

Detection of bactericidal activity of camel's milk compared with raw and processed cow's milk against pathogenic bacteria

Amal S. Othman

Department of Microbiology, Faculty of Applied Medical Science, October 6th University, 6th of October City, Egypt

Correspondence to Amal S. Othman, PhD., Central Axis, Part 1/1, 12588 Elsheikh Zayed, 6th of October City, Egypt
Tel: +20 100 622 8919;
e-mail: amalothman_1976@yahoo.com

Received 20 October 2015

Accepted 26 October 2015

Egyptian Pharmaceutical Journal

2016, 15:31–37

Objective

The aim of this study was to determine the antimicrobial activity of camel's milk compared with three types of raw and processed cow's milk as a natural safe way to overcome pathogenic bacteria instead of using chemotherapy, which leads to the phenomenon of microbial resistance.

Materials and methods

A total of 16 milk samples were collected; four samples from four healthy camels (4 years old), four fresh cow milk samples, four pasteurized milk samples, and four packed buttermilk samples. The camel's and cow's milk were boiled before the investigation. Bacterial isolation from these samples was carried out on specific media. The antibacterial activity for each milk type was assessed against seven Gram-positive and Gram-negative bacterial strains previously examined for its multidrug resistance activity. The minimal inhibitory concentration and minimal bactericidal concentration of each milk type was also determined. Transmission electron microscope was used for the highly affected bacterial strain for each of the milk types.

Results and conclusion

The four milk sample types were free from bacterial contamination. They all possessed antimicrobial activity but not for all seven examined bacterial strains. The strains affected were *Escherichia coli* and *Klebsiella pneumoniae* by camel's milk and *E. coli* and *Streptococcus faecalis* by the other three cow's milk types. Pasteurized milk and buttermilk showed the higher effects. Minimal inhibitory concentration and minimal bactericidal concentration for the milk types ranged between 10 and 80%. Transmission electron microscope studies on the bacteria affected revealed damage in bacterial cell wall and disturbance in cell protein content. It can be concluded that milk can be used *in vitro* as a natural safe way to overcome some pathogenic bacteria instead of using antibiotics.

Keywords:

antimicrobial activity, buttermilk, camel's milk, minimal inhibitory concentration, pasteurized, transmission electron microscope

Egypt Pharm J 15:31–37

© 2016 Division of Pharmaceutical and Drug Industries Research, National Research Centre 1687-4315

Introduction

The milk of mammals is protected to different extents against microbial contaminations by natural inhibitory systems, including the lactoperoxidase/thiocyanate/hydrogen peroxide system, lactoferrins, lysozyme, immunoglobulins, and free fatty acids [1–3]. The concentration and the activity of each of these antimicrobial systems/substances depend on the animal species and on the stage of lactation [4].

In particular, the levels of lysozyme and lactoferrins in camel milk are reported to be two and three times higher than those in cow's milk [2,5]. Camel milk can be kept for longer periods compared with cow's milk when refrigerated, and even with the desert heat it does not spoil shortly [6].

Camel milk also has valuable nutritional properties as it contains a high proportion of antibacterial substances and a higher concentration of vitamin C in comparison with cow milk [7,8]. In average, camel milk contains more proteins and whey protein compared with cow milk [9].

The ability of camel milk to inhibit the growth of pathogenic bacteria and its relation to whey lysozyme has been demonstrated in previous study [10].

Pasteurization of milk has been practiced as the most effective method of reducing the risk for contamination and spreading of disease. Although pasteurized milk is expected to have a shelf life of 14–20 days, the shelf life of pasteurized milk stored at ambient temperature is dependent upon the efficiency of pasteurization process [11].

This study was conducted to examine the purity of boiled raw camel's and cow's milk, pasteurized cow's milk, and packed buttermilk from bacterial contaminants and to determine the antimicrobial

This is an open access article distributed under the terms of the Creative Commons Attribution-NonCommercial-ShareAlike 3.0 License, which allows others to remix, tweak, and build upon the work non-commercially, as long as the author is credited and the new creations are licensed under the identical terms.

activity of these types of milk, which is considered as a natural safe way to overcome pathogenic bacteria instead of using chemotherapy, which leads to the phenomenon of microbial resistance.

