



1,2,4-Triazolo[1,5-a]pyrimidines as significant scaffold in drug discovery

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ABSTRACT

Over the years, 1,2,4-Triazolo[1,5-a]pyrimidine (TP) emerged as an important scaffold in medicinal chemistry due to its vast application in designing novel anti-cancer, antimicrobial, antiviral, antiparasitic, and CNS-modulating agents. Recently, publications of TP analogues with diverse biological activities have increased markedly. In this review, we focus on the most important applications of TP scaffold in medicinal chemistry

1. Introduction

The triazolopyrimidine derivatives are hetero-bicyclic organic compounds that gained considerable attention recently owing to their vast pharmacological activities. Four possible isomers of 1,2,4-triazole-fused pyrimidines may exist (Fig.1).[1–4] Among them, 1,2,4-triazolo[1,5-a]pyrimidine (TP) has gained the most attention due to its variety of applications in agriculture and medicinal chemistry. Some natural TPs have been identified such as essramycin 1(antibacterial drug) isolated from the broth of.

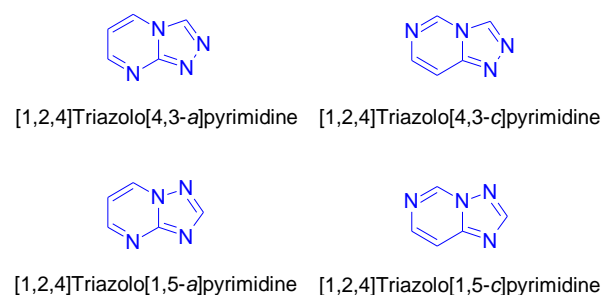


Fig. 1. Four representative isomers of 1,2,4-triazolopyrimidines

the marine species *Streptomyces*[5], in addition to some volatile oils **2-5** (Fig.2) isolated from *Polygonatum odoratum*, *Poncirus trifoliata*, *Paris polyphylla*, *Anchusa azurea*, respectively. However, the majority of 1,2,4-triazolo[1,5-*a*]pyrimidine derivatives were synthesized compounds. Interestingly, the first synthetic 1,2,4-triazolo[1,5-*a*]pyrimidine (TP) scaffold was reported by *Bulow & Hass* in 1909. [6]

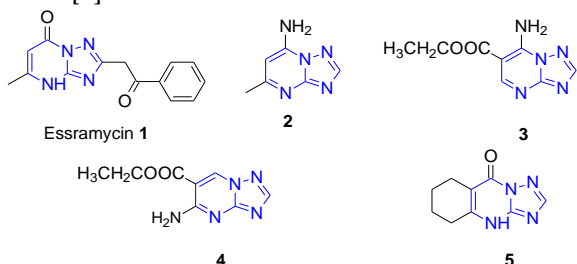


Fig. 2. Structures of TP derivatives 1-5.

2. Biological activities of 1,2,4-triazolo[1,5-*a*]pyrimidine derivatives

2.1. Anti-cancer activity

Cancer is the second leading cause of death in the world according to the World Health Organization (WHO), and till now, there is no complete cure effective against disseminated cancer. The TP scaffold has emerged as a prominent scaffold in developing promising target-based anti-cancer agents in the past few decades. These oncotargets include six categories: DNA damage repair system, signal transduction pathway, ubiquitin-proteasome pathway (UPP), epigenetic modification, tumor microenvironment (TME), and multidrug resistance (MDR).

2.1.1. DNA damage repair system target

DNA damage response (DDR) is a sophisticated process that cells develop to repair different DNA damage and preserve genome structure.[7,8] Although most damages undergo DNA repair, such repair isn't 100% efficient, and the accumulation of driver DNA mutations plays an important role in cancer cell growth.[9] Hence, targeting DDR became a promising approach to developing anti-cancer agents.[10]

2.1.1.1. Tubulin inhibitors

Heterodimers of globular tubulins are the main building blocks of the cylindrical skeleton of microtubules essential for many vital cellular functions[11]. Mutations and overexpression of tubulins leading to mitotic spindle formation and mitotic division, which disrupts this process, therefore tubulin inhibition is a major approach in developing chemotherapeutic agents [12–14].

A study made by *Zhang et al.*[15] led to the development of cevipabulin (TTI-237) **6** as a potent microtubular inhibitor that can inhibit the growth of several cancer cell lines. TTI-237 (Fig.3) acts by competing with a vinblastine binding site, not paclitaxel. However, TTI-237 inhibited depolymerization of microtubules like paclitaxel, proving that TTI-237 displayed mixed properties between vinblastine and paclitaxel. Unfortunately, the phase I clinical trial of TTI-237 was ended due to its toxicity.[16]

Additionally, in 2019, *Yang et al.*[17] reported a series of TP derivatives as restricted combretastatin-4 (CA-4) analogs showing marked anti-tubulin activity. Among them, compound **7** (Fig.3.) showed the most promising antitumor activity against the three tested cell lines, A549, HeLa, and HCT116, with IC₅₀ values of 1.02 μM, 0.75 μM, and 10.91 μM, respectively, compared to CA-4 with IC₅₀ values of 0.013 μM, 0.21 μM, and 6.10 μM, respectively. Moreover, compound **8** (Fig.3.) showed the highest tubulin inhibiting activity (IC₅₀ = 9.9 μM) compared to CA-4 (IC₅₀ = 4.22 μM). Furthermore, molecular modeling showed that the colchicine binding site was occupied by the TP derivative. Further structural modification of compound **7** by removing acetyl- and methyl- groups to obtain a series of TP analogs, among them compound **9** (Fig.3.) showed a 166-fold improvement in activity against the Hela cancer cell line.[18] Moreover, in 2022, a series of TP derivatives was reported by *Mohamed et al.*[19] as combretastatin CA-4 analogs. Notably, compound **10** (Fig.3.) showed a potent anti-tubulin activity (IC₅₀ = 3.84 μM) compared to CA-4 (IC₅₀ = 1.10 μM). Also, the mechanistic studies proved that compound **10** inhibited HCT-116 cells proliferation through inducing apoptosis and halting the cell cycle at the G2/M phase.

In the same year, *Chen et al.* [20] made a structural modification of compound **10** by removing the 5-aryl substitution and replacing the 2-free amino group with a substituted one, leading to the development of a series of TP derivatives. Interestingly, compound **11** (Fig.3) showed a five-fold improvement compared with compound **10** against the same three tested cell lines (A549, HeLa, and HCT116 cells). Moreover, compound **11** displayed potent tubulin polymerization inhibitor activity (IC₅₀ = 4.9 μM) compared to CA-4 (IC₅₀ = 4.20 μM).

At the same time, two studies of various 2,7-disubstituted-TP derivatives were conducted [21,22]. Among them, compounds **12** and **13** (Fig.3) were the most active and showed a marked tubulin-inhibiting activity with IC₅₀ values of 0.39 μM and 0.45 μM, respectively, compared to CA-4 (IC₅₀ = 0.75 μM). Moreover, compound **12** displayed potent antiproliferative activity against HT-29, MDA-MB-231, HeLa, A549, and Jurkat cancer cell lines. Interestingly,

compound **13** displayed potent antiproliferative activity against A549 and HeLa cancer cell lines with IC_{50} of 0.043 μ M and 0.038 μ M, respectively, compared to CA-4 (IC_{50} of 0.180 μ M and 0.004 μ M, respectively).

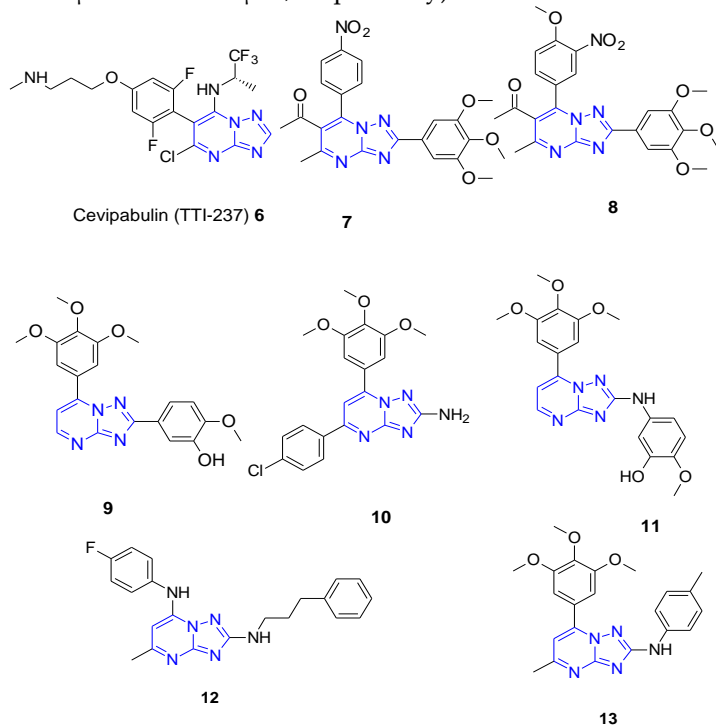


Fig.3. Structures of TP derivatives **6-13** having tubulin polymerization inhibitory activity.

2.1.1.2. Cyclin dependent kinase inhibitors (CDK inhibitors)

Cyclin-dependent kinases are a group of enzymes responsible for cell cycle activation, including metabolism, differentiation, cell division, and transcription through activating G1/M and G2/S transitions, making them a valuable target for developing anti-cancer agents.[23–26] Over the years, researchers have investigated TP derivatives as purine surrogates. For example, seliciclib (Roscovitine, CYC202)[27] **14** (**Fig.4**) is a purine derivative that inhibits multiple CDK enzymes (CDK-1, CDK-2, CDK-5 and CDK-7) through competition at ATP binding catalytic site with ATP. Now, Seliciclib **14** is in phase II clinical trial for treating Cushing disease and reverse hypercortisolism.[28] A medium throughput screening campaign of more than 3000 compounds led to the development of pyrazolopyrimidine derivative **15** as a potent CDK-2 inhibitor with IC_{50} = 1.8 μ M. Replacing C3 with nitrogen of compound **15** led to the developing of a TP derivative **16** with lower inhibitory activity against CDK-2. Optimization of SAR of compound **16** led to the development of TP derivative **17** with more potent activity against CDK-2 (IC_{50} = 1.2 μ M) and improved selectivity against GSK3- β (**Fig.4**).[29]

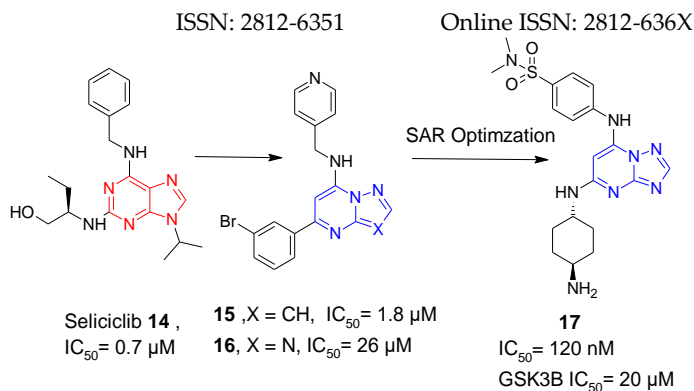


Fig.4. Structures of compounds **14-17** acting as potent CDK inhibitors.

