



Genetic Survey of the Most Distributed Commercial Rabbits Along The Nile in



Egypt: A Case Study by Microsatellite Markers

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Abstract

THE investigation of genetic diversity is vital for managing farm animals. Hence, the current study examined the genetic status of the most rabbit lines distributed commercially along 804 km of the surrounding villages on the Nile River. The results of survey showed that the percentages of the Verde line (VL), Hyplus line (HL), Alexandria line (AL), and New Zealand (NWZ) were 79, 70, 64, and 53%, respectively. According to previous result, 338 biological samples from previous lines (86 for VL, 84 for LA, 83 for HL, and 85 for NWZ) were collected from three populations for each rabbit line in Delta (D), Middle Egypt (M), and Upper Egypt (U) to determine the genetic variability using 26 microsatellite markers. According to results of microsatellite loci, the total number of recorded alleles was 166. Among rabbit lines, the mean number of alleles (MNa) was the lowest in NWZ (5.772) while it was highest in AL (7.154). Furthermore, the south populations (U) expressed high values of private alleles (Pa) and all populations (7, 5, 4, and 3 for AL, VL, HL, and NWZ, respectively). It was noticed that observed heterozygosity (H_o) > expected (H_e) among all lines and populations. The inbreeding coefficient (F_{IS}) recorded the highest value in NWZ (0.178) with bottleneck observation. In contrast, other lines were in negative values. Our findings show that the generated information is relevant for rabbit breeding and could be used as a guide in genetic improvement programs in Egypt.

Keywords: Commercial Rabbits, Egypt, Nile, Microsatellite, Genetic, rural areas.

Introduction

Rabbit husbandry projects meet the sustainable and alternative classifications [1]. It would likely succeed compared to other animal activity projects [2]. For farmers with little resources, it might promote food security, particularly in less developed nations (LDCs) like Cameroon, Egypt, Ghana, and Mexico [3]. The rabbit industry plays an important role in the agriculture sector in most Mediterranean countries [4]. Egypt is the third country in rabbit production by 75000 tyr^{-1} in 2021 [5]. It is considered the ideal project for improving the livelihood in rural areas for fast-growing and high productivity in rural areas [6].

The genetic variables of most farm animal species are suffering from a drastic decrease [7, 8]. According to FAO [9], 30-40% of farm animal genetic resources are necessary economically in the agriculture sector (FAGRs). The maintenance and documentation of FAGR's assistant decision-makers

in sustainable strategy design in developing countries [10]. In addition, it is considered a key to survival that could occur based on overall information regarding genetic variability and population structure [11]. Biodiversity plays an important role in the ecosystem by providing several services nutrition, water cycling, soil formation, and regulation of abiotic stress resistance [12]. Global warming affects genetically and physiologically animals, plants, and microorganisms [13]. Genetic markers are considered a wide method to identify species and to assess relationships among different populations which helps to estimate genetic status in a specific geographic area [12, 14].

Microsatellite markers are an important method for estimating genetic variability (novel mutation, genetic drift, bottleneck position, gene flow, genetic structure, and genetic distance) [15]. It contributes to enhance selection, particularly for the maintenance of

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animal breeds/ populations which ensures long-term food security [14]. This reflects on maximizing the productivity under several challenges such as changes in environmental hazards, marketing fluctuating, management, and vulnerability to certain diseases [16, 17].

Thus, the current study aims to evaluate the genetic variability, structure, and relationship among the commercial rabbit lines widely used in rabbit production along the Nile River in Egypt using 26 microsatellite markers.

Material and Methods

Rabbit survey

In this study, about 12.1 thousand bucks and does belonging to 89 commercial rabbit farms survived. These farms were in 47 rural centers along the Nile River as representative samples to determine rabbit lines distribution along the Nile in Egypt. The survey covered 3 main provinces that demonstrated 11 Egyptian governorates. They were Delta (Beheira, Monufia, Gharbia, Qalyubia, Dakahlia, Damietta, and Kafr El sheikh), Middle Egypt (Giza, Bani Suef, and Minya) and Upper Egypt (Asyut, Sohag, and Qena). Three governorates (Alexandria, Luxor, and Aswan) were excluded from the current study because the recording system was not available.

Samples Collection and DNA Extraction

A total of 338 biological samples (tissue specimens and hair bulbs) were collected from widely rabbit lines distributed in commercial farms along the Nile (Fig. 1). Rabbit samples in current study were not related, and if they were, the only relationship from one parent with only one individual was considered from the offspring. The diversity for each rabbit line was examined in 3 different populations: Delta (D), Middle Egypt (M), and Upper Egypt (U). The commercial farms that use a recording system were included in the study to guarantee the purity of breeding. The extraction of DNA from rabbit samples was carried out using the NaOH method [18].

Microsatellites DNA Markers Selection and Polymerase Chain Reaction (PCR)

Twenty-six microsatellite markers (Invitrogen, France) were classified into five multiplexes, were used in this study. The conditions of PCR reaction conditions for each multiplex were shown in Table S.1. The presence of PCR products was analyzed by a horizontal gel electrophoresis system (mini gel, Biometra®EU), using 2% agarose gel stained with ethidium bromide. The fragments size was determined using a genetic analyzer (ABI PRISM 3730 XL; Applied Biosystems).

