

## ORNITHOBACTERIUM RHINOTRACHEALE (ORT) ASSOCIATED WITH HATCHING PROBLEMS IN CHICKEN AND TURKEY EGGS.

A.A. EL-GOHARY

\*\* Faculty of Veterinary Medicine, Tanta University (Kafr El-Sheikh)

Received: 27.11.1997

Accepted :21.12.1997

### SUMMARY

Isolation and identification of *Ornithobacterium rhinotracheale* (ORT) from infertile eggs, dead embryos, dead in shell chicks and turkeys as well as day-old chicks and turkey poults obtained from native hatcheries suffering from increased embryonic mortality, dead in shell birds and high mortality in newly hatched chicks and turkey poults was undertaken for the first time. *Salmonella* give (*S. give* could also be concomitantly isolated in certain occasions. Pathogenicity testing of the isolated ORT and/or *S. give* was investigated in 3 day-old chicks. The clinicopathological picture in experimentally infected chicks was described.

from airsacculitis and pneumonia of meat turkeys and broilers (Hafex, 1994; van Beek et al., 1994; van Empe, 1996, Odor et al., 1997 and El-Gohary and Awaad 1997). ORT constitutes a distinct species and genus placed in taxonomic neighbourhood of the genera *Flavobacterium*, *Cytophaga*, *Capnocytophaga* and *Riemerella* within the rRNA super family V (Hinz et al., 1994).

The present work describes isolation and identification of ORT in native hatcheries in Egypt.

### MATERIAL AND METHODS

#### Specimens:

A total of 220 specimens constituting 55 infertile eggs, 110 dead embryos (55 early and 55 late deaths) as well as 55 dead in shell chicks and turkey poults were used. Additionally; samples of

### INTRODUCTION

*Ornithobacterium rhinotracheale* (ORT) gen. nov., sp. nov. has been described by Vandamme et al. (1994). ORT could repeatedly be isolated

lungs, trachea, liver, gall bladder, spleen, heart blood, pericardial fluid, and yolk sac contents were collected from 26 and 15; one-day-old hatched chicks and turkey poults respectively. All collected specimens were representing 9 native hatcheries complaining of low hatchability and high mortality in newly hatched chicks and turkey poults. The specimens were subjected to bacteriological examination.

#### **Bacteriological examination:**

Samples were cultured on MacConkey's agar, 10% sheep blood agar, brain heart infusion broth, peptone water, and Pasteurella broth. All cultured media were incubated at 37°C for at least 48 hours. Isolated bacteria were identified morphologically using Gram stain, biochemically using catalase test and API 20 E strip microtest system (Bio-Merieux, France). Serological identification of the isolates suspected to be *Salmonella* was conducted using polyvalent and monovalent salmonella antisera (Difco lab., Detroit, Michigan, USA). ORT suspected isolates were serologically identified with the agar-gel precipitation (AGPT) test against antisera of ORT serotypes A, B, C, D, E, G and H according to methods described by Van Empe et al. (1997) and Hafez and Sting (1997).

#### **Pathogenicity test:**

Eighty-five, day-old commercial broilers were floor reared and used for pathogenicity testing. Five out of these birds were randomly sacrificed and exposed for bacteriological examination to prove their freedom from bacterial infection. At 3

days of age the other 80 birds were divided into equal groups consisting of 20 each (1-4). Birds of group 1 were sprayed with 100 ml of peptone water containing  $10^9$  CFU/ml of ORT in a commercial sprayer (particle size 250 U). The developed mist was maintained in a commercial sprayer (particle size  $\geq 50$  U). The developed mist was maintained in the isolator for at least 15 minutes with the air circulation closed (Van Empe et al., 1996). Those of group 2 were inoculated with  $10^9$  CFU of the isolated bacteria per bird (El-Gohary, 1992). Chickens of group 3 were kept without infection as control. Birds of group 4 were kept for 3 weeks observation during which clinical signs and mortality were recorded. Dead as well as sacrificed birds at the end of observation period were subjected to post-mortem and bacteriological examination using a scoring system for the lesions post ORT infection was adopted after Van Empe et al. (1996).