2.1.1.3. Tyrosyl DNA Phosphodiesterase 2 (TDP2) Inhibitors

Topoisomerase II-mediated DNA damage is repaired by TDP2, making cancer cells more resistant to TOPII poisons (doxorubicin and etoposide).[30–32] Thus, TDP2 inhibitors increase cancer cells sensitivity towards TOPII poisons.[33] HTS of more than 1600 molecules led to identifying hit compound (P10A10) **18** as a promising candidate for TDP2 inhibition. Hit validation through the resynthesis of compound **19** led to unexpected results showing that this compound is utterly inactive at 100 μ M.[34] So, further structural optimization of compound **18** by shifting phenyl substitution from C-7 to C-5 led to the development of compound **19** with promising TDP2 inhibiting activity (IC_{50} = 22 μ M) (**Fig.5**).[35]

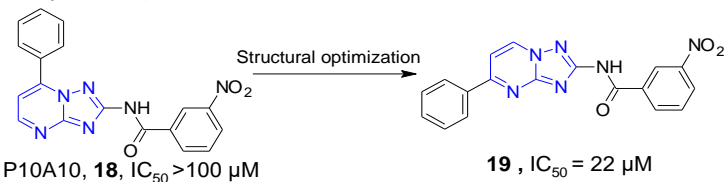
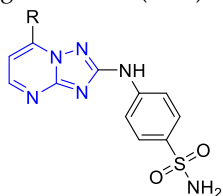


Fig.5. Structures of TP derivatives **18** and **19** acting as TDP2 inhibitors.

2.1.1.4. Carbonic Anhydrase inhibitors (CA inhibitors)

There are seven known CA families in nature, but only the alpha family is expressed in humans (hCAs).[36] Human alpha CA is a group of fifteen enzymes (CA I-CA XV) involved in different physiological processes.[37–40] Notably, CA IX and CA XII are overexpressed in various cancer cell lines but limited in normal tissue, making them valuable targets for developing anti-cancer agents.[36] Different CA inhibitors have been reported. Among them, sulfonamide derivatives were one of the most active classes.[40–42] Giampietro Viola et al.[43] identified a class of triazolopyrimidine-sulfanilamide hybrids as potent CA IX and CA XII inhibitors with K_i in a nanomolar range. Among them, TP derivatives **20** and **21** were the most active compared to acetazolamide (**Fig.6**).



20, R = 3-Pyridinyl, $K_i = 5.1$ nM (CA IX), $K_i = 8.8$ nM (CA XII)
21, R = 2,3,6-trifluorophenyl, $K_i = 8.6$ nM (CA IX), $K_i = 5.4$ nM (CA XII)
Acetazolamide, $K_i = 25$ nm (CA IX), $K_i = 5.7$ nM (CA XII)

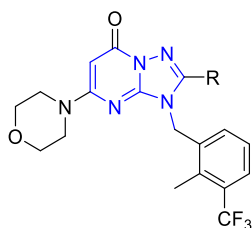
Fig.6. Structures of TP derivatives **20** and **21** acting as potent CA inhibitors.

2.1.2. Signal transduction pathway target

Modifying signaling transduction pathways that control cell growth, motility, division, and death is essential in cancer progression. Multiple inhibitors of signaling transduction pathways, phosphatidylinositol 3-kinases (PI3Ks), paired-box gene 2 (Pax2), and extracellular signal-regulated kinase 3 (ERK3) have been developed as effective anti-cancer treatment recently.

2.1.2.1. PI3K inhibitors

PI3Ks are a group of enzymes involved in inositol phosphorylation, cell metabolism, cell proliferation, and cell survival. Dysregulation of PI3K actions plays an active role in converting normal cells into cancerous ones and inactivating tumor suppressor proteins PTEN. Sanchez *et al.*[44] identified a series of triazolo[1,5-a]pyrimidinone derivatives as potent PI3K inhibitors with enhanced β isoform selectivity. Compounds **22a** and **22b** were the most active, with IC_{50} of 0.4 nM and 0.6 nM, respectively (**Fig.7**).



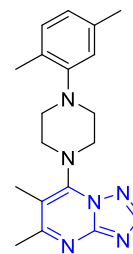
22a, R = -SCH₃, $IC_{50} = 0.4$ nM
22b, R = CH₃, $IC_{50} = 0.6$ nM

Fig.7. Structures of TP derivatives **22a** and **22b** acting as potent PI3K inhibitors.

2.1.2.2. Pax2 inhibitors

Pax proteins are a family of DNA-binding proteins essential for the growth of different organs, including the kidney, pancreas, ear, and CNS. Notably, Pax2 is expressed in kidney-developing nephrons but not in adult proximal or distal tubes. On the other hand, it is highly expressed in polycystic kidney epithelia and renal carcinoma. Based on the HTS campaign of more than 69000 compounds performed by Bradford and her colleagues to identify potent

Pax2 inhibitors, TP derivative (BG-1) **23** (**Fig.8**), was the most active, exhibiting a potent inhibition of Pax2-positive cancer cells with little effect on Pax2-negative ones.[45]

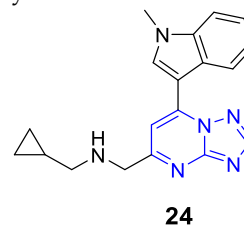


23, $IC_{50} = 1.5$ μ M

Fig.8. Structure of TP derivative **23** acting as potent Pax2 inhibitor.

2.1.2.3. ERK inhibitors

ERK signaling pathways play a significant role in cell growth regulation and are highly expressed in different types of cancers, making them promising therapeutic targets. Zhang *et al.* reported a novel TP series with potent activity against gastric cancer cells MGC-803. Compound **24** (**Fig.9**) was the most active with $IC_{50} = 13.1$ μ M and displayed potent inhibitory activities on the ERK signaling pathway, resulting in decreased ERK1/2, c-Raf, MEK1/2, and AKT phosphorylation levels.



24

Fig.9. Structure of TP derivative **24** acting as ERK inhibitor.

2.1.3. Ubiquitin-proteasome inhibitors

Hershko, Ciechanover, and Rose won the Nobel Prize in chemistry[46] for the identification of the ubiquitin-proteasome system (UPS). UPS is responsible for the ubiquitination, degradation, and turnover of various proteins necessary for various cellular functions, including cell division, migration, and death, through a triple cascade mechanism.[47] Since the UPS is important for different cell functions and disruption in the cell cycle can lead to oncogenesis, it became an attractive target for researchers to develop new antineoplastic agents.[48]

2.1.3.1. Targeting DCN1-UBC12 interaction

Ubiquitin-protein ligases E3 can be divided into U-box, HECT (homologous to E6-AP carboxy terminus), and RING (interesting new gene). Notably, the RING family contains two subclasses: cullin-containing RING-finger ligases

(CRLs) and other RINGs. Cullin-RING ligases (CRLs) account for nearly 20% of cellular protein degradation through the ubiquitin–proteasome system, and their dysfunction has been observed in many diseases, including cancer. There are five defective cullin neddylation (DCN) isoforms (DCN1–5) in the human genome. Among them, DCN1 is elevated in different types of cancers, including cervical cancer, prostate cancer, colorectal cancer, and laryngeal squamous cell carcinoma.[48] Wang *et al.* reported TP derivative **WS-291, 25** through HTRF as a hit compound for inhibition of DCN1-UBC12 interaction with an IC_{50} of 5.82 μ M. Structural optimization of **WS-291** led to the identification of a more potent inhibitor **WS-383, 26** with an IC_{50} of 0.011 μ M (**Fig.10**).[49]

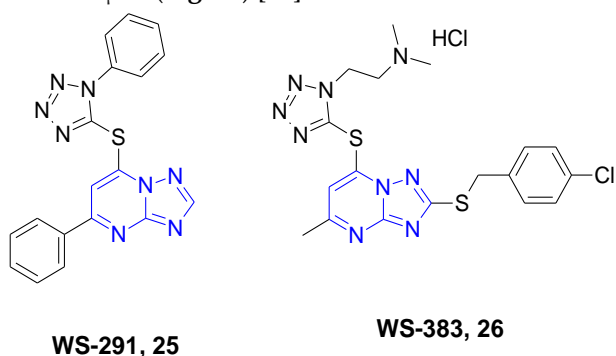


Fig.10. Structures of TP derivatives **25** and **26** targeting DCN1-UBC12 interaction.

2.1.3.2. Targeting RBX1-UBE2M interaction

In 2024, Ma *et al.*[50] identified TP derivative **WS-299, 27** (**Fig.11**) as a potent antiproliferative agent MGC-803 and HGC-27 cell lines with IC_{50} values of 1.0 μ M and 1.8 μ M, respectively. Compound **27** exerted its anti-cancer effect by targeting RBX1-UBE2M interaction and inhibiting the CUL3 and CUL5 neddylation modification.

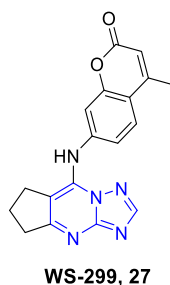


Fig.11. Structure of TP derivatives **27** targeting RBX1-UBE2M interaction.

2.1.4. Epigenetic modifier inhibitors

Epigenetic modification starts with modifying DNA and histone proteins that regulate accessibility and functions, remodeling the basic chromatin unit (nucleosome).[51] These epigenetic modifications control gene expression

and have proved crucial in maintaining the malignant process, making them important targets for cancer treatment. Many epigenetic regulators are approved by the FDA, including DNA methyltransferase, isocitrate dehydrogenase 1 or 2, histone methyltransferases, and histone deacetylases or under investigation, such as bromodomain proteins and histone demethylases.

2.1.4.1. LSD1 inhibitors

Lysine-specific demethylase 1 (LSD1) is an enzyme that removes the methyl group from mono-/di-methylated histone 3, lysine 4 (H3K4me1/2), and other methylated nonhistone proteins. Overexpression of LSD1 is noted in various cancers, making it an attractive target. Nine small molecules targeting LSD1 are under clinical investigation, but none of them approved for clinical usage yet owing to severe side effects. Wang *and* his colleagues reported two series of TP derivatives as potent LSD1/KDM1A inhibitors and evaluated their cytotoxic activity against MGC-803, EC109, A549, and PC-9 cell lines. Among the two series, compounds **28-31** (**Fig.12**) were the most potent LSD1 inhibitor with IC_{50} values of 0.882 μ M, 0.154 μ M, 1.19 μ M, and 0.557 μ M, respectively. Interestingly, compounds **28** and **29** enhanced the accumulation of H3K4me1/me2 and H3K9me2 (LSD1 substrates) and inhibited A549 cell migration in a concentration-dependent manner. Also, compounds **28** and **29** increased E-cadherin and claudin-1 expression (two epithelial indicators) and decreased expression of N-cadherin (mesenchymal marker), snail, and slug (transcription markers). On the other hand, compound **32** exhibited potent PC-9 growth inhibition with $IC_{50} = 0.59$ μ M, about 4-fold compared to 5FU, despite having less potent LSD1 inhibitory activity ($IC_{50} > 10$ μ M).[52–55]

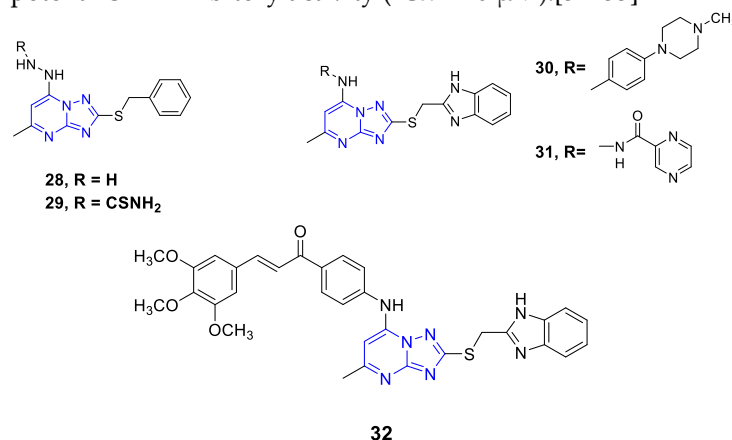


Fig.12. Structures of TP derivatives **28-32** acting as LSD1 inhibitors.