Data Analysis

The estimated number of alleles per microsatellite marker (N_a), observed at each microsatellite marker and the mean numbers of alleles (MNa), number of private alleles (Pa), observed and expected heterozygosity (H_o and H_e) were calculated by GENAIEX 6.4.1 program [19]. The calculation of allelic richness (Ar) and estimation of the inbreeding co-efficient within populations (F_{IS}) was carried out by FSTAT 2.9.3.2 software [20]. The situation of bottlenecks was detected by BOTTLENECK software v.1.2.02 [21] by 1000 simulations as the probability distribution (Wilcoxon- $P > 0.05$) through the two-phase model (TPM) and stepwise mutation model (SMM) methods. The mode shift indicator based on the shape of the allele frequency distribution was also performed. The neighbor-joining (NJ) tree was visualized in Mega tree explorer according to the Reynolds matrix [22]. The significance of the Hardy-Weinberg equilibrium (HWE) and polymorphism information content (PIC) calculation was carried out by Cervus 3.0.6 software [23]. The population structure evaluation was based on a Bayesian clustering analysis by the STRUCTURE 2.3.4 program [24] upon independent runs using 100000 Markov Chain Monte Carlo (MCMC) iterations and a burn-in of 30000 steps and was also performed for $2 \leq K \leq 12$ (K =number of assumed clusters). Evanno methods were used to compute the statistic ΔK was computed by graphic explanation [25].

Results

Data Survey

The percentages of rabbit lines distribution at commercial levels (Fig. 2). The survey results showed that the Verde Line (VL), Hyplus Line (HL), Alexandria Line (AL), and New Zealand (NWZ) were widely distributed commercially in survived rabbit farms (79, 70, 64, and 53%, respectively). Other commercial rabbits such as Flemish Giant (FG), California (CL), and Papillon (PAP) were found in lower percentages (36, 32, and 30%, respectively). We observed that other local Egyptian lines APRI, Moshtohour (ML), Red and black Baladi was limited distribution at commercial levels with 21%.

General population statistic

The summary of genetic variability for rabbit lines along the Nile is shown in Table 1. The mean values of MNa , Pa , and Ar were 6.379, 3.33, and 4.426, respectively. Moreover, the mean value of H_e is lesser than that of H_o (0.398 and 0.557, respectively). In this regard, AL showed the highest variability, while NWZ was the lowest. The value of MNa ranged from 5.772 in the NWZ to 7.313 in the AL. The mean values of Pa and Ar ranged from 2.000 and 3.999 in the NWZ and 4.333 and 5.068 in the AL, respectively. The mean values of H_e and H_o were 0.398 and 0.557, respectively. In all rabbit

lines, U populations expressed the highest genetic variability than others (D and M).

Inbreeding coefficient and bottleneck test

As shown in Fig. 3 a, the F_{IS} values for rabbit lines appeared in the significant negative (-0.190, -0.192, and -0.211 for AL, VL, and HL, respectively) except NWZ was in the positive values (0.178). In the same context, the F_{IS} was higher in Delta populations on all lines and reduced towards the south (Fig. 3 b). In Fig. 4 (a and b) the absence of bottleneck was found in VL, AL, and HL according to Wilcoxon signed-rank tests ($P > 0.05$), while it was observed in NWZ. The mean values of T_{MP}, S_{MM}, and I_{AM} varied from 0.462, 0.298, and 0.222 to 0.323, 0.254, and 0.179 (NWZ and AL, respectively).

Genetic parameters of Microsatellite markers

A total of 166 alleles were observed in the total samples at 26 microsatellite markers (Table 2). All microsatellites showed a polymorphism across the tested lines. The values of N_a and PIC varied from 0 to 14 and 0.274 to 0.865 for INRA140 and INRA205, respectively. About 73% of microsatellite markers were highly informative ($PIC > 0.5$), while 27% represented moderately informative ($0.25 < PIC < 0.5$). Moreover, 69.2% of microsatellite markers showed non significance of HWE, whilst 30.8% showed differences at the 3 levels ($P < 0.05$; $P < 0.01$ and $P < 0.001$).

Relationships among rabbit lines and structure

As shown in Fig. 5, a neighbour-joining tree is visualized the genetic relationships based on genetic distances among lines. The results showed two main clusters: the first cluster included AL, VL, and NWZ. Figure 6 illustrated that the structure analysis results depending on Bayesian approach and the number of clusters (K). The highest values of ΔK were obtained when $k=12$ (41.82) shows the clustering pattern arising from the analysis. The number of predefined clusters varied from K2 to K12.

Discussion

The wide distribution of exotic rabbit lines commercially is due to the higher productive and reproductive performance than Egyptian lines [26]. Moreover, the current survey showed that the Alexandria line (AL) was distributed by 64%, this may be due to the high body gain in 28-63 days and adapted under the Egyptian climate [27]. Although high residence of abiotic stresses, other Egyptian rabbit lines were limited distributed for low production and reproduction traits [28].

The highest genetic variability values were recorded in the AL line. The same observation was stated in the ML was synthesized recently as AL lines (29). However, the lowest was recorded in NWZ for underwent to several selection programs

that were carried out since 80's in Egypt (30, 31). The selective intense breeding programs decreased the genetic variability in commercial breeds (32). Among all populations, we observed that $H_o > H_e$, which might be attributed to an absence of inbreeding status in most populations is an indicator of the isolate-breaking effect inside each line population [33]. Furthermore, the current study showed that the highest genetic variability in all lines was expressed in the southward direction characterized by warm weather. The warm weather motivates mutations that cause genetic variability [34-37]. In the same context, it could be explained the increase of A_r values in Upper Egypt populations could be due to mutation richness.

