



## Shrimp Head Protein Hydrolysate as a Potential Feed Attractant for the Asian Redtail Catfish (*Hemibagrus nemurus*) Larvae

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### ABSTRACT

Protein hydrolysates are known for their feed attractant properties in various fish species; however, their efficacy in the Asian redbtail catfish (*Hemibagrus nemurus*) remains underexplored. This study evaluated the effects of shrimp head protein hydrolysate (SHPH) on feed attractiveness and palatability in the Asian redbtail catfish larvae. A completely randomized design was employed, incorporating four dietary treatments: a control (0% SHPH) and three experimental diets with 2.5, 5, and 7.5% SHPH supplementation, respectively. Larval responses were analyzed, including the attractiveness and palatability of the larval feed, the level of feed consumption, feed efficiency, feed rejection, and the time to first feed capture. Results demonstrated significant effects on feed attractiveness and palatability when SHPH was included in the diets. The treatment with 5% SHPH supplementation produced the most favorable outcomes across all measured parameters. These findings suggest that SHPH can significantly enhance the feeding responses of the Asian redbtail catfish larvae, with a 5% inclusion level in the diets being the most optimal dose. Incorporating SHPH into the larval diets of the Asian redbtail catfish has the potential to improve feed utilization and minimize waste, contributing to more sustainable aquaculture practices for this commercially important species.

### INTRODUCTION

The Asian redbtail catfish (*Hemibagrus nemurus*) is a highly valued freshwater fish species cultivated extensively throughout Indonesia and Southeast Asia (Agusnimar *et al.*, 2023; Saputra *et al.*, 2024). It is known for its desirable meat quality, characterized by a soft, firm texture, delicate flavor, and rich nutritional profile, particularly its high protein content at a range of 45-53% (Hasan *et al.*, 2019; Oktavian *et al.*, 2023). The

economic significance of the Asian redbtail catfish is evident in its market value, with prices in traditional Indonesian markets ranging from IDR 70,000–110,000/kg for fresh fish and IDR 325,000–350,000/kg for smoked products (**Heltonika & Karsih, 2017**).

The high demand for the Asian redbtail catfish is accompanied by an increase in fishing activities in the wild. This situation raises concerns that the Asian redbtail catfish population will decline and become extinct. One effort to maintain the Asian redbtail catfish population is through cultivation. Apart from reducing the intensity of fishing in natural waters, cultivation can also increase the production of good quality fish.

The Asian redbtail catfish hatchery requires feed that is suitable for larvae, easy to digest, and highly nutritious. Silk worms (*Tubifex* sp.) are often used due to their excellent protein, fat, and amino acid content, and they are preferred by fish larvae. However, *Tubifex* sp. can be expensive and difficult to obtain during certain seasons, and they also carry the risk of transmitting diseases and parasites.

To reduce dependence on *Tubifex* sp., alternative feeds, such as artificial feed in the form of pasta, have potential. According to **Siagian *et al.* (2022)**, this type of feed fits the larvae's mouth openings, is easy to digest, and has a soft texture.

Pasta-based feeds are promising alternative to live feeds such as *Tubifex* sp. in aquaculture due to their consistent availability, ease of production, customizable nutritional content, and cost-effectiveness. Previous studies have demonstrated the efficacy of pasta-based feeds in various fish species, including the Asian redbtail catfish, *Hemibagrus nemurus* (**Aryani *et al.*, 2013**), the African catfish, *Clarias gariepinus* (**Nurhayati *et al.*, 2014**; **Tjodi *et al.*, 2016**; **Mullah *et al.*, 2019**), the bonylip barb, *Osteochilus hasselti* (**Yusuf *et al.*, 2014**), the iridescent shark catfish, *Pangasianodon hypophthalmus* (**Jusadi *et al.*, 2015**), and the climbing perch, *Anabas testudineus* (**Rahmi *et al.*, 2016**). While these studies demonstrated a positive growth performance, enhancing feed attractiveness and palatability remains crucial for optimizing larval feed intake and maximizing growth potential.

Feed attractiveness, which is governed by visual and chemosensory cues, has an important role in the initial detection and location of food by fish (**Lokkeborg *et al.*, 2014**; **Olsen & Lundh, 2016**). Palatability, on the other hand, dictates the ultimate acceptance or rejection of food based on gustatory perception within the oral cavity (**Lokkeborg *et al.*, 2014**; **Olsen & Lundh, 2016**).

Shrimp head waste, often underutilized or discarded, is a potential source of valuable feed ingredients, including oil and protein hydrolysate, which can be extracted at low cost (**Decarli *et al.*, 2016**; **Silva *et al.*, 2017**; **Diana *et al.*, 2023**). Hydrolysis, a chemical process, breaks down complex proteins into smaller peptides and amino acids, enhancing their digestibility and bioavailability for fish (**Wisuthiphaet *et al.*, 2015**). These low molecular weight nitrogenous compounds, generated through hydrolysis, can act as potent feed attractants by stimulating the olfactory system of fish (**Broggi *et al.*, 2017**).