Materials and methods

Milk sample preparation

Camel's milk

Camel's milk samples were collected early in the morning from camel farm in Kerdasa, Giza, Egypt. Milk was collected from four healthy camels (4 years old) by means of hand milking in sterile screw bottles and kept in cool boxes until transported to the laboratory.

Cow's milk

Raw, pasteurized, and packed buttermilk samples were collected from a dairy shop, Giza, Egypt. Milk was collected (four samples each) in sterile screw bottles and also kept in cool boxes until transported to the laboratory.

Both camel's milk and raw cow's milk were boiled at 100°C for 10 min before usage.

Isolation and identification of bacteria from the examined milk types

Isolation and identification of Gram-negative bacterial pathogens were carried out following aseptic sampling techniques [12,13]. Briefly, a loopful (0.01 ml) of milk sample was streaked on 7% blood agar (base blood agar; Oxoid, Germany) and incubated aerobically at 37°C. The plates were checked for bacterial growth after 24, 48, and 72 h to rule out slow-growing microorganisms, and subcultured on blood agar at 37°C for 24 h to obtain pure culture. A single colony from a pure culture was then subjected to Gram's stain to observe morphological characteristics and transferred to brain heart infusion and MacConkey agar to be grown for further analysis. Identification of bacteria to the species level was carried out using a conventional method of biochemical reaction.

Bacterial strains

The following bacterial strains were used (ATCC, USA): *Pseudomonas aeruginosa* (ATCC: 27853), *Proteus vulgaris* (ATCC: 13315), *Salmonella typhi* (ATCC: 14028), *Escherichia coli* (ATCC: 25922), *Klebsiella pneumoniae* (ATCC: 13883), *Staphylococcus aureus* (ATCC: 25923), and *Streptococcus faecalis* (ATCC: 29212). They were subcultured on Nutrient agar (Lab M, UK) and incubated aerobically at 37°C. The organisms were maintained in the laboratory on nutrient agar slopes at 4°C [14].

Antimicrobial susceptibility test

Antimicrobial susceptibility was assessed using the disk diffusion method, and the results were interpreted using the Clinical and Laboratory Standards Institute (formerly National Committee for Clinical Laboratory Standards, NCCLS) break point criteria [15]. Antimicrobial drugs included the penicillin group (amoxicillin 30 µg), the glycopeptide group (vancomycin 30 µg), aminoglycosides (amikacin 30 µg), cephalosporin (cephradine 30 µg), and carbapenem (imipenem 10 µg). Multidrug resistant strains were detected and defined as strains that were nonsusceptible to at least one agent in three or more antimicrobial with group [16].

Screening for the milk's type antibacterial activity

Antibacterial activity was tested using the agar well diffusion method on nutrient agar media using milk dilutions (10–100%). The wells were made using a sterile borer and were filled with 0.45 µl of each concentration. The antibacterial assay plates were incubated at 37°C for 24 h. The diameter of the zones of inhibition around each well was taken as a measure of the antibacterial activity.

Each experiment was carried out in triplicate and the mean diameter of the inhibition zone was recorded [17].

Minimal inhibitory concentration and minimal bactericidal concentration

The minimal inhibitory concentration (MIC) was determined as the lowest concentration of an antimicrobial agent that prevents visible growth of a microorganism in a broth dilution susceptibility test. The minimal bactericidal concentration (MBC) was determined, after determining the results for the MIC, as the lowest concentration that achieved a 99.9% decrease in viable bacteria. The MBC can be determined from broth dilution MIC tests by subculturing on agar medium without disinfectant and incubating at 35°C for 16–20 h according to the macrodilution method described by the NCCLS [18]. The experiments were conducted in triplicate.

Transmission electron microscope examination

Conventional transmission electron microscope (TEM) is frequently selected to visualize the ultrastructural damage on both cell wall and cytoplasmic membrane of entire microbes [19].

At ultrastructural level, a simple negative staining for TEM of bacterial cells can report evidence on the mechanism of membrane disruption by antimicrobial proteins and peptides [20]. The highly affected bacterial

All concentrations of pasteurized milk affected *St. faecalis* and *E. coli* (Table 4). The 30% concentration was the highly effective concentration on both organisms.

All concentrations of buttermilk affected *St. faecalis* and *E. coli*, especially at concentrations 20 and 90%, respectively (Table 5).