2.1.4.2. BRD-4 inhibitor

Bromodomain containing protein-4 (BRD-4) is an epigenetic reader that is essential for gene expression during the G1 phase. BRD proteins have emerged to play a significant role in the occurrence of different diseases, including malignant tumors, making them attractive targets for researchers to discover new anti-cancer candidates. More than 20 BRD-4 inhibitors have been under clinical trials to treat different types of cancers.[56] Interestingly, TP scaffolds have emerged as possible bio-isostere to the N-acetyl fragment of ϵ -N-acetylated lysine. TP derivatives **33-35** (Fig.13.) have been reported as KAc mimetics and bind within the BRD-4 binding site, making them good inhibitors with IC_{50} of 24 μ M, 5 μ M, and 2.15 μ M, respectively, compared to JQ1 as a positive control (IC_{50} = 0.13 μ M). Compound **35** (WS-722) also induced cell apoptosis and enhanced caspase-3/7 and PARP cleavage of THP-1 cells (IC_{50} = 3.86 μ M).[57]

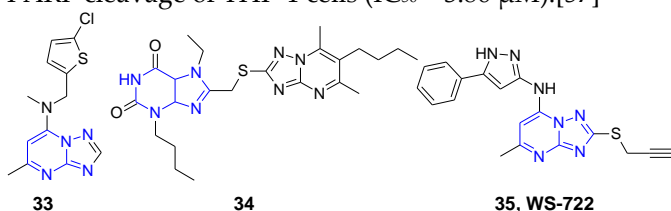


Fig.13. Structures of TP derivatives **33-35** acting as BRD-4 inhibitors.

2.1.5. Targeting tumor microenvironment

Tumor microenvironment (TME) has been reported as a crucial controller of cancer growth, metastasis, and multidrug resistance. TME includes tumor vasculature, immune cells, stromal cells, and extracellular matrix. Methionine aminopeptidase-2 (MetAP-2) is a key enzyme believed to play a crucial role in tumor angiogenesis and, therefore, tumor cell multiplication and spread. Screening campaign for MetAP-2 inhibitors led to identifying purine analogue **36** as a potent inhibitor of MetAP-2 with IC_{50} equal to 0.23 μ M. Further optimization of compound **36** by replacing the purine ring with TP scaffold led to the development of TP derivative, **37** (Fig.14.) which showed more potent inhibition activity against MetAP-2 (IC_{50} = 0.038 μ M) and good physicochemical profile.[58]

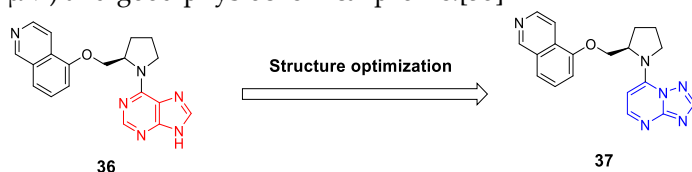


Fig.14. Structures of compounds **36** and **37** targeting tumor microenvironment.

2.1.6. Multidrug resistance (MDR)

A common challenging complication developed in most patients taking chemotherapeutic agents is multidrug

resistance (MDR). MDR occurs for different reasons, of which ATP-binding cassette (ABC) transporters that transport drugs outside tumor cells are believed to be the most prominent factor. TP derivatives have emerged as a promising candidate to reverse MDR by inhibiting ATP-binding cassette subfamily B member 1 (ABCB1), which enhances drug efflux outside cancer cells and reduces drug intake. *Liu et al.* reported different TP series acting as potent ABCB1 inhibitors; among them, compounds **38** and **39** (Fig.15.) were the most active with IC_{50} values equal to 22 nM and 3.67 nM, respectively.[59,60]

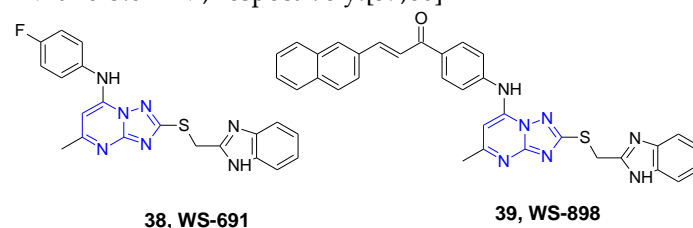


Fig.15. Structures of TP derivatives **38** and **39** acting as ABCB1 inhibitors.

2.2. Central nervous system

In recent years, triazolopyrimidine has emerged as a prominent scaffold in developing candidates for treating different CNS diseases. TP analogs exert their action through one of the following:

- Inhibition of Acetylcholine Esterase
- Stabilizing Microtubules
- Inhibition of PDE2A

2.2.1. Acetylcholine esterase inhibitors

Three studies conducted by *Jayaram et al.* led to the development of TP hybrids as a promising inhibitor of acetylcholine esterase for the treatment of Alzheimer's disease. These hybrids exert their action through dual targeting of the catalytic active site and the peripheral anionic site through π - π interaction with important residues (Tyr70 and Trp279), which play an important role in the regulation of the entry of substrate to the binding site of AChE. Among the synthesized three series, compounds **40-43** (Fig.16.) showed the most potent inhibiting activity of AChE with IC_{50} = 0.042 μ M, 0.065 μ M, 0.092 μ M, and 0.160 μ M, respectively, compared to reference compound tacrine (IC_{50} = 0.13 μ M). Also, *in silico* ADMET profiling showed that these derivatives possess drug-like properties as well as low toxicity. Based on these results, SAR revealed that the presence of the triazolopyrimidine or other isosteric groups is important for π - π interaction with the PAS residue of AChE, a linker (Piperazine) interacts with the aromatic gorge of AChE and a heterocyclic moiety intercalates

between sheets of amyloid β (quinoline or triazine or pyrimidine) are essential for inhibiting activity.[61–63]

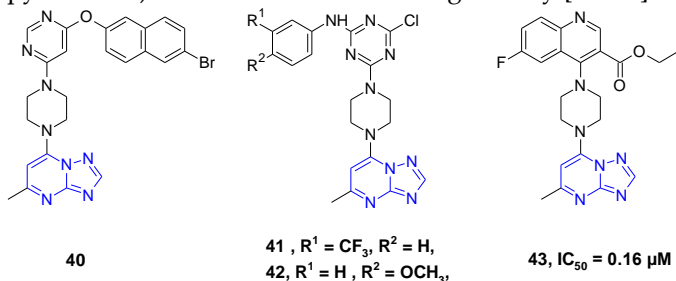


Fig.16. Structures of TP derivatives 40-43 acting as AChE inhibitors.

2.2.2. Microtubule stabilizing agents

Microtubule active agents have versatile applications in developing promising candidates for treating different diseases, including neuropathies and cancer. *Carlo Ballatore and his colleagues* conducted several studies to understand how MT active TPs either enhance the stabilization of MT or disrupt MT integrity. These studies revealed that TP analogs with an alkoxy side chain at position number 4 of the fluorinated phenyl ring destabilize MT assembly, whereas other TP analogs lacking the alkoxy side chain stabilize MT (compounds 44-46, Fig.17.). Alzheimer's disease and other neurodegenerative diseases are characterized by the presence of insoluble inclusions of tau protein, which is associated with axonal MT in the brain, leading to loss of axonal density and axonal dystrophy. Based on these facts, they examined MT stabilizing TP analog 44 (CNDR-51657) (Fig.17.) with a low dose (3-10 mg/kg) in the mouse tauopathy model, which is a low dose (3-10 mg/kg) for 3 months enhanced MT stabilization, decreased tau pathology, and reduced axonal defects.[64–67]

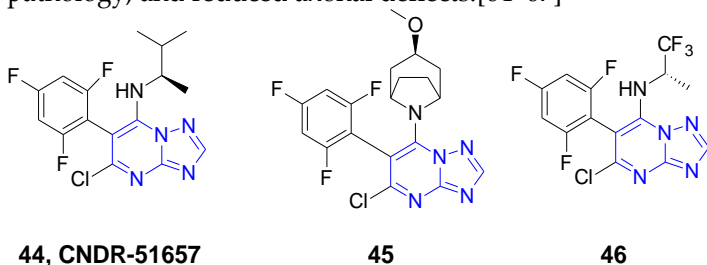


Fig.17. Structures of TP derivatives 44-46 acting as microtubule stabilizing agents.

2.2.3. Inhibition of PDEA2

Phosphodiesterase (PDE) enzymes are catabolic enzymes encoded by 21 genes and are classified into twelve families (PDE1-PDE12). PDEs are responsible for the hydrolysis of cAMP and cGMP to the corresponding inactive AMP and GMP, respectively. PDE2A is expressed in the brain,

especially in hippocampus, striatum and cortex. Thus, its inhibition led to an increase in intracellular cAMP and cGMP, which play a key role in neuroplasticity and correlate to learning and memory.[68] An important patent application of 566 (1,2,4)-triazolo(1,5-a)pyrimidine analogs was exemplified by Dart Neuroscience. Among them, 192 compounds were reported to have pIC_{50} less than 7 for PDE2A, but selectivity towards other PDEs wasn't disclosed in this patent (Compounds 47-49 (Fig.18) are representative examples of the most active TP derivatives). Further structural optimization led to the identification of the lead compound 50, DNS-8254, as a highly selective PDE2A inhibitor with IC_{50} of 8 nM and showed significant memory-enhancing effects and favorable pharmacokinetic properties *in vivo*.[69] In 2020, *Tresaden et al.* reported a hit to lead exploration of 1,2,4-triazolo(1,5-a)pyrimidine derivative 51 (Fig.18.) through HTS campaign (HTS) as a potent inhibitor of PDE2A enzyme ($IC_{50} = 1.3 nM$) and 100 fold selective over the other PDE enzymes.[70]

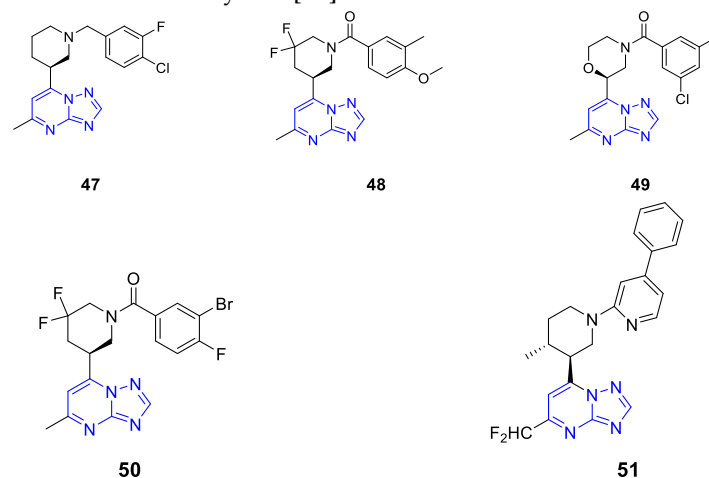


Fig.18. Structures of TP derivatives 47-51 acting as PDE2A inhibitors.