The efficacy of protein hydrolysates in improving feed attractiveness and palatability has been documented in several studies. For example, sardine protein hydrolysate enhanced food searching behavior in the silver catfish, *Rhamdia quelen* larvae (Broggi *et al.*, 2017). Similarly, poultry protein hydrolysate supplementation improved feed intake in the Nile tilapia, *Oreochromis niloticus* larvae (Alves *et al.*, 2019). Pig liver protein hydrolysate effectively increased feed consumption in the betta fish, *Betta splendens* larvae (Oliveira *et al.*, 2022), while feather protein hydrolysate demonstrated positive effects on feed attractiveness and palatability in the pacu, *Colossoma macropomum* × *Piaractus mesopotamicus* (Santos *et al.*, 2022). Therefore, this study aimed to evaluate the potential of shrimp head protein hydrolysate as a feed attractant and palatability enhancer for Asian redtail catfish larvae.

## MATERIALS AND METHODS

This study was conducted at the Microalgae and Fish Nutrition Laboratory, Faculty of Agriculture, Universitas Islam Riau, Pekanbaru, Riau, Indonesia, from May to August 2024.

### Production of shrimp head protein hydrolysate

The tiger shrimp head protein hydrolysate was prepared following the method described by Suparmi *et al.* (2019), with slight modifications. Briefly, 500g of finely ground shrimp head flour was mixed with 500mL of distilled water (1:1 w/v ratio) and homogenized for 2 minutes. The pH of the mixture was adjusted to 7.0 using 1 M NaOH or 1 M HCl to achieve the optimal pH for papain enzyme activity. Papain enzyme was added at a concentration of 15%, and the mixture was incubated at 60°C for 24 hours. Enzyme deactivation was achieved by boiling the mixture in a water bath at 85°C for 15 minutes. The supernatant (liquid phase) containing the shrimp head protein hydrolysate was separated from the precipitate (residue) by centrifugation at 3000rpm for 15 minutes, followed by filtration.

### Experimental diets

Four isonitrogenous (45%) and isoenergetic (5,000 kcal/kg) experimental diets were formulated following the nutritional recommendations proposed by Eguia *et al.* (2000). The diets evaluated were: SHPH 0 (control, diet with 0% shrimp head protein hydrolysate), SHPH 2.5 (diet with 2.5% shrimp head protein hydrolysate), SHPH 5 (diet with 5% shrimp head protein hydrolysate), and SHPH 7.5 (diet with 7.5% shrimp head protein hydrolysate). The composition of the diets is shown in Table (1).

**Table 1.** Composition of experimental diets used in evaluating the attractiveness and palatability of SHPH for the Asian redbtail catfish (*H. nemurus*)

Ingredient	Diet (% dry ingredients)			
	SHPH 0	SHPH 2.5	SHPH 5	SHPH 7.5
Fish meal	54	53.5	53	52.5
Soybean meal	28	26	24	22
Wheat flour	12	12	12	12
Fish oil	2	2	2	2
Vitamin mix <sup>a</sup>	2	2	2	2
Mineral mix <sup>b</sup>	2	2	2	2
SHPH <sup>c</sup>	0	2.5	5	7.5
Total	100	100	100	100

<sup>a</sup> Vitamin mix (mg/100 g diet): thiamine 5.0; riboflavin 5.0; Ca-pantothenate 10.0; niacin 2.0; pyridoxine 4.0; biotin 0.6; folic acid 1.5; cyanocobalamin 0.01; inositol 200;  $\rho$ -aminobenzoic acid 5.0; menadion 4.0; vitamin A palmitate 15.0; chole-calciferol 1.9;  $\alpha$ -tocopherol 20.0; choline chloride 900.0.

<sup>b</sup> Mineral mix (mg/100 g diet):  $\text{KH}_2\text{PO}_4$  412;  $\text{CaCO}_3$  282;  $\text{Ca}(\text{H}_2\text{PO}_4)$  618;  $\text{FeCl}_3 \cdot 4\text{H}_2\text{O}$  166;  $\text{ZnSO}_4$  9,99;  $\text{MnSO}_4$  6,3;  $\text{CuSO}_4 \cdot 2\text{H}_2\text{O}$  0,05;  $\text{CuSO}_4 \cdot 7\text{H}_2\text{O}$  0,05; KJ 0,15; Dekstrin 450; Selulosa 553,51.

<sup>c</sup> SHPH (Shrimp Head Protein Hydrolysate).

### Feed preparation and proximate analysis

Pasta-form artificial feeds were prepared by thoroughly mixing the primary ingredients (fish meal, soybean meal, and wheat flour). Fish oil, vitamin and mineral premix, and shrimp head protein hydrolysate were then added according to the designated treatments. Water was gradually incorporated and mixed until a homogenous paste consistency was achieved, as described by **Syamsunarno and Sunarno (2022)**. Proximate analysis (Table 2) and amino acid profiling (Table 3) of the prepared pasta feeds were conducted following the standard procedures outlined by **AOAC (2005)** and shown in Tables (2, 3). Prepared feeds were stored frozen to maintain freshness and thawed 15 minutes prior to feeding, as described by **Siagian *et al.* (2022)**.

**Table 2.** Chemical composition of the experimental diets used in evaluating the attractiveness and palatability of SHPH for the Asian redbtail catfish (*H. nemurus*)

Chemical composition	Diet (% dry matter)			
	SHPH 0	SHPH 2.5	SHPH 5	SHPH 7.5
Protein	45.49	45.78	45.73	45.79
Fat	16.46	12.78	11.47	10.68
Ash	12.81	11.94	12.78	11.98
NFE <sup>a</sup>	23.35	28.01	28.08	29.66
Energy (GE kcal/g) <sup>b</sup>	509.06	494.39	481.89	481.13

<sup>a</sup> NFE is calculated based on the difference (100 – protein – fat – ash) (**Hasan *et al.*, 2019**).