The F_{IS} values were found in negative values in three lines (AL, VL, and HL) which is an indication of inbreeding avoidance [38]. The values of bottleneck nearby agree with European commercial rabbits [39]. The negative values of F_{IS} with the absence of a bottleneck position concur [40].

In Table 2, the high percentage of non-significant differences in HWE confirms the large size of populations and random mating [41]. Most microsatellite markers showed high informative polymorphism (73%) which is evidence of a genetic polymorphism and linkage mapping [42].

Figure 5 shows the relationship among 4 Lines. In the first cluster, 3 Lines (AL, VL, and NWZ) were occupied. According to previous literature, AL contained 87.5% of VL and 12.5% of Black Baladi [26, 27, 43]. In addition, VL resulted after four specialized selections for the Spanish maternal A line that were synched from NWZ rabbits [44]. The second cluster included the HL line only. Because that HL line was bred from several other meat-type rabbits [45].

According to structure results (Fig.6), the stability of Upper Egypt populations was noticed. Also, the classification according to geographical areas was reported in the Algeri. We conclude that commercial rabbit farms are dependent on AL, VL, HL, and NWZ. Commercial rabbit lines along the Nile River are characterized by high genetic variability and stability with low inbreeding and the absence of genetic bottleneck in three lines (AL, VL, and HL) in contrast, NWZ is suffering from inbreeding and genetic bottleneck. In addition, these findings could be used as an initial guide to design further investigations for the improvement of rabbit lines' genetic improvement programs for commercial rabbit lines. The findings from this study can assist researchers in drawing a sustainable agriculture strategy that could contribute to the rabbit industry development in rural areas in Egypt. Also, it will meet the first and second goals of the United Nations' sustainable development agenda for 2030

(no poverty and zero hunger) which could achieve food security and improved nutrition of rabbits [36].

Conclusion

We conclude that commercial rabbit farms are dependent on AL, VL, HL, and NWZ. Commercial rabbit lines along the Nile River are characterized by high genetic variability and stability with low inbreeding and the absence of genetic bottleneck in three lines (AL, VL, and HL) in contrast, NWZ is suffering from inbreeding and genetic bottleneck. In addition, these findings could be used as an initial guide to design further investigations for the improvement of rabbit lines' genetic improvement programs for commercial rabbit lines. The findings from this study can assist researchers in drawing a sustainable agriculture strategy that could contribute to the rabbit industry development in rural areas in Egypt. Also, it will meet the first and second goals of the United Nations' sustainable development agenda

for 2030 (no poverty and zero hunger) which could achieve food security.

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Declaration of Conflict of Interest:

The authors declare that there is no conflict of interest.

Ethical Considerations

The present study has been conducted by the guidelines of the Ethics Committee of The International Animal Care and use Committee (IACUC) Faculty of science, Menoufia University, Egypt, Approval №: MUFAG/F/AP/2/21.



Fig. 1. Geographical sampling locations along the Nile River in Egypt.

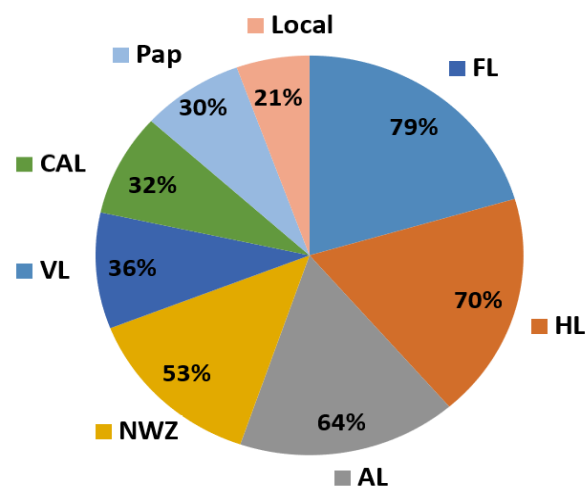


Fig. 2. The Percentages of Commercial Rabbits Distributions in Farms

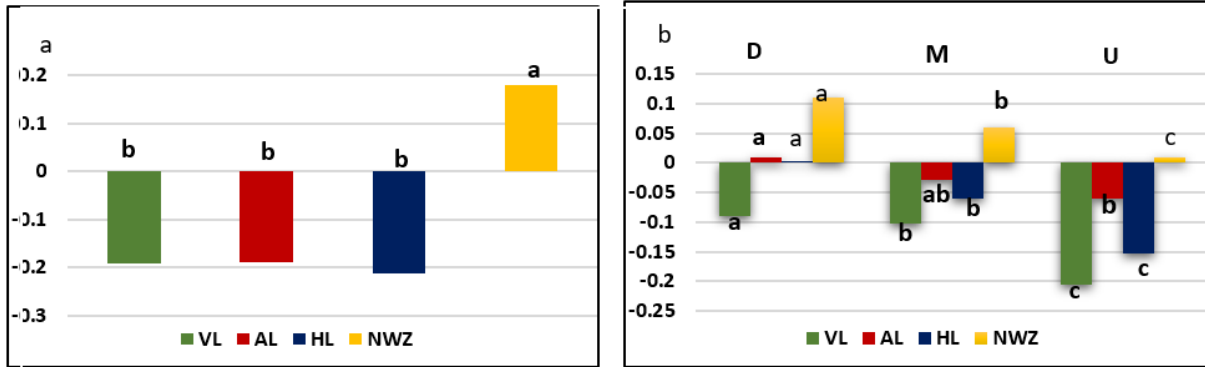


Fig. 3. The values of inbreeding coefficient (F_{IS}). a. The mean values of F_{IS} in rabbit lines. b. The comparative of F_{IS} in each geographical area. Verde Line (VL), Alexandria line (AL), Hyplus Line (HL), New Zeland (NWZ), Delta (D), Middle Egypt (M), and Upper Egypt (U).

