

HOSTED BY



ELSEVIER

Contents lists available at ScienceDirect

Egyptian Journal of Anaesthesia

journal homepage: www.elsevier.com/locate/egja

Research article

Lidocaine suppressed hyperinflammation in BALB/c mice model sterile injury via downregulation of toll-like receptor 4

Robert Hotman Sirait^a, Mochammad Hatta^{b,*}, Muhammad Ramli^c, Carmen Siagian^d, Bambang Suprayogi^e, Tigor Paniel Simanjuntak^f^a Department of Anesthesiology, Faculty of Medicine, Christian University of Indonesia, Jakarta, Indonesia^b Molecular Biology and Immunology Laboratory, Faculty of Medicine, University of Hasanuddin, Makassar, Indonesia^c Department of Anesthesiology, Faculty of Medicine, University of Hasanuddin, Makassar, Indonesia^d Department of Cliniton Nutrition, Faculty of Medicine, Christian University of Indonesia, Jakarta, Indonesia^e Department of Otorhinolaryngology, Faculty of Medicine, Christian University of Indonesia, Jakarta, Indonesia^f Department of Obstetric and Gynecology, Faculty of Medicine, Christian University of Indonesia, Jakarta, Indonesia

ARTICLE INFO

Keywords:

Lidocaine

TLR4

Sterile injury

ABSTRACT

Background: To study the efficacy of systemic lidocaine in suppressing toll-like receptor 4 (TLR4) protein level in BALB/c mouse with sterile injury.**Material and methods:** Twenty healthy adult male BALB/c mice were divided into lidocaine and control groups. The sterile injury was performed by breaking the left thigh bone of the mouse without laceration. Four hours after sterile injury the lidocaine group was treated with 2 mg/kg of lidocaine through tail vein injection. The same volume of distilled water was injected into control group instead of lidocaine. Blood was drawn from tail vein before injury, 4 h after sterile injury and 2 h after systemic lidocaine and distilled water administration. TLR4 protein level was examined by enzyme-linked immunosorbent assay (ELISA).**Results:** The TLR4 protein level in mice that sustained hyper inflammation due to sterile injury was significantly decreased in the lidocaine group. ($p < 0.00$).**Conclusion:** Systemic therapy of lidocaine effectively inhibits TLR4 protein in BALB/c mice that sustained hyperinflammation due to sterile injury.

1. Introduction

Toll-like receptors (TLRs) are identifying receptor initiating innate immune response against substances produced by pathogenic microbes, pathogen-associated molecular patterns (PAMPs) and endogenous molecules released by damaged cells, damage-associate molecular patterns (DAMPs) [1–6]. TLR4 is important to regulate immune system against inflammation caused by infection and trauma [7–11]. Previous studies have shown that when suppressed, TLR4 signaling pathway will provide global protection against sepsis-induced organ dysfunction [12–15]. In addition analgesic and anti-arrhythmia properties, lidocaine also is known to have anti-inflammatory properties and able to modulate inflammatory cascade while possessing protective effect against ischemic injuries on liver, lungs and heart on septic mouse model [16–18]. The anti-inflammatory effect of local anesthesia acts on various cells including monocytes, macrophages and neutrophils.

Although lidocaine is important for immune system and inflammation, the mechanisms involved in its action are less understood [19–21]. The aim of this study is to determine whether the injection lidocaine can suppress hyperinflammation response in BALB/c mouse with sterile injury via downregulation of the TLR4 signaling pathway (see Fig. 1).

2. Material and methods

This was a prospective laboratory experimental animal study using 20 healthy adult male BALB/c mice, age 10–12 weeks. Healthy BALB/c mice have glowing eyes no fainted fur, active and have a good appetite. Mice who died during the study were excluded. Mice were obtained from the maintenance and development unit of the experimental animal laboratory of Molecular Microbiology and Immunology Faculty of Medicine, Hasanuddin University, Makassar, Indonesia. The experiments were carried out according to procedures and principles of the

Peer review under responsibility of Egyptian Society of Anesthesiologists.

* Corresponding author.