2.3. Anti-infectious agents

TP scaffold represents a good pharmacophore for developing different candidates that have been acting as potent anti-infectious diseases over recent years. Different TP derivatives have been reported as antibacterial, antiparasitic, and antiviral agents.

2.3.1. Anti-bacterial agents

Despite essramycin 1[71] was the first TP derivative as an antibacterial agent, recent studies revealed that essramycin believed to be devoid of any antibacterial activities.[72,73] In literature, many TP derivatives have been reported as potent antibacterial agents with a broad spectrum against Gram-positive and Gram-negative bacteria.[74–80] Important results can be summarized that the most active

ring with a 1,2,4-triazole ring led to the developing of more potent compounds with improved *in vitro* and *in vivo* activities. Among other studies made by the same group of researchers, TP derivatives **60-64** (Fig.22.) showed the highest inhibitory activities of PA-PB1 with $IC_{50} = 1.1- 26 \mu M$, and a good safety profile.[81-84]

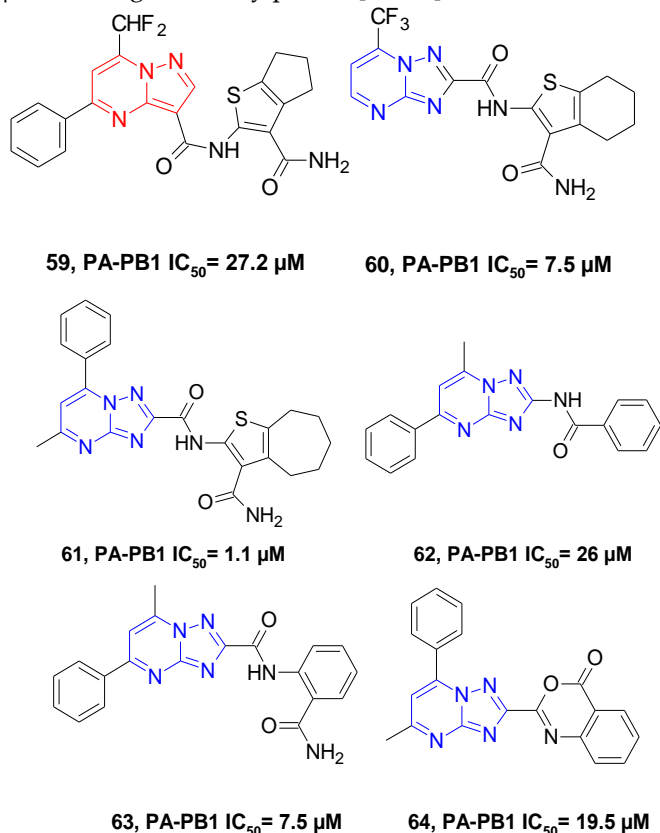


Fig.22. Structures of TP derivatives **59-64** acting as PA-PB1 inhibitors.

2.3.2.2.2. Anti-HCV activity

HCV is one of the main causes of chronic liver diseases and searching for safe and effective drug is significant. HCV nonstructural protein 5B (NS5B) is an RNA-dependent RNA polymerase that has the main role of viral replication. It differs from human DNA and RNA polymerases, making it an important therapeutic target for developing anti-HCV candidates. HTS campaign made by Pfizer for structural optimization of allosteric HCV NS5B dihydropyrone inhibitors led to the development of TP derivatives with marked potent HCV NS5B polymerase inhibition activity ($IC_{50} = 0.007 \mu M - 0.016 \mu M$). Among them, TP derivative **65** (Fig.23.) showed potent NS5B inhibition activity with $IC_{50} = 0.016 \mu M$, which then separated into its two enantiomers **65a**, **Filibuvir**, and **65b** with $IC_{50} = 0.007 \mu M$ and $0.012 \mu M$ respectively.[85] The R enantiomer **65a**, **Filibuvir**, reached phase II clinical trials for HCV treatment, but the investigation was discontinued in 2013 for strategic reasons.

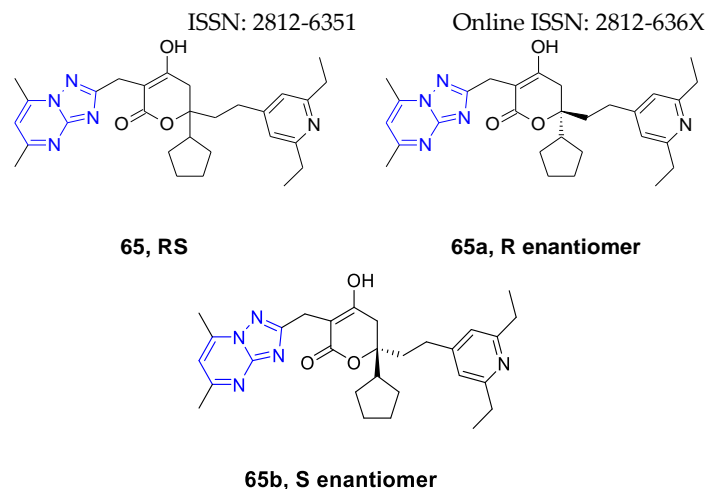


Fig.23. Structure of anti-HCV TP derivative **65** and its enantiomers **65a** and **65b**.

2.3.2.3. Anti-HIV activity

HIV-1 reverse transcriptase is the key enzyme involved in HIV replication, making it one of the crucial targets for the treatment of HIV/ AIDS. Three studies conducted by *Xin Yong Liu et al.* to identify non-nucleoside reverse transcriptase (NNRTI) led to the development of a series acting as a potent inhibitor of HIV.[86-88] The first study identified a series of pyrazolopyrimidine derivatives acting as potent anti-HIV-1 with EC_{50} from $0.07 \mu M$ to $5.98 \mu M$. Pyrazolopyrimidine derivative **66** (Fig.24.) was the most active with $EC_{50} = 0.07 \mu M$. [86]

In the other two studies, structure-guided core refining approaches and molecular docking studies led to the development of a series of TP derivatives acting as potent wild-type HIV-1 III_B inhibitors. TP derivatives **67** and **68** (Fig.24.) were the most active against wild-type HIV-1 III_B with $EC_{50} = 0.0200 mM$ and $0.0081 mM$, respectively, and showed weak to moderate activity against K103N/Y181C double resistant mutant strain of HIV-1 with $EC_{50} = 7.6 mM$ and $13 mM$ respectively. These compounds are believed to act by interacting with the diaryl pyrimidine allosteric site located 10 \AA away from the reverse transcriptase catalytic site.[87,88]

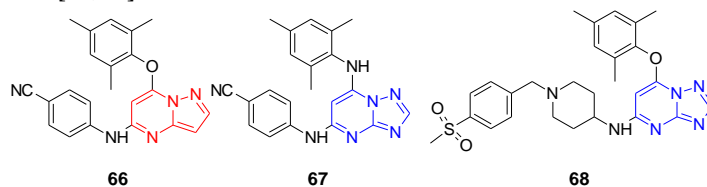


Fig.24. Structures of compounds **66-68** acting as anti-HIV.

2.3.3. Anti-parasitic agents

Recently, TP derivatives have been reported as a promising candidate for treating different infections caused by various parasites, mainly kinetoplastid and plasmodium parasites.

2.3.3.1. Activity against Kinetoplastid parasites

Many infections such as leishmaniasis, Chagas disease, and African sleeping sickness, are caused by *L. donovani*, *T. cruzi*, and *T. brucei* kinetoplastid parasites, respectively. Screening of more than 3 million compounds led to the development of a hit azabenzoxazole derivative **69** (GNF5343) (Fig.25.). Structural optimization of compound **69** by replacing azabenzoxazole with TP scaffold led to the development of a series of TP derivatives with broad spectrum activity against different kinetoplastid parasites with more favorable ADME-PK properties. TP derivatives **70-72** (Fig.25.) were the most active with $EC_{50} = 350 \mu\text{M}$, $71 \mu\text{M}$, and $15 \mu\text{M}$ against *L. donovani*, $EC_{50} = 55 \mu\text{M}$, $16 \mu\text{M}$, and $120 \mu\text{M}$ against *T. cruzi*, and $79 \mu\text{M}$, $22 \mu\text{M}$, and $70 \mu\text{M}$ against *T. brucei*, respectively.[89]

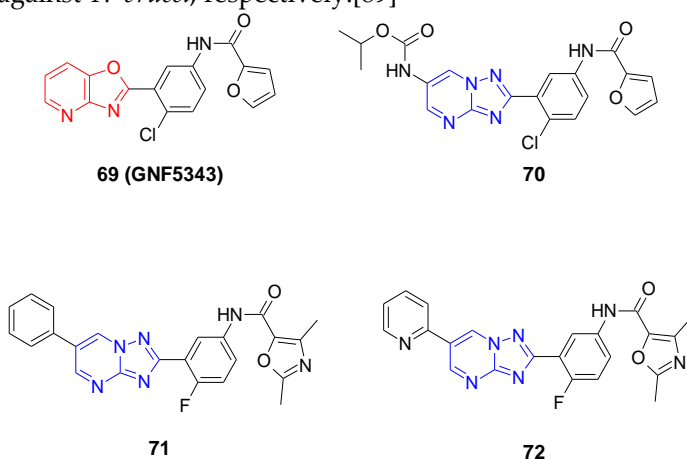


Fig.25. Structures of compounds **69-72** acting against kinetoplastid parasites.