<sup>b</sup> Gross energy was calculated according to **Chandan *et al.* (2021)** as 5.7, 9.5 and 4.0 kcal/g for protein, fat and NFE.

**Potential of Shrimp Head Protein for Attracting the Catfish Larvae**

**Table 3.** Amino acid profile of experimental diets used in evaluating the attractiveness and palatability of SHPH for the Asian redbtail catfish (*H. nemurus*)

Type of amino acid	Diet (%)			
	SHPH 0	SHPH 2.5	SHPH 5	SHPH 7.5
Essential amino acids				
Arginine	1.36	1.44	1.42	1.74
Histidine	0.33	0.71	0.83	0.61
Isoleucine	1.18	1.18	1.22	1.18
Leucine	2.37	2.44	2.47	2.49
Lysine	2.16	1.89	2.34	2.41
Methionine	0.66	0.67	0.66	0.68
Phenylalanine	1.03	1.11	1.26	1.22
Threonine	0.75	0.79	0.77	0.75
Valine	1.53	1.27	1.33	1.31
∑EAA	11.37	11.50	12.30	12.39
Non-essential amino acids				
Alanine	1.34	1.36	1.28	1.31
Aspartic acid	2.35	2.88	2.47	2.67
Cystine	0.89	0.74	0.87	0.91
Glutamic acid	3.27	3.55	3.26	3.54
Glycine	1.84	1.56	1.88	1.85
Proline	1.06	1.46	1.84	1.77
Serine	1.42	1.24	1.60	1.42
Tyrosine	1.02	0.95	0.99	0.88
∑NEAA	13.19	13.74	14.19	14.35
EAA/NEAA ratio	0.86	0.84	0.87	0.86

### Experimental units and fish acclimation

Twelve aquaria (60×33×35cm), each filled with 30L of freshwater and equipped with aeration, were used. 1800 healthy Asian redbtail catfish larvae (average weight 0.02g) were obtained from the Technical Service Unit of Fish Seed Center, Universitas Islam Riau. Upon arrival at the laboratory, the fish were acclimated in a holding tank (2×1×0.5m) for 30 minutes before being randomly distributed into each aquarium at a density of 150 fish per tank (5 fish/L), as described by **Nasution *et al.* (2021)**.

### Experimental diets and feeding regime

During a 10-day acclimation period, the fish were fed a combination of *Tubifex* sp. and pasta feed containing SHPH according to the assigned treatments. Following the adaptation period, a 10-day observation period commenced. During that time, fish were

fed the experimental diets five times daily (7:00 am, 11:00 am, 3:00 pm, 7:00 pm, and 11:00 pm) to apparent satiation (Nasution *et al.*, 2021).

### Water quality management

Water quality was maintained through daily filtering. Parameters including temperature, pH, dissolved oxygen, and ammonia were monitored at the beginning, middle, and end of the experiment. Temperature and pH were measured using a pH tester (H198108, Romania), DO using a DO meter (Lutron PDO-519, Taiwan), while ammonia was evaluated using an ammonia MR (HI715, Romania). The ranges of these parameters: temperature of 27.7-29.8°C, pH of 5.32-6.57, DO of 5.5-6.4mg/ L, and ammonia of 0.0607-0.1821mg/ L, and were within the optimal levels for growth and survival of the Asian redbtail catfish larvae.

### Experimental design and data analysis

A completely randomized design with three replicates per treatment was employed. Fish feeding behavior was recorded for three minutes after feed introduction using a camera. Observation of the fish responses lasted for ten days (Alves *et al.*, 2020). The palatability index was calculated based on the method described by Kasumyan and Sidorov (2012) and Alves *et al.* (2020) using this equation:

$$\text{Palatability Index} = \left( \frac{\text{consumption of test feed (g)} - \text{consumption of control feed (g)}}{\text{consumption of test feed (g)} + \text{consumption of control feed (g)}} \right) \times 100$$

The following feeding behaviors were analyzed: a) refusal of food after being caught (RFBC); b) not responding to food (NRF); c) time of first capture of food; level of feed consumption (LFC); and feed efficiency (FE) following these formulas:

$$\text{RFBC} = \left( \frac{\text{number of fish that refuse food after being caught (heads)}}{\text{number of sample fish (heads)}} \right) \times 100$$

$$\text{NRF} = \left( \frac{\text{number of fish that do not respond to food (heads)}}{\text{number of sample fish (heads)}} \right) \times 100$$

$$\text{LFC} = \text{weight of feed provided at the beginning (g)} - \text{weight of feed at the end (g)}$$

$$\text{FE} = \frac{(\text{final weight (g)} + \text{weight of dead fish (g)}) - \text{initial weight (g)}}{\text{total feed during rearing}} \times 100$$

Data were analyzed using one-way ANOVA, and significant differences between treatments were determined using Duncan's multiple range test at 95% confidence level using SPSS software.

## RESULTS

Diets supplemented with SHPH demonstrated greater palatability and feed consumption compared to the control diet (without SHPH) (Table 4). The SHPH 5 diet significantly enhanced both palatability index ( $P<0.05$ ) and feed consumption ( $P<0.05$ ) compared to all other treatments. Fish fed the SHPH 5 diet also exhibited significantly higher feed efficiency ( $P<0.05$ ) than those on other diets, with the exception of the SHPH 7.5 group. Furthermore, SHPH supplementation significantly impacted the time to first feed capture. Fish fed the SHPH 5 diet exhibited the fastest time (5.20 seconds), followed by SHPH 7.5 (6.60 seconds), SHPH 2.5 (10.20 seconds), and lastly, the control diet (13.80 seconds).