E-mail address: hattaram@yahoo.com (M. Hatta).<https://doi.org/10.1016/j.egja.2018.07.002>

Received 23 January 2018; Received in revised form 5 July 2018; Accepted 18 July 2018

Available online 11 October 2018

1110-1849/ © 2018 Egyptian Society of Anesthesiologists. Production and hosting by Elsevier B. V. All rights reserved. This is an open access article under the CC BY-NC-ND license (<http://creativecommons.org/licenses/by-nc-nd/4.0/>).

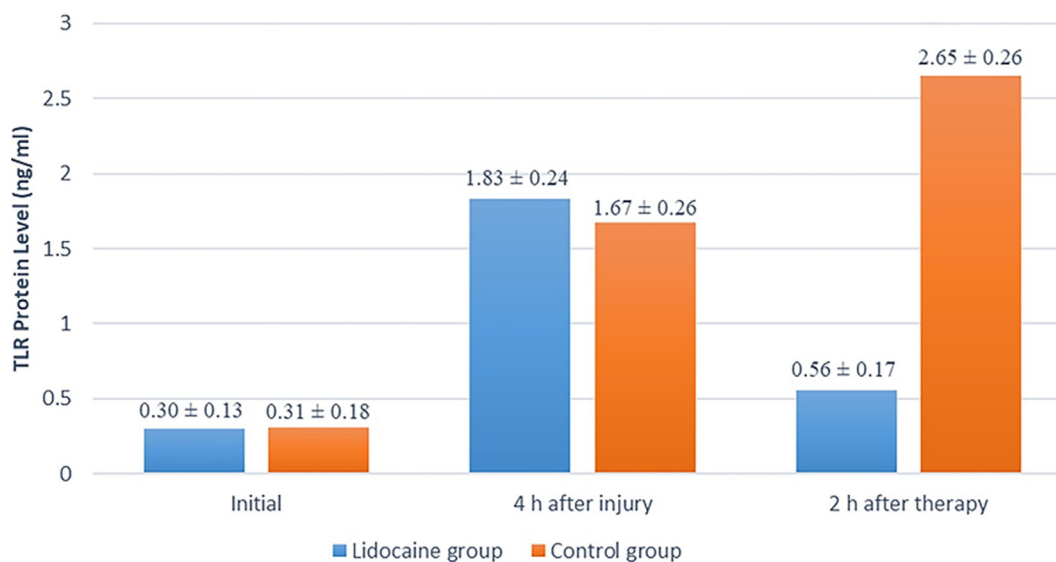


Fig. 1. TLR4 protein level of the lidocaine and control groups (n = 10 per group). Data was presented in form of mean and standard deviation. The p-value was tested with *t*-test and $p < 0.05$ was considered as significant.

Purpose of Control and Supervision of Experiments on Animal (CPCSEA). The number of research samples was determined by the ethical utilization of experimental animals in the healthcare sector using the principle of replacement, reduction and refinement. The research was conducted after obtaining the recommendation of ethical clearance from Medical Research Ethics Committee Faculty of Medicine, Hasanuddin University (Makassar, Indonesia) with registration number UH16050436 dated 28 October 2016. The study was conducted at the Laboratory of Molecular Microbiology and Immunology Faculty of Medicine, Hasanuddin University at the end of November 2016 until early December 2016.

Twenty healthy adult male BALB/c mice were divided into the following two groups: lidocaine and control group. Each group consisted of ten BALB/c mice. A blood sample (0.3 ml) was taken from the tail vein of each mouse for examination of initial TLR4 protein level. The mice were then anesthetized with 50 mg/kg of ketamine, intraperitoneally. A model of sterile injury was established by breaking the left thigh bone using two needle holders without laceration. Four hours after the mice underwent sterile injury, 0.3 ml of blood was taken from the tail vein (second blood test). The lidocaine group was then treated with 2 mg/kg of lidocaine (2% lidocaine, PT Kimia Farma, Jakarta, Indonesia) through tail vein injection once every 2 h continuously for 24 h. The control group was treated with the same volume of distilled water instead of lidocaine. Two hours after completion of the lidocaine and distilled water administrations, 0.3 ml of blood was drawn from the tail vein of both the lidocaine and control groups (third blood test). All blood samples were collected using centrifugation at 5000 rpm for 5 min and were kept in -80°C before used.