2.3.3.2. Antimalarial agents

Malaria is one of the most common causes of death around the world, especially in tropical and subtropical regions (> 90% of cases were found in Africa). Malaria is a mosquito-borne infection caused by one of four types of Plasmodium species: *P. falciparum*, *P. vivax*, *P. ovale*, and *P. malariae*, resulting in severe illness and even death if not treated well within one day of infection. Malaria is spread by the bite of female *Anopheles* mosquitoes. Plasmodium dihydroorotate dehydrogenase (*pDHODH*) is one of the key enzymes involved in *de novo* pyrimidine biosynthesis and has been validated clinically as a promising drug target for developing novel antimalarial agents. In this context, various *pDHODH* inhibitors based on different scaffolds have been reported in the last two decades and are known as **DSM** compounds. Among them, TP derivatives **73** (**DSM1**) and **74** (**DSM2**) were developed from the HTS campaign as potent *pDHODH* inhibitors with IC_{50} values of $0.047 \mu\text{M}$ and $0.056 \mu\text{M}$, respectively.[90] X-ray studies of

these compounds showed that the TP ring binds to a polar region (H-bond pocket) while the hydrophobic pocket is occupied by the naphthyl or anthranyl substituent. Structure-guided medicinal chemistry approaches and HTS efforts led to the development of different analogs of **DSM1** with more potent *pDHODH*-inhibiting activities and more favorable ADME properties. As examples, TP derivatives **75** (**DSM267**)[91], **76** (**DSM338**)[92], **77** (**DSM265**)[93], **78** (**DSM421**)[94], and **79** (**DSM422**)[95] with IC_{50} values equal to $0.038 \mu\text{M}$, $0.022 \mu\text{M}$, $0.033 \mu\text{M}$, $0.053 \mu\text{M}$, and $0.0046 \mu\text{M}$, respectively (Fig.26). Interestingly, **DSM265** (**77**) showed excellent *in vivo* anti-parasitic activity against both *P. falciparum* and *P. vivax* and reached phase IIa clinical trials.

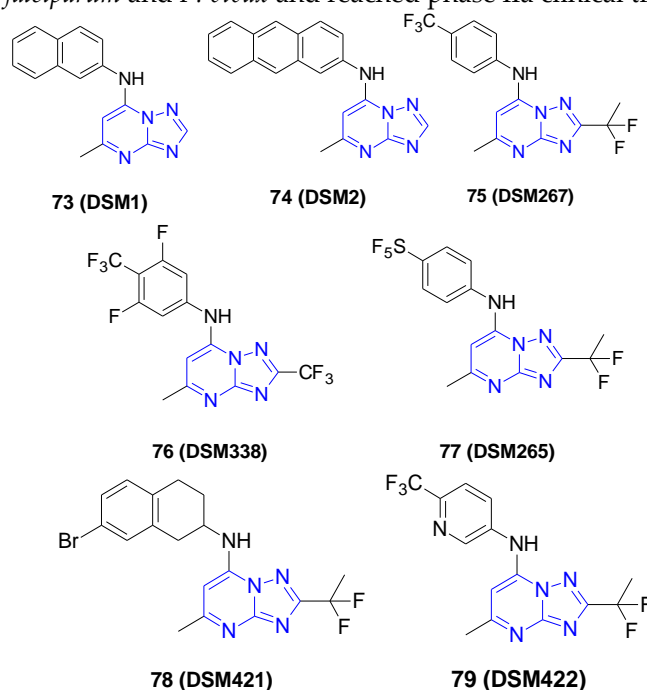


Fig.26. Structures of TP derivatives **73-79** acting as antimalarial agents.

3. Conclusion

The 1,2,4-Triazolo[1,5-a]pyrimidine scaffold emerged as a prominent scaffold in designing novel compounds acting as promising drug candidates for the treatment of several diseases.

Also, the development of pharmacologically active TP analogues has been accelerated recently, with many of these analogues displaying drug-like properties. This review summarizes the most important application of TP in medicinal chemistry and illustrates their interactions with several targets for their diverse biological activities.

Ethical consideration: All the participants in this study gave their informed permission.

Conflicts of Interest

4. References

- [1] G. Fischer, 1,2,4-Triazolo[1,5-a]pyrimidines, *Adv Heterocycl Chem* 57 (1993) 82–138. [https://doi.org/10.1016/S0065-2725\(08\)60887-9](https://doi.org/10.1016/S0065-2725(08)60887-9).
- [2] Fischer G, Recent progress in 1,2,4-triazolo[1,5-a]pyrimidine chemistry, *Adv Heterocycl Chem* 95 (2007) 143–219. [https://doi.org/10.1016/S0065-2725\(07\)95001-1](https://doi.org/10.1016/S0065-2725(07)95001-1).
- [3] G. Fischer, Recent advances in 1,2,4-triazolo[1,5-a]pyrimidine chemistry, in: *Adv Heterocycl Chem*, Academic Press Inc., (2019) 1–101. <https://doi.org/10.1016/bs.aihch.2018.10.002>.
- [4] P.K. Singh, S. Choudhary, A. Kashyap, H. Verma, S. Kapil, M. Kumar, M. Arora, O. Silakari, An exhaustive compilation on chemistry of triazolopyrimidine: A journey through decades, *Bioorg Chem* 88 (2019) 102919. <https://doi.org/10.1016/j.bioorg.2019.102919>.
- [5] M.M.A. El-Gendy, M. Shaaban, K.A. Shaaban, A.M. El-Bondkly, H. Laatsch, Essramycin: A First Triazolopyrimidine Antibiotic Isolated from Nature†, *J Antibiot (Tokyo)* 61 (2008) 149–157. <https://doi.org/10.1038/ja.2008.124>.
- [6] C. Bulow, K. Hass, Synthetische Versuche zur Darstellung von Derivaten des heterokondensierten, heterocyclischen 1.3-Triazo-7.0'-pyrimidins., *Berichte Der Deutschen Chemischen Gesellschaft*. 42 (1909) 4638–4644. <https://doi.org/10.1002/cber.19090420468>.
- [7] B.-B.S. Zhou, S.J. Elledge, The DNA damage response: putting checkpoints in perspective, *Nature* 408 (2000) 433–439. <https://doi.org/10.1038/35044005>.
- [8] S.P. Jackson, J. Bartek, The DNA-damage response in human biology and disease, *Nature* 461 (2009) 1071–1078. <https://doi.org/10.1038/nature08467>.
- [9] A. Tubbs, A. Nussenzweig, Endogenous DNA Damage as a Source of Genomic Instability in Cancer, *Cell* 168 (2017) 644–656. <https://doi.org/10.1016/j.cell.2017.01.002>.
- [10] T. Helleday, E. Petermann, C. Lundin, B. Hodgson, R.A. Sharma, DNA repair pathways as targets for cancer therapy, *Nat Rev Cancer* 8 (2008) 193–204. <https://doi.org/10.1038/nrc2342>.
- [11] M.A. Kristensson, The game of tubulins, *Cells* 10 (2021) 745. <https://doi.org/10.3390/cells10040745>.
- [12] C. D. Katsetos, P. Draber, Tubulins as Therapeutic Targets in Cancer: from Bench to Bedside, *Curr Pharm Des* 18 (2012) 2778–2792. <https://doi.org/10.2174/138161212800626193>.
- [13] O. Ebenezer, M. Shapi, J.A. Tuszynski, A Review of the Recent Developments of Molecular Hybrids Targeting Tubulin Polymerization, *Int J Mol Sci* 23 (2022) 4001. <https://doi.org/10.3390/ijms23074001>.
- [14] M.N. Peerzada, M.S. Dar, S. Verma, Development of tubulin polymerization inhibitors as anticancer agents, *Expert Opin Ther Pat* 33 (2023) 797–820. <https://doi.org/10.1080/13543776.2023.2291390>.
- [15] N. Zhang, S. Ayral-Kaloustian, T. Nguyen, R. Hernandez, C. Beyer, 2-Cyanoaminopyrimidines as a class of antitumor agents that promote tubulin polymerization, *Bioorg Med Chem Lett* 17 (2007) 3003–3005. <https://doi.org/10.1016/j.bmcl.2007.03.070>.
- [16] A. Wang-Gillam, S.M. Arnold, R.M. Bukowski, M.L. Rothenberg, W. Cooper, K.K. Wang, E. Gauthier, A.C. Lockhart, A phase I dose escalation study of TTI-237 in patients with advanced malignant solid tumors, *Invest New Drugs* 30 (2012) 266–272. <https://doi.org/10.1007/s10637-010-9506-3>.
- [17] F. Yang, L.Z. Yu, P.C. Diao, X.E. Jian, M.F. Zhou, C.S. Jiang, W.W. You, W.F. Ma, P.L. Zhao, Novel [1,2,4]triazolo[1,5-a]pyrimidine derivatives as potent antitubulin agents: Design, multicomponent synthesis and antiproliferative activities, *Bioorg Chem* 92 (2019) 103260. <https://doi.org/10.1016/j.bioorg.2019.103260>.
- [18] X. Sen Huo, X.E. Jian, J. Ou-Yang, L. Chen, F. Yang, D.X. Lv, W.W. You, J.J. Rao, P.L. Zhao, Discovery of highly potent tubulin polymerization inhibitors: Design, synthesis, and structure-activity relationships of novel 2,7-diaryl-[1,2,4]triazolo[1,5-a]pyrimidines, *Eur J Med Chem* 220 (2021) 113449. <https://doi.org/10.1016/j.ejmech.2021.113449>.
- [19] H.S. Mohamed, N.H. Amin, M.T. El-Saadi, H.M. Abdel-Rahman, Design, synthesis, biological assessment, and in-Silico studies of 1,2,4-triazolo[1,5-a]pyrimidine derivatives as tubulin polymerization inhibitors, *Bioorg Chem* 121 (2022) 105687. <https://doi.org/10.1016/j.bioorg.2022.105687>.
- [20] L. Chen, T.Y. Ji, X. Sen Huo, Z.Y. Zeng, W.X. Ye, C.C. Dai, Y.Q. Zhang, W.W. You, P.L. Zhao, Rational design, synthesis and biological evaluation of novel 2-(substituted amino)-[1,2,4]triazolo[1,5-a]pyrimidines as novel tubulin polymerization inhibitors, *Eur J Med Chem* 244 (2022) 114864. <https://doi.org/10.1016/j.ejmech.2022.114864>.
- [21] P. Oliva, R. Romagnoli, B. Cacciari, S. Manfredini, C. Padroni, A. Brancale, S. Ferla, E. Hamel, D. Corallo, S. Aveic, N. Milan, E. Mariotto, G. Viola, R. Bortolozzi, Synthesis and Biological Evaluation of Highly Active 7-Anilino Triazolopyrimidines as Potent Antimicrotubule Agents, *Pharmaceutics* 14 (2022) 1191. <https://doi.org/10.3390/pharmaceutics14061191>.
- [22] R. Romagnoli, P. Oliva, F. Prencipe, S. Manfredini, F. Budassi, A. Brancale, S. Ferla, E. Hamel, D. Corallo, S. Aveic, L. Manfreda, E. Mariotto, R. Bortolozzi, G. Viola, Design, synthesis and biological investigation of 2-anilino triazolopyrimidines as tubulin polymerization inhibitors with anticancer activities, *Pharmaceutics* 15 (2022) 1031. <https://doi.org/10.3390/ph15081031>.
- [23] U. Asghar, A.K. Witkiewicz, N.C. Turner, E.S. Knudsen, The history and future of targeting cyclin-dependent kinases in cancer therapy, *Nat Rev Drug Discov* 14 (2015) 130–146. <https://doi.org/10.1038/nrd4504>.
- [24] S. Tadesse, A.T. Anshabo, N. Portman, E. Lim, W. Tilley, C.E. Caldon, S. Wang, Targeting CDK2 in cancer: challenges and opportunities for therapy, *Drug Discov Today* 25 (2020) 406–413. <https://doi.org/10.1016/j.drudis.2019.12.001>.
- [25] P. Łukasik, M. Załuski, I. Gutowska, Cyclin-dependent kinases (Cdk) and their role in diseases development—review, *Int J Mol Sci* 22 (2021) 1–33. <https://doi.org/10.3390/ijms22062935>.
- [26] C. Borgo, C. D'Amore, S. Sarno, M. Salvi, M. Ruzzene, Protein kinase CK2: a potential therapeutic target for diverse human diseases, *Signal Transduct Target Ther* 6 (2021) 183. <https://doi.org/10.1038/s41392-021-00567-7>.
- [27] R.C. Jackson, A.L. Barnett, S.J. McClue, S.R. Green, Seliciclib, a cell-cycle modulator that acts through the inhibition of cyclin-dependent kinases, *Expert Opin Drug Discov* 3 (2008) 131–143. <https://doi.org/10.1517/17460441.3.1.131>.
- [28] N.-A. Liu, A. Ben-Shlomo, J.D. Carmichael, C. Wang, R.S. Swerdloff, A.P. Heaney, G. Barkhoudarian, D. Kelly, M. Nouredin, L. Lu, M. Desai, Y. Stolyarov, K. Yuen, A.N. Mamelak, J. Mirocha, M. Tighiouart, S. Melmed, Treatment of Cushing Disease With Pituitary-