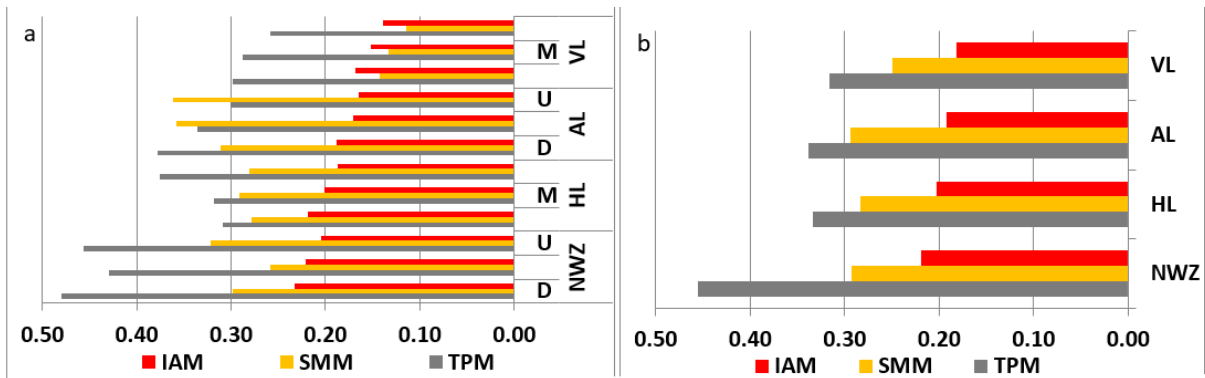


Fig. 4. a. Bottleneck test among lines, b. Bottleneck test among populations. VLine (VL), Alexandria line (AL), Hyplus Line(HL), New Zeland (NWZ), Delta (D), Middle Egypt (M), Upper Egypt (U), Infinite Allele Model (IAM)Two phase model (TMP), Step wise mutation model (SMM).

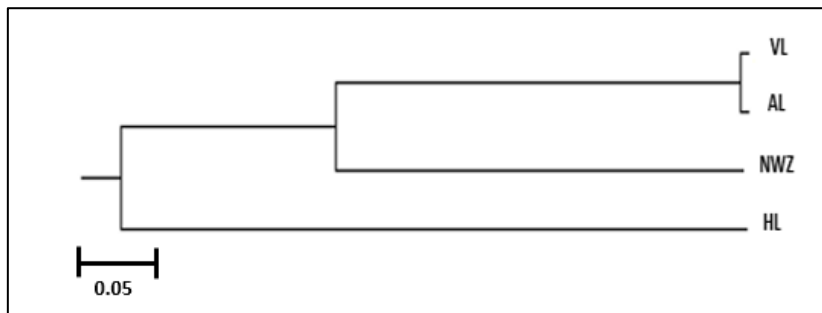


Fig. 5. Neighbor-joining tree for the rabbit populations. Verdi Line (VL), Alexandria line (AL), Hyplus Line(HL), New Zeland (NWZ).

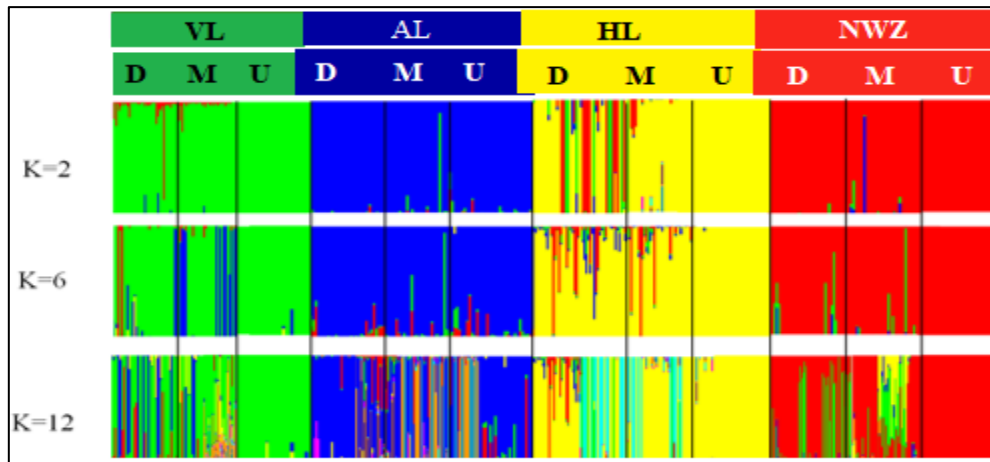


Fig. 6. Individual assignment of domestic rabbits inferred with Bayesian cluster analysis (STRUCTURE). Verde Line (V L), Alexandria line (AL), Hyplus Line (HL), New Zeland (NWZ), Delta (D), Middle Egypt (M), and Upper Egypt (U).

TABLE 1. Genetic variability for the most rabbit lines commercially distribution along the Nile River in Egypt.

