# NCIDENCE OF MOTILE AEROMONAS SPP IN RAW MILK AND SOME DAIRY PRODUCTS

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examined for the presence of presumptive

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total of 150 samples, 50 each of raw milk, asteurized milk and white soft cheese from lifferent dairy, farms; retail shops and upermarkets in Mansoura City, El-Dakahlia rovince, egypt., were examined for the presence if motile Aeromonas spp. The detection of the rganisms by quantitative methods providing nore reliable results compaed to qualitative nethod. Motile Aeromonas occurred in 5.3% and 6% of examined sampels using direct plating nethod and enrichement method respectively. 'he incidence of motile Aeromonas spp. was 6%; 6% and 16% sampels of raw milk; asteurized milk and white soft cheese, espectively using enrichement method. The most ommon Aeromonads isolated was Aeromonas ydrophila which comprised 46.2% of the solates, whilst, A. Caviae; A. Sobria and not lassified strains, represented 29.5%; 9.8% and 4.5% respectively. All Aeromonas isolates were niformly resistant to ampicillin; pincillin and rythromycin and sensitive to other tested ntibiotics with different activity percentage. The resence of motile Aeromonas spp in raw milk

and some diary products may represent an important vehicle of its transmission.

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# INTRODUCTION TO A STATE OF THE STATE OF THE

Aeromonas species have been strongly implicated in many countries as causative agents of gastroenteritis, particularly in children, the gelderly, and immunocompromised patients (Kirovand Hayward, 1993).

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Aeromonads are ubiquitous in nature, being readily isolated from a variety of sources. While Aeromonas species are generally thought of as aquatic organisms, aeromonas strains identical to diarrhaea-associated strains are readily isolated from a wide variety of retail foods (Fricker and tompsett, 1989; Palumbo et al., 1989 and Kirov et al., 1990) Moreover, many of these strains are able to grow at refrigeration temperatures (Palumbo and Buchanan, 1988 and Beuchat, 1991).

The purpore of the present study was to investigate the occurrence of motile Aeromonas

354

spp in rae milk, pasteurized milk and white soft cheese as well as search for its antibiotic sensetivity.

MATERIAL AND METHODS

## Sampling:

One hundred and fifty random samples of raw milk, pasteurized milk and white soft cheese (Each of 50) were collected from different dairy farms, retail dairy shops and supermarkts in the suburbs of Mansoura city, el-Dakahlia province, egypt. All samples were dispatched directly to the laboratory, where they were kept chilled and investigated for occurrence of motile Aeromonas spp.

Preparation and handling of samples were done according to Standard Methods for the examination of Dairy products (Richardson, 1985).

#### Isolation of Aeromonas spp.

1- Direct method: 0.1 ml of each sample after preparation was directly surface plated onto Starch ampicillin agar plates (SA) (Palumbo et al., 1985) and incubated at 28°C for 24 hours. Plates showed large (3-5mm) honey yellow to yellow

treatment with half-strength lugol's iodi, solution were picked up as presumptive Aeromonas spp.

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2- Enrichement method: 25 ml of each sample was added to 225 ml Ampicillin broth (TSB plants) and incubated at 28°C for 24h. A Loophul of the enrichement culture was streaked onto starch ampicillin agar plates and examined for the presence of presumptive Aeromonads colonies after 24 hour incubation at 28°C. The presumptive Aeromonas colonies were streaked onto blood agar for purification and verified as motile Aeromonas (Popoff, 1984). The isolates were identified according to Hickman-Brenner et al., 1987); and Kirov et al., (1983).

#### Antibiotic sensitivity:

Antibiotic snsitivity test of the Aeromonas isolates were performed by the disc method developed by Bauer et al., (1966). Antibiotic discs were obtained from Bio-ADWIC abuzaabal Egypt.

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#### RESULTS A SEW LIGHTING SHOWING TO SERVICE STATE OF THE SERVICE STATE OF

Results are presented in Tables (1 and 2).

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Results are presented in Tables (1 and 2).

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Table (1): Incidence of motile Aeromonas spp in raw milk and some milk products.

Dairy product	No. of samples	in A 2 man and Incidence version of the A					
		Direct platin	ig method	Enrichement method			
		No. positive	%	No. positive	%		
Raw milk	50	4	8	13	26		
Pasteurized milk	50	2 171102	4	3	6		
White soft cheese	50	2 180	10104021	W 80 8 4 0 6 3	16		
Total rugo inaupari	os 1001 the	stly 81 secon	5.31	24 9 118	16		

Table (2): Frequencey distribution of isolated motile Aeromonas spp.