- Targeting Seliciclib, *J Clin Endocrinol Metab* 108 (2023) 726–735. <https://doi.org/10.1210/clinem/dgac588>.
- [29] C.M. Richardson, D.S. Williamson, M.J. Parratt, J. Borgognoni, A.D. Cansfield, P. Dokurno, G.L. Francis, R. Howes, J.D. Moore, J.B. Murray, A. Robertson, A.E. Surgenor, C.J. Torrance, Triazolo[1,5-a]pyrimidines as novel CDK2 inhibitors: Protein structure-guided design and SAR, *Bioorg Med Chem Lett* 16 (2006) 1353–1357. <https://doi.org/10.1016/j.bmcl.2005.11.048>.
- [30] F.C. Ledesma, S.F. El Khamisy, M.C. Zuma, K. Osborn, K.W. Caldecott, A human 5'-tyrosyl DNA phosphodiesterase that repairs topoisomerase-mediated DNA damage, *Nature* 461 (2009) 674–678. <https://doi.org/10.1038/nature08444>.
- [31] Z. Zeng, F. Cortés-Ledesma, S.F. El Khamisy, K.W. Caldecott, TDP2/TTRAP is the major 5'-tyrosyl DNA phosphodiesterase activity in vertebrate cells and is critical for cellular resistance to topoisomerase II-induced DNA damage, *Journal of Biological Chemistry* 286 (2011) 403–409. <https://doi.org/10.1074/jbc.M110.181016>.
- [32] Y. Pommier, S. Yin N. Huang, R. Gao, B.B. Das, J. Murai, C. Marchand, Tyrosyl-DNA-phosphodiesterases (TDP1 and TDP2), *DNA Repair (Amst)* 19 (2014) 114–129. <https://doi.org/10.1016/j.dnarep.2014.03.020>.
- [33] S.S. Laev, N.F. Salakhutdinov, O.I. Lavrik, Tyrosyl-DNA phosphodiesterase inhibitors: Progress and potential, *Bioorg Med Chem* 24 (2016) 5017–5027. <https://doi.org/10.1016/j.bmc.2016.09.045>.
- [34] C.J.A. Ribeiro, J. Kankanala, K. Shi, K. Kurahashi, E. Kiselev, A. Ravji, Y. Pommier, H. Aihara, Z. Wang, New fluorescence-based high-throughput screening assay for small molecule inhibitors of tyrosyl-DNA phosphodiesterase 2 (TDP2), *European Journal of Pharmaceutical Sciences* 118 (2018) 67–79. <https://doi.org/10.1016/j.ejps.2018.03.021>.
- [35] C.J.A. Ribeiro, J. Kankanala, J. Xie, J. Williams, H. Aihara, Z. Wang, Triazolopyrimidine and triazolopyridine scaffolds as TDP2 inhibitors, *Bioorg Med Chem Lett* 29 (2019) 257–261. <https://doi.org/10.1016/j.bmcl.2018.11.044>.
- [36] S. Kumar, S. Rulhania, S. Jaswal, V. Monga, Recent advances in the medicinal chemistry of carbonic anhydrase inhibitors, *Eur J Med Chem* 209 (2021) 112923. <https://doi.org/10.1016/j.ejmech.2020.112923>.
- [37] R.P. Henry, Multiple roles of carbonic anhydrase in cellular transport and metabolism, *Annu. Rev Physiol.* 58 (1996) 5–52. <https://doi.org/10.1146/annurev.ph.58.030196.002515>
- [38] C. Geers, G. Gros, Carbon Dioxide Transport and Carbonic Anhydrase in Blood and Muscle, *Physiol Rev* 80 (2000) 715. <https://doi.org/10.1152/physrev.2000.80.2.681>
- [39] E. Kupriyanova, N. Pronina, D. Los, Carbonic anhydrase – a universal enzyme of the carbon-based life, *Photosynthetica* 55 (2017) 3–19. <https://doi.org/10.1007/s11099-017-0685-4>.
- [40] C.T. Supuran, Advances in structure-based drug discovery of carbonic anhydrase inhibitors, *Expert Opin Drug Discov* 12 (2017) 61–88. <https://doi.org/10.1080/17460441.2017.1253677>.
- [41] C.T. Supuran, Carbonic anhydrases: Novel therapeutic applications for inhibitors and activators, *Nat Rev Drug Discov* 7 (2008) 168–181. <https://doi.org/10.1038/nrd2467>.
- [42] M.A. Said, W.M. Eldehna, A. Nocentini, A. Bonardi, S.H. Fahim, S. Bua, D.H. Soliman, H.A. Abdel-Aziz, P. Gratteri, S.M. Abou-Seri, C.T. Supuran, Synthesis, biological and molecular dynamics investigations with a series of triazolopyrimidine/triazole-based benzenesulfonamides as novel carbonic anhydrase inhibitors, *Eur J Med Chem* 185 (2020) 111843. <https://doi.org/10.1016/j.ejmech.2019.111843>.
- [43] R. Romagnoli, T. De Ventura, S. Manfredini, E. Baldini, C.T. Supuran, A. Nocentini, A. Brancale, R. Bortolozzi, L. Manfreda, G. Viola, Design, synthesis, and biological investigation of selective human carbonic anhydrase II, IX, and XII inhibitors using 7-aryl/heteroaryl triazolopyrimidines bearing a sulfanilamide scaffold, *J Enzyme Inhib Med Chem* 38 (2023) 2270180. <https://doi.org/10.1080/14756366.2023.2270180>.
- [44] R.M. Sanchez, K. Erhard, M.A. Hardwicke, H. Lin, J. McSurdy-Freed, R. Plant, K. Raha, C.M. Rominger, M.D. Schaber, M.D. Spengler, M.L. Moore, H. Yu, J.I. Luengo, R. Tedesco, R.A. Rivero, Synthesis and structure-activity relationships of 1,2,4-triazolo[1,5-a] pyrimidin-7(3H)-ones as novel series of potent β isoform selective phosphatidylinositol 3-kinase inhibitors, *Bioorg Med Chem Lett* 22 (2012) 3198–3202. <https://doi.org/10.1016/j.bmcl.2012.03.039>.
- [45] S.T.J. Bradford, E. Grimley, A.M. Laszczyk, P.H. Lee, S.R. Patel, G.R. Dressler, Identification of Pax protein inhibitors that suppress target gene expression and cancer cell proliferation, *Cell Chem Biol* 29 (2022) 412–422.e4. <https://doi.org/10.1016/j.chembiol.2021.11.003>.
- [46] K.D. Wilkinson, Ubiquitin: A nobel protein, *Cell* 119 (2004) 741–745. <https://doi.org/10.1016/j.cell.2004.12.001>.
- [47] E. Reinsteint, A. Ciechanover, Narrative Review: Protein Degradation and Human Diseases: The Ubiquitin Connection, *Ann. Intern Med* 145 (2006) 676–684. www.annals.org.
- [48] L.N. Micel, J.J. Tentler, P.G. Smith, S.G. Eckhardt, Role of ubiquitin ligases and the proteasome in oncogenesis: Novel targets for anticancer therapies, *Journal of Clinical Oncology* 31 (2013) 1231–1238. <https://doi.org/10.1200/JCO.2012.44.0958>.
- [49] S. Wang, L. Zhao, X.J. Shi, L. Ding, L. Yang, Z.Z. Wang, D. Shen, K. Tang, X.J. Li, M.A.A. Mamun, H. Li, B. Yu, Y.C. Zheng, S. Wang, H.M. Liu, Development of Highly Potent, Selective, and Cellular Active Triazolo[1,5-a]pyrimidine-Based Inhibitors Targeting the DCN1-UBC12 Protein-Protein Interaction, *J Med Chem* 62 (2019) 2772–2797. <https://doi.org/10.1021/acs.jmedchem.9b00113>.
- [50] T. Ma, Q. Song, B. Cheng, E. Guo, X. Wang, M. Li, M. Dai, S. Li, S. Feng, B. Yu, Proapoptotic effect of WS-299 induced by NOXA accumulation and NRF2-counterbalanced oxidative stress damage through targeting RBX1-UBE2M interaction in gastric cancers, *Bioorg Chem* 144 (2024) 107142. <https://doi.org/10.1016/j.bioorg.2024.107142>.
- [51] Z.R. Li, S. Wang, L. Yang, X.H. Yuan, F.Z. Suo, B. Yu, H.M. Liu, Experience-based discovery (EBD) of aryl hydrazines as new scaffolds for the development of LSD1/KDM1A inhibitors, *Eur J Med Chem* 166 (2019) 432–444. <https://doi.org/10.1016/j.ejmech.2019.01.075>.
- [52] S. Wang, L.J. Zhao, Y.C. Zheng, D.D. Shen, E.F. Miao, X.P. Qiao, L.J. Zhao, Y. Liu, R. Huang, B. Yu, H.M. Liu, Design, synthesis and biological evaluation of [1,2,4]triazolo[1,5-a]pyrimidines as potent lysine specific demethylase 1 (LSD1/KDM1A) inhibitors, *Eur J Med Chem* 125 (2017) 940–951. <https://doi.org/10.1016/j.ejmech.2016.10.021>.
- [53] S. Wang, Z.R. Li, F.Z. Suo, X.H. Yuan, B. Yu, H.M. Liu, Synthesis, structure-activity relationship studies and biological characterization of new [1,2,4]triazolo[1,5-a]pyrimidine-based LSD1/KDM1A