Line	Population	N	MNa± SD	Pa	Ar± SD	H _e ± SD	H _o ± SD
VL	D	30	5.885 ±0.274	2	3.711 ±0.220	0.350 ±0.033	0.510 ±0.025
	M	25	6.000 ±0.200	4	3.982±0.194	0.380 ±0.033	0.529 ±0.027
	U	31	6.231 ±0.339	5	4.193 ±0.204	0.385 ±0.030	0.582 ±0.032
	Mean Values		6.038 ±0.271	3.667	3.962±0.366	0.372 ±0.032	0.540 ±0.028
AL	D	32	7.253 ±0.330	2	5.866 ±0.361	0.435 ±0.027	0.604 ±0.030
	M	25	7.196 ±0.304	4	4.365±0.301	0.440 ±0.035	0.604 ±0.030
	U	27	7.492 ±0.460	7	4.981 ±0.307	0.445 ±0.042	0.578 ±0.035
	Mean Values		7.154 ±0.364	4.333	5.068±0.523	0.442 ±0.034	0.595 ±0.035
HL	D	31	6.462 ±0.237	3	4.698±0.303	0.445 ±0.042	0.588 ±0.035
	M	24	6.557 ±0.278	3	4.512 ±0.333	0.363±0.033	0.561 ±0.033
	U	28	6.642 ±0.304	4	4.82 2±0.393	0.386 ±0.031	0.602 ±0.029
	Mean Values		6.553 ±0.273	3.333	4.677±0.443	0.398 ±0.035	0.583 ±0.032
NWZ	D	28	5.145 ±0.190	1	3.391 ±0.239	0.350 ±0.040	0.397 ±0.034
	M	27	5.885 ±0.224	2	3.696 ±0.255	0.380 ±0.037	0.595 ±0.023
	U	30	6.308 ±0.336	3	4.392 ±0.301	0.355 ±0.044	0.584 ±0.033
	Mean Values		5.772 ±0.250	2.000	3.999 ±0.265	0.379 ±0.046	0.509 ±0.033
Overall mean			6.379±0.284	3.333	4.426±0.301	0.398±0.033	0.557±0.038

Verde Line (VL), Alexandria line (AL), Hyplus Line (HL), New Zealand (NWZ), Delta (D), Middle Egypt (M), Upper Egypt (U), Number of samples (N), Mean number of observed alleles (MNa), Stander deviation (SD), number of private alleles (Pa), Allelic richness (Ar) mean observed and expected heterozygosity (H_e and H_o).

TABLE 2. The characteristics of 26 microsatellite markers used for rabbit lines along the Nile.

Multiplex	Microsatellite loci	Na	Allele range	H _e ± SD	H _o ± SD	Mean PIC	HWE
1	INRA106	6	122-134	0.353 ±0.039	0.582 ±0.028	0.645	Ns
	INRA176	6	227-240	0.118 ±0.022	0.323 ±0.064	0.388	***
	INRA203	6	204-216	0.357 ±0.053	0.630 ±0.036	0.650	Ns
2	INRA087	12	190-216	0.596 ±0.073	0.725 ±0.051	0.852	Ns
	INRA119	2	229-243	0.342 ±0.054	0.486 ±0.048	0.518	*
	INRA140	0	183-189	0.458 ±0.036	0.490 ±0.023	0.274	Ns
	INRA157	7	138-144	0.401 ±0.033	0.585 ±0.039	0.609	***
	INRA190	1	200-210	0.126 ±0.034	0.365 ±0.031	0.455	Ns
	INRA201	9	133-143	0.371 ±0.032	0.512 ±0.039	0.498	Ns
	SAT12	6	144-164	0.276 ±0.037	0.584 ±0.028	0.582	***

Multiplex	Microsatellite loci	Na	Allele range	H _e ± SD	H _o ± SD	Mean PIC	HWE
3	SAT 03	10	146-162	0.508 ±0.037	0.595 ±0.027	0.687	***
	SAT 04	8	195-241	0.426 ±0.054	0.642 ±0.051	0.701	Ns
	SAT 05	7	206-232	0.338 ±0.044	0.505 ±0.047	0.501	Ns
	SAT 07	7	184-194	0.388 ±0.044	0.573 ±0.041	0.675	Ns
	SAT 08	3	136-156	0.309 ±0.024	0.470 ±0.036	0.471	**
4	INRA102	6	128-136	0.542 ±0.033	0.669 ±0.037	0.610	Ns
	INRA104	7	117-127	0.410 ±0.028	0.606 ±0.036	0.701	Ns
	INRA169	8	174-180	0.494 ±0.049	0.623 ±0.048	0.699	Ns
	INRA185	5	166-175	0.366 ±0.045	0.420 ±0.094	0.438	Ns
	INRA192	8	114-132	0.607 ±0.029	0.643 ±0.026	0.699	***
	INRA205	14	178-190	0.566 ±0.044	0.719 ±0.037	0.865	***
	INRA228	5	228-232	0.356 ±0.063	0.415 ±0.052	0.423	Ns
	SAT13	4	114-126	0.411 ±0.029	0.530 ±0.036	0.575	Ns
5	INRA259	6	173-195	0.518 ±0.054	0.609 ±0.039	0.666	Ns
	INRA313	7	239-275	0.450 ±0.026	0.619 ±0.020	0.695	Ns
	INRA342	6	170-196	0.458 ±0.046	0.581 ±0.043	0.594	Ns

Number of observed alleles (Na), mean observed and expected heterozygosity (H_o and H_e), Stander deviation (SD), mean PIC, mean polymorphism information content per microsatellite marker and Hardy-Weinberg Equilibrium (HWE). *P<0.05; **P<0.01, *** P<0.001, NS: non-significant.

