-3787 (2016).
47. Amirkolaie, A.K., Dadashi, F., Ouraji, H. and Khalili, K.J., The potential of tomato pomace as a feed ingredient in common carp (*Cyprinus carpio* L.) diet. *Journal of Animal and Feed Sciences*, **24** (2), 153-159 (2015).
48. Kavitha, P., Ramana, J., Rama Prasad, J., Reddy, P. and Reddy, P., Nutrient utilization in broilers fed dried tomato pomace with or without enzyme supplementation. *Indian Journal of Animal Nutrition*, **21**(1), 17-21 (2004).
49. Sahin, N., Orhan, C., Tuzcu, M., Sahin, K. and Kucuk, O., The effects of tomato powder supplementation on performance and lipid peroxidation in quail. *Poultry Science*, **87**(2), 276-283 (2008).
50. Rahmatnejad, E., Pour, M.B., Mamuel, M., Mirzadeh, K. and Perai, A.H., The effects of dried tomato pomace and a multipleenzyme mixture supplementation (Rovabio Excel™) on performance and carcass quality of broiler chickens. *African Journal of Biotechnology*, **10** (45), 9207-9212 (2011).
51. King, A.J. and Zeidler, G., Tomato pomace may be a good source of vitamin E in broiler diets. *California Agriculture*, **58**(1), 59-62 (2004).
52. Wen, Z.P., Zhou, X.Q., Feng, L., Jiang, J. and Liu, Y., Effect of dietary pantothenic acid supplement on growth, body composition and intestinal enzyme activities of juvenile Jian carp (*Cyprinus carpio* var. Jian). *Aquaculture Nutrition*, **15**(5), 470-476 (2009).
53. Liaqat, R., Fatima, S., Komal, W., Minahal, Q. and Hussain, A.S., Dietary supplementation of methionine, lysine, and tryptophan as possible modulators of growth, immune response, and disease resistance in striped catfish (*Pangasius hypophthalmus*). *PLOS ONE*, **19**(4), e0301205 (2024).
54. Kesbiç, O.S., Effects of the cinnamon oil (*Cinnamomum verum*) on growth performance and blood parameters of rainbow trout (*Oncorhynchus mykiss*). *Turkish Journal of Agriculture-Food Science and Technology*, **7**(2), 370-376 (2019).
55. Elazab, M., Zahran, S., Ahmed, M. and Elkom, A., Productive performance of growing rabbits fed diet containing different levels of tomato pomace. *Benha Vet. Med. J.*, **22**(2), 46-57 (2011).
56. Elsabagh, M., Mohamed, R., Moustafa, E.M., Hamza, A., Farrag, F., Decamp, O., Dawood, M.A. and

- Eltholth, M., Assessing the impact of *Bacillus* strains mixture probiotic on water quality, growth performance, blood profile and intestinal morphology of Nile tilapia, *Oreochromis niloticus*. *Aquaculture Nutrition*, **24**(6), 1613-1622 (2018).
57. Khojasteh, S.B., The morphology of the post-gastric alimentary canal in teleost fishes: a brief review. *International Journal of Aquatic Science*, **3**(2), 71-88 (2012).
58. Lauriano, E., Pergolizzi, S., Capillo, G., Kuciel, M., Alesci, A. and Faggio, C., Immunohistochemical characterization of Toll-like receptor 2 in gut epithelial cells and macrophages of goldfish *Carassius auratus* fed with a high-cholesterol diet. *Fish & Shellfish Immunology*, **59**, 250-255 (2016).
59. Kim, M., Ingale, S.L., Hosseindoust, A., Choi, Y., Kim, K. and Chae, B., Synergistic effect of exogenous multi-enzyme and phytase on growth performance, nutrients digestibility, blood metabolites, intestinal microflora and morphology in broilers fed corn-wheat-soybean meal diets. *Animal Bioscience*, **34**(8), 1365 (2021).
60. Mun, S.H., You, J.H., Oh, H.J., Lee, C.H., Baek, H.J., Lee, Y.-D. and Kwon, J.Y., Expression patterns of growth related genes in juvenile red spotted grouper (*Epinephelus akaara*) with different growth performance after size grading. *Development & Reproduction*, **23**(1), 59-62 (2019).
61. Hassaan, M.S., Mohammady, E.Y., Soaudy, M.R., Elashry, M.A., Moustafa, M.M., Wassel, M.A., El-Garhy, H.A., El-Haroun, E.R. and Elsaied, H.E., Synergistic effects of *Bacillus pumilus* and exogenous protease on Nile tilapia (*Oreochromis niloticus*) growth, gut microbes, immune response and gene expression fed plant protein diet. *Animal Feed Science and Technology*, **275**, 114892 (2021).
62. Hassaan, M.S., Mohammady, E.Y., Soaudy, M.R., El-Garhy, H.A., Moustafa, M.M., Mohamed, S.A. and El-Haroun, E.R., Effect of *Silybum marianum* seeds as a feed additive on growth performance, serum biochemical indices, antioxidant status, and gene expression of Nile tilapia, *Oreochromis niloticus* (L.) fingerlings. *Aquaculture*, **509**, 178-187 (2019).
63. A El-ashry, M., Effect of dietary xylanase on growth performance, digestive enzymes and physiological responses of Nile tilapia, *Oreochromis niloticus* fingerlings fed plant-based diets. *Annals of Agricultural Science, Moshtohor*, **59**(5), 71-80 (2021).
64. Del Vesco, A., Gasparino, E., Oliveira Neto, A., Guimarães, S., Marcato, S. and Voltolini, D., Dietary methionine effects on IGF-I and GHR mRNA expression in broilers. *Genet. Mol. Res.*, **12**(4), 6414-6423 (2013).
65. Abedalhammed, H.S., Naser, A.S., Al-Maathedy, M.H., Mohammed, T.T., Jaber, B.T. and Al-Asha'ab, M.H., The effect of vitamin E as an antioxidant with different levels of dried tomato pomace supplementation on diets of common carp (*Cyprinus carpio* L.) on blood indices. *Biochemical and Cellular Archives*, **20**(2), 5173-5176 (2020).