- inhibitors, *Eur J Med Chem* 167 (2019) 388–401. <https://doi.org/10.1016/j.ejmech.2019.02.039>.
- [54] S. Wang, X. Bin Ma, X.H. Yuan, B. Yu, Y.C. Xu, H.M. Liu, Discovery of new [1,2,4] Triazolo[1,5-a]Pyrimidine derivatives that Kill gastric cancer cells via the mitochondria pathway, *Eur J Med Chem* 203 (2020) 112630. <https://doi.org/10.1016/j.ejmech.2020.112630>.
- [55] J.L. Huo, S. Wang, X.H. Yuan, B. Yu, W. Zhao, H.M. Liu, Discovery of [1,2,4]triazolo[1,5-a]pyrimidines derivatives as potential anticancer agents, *Eur J Med Chem* 211 (2021) 113108. <https://doi.org/10.1016/j.ejmech.2020.113108>.
- [56] T. Lu, W. Lu, C. Luo, A patent review of BRD4 inhibitors (2013-2019), *Expert Opin Ther Pat* 30 (2020) 57–81. <https://doi.org/10.1080/13543776.2020.1702645>.
- [57] S. Wang, D. Shen, L. Zhao, X. Yuan, J. Cheng, B. Yu, Y. Zheng, H. Liu, Discovery of [1,2,4]triazolo[1,5-a]pyrimidine derivatives as new bromodomain-containing protein 4 (BRD4) inhibitors, *Chinese Chemical Letters* 31 (2020) 418–422. <https://doi.org/10.1016/j.ccl.2019.08.029>.
- [58] T. Heinrich, H.P. Buchstaller, B. Cezanne, F. Rohdich, J. Bomke, M. Friese-Hamim, M. Krier, T. Knöchel, D. Musil, B. Leuthner, F. Zenke, Novel reversible methionine aminopeptidase-2 (MetAP-2) inhibitors based on purine and related bicyclic templates, *Bioorg Med Chem Lett* 27 (2017) 551–556. <https://doi.org/10.1016/j.bmcl.2016.12.019>.
- [59] L. Chang, M. Xiao, L. Yang, S. Wang, S.Q. Wang, A. Bender, A. Hu, Z.S. Chen, B. Yu, H.M. Liu, Discovery of a non-toxic [1,2,4]triazolo[1,5-a]pyrimidin-7-one (WS-10) that modulates ABCB1-mediated multidrug resistance (MDR), *Bioorg Med Chem* 26 (2018) 5006–5017. <https://doi.org/10.1016/j.bmc.2018.08.021>.
- [60] S. Wang, S.Q. Wang, Q.X. Teng, L. Yang, Z.N. Lei, X.H. Yuan, J.F. Huo, X.B. Chen, M. Wang, B. Yu, Z.S. Chen, H.M. Liu, Structure-Based Design, Synthesis, and Biological Evaluation of New Triazolo[1,5- a]Pyrimidine Derivatives as Highly Potent and Orally Active ABCB1 Modulators, *J Med Chem* 63 (2020) 15979–15996. <https://doi.org/10.1021/acs.jmedchem.0c01741>.
- [61] J. Kumar, P. Meena, A. Singh, E. Jameel, M. Maqbool, M. Mobashir, A. Shandilya, M. Tiwari, N. Hoda, B. Jayaram, Synthesis and screening of triazolopyrimidine scaffold as multi-functional agents for Alzheimer's disease therapies, *Eur J Med Chem* 119 (2016) 260–277. <https://doi.org/10.1016/j.ejmech.2016.04.053>.
- [62] E. Jameel, P. Meena, M. Maqbool, J. Kumar, W. Ahmed, S. Mumtazuddin, M. Tiwari, N. Hoda, B. Jayaram, Rational design, synthesis and biological screening of triazine-triazolopyrimidine hybrids as multitarget anti-Alzheimer agents, *Eur J Med Chem* 136 (2017) 36–51. <https://doi.org/10.1016/j.ejmech.2017.04.064>.
- [63] J. Kumar, A. Gill, M. Shaikh, A. Singh, A. Shandilya, E. Jameel, N. Sharma, N. Mrinal, N. Hoda, B. Jayaram, Pyrimidine-Triazolopyrimidine and Pyrimidine-Pyridine Hybrids as Potential Acetylcholinesterase Inhibitors for Alzheimer's Disease, *Chemistry Select* 3 (2018) 736–747. <https://doi.org/10.1002/slct.201702599>.
- [64] B. Zhang, Y. Yao, A.S. Cornec, K. Oukoloff, M.J. James, P. Koivula, J.Q. Trojanowski, A.B. Smith, V.M.Y. Lee, C. Ballatore, K.R. Brunden, A brain-penetrant triazolopyrimidine enhances microtubule-stability, reduces axonal dysfunction and decreases tau pathology in a mouse tauopathy model, *Mol Neurodegener* 13 (2018) 1–15. <https://doi.org/10.1186/s13024-018-0291-3>.
- [65] K. Oukoloff, J. Kovalevich, A.S. Cornec, Y. Yao, Z.A. Owyang, M. James, J.Q. Trojanowski, V.M.Y. Lee, A.B. Smith, K.R. Brunden, C. Ballatore, Design, synthesis and evaluation of photoactivatable derivatives of microtubule (MT)-active [1,2,4]triazolo[1,5-a]pyrimidines, *Bioorg Med Chem Lett* 28 (2018) 2180–2183. <https://doi.org/10.1016/j.bmcl.2018.05.010>.
- [66] K. Oukoloff, G. Nzou, C. Varricchio, B. Lucero, T. Alle, J. Kovalevich, L. Monti, A.S. Cornec, Y. Yao, M.J. James, J.Q. Trojanowski, V.M.Y. Lee, A.B. Smith, A. Brancale, K.R. Brunden, C. Ballatore, Evaluation of the Structure-Activity Relationship of Microtubule-Targeting 1,2,4-Triazolo[1,5- a]pyrimidines Identifies New Candidates for Neurodegenerative Tauopathies, *J Med Chem* 64 (2021) 1073–1102. <https://doi.org/10.1021/acs.jmedchem.0c01605>.
- [67] T. Alle, C. Varricchio, Y. Yao, B. Lucero, G. Nzou, S. Demuro, M. Muench, K.D. Vuong, K. Oukoloff, A.S. Cornec, K.R. Francisco, C.R. Caffrey, V.M.Y. Lee, A.B. Smith, A. Brancale, K.R. Brunden, C. Ballatore, Microtubule-Stabilizing 1,2,4-Triazolo[1,5- a]pyrimidines as Candidate Therapeutics for Neurodegenerative Disease: Matched Molecular Pair Analyses and Computational Studies Reveal New Structure-Activity Insights, *J Med Chem* 66 (2023) 435–459. <https://doi.org/10.1021/acs.jmedchem.2c01411>.
- [68] A.A. Trabanco, P. Buijnsters, F.J.R. Rombouts, Towards selective phosphodiesterase 2A (PDE2A) inhibitors: a patent review (2010 - present), *Expert Opin Ther Pat* 26 (2016) 933–946. <https://doi.org/10.1080/13543776.2016.1203902>.
- [69] L. Gomez, M.E. Massari, T. Vickers, G. Freestone, W. Vernier, K. Ly, R. Xu, M. McCarrick, T. Marrone, M. Metz, Y.G. Yan, Z.W. Yoder, R. Lemus, N.J. Broadbent, R. Barido, N. Warren, K. Schmelzer, D. Neul, D. Lee, C.B. Andersen, K. Sebring, K. Aertgeerts, X. Zhou, A. Tabatabaei, M. Peters, J.G. Breitenbucher, Design and Synthesis of Novel and Selective Phosphodiesterase 2 (PDE2a) Inhibitors for the Treatment of Memory Disorders, *J Med Chem* 60 (2017) 2037–2051. <https://doi.org/10.1021/acs.jmedchem.6b01793>.
- [70] G. Tresadern, I. Velter, A.A. Trabanco, F. Van Den Keybus, G.J. MacDonald, M.V.F. Somers, G. Vanhoof, P.M. Leonard, M.B.A.C. Lamers, Y.E.M. Van Roosbroeck, P.J.J.A. Buijnsters, Triazolo[1,5-a]pyrimidine Phosphodiesterase 2A Inhibitors: Structure and Free-Energy Perturbation-Guided Exploration, *J Med Chem* 63 (2020) 12887–12910. <https://doi.org/10.1021/acs.jmedchem.0c01272>.
- [71] U. Battaglia, C.J. Moody, A short synthesis of the triazolopyrimidine antibiotic essramycin, *J Nat Prod* 73 (2010) 1938–1939. <https://doi.org/10.1021/np100298m>.
- [72] E.H.L. Tee, T. Karoli, S. Ramu, J. Huang, M.S. Butler, M.A. Cooper, The Synthesis of Essramycin and Comparison of its Anti-bacterial Activity, *J Nat Prod* 73 (2010) 1940–1942. <https://doi.org/10.1021/np100648q>.
- [73] H. Wang, D. Heseck, M. Lee, E. Lastochkin, A.G. Oliver, M. Chang, S. Mobashery, The Natural Product Essramycin and Three of Its Isomers Are Devoid of Antibacterial Activity HHS Public Access, *J Nat Prod* 79 (2016) 1219–1222. <https://doi.org/10.1021/acs.jnat-prod>.
- [74] V. Patil, M. Kale, A. Raichurkar, B. Bhaskar, D. Prahlad, M. Balganes, S. Nandan, P. Shahul Hameed, Design and synthesis of triazolopyrimidine acylsulfonamides as novel anti-mycobacterial leads acting through inhibition of acetohydroxyacid synthase, *Bioorg Med Chem Lett* 24 (2014) 2222–2225. <https://doi.org/10.1016/j.bmcl.2014.02.054>.