The level of TLR4 in the serum was determined with ELISA kits (Life Span Bioscience, Inc. Seattle, North America) according to the manuals from the manufacturer.

The data were analyzed using SPSS software version 20. The normally distributed data were tested with Kolmogorov-Smirnov test. The data was then presented as mean \pm SD and tested with *t*-test. A value of $p < 0.05$ was considered significant.

3. Results

The mean weights of BALB/c mice in the lidocaine and control groups were 39.30 g and 39.34 g, respectively. There was no significant difference between the two experimental groups ($p > 0.05$).

The initial level of TLR4 protein in the lidocaine group was

0.30 ± 0.13 (ng/ml). Four hours after sterile injury, the protein level increased to 1.83 ± 0.24 (ng/ml), and 2 h after systemic lidocaine treatment, the level decreased to 0.56 ± 0.17 (ng/ml), $p < 0.05$. The initial level of TLR4 protein in the control group 0.31 ± 0.18 (ng/ml). Four hours after the sterile injury, the level increased to 1.67 ± 0.26 (ng/ml), and 2 h after systemic distilled water administration, the level increased to 2.65 ± 0.26 (ng/ml), $p < 0.05$.

4. Discussion

Toll-like receptors are a large family of type I transmembrane protein, function as pattern recognition receptors of the innate immune system [2–4]. TLRs are able to recognize microbes product or pathogen associated molecular patterns (PAMPs) and endogenous ligand related to inflammation or damage associated molecular patterns (DAMPs) [3–5]. TLR4 are extracellular TLRs that first found on mammals, presented mainly by polymorphonuclear leucocytes, monocytes, macrophages, dendritic cells, and any other cells including epithelial and endothelial cells [3,6,7]. TLR4 is important for regulation of immunologic and inflammatory response as it utilized Toll/IL-1 receptor (TIR) domain-containing adapter protein (TIRAP) and MyD88 adapter-like (Mal) to “bridge” myeloid differentiation primary response gene 88 (MyD88) to the receptors and thus activate nuclear factor kappa B (NF- κ B). TLR4 transduction signaling used mainly MyD88-dependent pathway, utilizing TIRAP to bridge TLR4 and MyD88 [1,3].

Our research showed that TLR4 protein level were present in normal BALB/c mouse blood. Four hours after sterile injury, the TLR4 protein level increased 6.1 fold in lidocaine group and 5.39 fold in control group. The increased TLR4 protein level showed that sterile injury inflicted substantial sterile hyperinflammation in BALB/c mice. After treatment with 2 mg/kg of lidocaine through the tail vein, once every 2 h continuously for 24 h the level of TLR4 protein decreased from 1.83 ± 0.24 to 0.56 ± 0.15 ($p < 0.00$). Our results showed that intravenous administration of 2 mg/kg of lidocaine, effectively suppressed of TLR4 protein level in BALB/c mouse with a sterile injury [19,20]. In contrast, the level of TLR4 protein in control group continued to rise from before the injury to after distilled water administration. The increased levels of TLR4 protein in control group were statistically significant ($p < 0.05$), revealing that systemic distilled water treatment does not effectively suppress sterile inflammation [20–22].

Previous study showed that systemic lidocaine therapy possessed

anti-inflammatory effect on various diseases or septic model and organ failure on experimental animals via downregulation of TLR4 [3,22,25]. Research conducted by Liu et al. [3], showed that systemic lidocaine therapy can inhibit production of inflammatory mediators including interleukin-6 (IL-6), interleukin-1 β (IL-1 β), γ interferon, tumor necrosis factor α (TNF α) induced by LPS and down regulation of TLR4 dan NF- κ B [3,25–27]. Activation of NF- κ B is inhibited by systemic lidocaine administration and possess protective effect during sepsis [3].