**Table 4.** Average value of attractiveness and palatability of SHPH for the Asian redbtail catfish (*H. nemurus*)

Treatments (%)	Palatability index (%)	Feed consumption (g)	Feed efficiency (%)	Feed refusal after being caught (%)	Fish not responding to feed (%)	First feed capture time (second)
SHPH 0 (Control)	0.00 ± 0.36 <sup>a</sup>	40.38 ± 0.29 <sup>a</sup>	53.22 ± 2.56 <sup>a</sup>	1.67 ± 0.14 <sup>c</sup>	3.02 ± 0.16 <sup>c</sup>	13.80 ± 0.60 <sup>d</sup>
SHPH 2.5	14.01 ± 0.93 <sup>b</sup>	53.55 ± 1.01 <sup>b</sup>	56.14 ± 2.34 <sup>a</sup>	1.20 ± 0.27 <sup>ab</sup>	2.53 ± 0.24 <sup>b</sup>	10.20 ± 0.60 <sup>c</sup>
SHPH 5	20.64 ± 0.54 <sup>d</sup>	61.38 ± 0.69 <sup>d</sup>	67.38 ± 0.95 <sup>b</sup>	1.11 ± 0.16 <sup>a</sup>	1.93 ± 0.17 <sup>a</sup>	5.20 ± 0.35 <sup>a</sup>
SHPH 7.5	17.82 ± 0.36 <sup>c</sup>	57.89 ± 0.44 <sup>c</sup>	65.82 ± 1.52 <sup>b</sup>	1.54 ± 0.12 <sup>bc</sup>	2.46 ± 0.12 <sup>b</sup>	6.60 ± 0.00 <sup>b</sup>

Description: SHPH (shrimp head protein hydrolysate). The values shown are the mean ± SD. Different superscript letters in the same column indicate significant differences between treatments ( $P<0.05$ ).

## DISCUSSION

Supplementation of SHPH in the feed for the Asian redbtail catfish larvae led to better outcomes compared to the control group without SHPH supplementation. This result can be attributed to the presence of low molecular weight peptide chains in SHPH (Ribeiro *et al.*, 2017; Resende *et al.*, 2023; Nikoo *et al.*, 2024; Saputra *et al.*, 2024). These peptides are known to provide essential amino acids (Mach & Nortvedt, 2011; Santos *et al.*, 2022), improve protein digestibility (Khosravi *et al.*, 2015; Sandbakken *et al.*, 2023), and enhance feed consumption (Alves *et al.*, 2019; Pham *et al.*, 2021). These findings are consistent with other studies that have demonstrated the positive effects of protein hydrolysate on palatability in farmed fish, including the Atlantic salmon, *Salmo salar* (Refstie *et al.*, 2004), the barramundi, *Lates calcarifer*

(**Chotikachinda *et al.*, 2013**), and the European seabass, *Dicentrarchus labrax* (**Leduc *et al.*, 2018**).

The feeding behavior of fish is strongly influenced by low molecular weight compounds including amino acids, peptides, and nitrogen compounds (**Alves *et al.*, 2020**). These compounds are detected by the fish's senses of smell and taste, and play critical roles in controlling feeding behavior by assessing the attractiveness and palatability of food (**Siikavuopio *et al.*, 2017**; **Oliveira *et al.*, 2022**). In addition, various fish species exhibit different degrees of sensitivities and preferences for specific amino acids, resulting in differences in their acceptance or rejection of food (**Olsen & Lundh, 2016**; **Alves *et al.*, 2019**; **Oliveira *et al.*, 2022**).

There are certain amino acids such as cysteine, betaine, glutamic acid, serine, glycine, alanine, proline, methionine, phenylalanine, arginine, tyrosine, valine, leucine, and glutamine, which act as feeding stimulants in fish. These amino acids provide chemical signals to the taste system of this fish, enhancing the perceived attractiveness of food (**Santos *et al.*, 2022**). Numerous studies have demonstrated the stimulatory effects of low molecular weight substances, including amino acids, peptides, and nitrogen compounds, on fish feeding behavior, feed consumption, digestive enzymes, and antioxidant enzyme activity (**Sudagar *et al.*, 2010**; **Silva *et al.*, 2017**; **Giri *et al.*, 2021**; **Sandbakken *et al.*, 2023**; **Santos *et al.*, 2023**; **Fan *et al.*, 2024**; **Nikoo *et al.*, 2024**).

Palatability is a critical factor in food selection for fish, as these influences whether the fish accepts or rejects the feed. When food enters the mouth of the fish, the taste receptors detect and recognize the taste-providing substances. Based on this sensory information, the fish decides whether to swallow or reject the food and determines the amount that will be consumed (**Lokkeborg *et al.*, 2014**; **Olsen & Lundh, 2016**; **Olsen & Toppe, 2017**).

The results obtained in this study (Table 4) indicate that a 5% SHPH supplementation level is the optimum dose for enhancing palatability and feed consumption, which in turn improves the growth of the Asian redbtail catfish larvae compared to other treatments. This finding can be attributed to the ability of this dosage of supplementation to fulfill the free amino acid requirements of the test feed (Table 3). This optimal supplementation level enhanced the attractiveness and palatability of the feed, resulting in improved feed efficiency. These observations are consistent with the findings of **Ndobe *et al.* (2022)**, who reported that an optimal attractant dosage in fish feed can enhance the feed conversion ratio by positively influencing feed intake, growth, and palatability. Research on various fish species, such as freshwater eels, gourami, and seabass, have shown that attractants such as squid meal, garlic extract, turmeric extract, and other natural ingredients can significantly increase feed consumption. In addition, **Afifah *et al.* (2021)** reported that attractants in optimal amounts can stimulate fish appetite, make food more attractive, and increase the speed at which fish find and consume food.