TABLE S1. Basic information of microsatellite markers and PCR reaction

Multi-plex	Locus	Accession №	Repeat Pattern	Chromo-some	Temperature	Time	Cycles
1	INRA106	AJ874448	(CA) ₁₄	13	95	15 m	
	INRA176	AJ874514	(TC) ₁₁ (TG) ₁₃ AG(TG) ₃	12	95	20 s	7X
	INRA203	AJ874540	(GT) ₁₆	3	63 - 56 (↓10)	1 min	
					72	45 s	
					95	30 s	35X
					55	30 s	
					72	45 s	
					60	15 m	
2	INRA087	AJ874430	(TG) ₁₄	8	95	15 m	
	INRA119	AJ874461	(GT) ₁₆	18	95	30 s	7X
	INRA140	AJ874480	(TG) ₁₄	14	61 - 55 (↓10)	45 s	
	INRA157	AJ874497	(GT) ₁₂	8	72	45 s	
	INRA190	AJ874528	(TG) ₁₂	18	95	30 s	37X
	INRA201	AJ874538	(TG) ₁₄ (AG) ₁₀	12	55	45 s	
	SAT12	X99891	(CTAT) ₁₀	7	72	1 min	
				60	20 m		
				10	∞		
3	SAT 03	J03744	(TC) ₂₂	3	95	15 min	
	SAT 04	M33582	(TC) ₁₃ (N) ₂ (TC) ₂ TG(TC) ₇	15	95	30 sec	32x
	SAT 05	X99887	(TC) ₂₃ TT(CT) ₅	3	60	30 sec	
	SAT 07	X99889	(CT) ₁₄ (GT) ₈ TT(GT) ₅	17	72	45 sec	8x
	SAT 08	X99888	(TG) ₁₄	10	95	30 sec	
					53	30 sec	
					72	45 sec	
					60	20 min	
				12	∞		

Multi-plex	Locus	Accession №	Repeat Pattern	Chromosome	Temperature	Time	Cycles
4	INRA102	AJ874444	(AC) ₁₈	19	95	15 m	
	INRA104	AJ874448	(GT) ₁₆	108	54	45 s	
	INRA169	AJ874508	(CA) ₁₇	1	95	30 s	8x
	INRA185	AJ874523	(AC) ₁₃	4	72	45 s	
	INRA192	AJ874530	(TG) ₁₁	96	95	30 s	31x
	INRA205	AJ874542	(TG) ₁₇	4	60	15 m	60
	INRA228	AJ874561	(TG) ₁₂	-			
	SAT13	X99892	(GT) ₁₃	1			
5	INRA259	AJ874589	(GT) ₁₄ (GA) ₉	1	95	15 m	
	INRA313	AJ874634	(TG) ₇ (AC) ₁₀ GC(AC) ₆ GC(AC) ₇	14	95	30 s	8X
	INRA342	AJ874659	(GA) ₂₂	13	63 - 56 (↓10)	45 s	
					72	45 s	
					95	30 s	28X
					56	30 s	
					72	45 s	
60	20 m						
10	∞						

References

- Siddiqui, S.A., Gerini, F., Ikram, A., Saeed, F., Feng, X. and Chen, Y. Rabbit meat production, consumption and consumers' attitudes and behavior. *Sustainability*, **15** (3), 1 (2023).
- Crovato, S., Pinto, A., Di Martino, G., Mascarello, G., Rizzoli, V., Marcolin, S. and Ravarotto, L. Purchasing habits, sustainability perceptions, and welfare concerns of Italian consumers regarding rabbit meat. *Foods*, **11** (9), 1205 (2022).
- Lukefahr, S. Sustainable and alternative systems of rabbit production. Proceedings of the 8th world rabbit congress, Puebla, Mexico, 7-10 (2004).
- Mancin, E., Sosa-Madrid, B.S., Blasco, A. and Ibáñez-Escriche, N. Genotype Imputation to Improve the Cost-Efficiency of Genomic Selection in Rabbits. *Animals*, **11**, 803 (2021).
- Helgi-Library. Which country produces the most rabbit meat?. <https://www.helgilibrary.com/charts/which-country-produces-the-most-rabbit> (Aug-28), Web-Page, 1 (2024).
- Ziege, M., Babitsch, D., Brix, M., Kriesten S., Straskraba, S., Wenninger, S., Wronski, T. and Plath, M.. Extended diurnal activity patterns of European rabbits along a rural-to-urban gradient. *Mammalian Biology*, **81** (5), 534 (2016).
- Quigley, K.M., Bay, L.K. and van Oppen, M.J. Genome- wide SNP analysis reveals an increase in adaptive genetic variation through selective breeding of coral. *Molecular Ecology*, **29** (12), 2176 (2020).
- Sharma, R., Ahlawat, S., Sehrawat, R., Aggarwal, R., Chandran, P., Kamal, R. K., Dey, A. and Tantia, M. Morphometric characteristics and microsatellite markers based diversity and differentiation recognizes the first prospective cattle breed from the Jharkhand state of India. *Animal Biotech.*, **34** (7), 2017 (2023).
- FAO. Biodiversity and the livestock sector Guidelines for quantitative assessment, FAO (1), 1 (2019).
- Nery, L.M., da Cunha e Silva, D.C. and Sabonaro, D.Z. Agriculture technology transfer: A multicriteria analysis for decision making. *Environment, Development and Sustainability*, **26** (6), 155 (2024).
- Sheriff, O. and Alemayehu, K. Genetic diversity studies using microsatellite markers and their contribution in supporting sustainable sheep breeding programs: A review. *Cogent Food & Agriculture*, **4** (1), 145 (2018).
- Sadguru, P. and Seema, S. Impact of Climate Change on Biodiversity: An Overview. *International Journal of Biological Innovations*, (2), 60 (2019).
- Verma, A. K. Sustainable development and environmental ethics. *International Journal on Environmental Sciences*, **10** (1), 1 (2019).
- D'ambrosio, J., Phocas, F., Haffray, P., Bestin, A., Brard-Fudulea, S., Poncet, C., Quillet, E., Dechamp, N., Fraslin, C. and Charles, M. Genome-wide estimates of genetic diversity, inbreeding and effective size of experimental and commercial rainbow trout lines undergoing selective breeding. *Genetics Selection Evolution*, **51**, 1 (2019).
- Piro, M., Mabsoute, F., El Khattaby, N., Laghouaouta, H. and Boujenane, I. Genetic variability of dromedary camel populations based on microsatellite markers. *Animal*, **14** (12), 2452 (2020).
- Verma, N., Sharma, R., Aggarwal, R. and Dangi, P. Evaluation of Genetic Diversity in Goats of Telangana and Andhra Pradesh States of India. *Indian Journal of Animal Research*, **56** (5), 536 (2022).
- De Kort, H., Prunier, J.G., Ducatez, S., Honnay, O., Baguette, M., Stevens, V. M. and Blanchet, S. Life