The finding of this study showed that injection of 2 mg/kg lidocaine, once every 2 h continuously for 24 h effectively suppressed TLR4 protein in sterile hyperinflammation model when compared with the control group. The results of this study were consistent with the results of previous research that systemic lidocaine therapy has anti-inflammatory properties by suppressing hyperinflammation caused by pathogenic infection and sterile injury [20,27].

5. Conclusion

Systemic therapy of 2 mg/kg of lidocaine, once every 2 h continuously for 24 h, effectively suppressed hyperinflammation on BALB/c mouse that underwent sterile injury via downregulation of TLR4 protein level.

6. Author contribution

Following authors have made substantial contribution to the manuscript as under:

Sirait R H: Concept, Data Collection and writing

Hatta M: Review, concept, data analysis

Ramli M: Data analysis, data collection

Siagian C: Bibliography, writing

Suprayogi B: Data collection, data analysis

Simanjutak TP: writing, data collection

Authors agree to be accountable for all aspects of the work in ensuring that question related to the accuracy or integrity of any part of the work are appropriately investigated and resolved.

Acknowledgement

We would like to thank Rommy Usman and Mus (Molecular Biology and Immunology Laboratory, Faculty of Medicine, Hasanuddin University, Makassar, Indonesia who helped in the implementation of our research activities.

Financial support and sponsorship

Nil.

Conflict of interest

The authors declare there is no conflict of interest.

References

- [1] Couture LA, Piao W, Ru LW, Vogel SN, Toshchakov VY. Targeting Toll-Like Receptor (TLR) Signaling By Toll/Interleukin-1 Receptors (TIR) Domain-containing Adaptor Protein/MYD88 Adaptors-Like (TIRAP/Mal)-derived Decoy Peptides. *J Biol Chem* 2012;287:24641–8.
- [2] Abbas KA, Lichtman AH, Pillai S. Cellular and Molecular IMMUNOLOGY Chapter 4 9th ed. Philadelphia: Elsevier Saunders; 2017. p. 51–85.