## ثقل الطماطم مع إنزيمات خارجية المنشأ و/أو أحماض أمينية تعزز أداء النمو، والمعايير النسيجية والتعبير الجيني للنمو في أسماك البلطي النيلي

عزه احمد حافظ<sup>1</sup>، الدسوقي السعيد ناصف<sup>1</sup>، عبدالناصر عبداللطيف بكر<sup>1</sup>، ايمان مصطفى مصطفى<sup>2</sup>، وليد صبحي عبدو<sup>3</sup> و السيد محمد حجازي<sup>1</sup>

<sup>1</sup> قسم التغذية والتغذية الاكلينيكية، كلية الطب البيطري، جامعة كفر الشيخ، مصر.

<sup>2</sup> قسم امراض ورعاية الاسماك، كلية الطب البيطري، جامعة كفر الشيخ، مصر

<sup>3</sup> قسم امراض الباثولوجي، كلية الطب البيطري، جامعة كفر الشيخ، مصر.

### الملخص

قامت هذه الدراسة بتقييم تأثير تغذية ثقل الطماطم (TP15%) مع أو بدون إنزيمات وأحماض أمينية لأسماك البلطي النيلي لمدة 8 أسابيع على أداء النمو والإنزيمات الهاضمة والبارامترات النسيجية وبعض التعبيرات الجينية للكبد. تم إعداد خمس وجبات غذائية تجريبية بمحتوى بروتيني مماثل (28%) (1) نظام غذائي ضابطة بدون بروتين TP، (2) بروتين TP 15%، (3) بروتين TP 15% مع ليسين (1.79%) وميثيونين (0.94%)، (4) بروتين TP 15% مع إنزيمات خارجية (0.2%) و (5) بروتين TP 15% مع ليسين وميثيونين وإنزيمات خارجية. وقد كشفت النتائج عن تحسن ملحوظ في أداء النمو، خاصة زيادة وزن الجسم بنسبة 31.05% و 33.03% ومعامل التحويل الغذائي بنسبة 21.83% و 23.35% في المجموعتين الرابعة والخامسة على التوالي مقارنةً بالمجموعة الثانية. زادت الإنزيمات وحدها أو الإنزيمات مع الأحماض الأمينية بشكل ملحوظ من طول الزغابات وعدد الخلايا الكأسية في أمعاء أسماك البلطي. لم يكن هناك فرق كبير في مستويات الليباز والأميلاز والبروتين الكلي والألبومين والجلوبولين في جميع المجموعات. كانت مستويات التعبير النسبي لعامل النمو IGF-1 وهرمون النمو في كبد أسماك البلطي مرتفعة بشكل ملحوظ ( $p < 0.05$ ) في المجموعات الثلاث الأخيرة. أخيرًا، أدت إضافة الإنزيمات مع أو بدون الأحماض الأمينية إلى الوجبات الغذائية لأسماك البلطي النيلي إلى تحسين معايير أداء النمو والتعبير الجيني.

**الكلمات المفتاحية:** ثقل الطماطم، الإنزيمات الخارجية، الأحماض الأمينية، التعبير الجيني.