Dairy Dairy	No. of	A. Hydrophila		A.Caviae		A.Sobria		Not classified	
products	isolates	No	%	No	%	No	%	No	%
Raw milk	65	34	52.3	21	32.3	5	7.7	5	7.7
Pasteurized milk	25	9	36	12	48	4	16.	0	0
White soft cheese	42	18	42.9	6	14.3	4	9.5	14	33.3
Total	132	61	46.2	39	29.5	13	9.8	19	14.5

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Table (3): Antibiogram of A. hydrophila, A. sobria and A. caviae isolated from milk and some dairy products.

Antibiotic discs	A.hydrophil (20)		A.sobi	ria(6)	A.caviae(12)		
and their potency	Sensitive isolates	Activity percent	Sensitive isolates	Activity percent	Sensitive isolates	Activity percent	783
Chlormphincol (30ug)	13 \20 \1 v	1 be 100 to	ten 0 yel	0	10	83.3	
Neomycin (30ug)	7	35	18703 (61)	50	12	100	Alk
Gentamycin (10ug)	13	65 A	14.6	100	red 10 olg	83.3	WEL
Erythromycin (15ug)	0	0	0	0	0	iban One	hedi
Trimethoprim- sulphame-thoxazole	10	50	5	83.3	Malponi	91.6	e)gja
(1.25ug-23.75ug)	16 M	70 PO CONTRACTOR (T. 08 21)	0.13	101 State 190	2 12 19 1	16.6	708
Streptomycin (30ug) pencillin (10ug)	0	0	0	0	0	0	6 (26
Ampicillin (10ug)	0	0	0	0	. 0	0	763
Naladixic acid (30ug)	14	70	4	66.6	9	75	1.62
Tetracyclin (30ug)	NE6 12 61	60	50640	83.3	80 S/3 BQ	25	ig vo
Amoxycillin (25ug)	0	0 0	- bod asod	16.6	an h Onton	01,000,0	LAR (

<sup>\*</sup> Nermber of tested isolates.

gales) nijom. Results given in Table (1) indicate that the incidence of motile Aeromonas spp in relation to the method of isolation was greatly varied. The detection of the organisms by enrichements method providing more reliable results compared to direct plating method. Out of 150 samples of dairy products 8 (5.3%) were positive by direct plating method, meanwhile with enrichement method 24 (16%) sample were positive. Nearly similar results were given by Schweizer et al., (1995) who found that the incidence of Aeromonas hydrophila in raw milk was higher by using enrichement method than by direct plating method Moreover, Palumbo et al., (1985) mentioned that motile Aeromonas spp may be present in milk at levels not initially detectable and subsequently out grow after enrichement or prolonged refrigerated storage.

It is likely that the use of enrichement method for isolation of motile Aeromonas spp will be required for optimal recovery of this bacteria.

#### 1- Raw milk:

Motile Aeromonas spp was detected in (13) 26% of raw milk samples by using enrichement method. This finding finding nearly simulate that obtained by Ibrahim and Macrae (1991). while lower findings were reported by Kielwein et al. (1969), Schweizer er al., (1995) and El-Said (1996) and higher findings were reported by Food and Drug Administration "FDA" (1985) and Kirov et al., (1993). On the contorary Krovacek et al. (1992) failed to isolated motile Aeromonas spp

The distribution of isolated Aeromonas spin isolates) from raw milk samples were identified. A. hydrophila 34 (52.3%); A. caviae 21 (32; A. sobria 7 (17.7%) and not classified strain (7.5%) (Table 2) these findings substantiate in have been reported by Ibrahim and Mac ae (19 and Kirov et al., (1993).

It seems that the frequent ocurrence of hydrophila and A. caviae in the environme water (Neves et al., 1990 and Schubert 1991 animal faeces containing organisms (Gray et al., 1990) or milker handlers symptomatic (asymptomatic (Kirov 1993), give ampliopportunity for contamination of milk.

#### 2- Pasteurized milk:

It is evident from table (1) that the incidence of motile Aeromonas spp in pasteurized milk samples was 6% using enrichement method which coincides with that of Walker and Brooks (1993). Greenaway (1988) and Freitas et al. (1993) recorded high results, while loer incidence was obtained by Kirov et al. (1993).