- [3] Liu J, Zhang H, Qi Z, Zheng X. Lidocaine protects against renal and hepatic dysfunction in septic rats via downregulation of Toll-like receptor 4. *Mol Med Reports* 2014;9:118–24.
- [4] Nystrom S, Antoine DJ, Lundback P, Lock JG, Nita AF, Hoghstrand K, et al. TLR activation regulates damage-associated molecular pattern isoforms released during pyroptosis. *EMBO J*. 2013;32:86–99. [PMC free article] [PubMed].
- [5] Hollmann MW, Durieux ME. Local anesthetics and the inflammatory response: a new therapeutic indication? *Anesthesiology* 2000;93:858–75.
- [6] Fitzgerald KA, Palsson-McDermott EM, Bowie AG, Jefferies CA, Mansell AS, Brady G, et al. Mal (MyD88 adapter-like) is required for Toll-like receptor 4 signal transduction. *Nature* 2001;413:78–83.
- [7] Kenny EF, Talbot S, Gong M, Golenbock DT, Bryant CE, O'Neill LA. MyD88 adapter-like is not essential for TLR2 signaling and inhibits signaling by TLR3. *J Immunol* 2009;183:3642–51.
- [8] Nyman T, Stenmark P, Flodin S, Johanson I, Hammarström M, Nordlund P. The crystal structure of the human Toll-like receptor 10 cytoplasmic domain reveals a putative signaling dimer. *J Biol Chem* 2008;283:11861–5.
- [9] Kagan JC, Su T, Horng T, Chow A, Akira S, Medzhitov R. TRAM couples endocytosis of Toll-like receptor 4 to the induction of interferon-beta. *Nat Immunol* 2008;9:361–8.
- [10] Verstak B, Nagpal K, Bottomley SP, Golenbock DT, Hertzog PJ, Mansell A. MyD88 adapter-like (Mal)-TIRAP interaction with TRAF6 is critical for TLR2- and TLR4-mediated NF- κ B proinflammatory responses. *J Biol Chem* 2009;284:24192–203.
- [11] Li CY, Tsai CS, Hsu PC, Chueh SH, Wong CS, Ho ST. Lidocaine attenuates monocytes chemoattractant protein-1 production and chemotaxis in human monocytes: possible mechanism for its effect on inflammation. *Anesth Analg* 2003;97:1312–6.
- [12] Botos I, Segal DM, Davies DR. The structural biology of toll-like receptors. *Structure* 2011;19:447–59.
- [13] Núñez Miguel R, Wong J, Westoll JF, Brooks HJ, O'Neill LA, Gay NJ, et al. A dimer of the Toll-like receptor 4 cytoplasmic domain provides a specific scaffold for the recruitment of signaling adapter proteins. *PLoS ONE* 2007;2:e788.
- [14] Jin MS, Lee JO. Structures of the toll-like receptor family and its ligand complexes. *Immunity* 2008;29:182–91.
- [15] Wang H, Ma S. The cytokine storm and factors determining the segments and severity of organ dysfunction in multiple organ dysfunction syndrome. *Am J Emerg Med* 2008;26:711–5.
- [16] Caracas HC, Maciel JV, Martins PM, de Souza MM, Maia LC. The use of lidocaine as an anti-inflammatory substance: a systematic review. *J Dent* 2009;37:937.
- [17] Gallos G, Jones DR, Nasr SH, Emala CW, Lee HT. Local anesthetics reduce mortality and protect against renal and hepatic dysfunction in murine septic peritonitis. *Anesthesiology* 2004;101:902–11.
- [18] Sirait RH, Hatta M, Ramli M, Simanjuntak TP, Suprayogi P, Islam AA, et al. The analysis of the effective systemic lidocaine dosage on the expression of HMGB1 mRNA in mice with sterile musculoskeletal injury. *Open J Anesthesiol* 2017;7:35–41.
- [19] Takao Y, Mikawa K, Nishina K, Maekawa N, Obara H. Lidocaine attenuates hyperoxic lung injury in rabbits. *Acta Anesthesiol Scand* 1996;40:318–25.
- [20] Faure E, Equils O, Sieling PA, Thomas L, Zhang FX, Kirschning CJ, et al. Bacterial lipopolysaccharide activates NF- κ B through toll-like receptors 4 (TLR-4) in cultured human dermal endothelial cells. Differential expressions of TLR-4 and TLR-2 in endothelial cells. *J Biol Chem* 2000;275:11058–63.
- [21] Wang HL, Ying YQ, Yu YX, Rong F, Lei WF, Zhang WH. The protective effect of lidocaine on septic rats via the inhibition of high mobility group box 1 expression and NF- κ B activation. *Mediators Inflamm* 2013;570:370.
- [22] Toshchakov VY, Szmajdzinski H, Couture LA, Lakowicz JR, Vogel SN. Targeting TLR4 signaling by TLR4 Toll/IL-1 receptor domain-derived decoy peptides: identification of the TLR4 Toll/IL-1 receptor domain dimerization interface. *J Immunol* 2011;186:4819–27.
- [25] Muzio M, Bosisio D, Polenttaruti N, D'amico G, Stoppacciaro A, Mancinelli R, et al. Differential expression and regulation of toll-like receptors (TLR) in human leukocytes: selective expressions of TLR3 in dendritic cells. *J Immunol* 2000;164:5998–6004.
- [26] O'Neill LAJ, Bowie AG. The family of five. TIR-domain-containing adaptor in toll-like receptors signaling. *Nat Rev Immunol* 2007;7:353–64.
- [27] Johnson GB, Brunn GJ, Platt JL. Cutting edge: an endogenous pathway to systemic inflammatory response syndrome (SIRS)-like reactions through Toll-like receptor 4. *J Immunol* 2004;172:20–4.

Further reading

- [23] Dewar DC, Balogh ZJ. The epidemiology of multiple organ failure: a definition controversy. *Acta Anaesthesiol Scand* 2011;55:248–9.
- [24] Kawai T, Akira S. The role of pattern-recognition receptors in innate immunity: update in toll-like receptors. *Nat Immunol* 2010;11:373–84.