#### **RESULTS**

Obtained results are shown in Tables 1-4. Bacteriological examination revealed isolation of a non-lactose fermenter gram negative bacillus suspected to be *Salmonella* species from yolk sac, heart blood and liver of day-old chickens and turkeys and from dead embryos of turkeys. The pleomorphic gram negative rods on blood agar (Tables 1 and 2). The colonial morphology of the latter was 1-2 mm, grey, convex, circular, point shaped colonies. They were able to grow on peptone water and Pasteurella broth and triple sugar iron agar (TSI) slant with no change.

in the but portion. They also were unable to grow on MacConkey's, S.S., Gassner agar or brain heart infusion broth or agar. the colonies have a distinct butyric acid odour. Biochemical characters of the isolated bacteria are shown in Table (3) Serological examination of all ORT and Salmonella isolates clarified that they belong to serotype A and S. give (El; 3, 10; I, V; 7) respectively.

Pathogenicity testing of ORT to 3-day-old chicks revealed only mild airsacculitis at 2<sup>nd</sup>. week post infection (PI). Mild airsacculitis, mild tracheitis and unilateral pneumonia were recorded at the 3<sup>rd</sup>. and 4<sup>th</sup> week PI with 15% mortality and 35%

rate of reisolation.

Pathogenicity testing of S. give showed slight airsacculitis at 2<sup>nd</sup> week PI. Unilateral pneumonia and mild tracheitis at 3<sup>rd</sup>. week PI. While on 4<sup>th</sup>. week mild tracheitis, mild airsacculitis and unilateral pneumonia were recorded accompanied with 20% mortality and 85% rate of reisolation.

Pathogenicity testing of both ORT and S. give resulted in slight airsacculitis at the 1<sup>st</sup>. week PI. On the 2<sup>nd</sup> week; there was slight tracheitis with unilateral pneumonia and moderate airsacculitis. On the 3<sup>rd</sup> and 4<sup>th</sup> weeks; there was moderate tracheitis, moderate airsacculitis and bilateral pneumonia. The mortality reached 25%. The rate of reisolation of ORT and S. give was 30% and 80 respectively (Table 4).

Table (1): Isolation and identification of ORT and S.give from hatching eggs and newly hatched chickens and turkeys.

No. of hatch	species	Samples					Total	Rate of isolation	
		I.E	ED	LD	D.Sh	D-O		ORT	S. give
1	Turkey	10	10	10	10	10	50	9	-
2 a	Turkey	5	5	5	5	5	25	2	1
2 b	Chicken	5	5	5	5	5	25	5	10
3	Chicken	5	5	5	5	3	23	1	-
4	Chicken	5	5	5	5	3	23	-	4
5	Chicken	5	5	5	5	3	23	-	-
6	Chicken	5	5	5	5	3	23	-	-
7	Chicken	5	5	5	5	3	23	2	4
8	Chicken	5	5	5	5	3	23	-	2
9	Chicken	5	5	5	5	3	23	-	-
Total	9 Turkey & chicken	55	55	55	55	41	261	19	21

I.E= Infertile eggs.  
E.D= Early deaths.  
L.D= Late deaths.

D.Sh = Dead in shell embryo.  
D-O = Day-old chicks.

Table(2): Frequency of isolation of ORT and S.give from day old chicks and turkey poults.

Organs	Frequency of isolation			
	Day-old chicks		Day-old turkey poults	
	ORT	S.give	ORT	S.give
Yolk sac	2	6	3	1
Heart blood	-	3	1	-
Pericardium	-	-	-	-
Liver	-	3	-	-
Gall bladder	-	-	-	-
Spleen	-	-	-	-
Lung	1	-	1	-
Trachea	-	-	-	-
Intestine	-	1	-	-
Total	3	13	5	1

Table (3) : Biochemical characteristics of the isolated bacteria from hatching eggs and newly hatched chickens and turkeys.