Several studies show that the use of hydrolysate as a food attractant for fish yielded positive results. **Broggi et al. (2017)** reported that the use of sardine protein hydrolysate in the feeds for the South American catfish, *Rhamdia quelen* increased the feed intake of this species. Furthermore, **Alves et al. (2019)** evaluated the palatability of protein hydrolysates from industrial products for tilapia, *Oreochromis niloticus*. The findings in the study were that all diets containing poultry by-products exhibited higher palatability indices, feed consumption rates, and reduced feed rejection behavior. **Oliveira et al. (2022)** found that protein hydrolysates from agro-industrial by-products, such as chicken protein hydrolysate, pig liver protein hydrolysate, feather protein hydrolysate, and pig mucosa protein hydrolysate, have good attractiveness and palatability indices to replace fish meal in the feeds for the betta fish, *Betta splendens*. Additionally, **Santos et al. (2022)** found that feather protein hydrolysate is proven to have good attractiveness and palatability for the mondfish, *Colossoma macropomum* × *Piaractus mesopotamicus*. Furthermore, recent research by **Santos et al. (2023)** elucidated that feather protein hydrolysate can be used to replace fish meal in the diets of the Nile tilapia, *Oreochromis niloticus* without changing the taste and eating behavior of the fish.

The decreased feed palatability index observed at a 7.5% SHPH supplementation level suggests that this concentration exceeded the optimal limit for SHPH as an attractant. This over-supplementation likely reduced the palatability of the feed, leading to decreased consumption. According to **Peng et al. (2022)**, excessive attractant supplementation can cause feed to become unpalatable and cause nutritional imbalances, which can reduce appetite. Similarly, **Yue et al. (2022)** noted that exceeding the optimal attractant supplementation level can reduce the appeal of the feed to fish, resulting in decreased feed consumption and growth.

Feed rejection in fish is closely linked to their ability to detect and recognize favorable or undesirable substances in their food, ultimately influencing their decision to accept or reject the feed (**Lokkeborg et al., 2014; Moraes, 2016; Olsen & Lundh, 2016**). Increasing the supplementation percentage of SHPH (2.5, 5, and 7.5%) in the feed led to a decrease in both the rejection rate and the number of fishes exhibiting no response to the feed ( $P < 0.05$ ). The control feed (SHPH 0%) had the highest values for rejection and non-response, reaching 1.67 and 3.02%, respectively. This outcome highlights the importance of SHPH in enhancing feed palatability and, consequently, acceptance by the Asian redbtail catfish larvae. These findings are congruent with those of **Alves et al. (2019)**, who observed a similar impact on feed rejection and non-response in tilapia when industrial by-product protein hydrolysate supplementation was absent in the diets.

Rapid food perception is a crucial aspect of fish feeding behavior, and it can be effectively triggered by providing highly attractive food. This attraction stimulates an extra-oral response, so that the fish will try to find food upon sensing that stimulus (**Kasumyan & Doving, 2003; Moraes, 2016**). To reduce leaching of nutrients into the

water, rapid capture of feed is also important to ensure complete and balanced consumption and to reduce the impact of nutrient losses to the environment due to exposure to feed in water (Cyrino *et al.*, 2010; Oliveira *et al.*, 2022). Statistical analysis revealed a significant difference in the time taken to capture the first feed among the treatment groups ( $P < 0.05$ ). The optimal result was achieved with 5% SHPH supplementation, where the time to first feed capture was 5.20 seconds. Research by Alves *et al.* (2019) showed that the Nile tilapia, *O. niloticus* fed chicken protein hydrolysate, liver and sea urchins had an average time to catch the first feed at 0.82 seconds. Santos *et al.* (2022) attributed this rapid response to a well-developed extra-oral gustatory system, which enables fish to efficiently locate or avoid food by exhibiting various movements, including stopping, returning, swimming backward, turning sideways, and zigzagging.

## CONCLUSION

This study demonstrates that dietary SHPH supplementation can significantly enhance both the attractiveness and palatability of feed for the Asian redbtail catfish larvae. A 5% SHPH inclusion rate proved optimal, yielding significantly better results than other tested doses. These findings suggest that SHPH could potentially be used as a feed attractant for the Asian redbtail catfish larvae.