- history, climate and biogeography interactively affect worldwide genetic diversity of plant and animal populations. *Nature Communications*, **12** (1), 516 (2021).
18. Cinelli, P., Rettich, A., Seifert, B., Bürki, K. and Arras, M. Comparative analysis and physiological impact of different tissue biopsy methodologies used for the genotyping of laboratory mice. *Laboratory Animals*, **41** (2), 174 (2007).
 19. Smouse, R.P.P. and Peakall, R. GenAlEx 6.5: genetic analysis in Excel. Population genetic software for teaching and research. *Bioinformatics*, **28** (19), 2537 (2012).
 20. Goudet, J. Fst v. 2.9.3.2.: A computer program to calculate F-statistics. *J. Hered*, (2002).
 21. Piry, S., Luikart, G. and Cornuet, J. M. Bottleneck: a computer program for detecting recent reductions in the effective population size using allele frequency data, *Journal of Heredity*, **90** (4), 502 (1999).
 22. Tamura, K., Stecher, G., Peterson, D., Filipowski, A. and Kumar, S. MEGA6: molecular evolutionary genetics analysis ver 6. *J. Molecular Biology Evolution*, **30** (12), 272 (2013).
 23. Kalinowski, S. T., Taper, M. L. and Marshall, T. C. Revising how the computer program accommodates genotyping error increases success in paternity assignment. *Molecular Ecology*, **16** (5), 1099 (2007).
 24. Pritchard, J.K., Stephens, M. and Donnelly, P. Inference of population structure using multilocus genotype data. *J. Genetics*, **155** (2), 945 (2000).
 25. Evanno, G., Regnaut, S. and Goudet, J. Detecting the number of clusters of individuals using the software: a simulation study. *Molecular Ecology*, **14** (8), 2611 (2005).
 26. Youssef, Y., Emam, A. and Abou-Khadiga, G. Rabbit Breeding Situation in Egypt-A Case Study. World Rabbit Science Association, 12th World Rabbit Congress, Nov. 3-5- Nantes, France, Communication, 1 (2021).
 27. Khalil, M. and Al-Saef, A. Methods, criteria, techniques and genetic responses for rabbit selection: a review. Proc 9th World Rabbit Congress, 1 (2008).
 28. Mostafa, A.R., Emam, A.M., Dorina, M., Mohamed, S., Ayman, A. and Monica, M. Rabbits Meat Production in Egypt and its Impact on Food Security, Small Holders Income and Economy. *J. Agricultural Research Technology*, **24** (1), 81 (2020).
 29. El-Aksher, S.H., Sherif, H., Khalil, M., El-Garhy, H.A. and Ramadan, S. Molecular analysis of a new synthetic rabbit line and their parental populations using microsatellite and SNP markers. *Gene Reports*, **8**, 17 (2017).
 30. Khalil, M., Owen, J. and Afifi, E. A review of phenotypic and genetic parameters associated with meat production traits in rabbits, *Biological Science*, **54** (9), 725 (1986).
 31. Elfadil, G., Gouda, G., Rashed, M. and Shemeis, A. Selection indexes for efficient meat production capacity from New Zealand white rabbits. *Advances in Animal and Vet Sciences*, **11** (9), 1557 (2023).
 32. Justinski, C., Wilkens, J. and Distl, O. Genetic diversity and trends of ancestral and New inbreeding in German sheep breeds by pedigree data. *Animals*, **13** (4), 623 (2023).
 33. Sudina, K., Anjaly, K., Arun, R. and Jayadevi, V.E. Genetic diversity analysis of class II MHC in indigenous poultry-a comparison with commercial breed, *J. Indian Vet. Assoc.*, **20**(2), 36 (2022).
 34. Teixeira, J. C. and Huber, C.D. The inflated significance of neutral genetic diversity in conservation genetics. *Proce. of the National Academy of Sciences*, **118** (10), e2015096118 (2021).
 35. Hasan, M.U., Kozakli, Ö. and Ceyhan, A. Possibilities of using CRISPR-based genome editing Technologies in Livestock. *J. of Agriculture, Food, Environment and Animal Sciences*, **3** (1), 56 (2022).
 36. Bouhali, A., Homrani, A., Ferrand, N., Lopes, S. and Emam, A.M. Assessment of genetic diversity among native Algerian rabbit populations using microsatellite markers. *Archives Animal Breeding*, **66** (3), 207 (2023).
 37. Emam, A.M., Mourad, R.S. and Faid-Allah, E. Egyptian Native Rabbits Along The Nile River: A Microsatellite Marker-Based Genetic Field Study. *Journal of Applied Molecular Biology (JAMB)*, **2** (2), 127 (2024).
 38. Mihailova, Y., Rusanov, K., Rusanova, M., Vassileva, P., Atanassov, I., Nikolov, V. and Todorovska E. G. Genetic diversity and population structure of Bulgarian autochthonous sheep breeds revealed by microsatellite analysis. *Animals*, **13** (11), 1878 (2023).
 39. Ziege, M., Theodorou, P., Jüngling, H., Merker, S., Plath, M., Streit, B. and Lerp, H. Population genetics of the European rabbit along a rural-to-urban gradient. *Scientific Reports*, **10** (1), 2448 (2020).
 40. Radhika, G., Aravindakshan, T., Anilkumar, K., Manoj, M. and Thomas, S. Genetic diversity analysis of cattle genetic groups of Kerala state using microsatellite data. *Animal Biotechnology*, **34** (4), 1154 (2023).
 41. Adeolu, A.I., Wheto, M., Oleforuh-Okoleh, V.U., Nwose, R. N., Adenaike, A. S., Yakubu, A., Abiola, E. M. and Mohammed B. G. Genetic Diversity of Rabbit (*Oryctolagus cuniculus*) Population in South Eastern Nigeria Using Microsatellite Markers. *Tropical Animal Science Journal*, **44** (3), 280 (2021).
 42. Shihabi, M. Inbreeding and selection on The X-chromosome in domestic animal populations. *PhD Thesis*, Zagreb Univ., Faculty of Agriculture, (2024).
 43. El-Raffa, A. Formation of a rabbit synthetic line (Alexandria line) and primary analysis of its productive and reproductive performance. *Egypt Poult. Sci.*, **27**(2), 321 (2007).
 44. Estany, J., Baselga, M., Blasco, A. and Camacho, J. Mixed model methodology for the estimation of genetic response to selection in litter size of rabbits. *Livestock Production Science*, **21**(1), 67 (1989).
 45. Vostrý, L., Mach, K., Jakubec, V., Dokoupilová, A. and Majzlík, I. The influence of weaning weight on growth of the hyplus broiler rabbit. *Proceeding of the 9th World Rabbit Congress Verona, Italy*, 255 (2008).