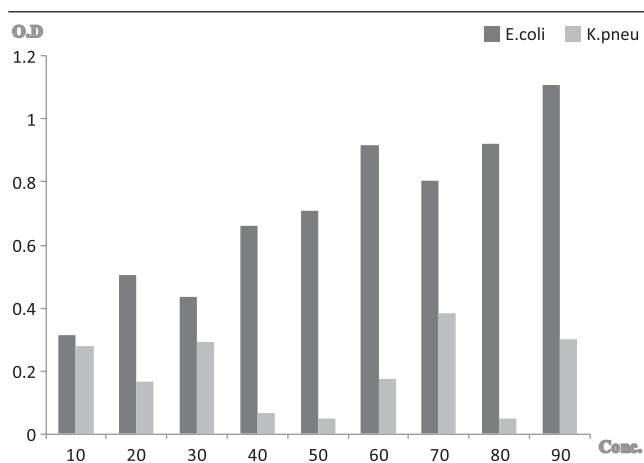
From Table 2, it is clear that camel's milk affected *E. coli* and *K. pneumoniae*. The growth rate of both organisms were also measured using a spectrophotometer (optical density), and then the MIC and MBC were determined for each of them (Fig. 1).

K. pneumoniae was highly affected by camel's milk compared with *E. coli* (Fig. 1). The 10% concentration

was the MIC of camel's milk to *E. coli*, whereas it was 80% for *K. pneumoniae*. To determine the MBC for both of the MIC, they were subcultured on nutrient broth media and it was found that the MBC was 10% for camel's milk to *E. coli* and 80% for *K. pneumoniae*.

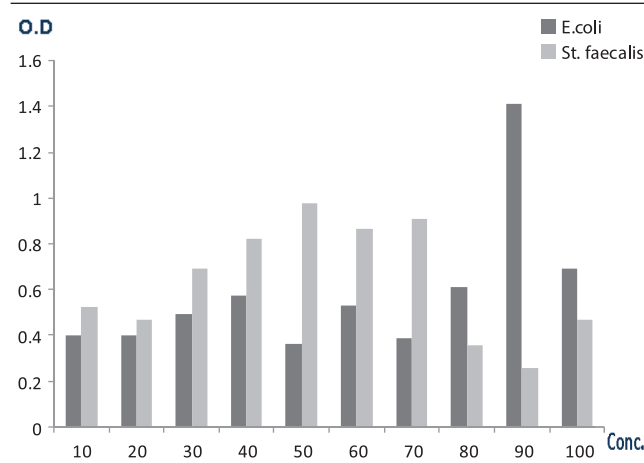
Raw cow's milk, pasteurized milk, and packed buttermilk affected only *E. coli* and *St. faecalis*, and so they were chosen to investigate the effect of these three milk types on them measured using a spectrophotometer (optical density), and the MIC and MBC for both of them were also determined (Tables 3–5 and Figs 2–4).

Figure 1



Determination of the effect of camel's milk on *Escherichia coli* and *Klebsiella pneumoniae* at different concentrations to detect the minimal inhibitory concentration. OD, optical density.

Figure 2



Determination of the effect of raw cow's milk on *Escherichia coli* and *Streptococcus faecalis* at different concentrations to detect the minimal inhibitory concentration. OD, optical density.

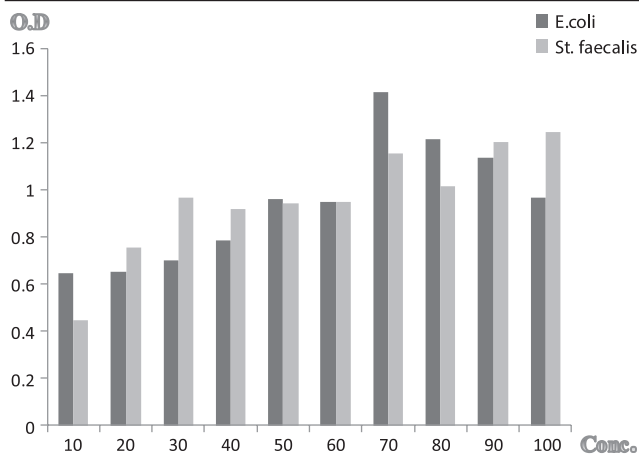
Table 4 Determination of the effect of pasteurized milk on the tested bacterial strains