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## REFERENCES

- Afifah, D.; Arief, M. and Al-Arif, M.A. (2021). The effect of garlic (*Allium sativum*) and turmeric (*Curcuma longa*) extract addition in commercial feed on feeding rate, feed efficiency and feed conversion ratio of gouramy fish (*Osphronemus gouramy*). IOP Conf. Series: Earth and Environmental Science, 679: 012073.
- Agusnimar; Setiaji, J.; Sadikin, K.; Marliana, D.; Cahyo, F.E. and Hadi, K. (2023). The effects of feeding different percentages of fermented sago dregs and anchovy head meal feed on survival and growth of asian redbtail catfish (*Hemibagrus nemurus*). Jurnal Riset Akuakultur, 18(2): 71-80. [In Indonesian].
- Alves, D.R.S.; Oliveira, S.R.D.; Luczinski, T.G.; Boscolo, W.R.; Bittencourt, F.;

- Signor, A. and Detsch, D.T.** (2020). Attractability and palatability of liquid protein hydrolysates for Nile tilapia juveniles. *Aquaculture Research*, 00: 1-8.
- Alves, D.R.S.; da Silva, T.C.; Rocha, J.D.M.; de Oliveira, S.R.; Señor, A. and Boscolo, W.R.** (2019). Compelling palatability of protein hydrolysates for Nile tilapia juveniles. *Latin American Journal of Aquatic Research*, 47(2): 371-376.
- AOAC.** (2005). Official method of analysis. 18<sup>th</sup> Edition, Association of Officiating Analytical Chemists, Washington DC, Method 935.14 and 992.24.
- Aryani, N.; Pamukas, N.A. and Adelina.** (2013). Growth of green catfish seed fed on sludge worm and artificial feed combination. *Jurnal Akuakultur Indonesia*, 12(1): 18–24. [In Indonesian].
- Broggi, J.A.; Wosniak, B.; Uczay, J.; Pessatti, N.A. and Fabregat, T.E.H.P.** (2017). Sardine waste protein hydrolysate as feeding stimulant for silver catfish juveniles. *Arquivo Brasileiro de Medicina Veterinaria e Zootecnia*, 69(2): 505-512.
- Chandan, C.S.S.; Roy, P.; Khatun, F. and Roy, N.C.** (2021). Effect of dietary protein on growth, survival and cannibalism of larval striped snakehead, *Channa striata* (Bloch, 1793). *Asian Fisheries Science*, 34: 236–242.
- Chotikachinda, R.; Tantikitti, C.; Benjakul, S.; Rustad, T. and Kumarnsit, E.** (2013). Production of protein hydrolysates from skipjack tuna (*Katsuwonus pelamis*) viscera as feeding attractants for Asian seabass (*Lates calcarifer*). *Aquaculture Nutrition*, 19(5): 773–784.
- Cyrino, J.E.P.; Bicudo, Á.J.D.A.; Sado, R.Y.; Borghesi, R. and Dairiki, J.K.** (2010). Fish farming and the environment - the use of environmental, friendly feeds in fish culture. *Revista Brasileira de Zootecnia*, 39: 68-87.
- Decarli, J.A.; Pedron, F.A.; Lazzari, R.; Signor, A.; Boscolo, W.R. and Feiden, A.** (2016). Protein hydrolysates for the feeding of catfish (*Rhamdia voulezi*) in cages. *Revista Brasileira de Ciência Veterinária*, 23: 168–173.
- Diana, M.; Hartati; Teguh; Anthony, S. and Gunarti, T.S.** (2023). Utilization of shrimp waste (Shells and Heads) as a MSG substitute for flavor enhancement in Muara Telang Marga Village, Banyuasin Regency. National Seminar on Community Service, Universitas Terbuka, 637–644. [In Indonesian].
- Eguia, R.V.; Kamarudin, M.S. and Santiago, C.B.** (2000). Growth and survival of river catfish *Mystus nemurus* (Cuvier & Valenciennes) larvae fed isocaloric diets with different protein levels during weaning. *Journal of Applied Ichthyology*, 16: 104–109.
- Fan, Z.; Wu, D.; Li C.; Zhou, M.; Wang, L.; Zhang, H.; Li J.; Rong, X.; Miao, L.; Zhao, D. and Wang, J.** (2024). Application of fish protein hydrolysates in common carp (*Cyprinus carpio*) diets for fish meal sparing: Evidence from growth, intestinal

- health and microflora composition. *Aquaculture Reports*, 36: 102160.
- Giri, N.A.; Astuti, N.W.W.; Sudewi; Marzuqi, M. and Asih, Y.N.** (2021). Fish hydrolysate supplemented diet improved feed efficiency and growth of coral trout (*Plectropomus leopardus*). *IOP Conf. Series: Earth and Environmental Science*, 890: 012024.
- Hasan, B.; Putra, I.; Suharman, I.; Iriani, D. and Muchlisin, Z.A.** (2019). Growth performance and carcass quality of river catfish *Hemibagrus nemurus* fed salted trash fish meal. *Egyptian Journal of Aquatic Research*, 45(3): 259–264.
- Heltonika, B. and Karsih, O.R.** (2017). Maintenance of Asian redbtail catfish (*Hemibagrus nemurus*) fry with photoperiod technology. *Jurnal Berkala Perikanan Terubuk*, 45(1): 125–137. [In Indonesian].
- Jusadi, D.; Anggraini, R.S. and Suprayudi, M.A.** (2015). Combination of *Tubifex* and artificial diet for catfish *Pangasianodon hypophthalmus* larvae. *Jurnal Akuakultur Indonesia*, 14(1): 30–37. [In Indonesian].
- Kasumyan, A.O. and Doving, K.B.** (2003). Taste preferences in fishes. *Fish and Fisheries*, 4 (4): 289-347.
- Kasumyan, A.O. and Sidorov, S.S.** (2012). Effects of the long-term anosmia combined with vision deprivation on the taste sensitivity and feeding behaviour of the rainbow trout *Parasalmo (=Oncorhynchus) mykiss*. *Journal of Ichthyology*, 52: 109–119.
- Khosravi, S.; Rahimnejad, S.; Herault, M.; Fournier, V.; Lee, C.R.; Bui, H.T.D.; Jeong, J.B. and Lee, K.J.** (2015). Effects of protein hydrolysates supplementation in low fish meal diets on growth performance, innate immunity and disease resistance of red sea bream *Pagrus major*. *Fish & Shellfish Immunology*, 45(2): 858-868.
- Leduc, A.; Atylny-Gaudin, C.Z.; Robert, M.; Corre, E.; Corguille, G.L.; Castel, H.; Lefevre-Scelles, A.; Fournier, V.; Gisbert, E.; Andree, K.B. and Henry, J.** (2018). Dietary aquaculture by-product hydrolysates: impact on the transcriptomic response of the intestinal mucosa of European seabass (*Dicentrarchus labrax*) fed low fish meal diets. *BMC Genomics*, 19: 396.
- Lokkeborg, S.; Siikavuopio, S.I.; Humborstad, O.B.; Palm, A.C.U. and Ferter, K.** (2014). Towards more efficient longline fisheries: Fish feeding behavior, bait characteristics and development of alternative baits. *Reviews in Fish Biology and Fisheries*, 24: 985–1003.
- Mach, D.T.N. and Nortvedt, R.** (2011). Free amino acid distribution in plasma and liver of juvenile cobia (*Rachycentron canadum*) fed increased levels of lizardfish silage. *Aquaculture Nutrition*, 17: 644-656.
- Moraes, S.** (2016). The physiology of taste in fish: potential implications for feeding