Tests		Pin point non haemolytic colonies on blood agar	Non-Lactose fermenting colonies on S.S. agar
API 20 E*	ONPG	+	+
	ADH	+	-
	LDC	-	-
	ODC	-	+
	CIT	-	-
	H <sub>2</sub> S	-	-
	URE	+	-
	TDA	-	-
	IND	-	+
	VP	-	-
	GEL	-	+
	GLU	-	+
	MAN	+	+
	INO	+	-
	SOR	-	+
	RHA	+	+
	SAC	+	+
	MEL	-	+
	AMY	+	+
	ARA	+	+
OX	+	-	
Catalase test	-	+	
The isolated pathogens	Ornithobacterium rhinotracheale	S. give	

\* Code number : 4155159 ( 40 )

ONPG = Beta - galactocidase .

ADH = Arginine hydrolase .

LDC = Lysine decarboxylase .

ODC = Ornithine decarboxylase .

CIT = Citrate utilization .

OX = Cytochrome oxidase .

H<sub>2</sub>S = H<sub>2</sub>S production .

URE = Urease .

TDA = Tryptophane desaminase .

IND = Indole production .

VP = Acetoin production .

GEL = Gelatinase .

GLU = Glucose [ fermentation / oxidation ] .

MAN = Mannitol [ fermentation / oxidation ] .

INO = Inositol [ fermentation / oxidation ] .

SOR = Sorbitol [ fermentation / oxidation ] .

RHA = Rhamnose [ fermentation / oxidation ] .

ARA = Arabinose [ fermentation / oxidation ] .

SAC = Sucrose [ fermentation / oxidation ] .

MEL = Meliose [ fermentation / oxidation ] .

AMY = Amygdalin [ fermentation / oxidation ] .

Table (4) : Experimental challenge of three days-old chicks with the isolated strains of Ornithobacterium rhinotracheale (ORT) and S.give.

Group No.	Lesion scores *												Mortalities				Reisolation		
	1st. week P.I.		2 nd. week P.I.		3 rd. week P.I.		4 th. week P.I.		1st. week P.I.	2 nd. week P.I.	3 rd. week P.I.	4 th. week P.I.	Total No.	%	ORT P.I.	%	S.give P.I.	%	
Tr.	L.	AS	Tr.	L.	AS	Tr.	L.	AS	Tr.	L.	AS	No.							%
1	0	0	0	0	1	1	1	1	1	1	1	3	15	7/20	35	0/20	0		
2	0	0	0	0	1	0	1	1	1	1	1	4	20	0/20	0	17/20	85		
3	0	0	1	1	1	2	2	2	2	2	2	5	25	6/20	30	16/20	80		
4	0	0	0	0	0	0	0	0	0	0	0	0	0	0/20	0	0/20	0		

1 = Challenge with Ornithobacterium rhinotracheale isolates.  
 2 = Challenge with S. give isolates.  
 3 = Challenge with Ornithobacterium rhinotracheale + S. give isolates  
 4 = Blank control  
 Tr. = Trachea. L. = Lung. AS. = Air sac  
 Ω No. of positive / total No. of examined birds

\* Lesion scores are calculated After van Empe et al. (1996) as follows:  
 Trachea : 0 = No abnormality, 1 = Slight exudate. 2 = moderate exudate. 3 = Lumen filled with exudate.  
 Lungs : 0 = No abnormality, 1 = Unilateral pneumonia. 2 = Bilateral pneumonia. 3 = Consolidation.  
 Air sacs : 0 = No abnormality, 1 = Slight airsacculitis. 2 = Moderate airsacculitis with limited pinheaded foci of fibrinous exudate. 3 = Severe fibrinous airsacculitis.



## DISCUSSION

*Ornithobacterium rhinotracheale* (ORT) was isolated in different areas all over the world from turkeys and chickens suffering from respiratory infection (Charlton et al., 1993; Hafez et al., 1993; Hinz et al., 1994; van Beek et al., 1994; Tanyi et al., 1995, Odor et al. (1997) and El-Gohary and Awaad 1997).

Nine hatcheries suffered from a problem of increased embryonic mortality (early and late deaths) as well as increased number of dead in shell chicks and turkey poults with high mortality in newly hatched birds were subjected to the present investigation.

Two out of the 9 examined hatcheries were incubating turkey eggs, one of them was also incubating chicken eggs in the same hatchery. The remaining 7 hatcheries were incubating chicken eggs.