- [75] H. Wang, M. Lee, Z. Peng, B. Blázquez, E. Lastochkin, M. Kumarasiri, R. Bouley, M. Chang, S. Mobashery, Synthesis and evaluation of 1,2,4-triazolo[1,5-a]pyrimidines as antibacterial agents against *Enterococcus faecium*, *J Med Chem* 58 (2015) 4194–4203. <https://doi.org/10.1021/jm501831g>.
- [76] I.P. Jung, N.R. Ha, S.C. Lee, S.W. Ryoo, M.Y. Yoon, Development of potent chemical antituberculosis agents targeting *Mycobacterium tuberculosis* acetohydroxyacid synthase, *Int J Antimicrob Agents* 48 (2016) 247–258. <https://doi.org/10.1016/j.ijantimicag.2016.04.031>.
- [77] R.H. Abd El-Aleam, R.F. George, G.S. Hassan, H.M. Abdel-Rahman, Synthesis of 1,2,4-triazolo[1,5-a]pyrimidine derivatives: Antimicrobial activity, DNA Gyrase inhibition and molecular docking, *Bioorg Chem* 94 (2020) 103411. <https://doi.org/10.1016/j.bioorg.2019.103411>.
- [78] A.A. Abu-Hashem, M. El-Shazly, Synthesis and Antimicrobial Evaluation of Novel Triazole, Tetrazole, and Spiropyrimidine-Thiadiazole Derivatives, *Polycycl Aromat Compd* 41 (2021) 478–497. <https://doi.org/10.1080/10406638.2019.1598448>.
- [79] M.A.A. Mohamed, A.A. Bekhit, O.A.A. Allah, A.M. Kadry, T.M. Ibrahim, S.A. Bekhit, K. Amagase, A.M.M. El-Saghier, Synthesis and antimicrobial activity of some novel 1,2-dihydro-[1,2,4]triazolo[1,5-a]pyrimidines bearing amino acid moiety, *RSC Adv* 11 (2021) 2905–2916. <https://doi.org/10.1039/d0ra08189b>.
- [80] N.H. Amin, M.T. El-Saadi, A.A. Ibrahim, H.M. Abdel-Rahman, Design, synthesis and mechanistic study of new 1,2,4-triazole derivatives as antimicrobial agents, *Bioorg Chem* 111 (2021) 104841. <https://doi.org/10.1016/j.bioorg.2021.104841>.
- [81] S. Massari, G. Nannetti, J. Desantis, G. Muratore, S. Sabatini, G. Manfroni, B. Mercorelli, V. Cecchetti, G. Palù, G. Cruciani, A. Loregian, L. Goracci, O. Tabarrini, A Broad anti-influenza hybrid small molecule that potently disrupts the interaction of polymerase acidic protein-basic protein 1 (PA-PB1) subunits, *J Med Chem* 58 (2015) 3830–3842. <https://doi.org/10.1021/acs.jmedchem.5b00012>.
- [82] S. Massari, J. Desantis, G. Nannetti, S. Sabatini, S. Tortorella, L. Goracci, V. Cecchetti, A. Loregian, O. Tabarrini, Efficient and regioselective one-step synthesis of 7-aryl-5-methyl- and 5-aryl-7-methyl-2-amino-[1,2,4]triazolo[1,5-a] pyrimidine derivatives, *Org Biomol Chem* 15 (2017) 7944–7955. <https://doi.org/10.1039/c7ob02085f>.
- [83] S. Massari, C. Bertagnin, M.C. Pismataro, A. Donnadio, G. Nannetti, T. Felicetti, S. Di Bona, M.G. Nizi, L. Tensi, G. Manfroni, M.I. Loza, S. Sabatini, V. Cecchetti, J. Brea, L. Goracci, A. Loregian, O. Tabarrini, Synthesis and characterization of 1,2,4-triazolo[1,5-a]pyrimidine-2-carboxamide-based compounds targeting the PA-PB1 interface of influenza A virus polymerase, *Eur J Med Chem* 209 (2021) 112944. <https://doi.org/10.1016/j.ejmech.2020.112944>.
- [84] M.C. Pismataro, T. Felicetti, C. Bertagnin, M.G. Nizi, A. Bonomini, M.L. Barreca, V. Cecchetti, D. Jochmans, S. De Jonghe, J. Neyts, A. Loregian, O. Tabarrini, S. Massari, 1,2,4-Triazolo[1,5-a]pyrimidines: Efficient one-step synthesis and functionalization as influenza polymerase PA-PB1 interaction disruptors, *Eur J Med Chem* 221 (2021) 113494. <https://doi.org/10.1016/j.ejmech.2021.113494>.
- [85] H. Li, J. Tatlock, A. Linton, J. Gonzalez, T. Jewell, L. Patel, S. Ludlum, M. Drowns, S. V. Rahavendran, H. Skor, R. Hunter, S.T. Shi, K.J. Herlihy, H. Parge, M. Hickey, X. Yu, F. Chau, J. Nonomiya, C. Lewis, Discovery of (R)-6-cyclopentyl-6-(2-(2,6-diethylpyridin-4-yl)ethyl)-3-((5, 7-dimethyl-[1,2,4]triazolo[1,5-a]pyrimidin-2-yl)methyl)-4-hydroxy-5, 6-dihydropyran-2-one (PF-00868554) as a potent and orally available hepatitis C virus polymerase inhibitor, *J Med Chem* 52 (2009) 1255–1258. <https://doi.org/10.1021/jm8014537>.
- [86] Y. Tian, D. Du, D. Rai, L. Wang, H. Liu, P. Zhan, E. De Clercq, C. Pannecouque, X. Liu, Fused heterocyclic compounds bearing bridgehead nitrogen as potent HIV-1 NNRTIs. Part 1: Design, synthesis and biological evaluation of novel 5,7-disubstituted pyrazolo[1,5-a]pyrimidine derivatives, *Bioorg Med Chem* 22 (2014) 2052–2059. <https://doi.org/10.1016/j.bmc.2014.02.029>.
- [87] L. Wang, Y. Tian, W. Chen, H. Liu, P. Zhan, D. Li, H. Liu, E. De Clercq, C. Pannecouque, X. Liu, Fused heterocycles bearing bridgehead nitrogen as potent HIV-1 NNRTIs. Part 2: Discovery of novel [1,2,4]Triazolo[1,5-a]pyrimidines using a structure-guided core-refining approach, *Eur J Med Chem* 85 (2014) 293–303. <https://doi.org/10.1016/j.ejmech.2014.07.104>.
- [88] B. Huang, C. Li, W. Chen, T. Liu, M. Yu, L. Fu, Y. Sun, H. Liu, E. De Clercq, C. Pannecouque, J. Balzarini, P. Zhan, X. Liu, Fused heterocycles bearing bridgehead nitrogen as potent HIV-1 NNRTIs. Part 3: Optimization of [1,2,4]triazolo[1,5-a]pyrimidine core via structure-based and physicochemical property-driven approaches, *Eur J Med Chem* 92 (2015) 754–765. <https://doi.org/10.1016/j.ejmech.2015.01.042>.
- [89] S. Khare, A.S. Nagle, A. Biggart, Y.H. Lai, F. Liang, L.C. Davis, S.W. Barnes, C.J.N. Mathison, E. Myburgh, M.Y. Gao, J.R. Gillespie, X. Liu, J.L. Tan, M. Stinson, I.C. Rivera, J. Ballard, V. Yeh, T. Groessl, G. Federe, H.X.Y. Koh, J.D. Venable, B. Bursulaya, M. Shapiro, P.K. Mishra, G. Spraggon, A. Brock, J.C. Mottram, F.S. Buckner, S.P.S. Rao, B.G. Wen, J.R. Walker, T. Tuntland, V. Molteni, R.J. Glynn, F. Suppek, Proteasome inhibition for treatment of leishmaniasis, Chagas disease and sleeping sickness, *Nature* 537 (2016) 229–233. <https://doi.org/10.1038/nature19339>.
- [90] X. Deng, R. Gujjar, F. El Mazouni, W. Kaminsky, N.A. Malmquist, E.J. Goldsmith, P.K. Rathod, M.A. Phillips, Structural plasticity of malaria dihydroorotate dehydrogenase allows selective binding of diverse chemical scaffolds, *Journal of Biological Chemistry* 284 (2009) 26999–27009. <https://doi.org/10.1074/jbc.M109.028589>.
- [91] J.M. Coteron, M. Marco, J. Esquivias, X. Deng, K.L. White, J. White, M. Koltun, F. El Mazouni, S. Kokkonda, K. Katneni, R. Bhamidipati, D.M. Shackleford, I. Angulo-Barturen, S.B. Ferrer, M.B. Jiménez-Díaz, F.J. Gamó, E.J. Goldsmith, W.N. Charman, I. Bathurst, D. Floyd, D. Matthews, J.N. Burrows, P.K. Rathod, S.A. Charman, M.A. Phillips, Structure-guided lead optimization of triazolopyrimidine-ring substituents identifies potent plasmodium falciparum dihydroorotate dehydrogenase inhibitors with clinical candidate potential, *J Med Chem* 54 (2011) 5540–5561. <https://doi.org/10.1021/jm200592f>.
- [92] X. Deng, S. Kokkonda, F. El Mazouni, J. White, J.N. Burrows, W. Kaminsky, S.A. Charman, D. Matthews, P.K. Rathod, M.A. Phillips, Fluorine modulates species selectivity in the triazolopyrimidine class of Plasmodium falciparum dihydroorotate dehydrogenase inhibitors, *J Med Chem* 57 (2014) 5381–5394. <https://doi.org/10.1021/jm500481t>.
- [93] M.A. Phillips, J. Lotharius, K. Marsh, J. White, A. Dayan, K.L. White, J.W. Njoroge, F. El Mazouni, Y. Lao, S. Kokkonda, D.R. Tomchick, X. Deng, T. Laird, S.N. Bhatia, S. March, C.L. Ng, D.A. Fidock, S. Wittlin, M. Lafuente-Monasterio, F. Javier, G. Benito, L. Maria, S. Alonso, M. Santos Martinez, M. Belen Jimenez-Diaz, S. Ferrer Bazaga, I. Angulo-

Barturen, J.N. Haselden, J. Louttit, Y. Cui, A. Sridhar, A.-M. Zeeman, C. Kocken, R. Sauerwein, E. Riccio, J. Mirsalis, I. Bathhurst, T. Rueckle, X. Ding, B. Campo, D. Leroy, M.J. Rogers, P.K. Rathod, J.N. Burrows, S.A. Charman, A long-duration dihydroorotate dehydrogenase inhibitor (DSM265) for prevention and treatment of malaria, *Sci Transl Med* 7 (2015) 1–12. <https://doi.org/10.1126/scitranslmed.aaa6645>.

- [94] M.A. Phillips, K.L. White, S. Kokkonda, X. Deng, J. White, F. El Mazouni, K. Marsh, D.R. Tomchick, K. Manjaganagara, K.R. Rudra, G. Wirjanata, R. Noviyanti, R.N. Price, J. Marfurt, D.M. Shackleford, F.C.K. Chiu, M. Campbell, M.B. Jimenez-Diaz, S.F. Bazaga, I. Angulo-Barturen, M.S. Martinez, M. Lafuente-Monasterio, W. Kaminsky, K. Silue, A.M. Zeeman, C. Kocken, D. Leroy, B. Blasco, E. Rossignol, T. Rueckle, D. Matthews, J.N. Burrows, D. Waterson, M.J. Palmer, P.K. Rathod, S.A. Charman, A Triazolopyrimidine-Based Dihydroorotate Dehydrogenase Inhibitor with Improved Drug-like Properties for Treatment and Prevention of Malaria, *ACS Infect Dis* 2 (2016) 945–957. <https://doi.org/10.1021/acsinfecdis.6b00144>.
- [95] S. Kokkonda, X. Deng, K.L. White, J.M. Coteron, M. Marco, L. De Las Heras, J. White, F. El Mazouni, D.R. Tomchick, K. Manjaganagara, K.R. Rudra, G. Chen, J. Morizzi, E. Ryan, W. Kaminsky, D. Leroy, M.S. Martínez-Martínez, M.B. Jimenez-Diaz, S.F. Bazaga, I. Angulo-Barturen, D. Waterson, J.N. Burrows, D. Matthews, S.A. Charman, M.A. Phillips, P.K. Rathod, Tetrahydro-2-naphthyl and 2-Indanyl Triazolopyrimidines Targeting Plasmodium falciparum Dihydroorotate Dehydrogenase Display Potent and Selective Antimalarial Activity, *J Med Chem* 59 (2016) 5416–5431. <https://doi.org/10.1021/acs.jmedchem.6b00275>.