المسح الوراثي للأرانب التجارية الأكثر انتشارا على طول نهر النيل في مصر: دراسة حالة باستخدام الواسمات الوراثية

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الملخص

تعد دراسة التنوع الوراثي أمر هام لإدارة الحيوانات المزارعية. الدراسة الحالية اختبرت الوضع الوراثي لخطوط الأرانب الأكثر توزيعا تجاريا على طول 804 كم من القرى المحيطة على نهر النيل. وأظهرت نتائج المسح أن النسب المئوية لانتشار خط الفي لاين (VL) وخط الهاي بلاس (HL) وخط الاسكندرية (AL) و الأرانب النيوزيلندي (NWZ) كانت 79 و 70 و 64 و 53% على التوالي. ووفقا للنتيجة السابقة، تم جمع 338 عينة بيولوجية من الخطوط السابقة (86 لـ VL و 84 لـ LA و 83 لـ HL و 85 لـ NWZ) من ثلاث عشائر لكل خط في الدلتا (D) ومصر الوسطى (M) وصعيد مصر (U) لتحديد التباين الوراثي باستخدام 26 واسم وراثي (ميكروساتلايت). وفقا لنتائج الواسمات الوراثية، بلغ العدد الإجمالي للأليلات 166. ومن بين خطوط الأرانب، كان متوسط عدد الأليلات (MNa) هو الأدنى في ال NWZ (5.772) بينما كانت الأعلى في ال AL (7.154). علاوة على ذلك، عبرت عشائر الجنوب (U) عن قيم عالية للأليلات الخاصة (Pa) لجميع الخطوط (7 و 5 و 4 و 3 لـ AL و VL و HL و NWZ، على التوالي). وقد لوحظ أن التباين الريبوسومي الملحوظ (H_o) المتوقع (H_e) بين جميع الخطوط والعشائر. وسجل معامل التزاوج الداخلي (F_{IS}) أعلى قيمة في NWZ (0.178) مع ملاحظة وضع عنق الزجاجة الوراثي. وعلى النقيض من ذلك، كانت السلالات الأخرى بقيم سلبية. وتُظهر نتائجنا أن المعلومات الناتجة ذات صلة بتربية الأرانب ويمكن استخدامها كدليل في برامج التحسين الوراثي في مصر.

الكلمات الدالة: الأرانب التجارية، مصر، النيل، الواسمات الوراثية، المناطق الريفية.