As one out of 9 examined native hatcheries were incubating both chicken and turkey eggs together; ORT could be isolated from both species. Moreover, it was not only isolated from dead embryos but also from hatched day old chicks and turkey poults.

The obtained results clarified that 11 out of 75 examined turkey specimens (14.7%) were ORT positive, while only 8 out of 186 examined chicken specimens (4.3%) were ORT positive. This result indicated the magnified role played by ORT in turkey eggs hatchability problems.

All ORT isolates belonged to the serotype A. Serotype A appears to be the most common ORT serotype in chickens and turkeys (Van Empel, 1994 and Hafez and Beyer 1997).

Several infectious diseases appeared to be involved in drop of egg production. In turkey flocks Chin (1996) reported on drop in egg production after ORT infection in turkey breeder flock. Same observation was made by Hafez (1997) in broiler breeder flocks with antibodies to both ORT and avian pneumovirus (Turkey rhinotracheitis).

*S. give* could concurrently be isolated with ORT from 4 out of the 9 examined hatcheries.

Experimental infection of 3 days-old chicks with the isolated ORT and/or *S. give* developed respiratory disease with at least in part the same characteristics as seen in the natural outbreak described by aforementioned authors.

It is apparent that concomitant *S. give* infection increased the severity of the lesion associated with ORT (Table 3). Traver (1996) reported that concomitant ORT and ND infection led to a significantly more severe respiratory disease syndrome in the affected broilers, El-Gohary and Awaad (1997) concluded that experimental infection of *E. coli* aggravated ORT post-mortem lesions. The most prominent clinical signs after experimental infection appeared to be mild respiratory signs. The most prominent lesion after experimental infection of either ORT or *S. give* was the development of mild airsacculitis.

Moreover; concomitant infection with both pathogens yielded lesions of mild to moderate airsacculitis with unilateral or bilateral pneumonia. Similar postmortem features had also been observed in the natural outbreaks and in experimental infections of ORT (Hafez et al.; 1993, and El-Gohary and Awaad; 1997).

Contrary to our results, Ryll et al. (1997) failed to reproduce respiratory score lesions in chickens experimentally infected with ORT and concluded that it does not appear to be a primary, but secondary respiratory pathogen in broilers.

Eventually; in the light of the obtained result, critical questions are emerging such as what is ORT status of SPF birds? and are living vaccines free from bacterial (ORT) contamination?. Such questions are urgently in need of further investigations.

## REFERENCES

- Charlton, B.R., S.E., Channing-Santiago, A.A. Brickford, C.J. Cardona, R.L. Walker (1993): Preliminary characterisation of a pleomorphic gram-negative rod associated with avian respiratory disease. *J. vet. Diagn. Invest.*, 5: 47-51.
- Chin, R.P. (1996): *Ornithobacterium rhinotracheale* infection in turkeys. Hoffmann LaRoche. 2<sup>nd</sup> International turkey Advisory Board Conference. Raleigh, North Carolina.
- El-Gohary, A.A. (1992): Some studies on paratyphoid infections problem in poultry Ph.D. thesis, Faculty of veterinary Medicine, Cairo University.

El-Gohary, A.A. and M.H.H. Awaad (1997): *Ornithobacterium rhinotracheale* (ORT) infection in chicken broilers. *Vet. Med. J., Giza, Egypt*, in Press.

Hafez, H.M.; W. Kruse; J. Emele; and R. Sting (1997): Atemwegsinfektion bei Mast puten durch einen ähnlichen Erreger: Klinik, Diagnostik und Therapie. *Internationale Fachtagung über Geflügel Krankheiten*. Deutsche Veterinär Medizinische Gesellschaft, Frankfurt strasse 8, D- 35392 Postdam, 28 pp 105-112.

Hafez, H.M. (1994): Respiratory disease conditions in turkeys caused by *Ornithobacterium rhinotracheale*. Clinical signs, diagnostics and therapy. *Western Poultry Disease Conference*, Sacramento, California.

Hafez, H.M. and W. Beyer (1997): Preliminary investigation on *Ornithobacterium rhinotracheale* "ORT" using PCR-Fingerprints. *Proceedings of the International Congress of the World Veterinary Association*, Budapest, August 1997. P. 158.