- stimulation and gut chemical sensing. *Reviews in Fisheries Science & Aquaculture*, 25(2): 133–149.
- Mullah, A.; Diniarti, N. and Astriana, B.H.** (2019). The effect of sludge worms (*Tubifex*) combined with artificial feed on feed efficiency and growth of African catfish larvae (*Clarias gariepinus*). *Jurnal Perikanan*, 9(2): 160–171. [In Indonesian].
- Nasution, M.R.; Aryani, N. and Nuraini** (2021). The effect of stocking density and different feeding frequencies on growth and survival of Asian redbtail catfish (*Hemibagrus nemurus*) larvae. *Jurnal Ilmu Perairan (Aquatic Science)*, 9(3): 173–179. [In Indonesian].
- Ndobe, S.; Muamar; Rosyida, E.; Widiastuti, I.M.; Mansyur, K.; Fadly, Y. and Tantu** (2022). Effect of adding squid meal as an attractant to freshwater eel (*Anguilla marmorata*) feed on elver growth, feed palatability, efficiency and conversion. *IOP conference series*, 1075: 012016.
- Nikoo, M.; Mozanzadeh, M.T.; Noori, F.; Imani, A.; Houshmand, H.; Sam, M.R. and Jafari, F.** (2024). The effects of protein hydrolysates from rainbow trout by-products on growth, digestive and antioxidant enzymes, and liver lysozyme activity in sobaity (*Sparidentex hasta*) and Arabian yellowfin (*Acanthopagrus arabicus*) seabream juveniles. *Aquaculture Reports*, 37: 102229.
- Nurhayati.; Utomo, N.B.P. and Setiawati, M.** (2014). Development of digestive enzymes and growth of fish larvae of African catfish, *Clarias gariepinus* Buchell 1822, that given of silk worms and artificial feed combination. *Jurnal Iktiologi Indonesia*, 14(3): 167–178. [In Indonesian].
- Oktavian, T.; Rosyadi and Hadi, K.** (2023). The influence of different salinities on growth and survival of asian redbtail catfish fry (*Hemibagrus nemurus*). *Jurnal Dinamika Pertanian*, 39(3): 283-292. [In Indonesian].
- Oliveira, S.R.D.; Alves, D.R.S.; Gomes, R.L.M.; Hattori, J.F.D.A.; Signor, A.; Boscolo, W.R. and Bittencourt, F.** (2022). Attractivity and palatability of different hydrolysed proteins for the ornamental species *Betta splendens* (Regan, 1910). *Aquaculture Research*, 16: 15798.
- Olsen, K.H. and Lundh, T.** (2016). Feeding stimulants in an omnivorous species, crucian carp *Carassius carassius* (Linnaeus 1758). *Aquaculture Reports*, 4: 66–73.
- Olsen, R.L. and Toppe, J.** (2017). Fish silage hydrolysates: not only a feed nutrient, but also a useful feed additive. *Trends in Food Science and Technology*, 66: 93-97.
- Peng, D.; Peng, B.; Li J.; Zhang, Y.; Luo, H.; Xiao, Q.; Tang, S. and Liang, X.** (2022). Effects of three feed attractants on the growth, biochemical indicators, lipid metabolism and appetite of Chinese perch (*Siniperca chuatsi*). *Aquaculture Reports*,

23: 101075.