Aeromonas caviae was the predominant species found in pasteurized milk which comprised 48% of isolates, whilst A. hydrophila and a. sobria represented 36% and 16% of isolates respectively. (Table 2). Nearly similar results were given by Freitas et al. (1993). On the contorary Kirov et al. (1993) mentioned that the predominant species recovered from pasteurized milk was A. sobria 42.9%).

Vet.Med.J., Giza. Vol. 45, No. 3(1997)

Aeromonads are not heat-resistant and has adecimal reduciton time (Dvalue) of between 2.2 and 6.6 min at 48°C (Palumbo et al., 1987) or 0.17 min at 55°C (condon et al., 1992). Consequently pasteurization can be relied upon to destroy the strains of Aeromonads: Thereefore the presence of these species in the pasteurized milk imply post pasteurization contamination.

# 3- White soft chees:

The incidence of motile Aeromonas spp in examined white soft cheese was 16% using enrichement method. (Table 1). Relatively few survey have verified by incidence of Aeromonads in white soft cheese. Freitas et al. (1993) could isolat motile Aeromonas spp from 32% samples of not aged white cheese. While a lower contamination rate 2% was reported in study of Walker and Brooks (1993). In egypt, Abd El-Rahman and Ahmed (1988) succeeded to isolate Aeromonas hydrophila from soft cheese.

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Aeromonas hydrophila was the major species found in white soft chees which represented 42.9% of isolates, followed by non classified Strains 33.3%, A. Caviae 14.3% and A. Sobria 9.5% (Table 2). Contrast result was reported by Freitas et al. (1993) who found that non classified strains were the most frequent isolates 61.5%.

### Antibiotic sensitivity of isolates:

The results recorded in Table (3) reveal that all areomonads isolates were showed uniform resistant to ampicillin; pencillin and erythromycin. These findings are similar to those

reported by Gilardi (1983); Villuendas et al. (1991) and Krovacek t al. (1992). Conversely Rahim and Kay (1988) found that 7.5% of52 aeromonas isolates were sensitivite to ampicllin.

The obtained data revealed that A. hydrophila was sensitive to chlormphenical, neomycin, gentamycin trimethoprim sulpha-thoxazole, streptomycin, naladixic acid and tetracyclin with activity percentage of 100%; 35%; 65%; 50%; 80%; 70% and 60% respectively. While it was resistant to erythromycin; ampcillin pencillin and amoxycillin. These findings agree with those reported by Overman (1980); Fass and Barnishan, (1981) who reported that all tested isolates were susceptible to chlormphenical and they were resistant to pencillin; erythromycin; cephalosporins and polymxins. In contrary. Altwegg and Geiss (1989) indicated that the environmental isolates of A. hydrophila were resistant to chlormphenicol and tetracyclin.

With regard to antibiotic susceptability of A. Sobria, the obtained results showed that it was sensitive to neomycin; gentamycin; trimethoprim-sulpha thoxazole; naladixic acid, tetracyclin and amoxycillin with activity percentage of 50%, 100%, 83%, 83.3%, 66.6<sup>A</sup>, 83.3% and 16.6% respectively. While it was resistant to chlormphenical erythromycin, pencillin, ampicillin and streptomycin which are inagreement with that of Krovacek et al. (1992).

Conversely results were reported by Bornemann (1989) who found that 72% of 50 strains of A. Sobria were resistant to ampicillin and 8% to chlormphenical. Regarding to antibiotic

Vet.Med.J., Giza. Vol. 45, No. 3(1997)

357

sensitivity of A. Caviae, the obtained data agree with the finding reported by Hassan (1991).

#### Public health significance of isolates:

It is generally accepted that some strains of motile Aeromonas species afe enteropathogens. Such strains possess virulence associated properties such as the ablity to produce enterotoxin, cytotoxin, haemolysins and an array of proteases and/or invasive ability (Kirov 1993) Some of these strains are able to grow in food under refrigeration (Beuchat 1991) Two types of gastrointestnal illness have been attributed to A. hydrophia and A. sobria. The first and most common is cholera like illness. Both types of diarrhoea are usually mild and self-limited (Stelma, 1989). Janda (1991) and Kirov (1993) reported that A. sobiria was the most spp. often associated with more sever gastrointestinal illness.

In conclusion, the present study shows that species of Aeromonas can be isolated from raw milk, pasteurized milk and white soft cheese with various isolation rates. Furthermore its presence in raw milk and some dairy products shold be regarded as potential health threat particularly for children and immunocompromised individuals.

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Vet.Med.J., Giza. Vol. 45, No. 3(1997)

358

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