Hafez, H.M. (1997): Serological surveillance of *Ornithobacterium rhinotracheale* "ORT" in breeder flocks. *Proceedings of Xith International Congress of the World Veterinary Poultry Association*, Budapest, August 1997. P. 311.

Hafez, H.M.; and R. Sting (1997): Comparative investigation on different *Ornithobacterium rhinotracheale* "ORT" isolates (In Press).

Hinz, K.H., C. Blome, and M. Ryll (1994): Acute pneumonia and air sacculitis associated with *Ornithobacterium rhinotracheale* in turkeys. *Vet. Rec.* 135:233-234.

Odor E.M.; M. Salem; B. Sample; C. Pope Jr.; and J. Murphy (1997): Diagnostic dilemma of *Ornithobacterium rhinotracheale* Presented October 1997, at the 32<sup>nd</sup> national Meeting on poultry health processing, Ocean City Maryland.

*Vet. Med. J., Giza. Vol. 46, No. 2 (1998)*



Ryll, M., K.H. Hinz, V. Neumann and H. Salisch (1997): Pathogenicity of *Ornithobacterium rhinotracheale* for chicken under experimental conditions. XI the international congress of the world Veterinary Poultry Association pp: 47.

Tanyi, J.; A. Bistyak; E. Kaszanyitzky; F. Vetesi and M. Dobos-Kovacs (1995): Isolation of *ornithobacterium rhinotracheale* from chickens, hens and turkeys showing respiratory symptoms. Magyar Allatrovosk Lapja, 50 (6): 328-330.

Traver, F. (1996): Cited from Chin, R.; R. Droual and B.R. Charlton (1966): *Ornithobacterium rhinotracheale* infection in turkeys. Proc. of the Turkey ORT Symposium, September 4-6, 1996 Minneapolis, Minnesota, pp: 68.

Van Beek, P.N.G.M., P.C.M. Van empel, G. Van den Bosch, P.K. Storm, J.H. Bongers, and J.H. Dupreez (1994): Ademhalings problem groeivertaging en gewrichtssontssteking Bij Kalkoenen en Vleeskuikens door een Pasteurella-achtige bacterie *Ornithobacterium*

*rhinotracheale* of "Taxon 28". Tijdschr. Diergeneeskd. 119: 99-101.

Vandamme, P., P. Segers, M. VanCanneyt, K. Van Hove, r. Mutters, J. Hommez, F. Dewhirst, B. Paster, K. Kersters, E. Falsen, L.A. Devvieste, M. bisgaard, K.H. Hinz, and W. (1994): Mannheim *Ornithobacterium rhinotracheale* gen. nov., sp. nov., isolated from the avian respiratory tract. Int. J. Syst. Bacteriol. 44: 24-37.

Van Empel, P. (1994): *Ornithobacterium rhinotracheale*: Isolation, identification and experimental infection results. Paper given at Poultry Veterinarian study group of the EU held in Amsterdam 11th November, 1994.

Van Empel, P.; H. Van Den Bosch; D. Goovaerts, and P. Storm (1996): Experimental infection in turkeys and chickens with *Ornithobacterium rhinotracheale*. Avian Dis. 40:858-864.

Van Empel, P.; H. Van Den Bosch; P. Loeffen, and P. Storm (1997): Identification and serotyping of *Ornithobacterium rhinotracheale*. Avian Dis. (In Press.

INTRODUCTION

The earliest studies on parasites of the Red Sea fishes were reported by Jagerskiold and Odhner (1901) and Wilson (1928) who collected cestode parasites during the Swedish Zoological Expedition to the White Nile and Red Sea. Troquist (1931) described a nematode, *Trichostrongylus sabarenonchus*, from both

fishes and waterbirds (Banaqa and Hassan (1983) carried out a general survey on helminthic parasites of marine elasmobranchs from Egyptian coastal waters of both the Mediterranean and Red Sea.

The objectives of this preliminary study were to identify parasites of P.O.S. fishes caught from Saudi Arabian waters. This study was dictated by the importance and development of Red Sea fisheries and because fish are anticipated to play a major part in a source of animal protein for the growing population of Saudi Arabia.