- Pham, H.D.; Siddik, M.A.B.; Phan, U.V.; Le, H.M. and Rahman, M.A.** (2021). Enzymatic tuna hydrolysate supplementation modulates growth, nutrient utilisation and physiological response of pompano (*Trachinotus blochii*) fed high poultry-by-product meal diets. *Aquaculture Reports*, 21: 100875.
- Rahmi, I.; Yulisman and Muslim** (2016). Survival rate and growth of climbing perch larvae (*Anabas testudineus*) fed with combination silk worms and artificial feed. *Jurnal Akuakultur Rawa Indonesia*, 4(2): 128–139. [In Indonesian].
- Refstie, S.; Olli, J.J. and Standal, H.** (2004). Feed intake, growth, and protein utilisation by post-smolt Atlantic salmon (*Salmo salar*) in response to graded levels of fish protein hydrolysate in the diet. *Aquaculture*, 239(1–4): 331–349.
- Resende, D.; Pereira, R.; Domínguez, D.; Pereira, M.; Pereira, C.; Pintado, M.; Valente, L.M.P. and Velasco, C.** (2023). Stress response of European seabass (*Dicentrarchus labrax*) fed plant-based diets supplemented with swine blood hydrolysates. *Aquaculture Reports*, 30: 101600.
- Ribeiro, M.D.S.; Fonseca, F.A.L.D.; Queiroz, M.N.D.; Affonso, E.G.; Conceição, L.E.C.D. and Gonçalves, L.U.** (2017). Fish protein hydrolysate as an ingredient in diets for *Arapaima gigas* juveniles. *Boletim Do Instituto De Pesca*, 43(especial): 85–92.
- Sandbakken, I.S.; Five, K.K.; Bardal, T.; Knapp, J.L. and Olsen, R.E.** (2023). Salmon hydrolysate as a protein source for Atlantic salmon; prion content and effects on growth, digestibility and gut health. *Aquaculture*, 576: 739863.
- Santos, R.A.D.; Brisqueleal, J.C.P.; Piovesan, M.R.; Souza, O.J.D. and Boscolo, W.R.** (2023). Attractiveness and palatability of feather protein hydrolysate for juvenile Nile tilapia (*Oreochromis niloticus*). *Revista Observatorio De La Economia Latinoamericana*, 21(9): 11300–11317.
- Santos, R.A.D.; Piovesan, M.R.; Oliveira, S.R.D.; Hattori, J.F.D.A.; Souza, O.J.D.; Boscolo, W.R.; Signor, A. and Bittencourt, F.** (2022). Attractiveness and palatability of hydrolyzed feather protein for juvenile tambacu (*Colossoma macropomum* × *Piaractus mesopotamicus*). *Research, Society and Development*, 11(16): 19111637352.
- Saputra, I.; Lee, Y.N. and Fotedar, R.** (2024). The effect of supplementation of fish protein hydrolysate to the bsf-based aquafeed on the growth, survival, fatty acids, and histopathology of juvenile lobster (*Panulirus ornatus*). *Aquaculture Nutrition*, 2024: 1–11.
- Saputra, A.; Suryaningrum, L. H.; Sari, R. W.; Rahardjo, S.; Siti Murniasih, S. and Edy Farid Wadjdy, E. F.** (2024). Optimizing Water Flow Rate to Improve Growth,

Feed Efficiency, and Stress Response in Sustainable *Hemibagrus nemurus* Farming. Egyptian Journal of Aquatic Biology & Fisheries, 28(4): 1993 – 2010

- Siagian, D.R.; Aryani, N.; Heltonika, B. and Tartila, S.S.Q.** (2022). Evaluation of feeding periods of combination between tubificid worms and formulated feed paste on growth and survival of giant gourami larvae. Jurnal Riset Akuakultur, 17(4): 265–277. [In Indonesian].
- Siikavuopio, S.I.; James, P.; Stenberg, E.; Evensen, T. and Saether, B.S.** (2017). Evaluation of protein hydrolysate of by-product from the fish industry for inclusion in bait for longline and pot fisheries of Atlantic cod. Fisheries Research, 188: 121–124.
- Silva, T.C.; Rocha, J.D.M.; Moreira P.; Signor, A. and Boscolo, W.R.** (2017). Fish protein hydrolysate in diets for Nile tilapia post-larvae. Pesquisa Agropecuária Brasileira, 52: 485–492.
- Sudagar, M.; Zelti, H. and Hosseini, A.** (2010). The use of citric acid as attractant in diet of grand sturgeon *Huso huso* fry and its effects on growing factors and survival rate. AACL Bioflux, 3(4): 311-316.
- Suparmi; Harahap; Nursyirwani; Efendi, I. and Dewita** (2019). Production and characteristics of rebon shrimp (*Mysis relicta*) protein hydrolysate with different concentrations of papain enzymes. International Journal of Oceans and Oceanography, 13(1): 189–198.
- Syamsunarno, M.B. and Sunarno, M.T.D.** (2022). Response of post-larva of snakehead (*Channa striata*) to feeding dried silkworm (*Tubifex* sp.) and artificial diet. DEPIK Journal of Aquatic, Coastal and Fishery Sciences, 11(1): 16–22.
- Tjodi, R.; Kalesaran, O.J. and Watung, J.C.** (2016). The effect of feed combination on growth and survival of catfish larvae, *Clarias gariepinus*. Budidaya Perairan, 4(2): 1–7. [In Indonesian].
- Wisuthiphaet, N.; Kongruang, S. and Chamcheun, C.** (2015). Production of fish protein hydrolysates by acid and enzymatic hydrolysis. Journal of Medical and Bioengineering, 4: 466–470.
- Yue, Y.; Chen, M.; Bao, X.; Yu, Y.; Shi, W.; Kumkhong, S.; Liu, Y.; Yang, Y. and Yu, H.** (2022). Effects of three feed attractants on the growth performance and meat quality of the largemouth bass (*Micropterus salmoides*). Frontiers In Marine Science, 9: 1029969.
- Yusuf, D.H.; Sugiharto and Wijayanti, G.E.** (2014). Development of post-larval *Osteochilus hasselti* C.V. with different feeding regimens. Scripta Biologica, 1(3): 185–192. [In Indonesian].