Bacterial strains	Milk dilutions [inhibition zone diameter (mm)]									
	10%	20%	30%	40%	50%	60%	70%	80%	90%	100%
<i>Pseudomonas aeruginosa</i>	00	00	00	00	00	00	00	00	00	00
<i>Proteus vulgaris</i>	00	00	00	00	00	00	00	00	00	00
<i>Salmonella typhi</i>	00	00	00	00	00	00	00	00	00	00
<i>Escherichia coli</i>	25	21	32	30	26	24	24	22	22	30
<i>Klebsiella pneumonia</i>	00	00	00	00	00	00	00	00	00	00
<i>Staphylococcus aureus</i>	00	00	00	00	00	00	00	00	00	00
<i>Streptococcus faecalis</i>	21	18	24	11	14	15	14	16	13	13

Table 5 Determination of the effect of packed buttermilk on the tested bacterial strains

Bacterial strains	Milk dilutions [inhibition zone diameter (mm)]									
	10%	20%	30%	40%	50%	60%	70%	80%	90%	100%
<i>Pseudomonas aeruginosa</i>	00	00	00	00	00	00	00	00	00	00
<i>Proteus vulgaris</i>	00	00	00	00	00	00	00	00	00	00
<i>Salmonella typhi</i>	00	00	00	00	00	00	00	00	00	00
<i>Escherichia coli</i>	23	20	23	23	20	24	22	22	25	24
<i>Klebsiella pneumonia</i>	00	00	00	00	00	00	00	00	00	00
<i>Staphylococcus aureus</i>	00	00	00	00	00	00	00	00	00	00
<i>Streptococcus faecalis</i>	19	22	15	20	16	18	20	18	12	12

Figure 3



Determination of the effect of pasteurized milk on *Escherichia coli* and *Streptococcus faecalis* at different concentrations to detect the minimal inhibitory concentration. OD, optical density.

E. coli was the highly affected strain compared with *St. faecalis* except at high concentrations of 70–100% (Fig. 2). MIC of pasteurized milk on both *E. coli* and *St. faecalis* was found to be 10 and 80%, respectively, and they were detected as the MBC also for both organisms.

The effect of pasteurized milk on both *E. coli* and *St. faecalis* was nearly the same (Fig. 3). The MIC and MBC for both of them were found to be 10%.

The effect of packed buttermilk on both *E. coli* and *St. faecalis* was nearly the same (Fig. 4). MIC and MBC were found to be 10% for both organisms.

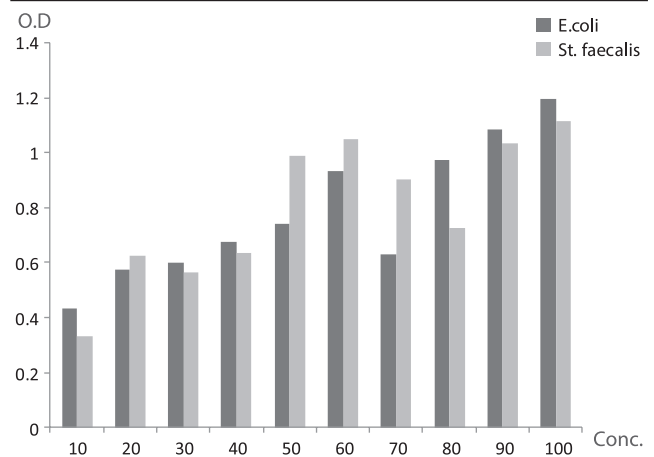
E. coli was chosen as one of the strains affected by the three cow milk types. It was scanned using TEM to see the effect of each milk type on the bacterial cell structure. *K. pneumoniae* was also chosen as one of the affected strains to be scanned before and after treatment with camel's milk (Fig. 5).

There was high effect of different milk types on internal and cell wall structure of both scanned organisms (Fig. 5).

Discussion

There is an urgent need to find new antimicrobials to treat bacterial pathogens. It is generally well established that the food constituents can be used to reduce the risk of developing or aggravating human disease conditions. In this regard, functional foods had emerged as adjuvant or alternative to chemotherapy, especially in the prevention and management of human diseases and for maintaining optimum health

Figure 4



Determination of the effect of packed buttermilk on *Escherichia coli* and *Streptococcus faecalis* at different concentrations to detect the minimal inhibitory concentration. OD, optical density.

state [21]. Interest in camel milk usage for human nutrition is increasing due to its distinct composition and unique biofunctional properties [22].

In this study the boiled camel and cow milk and packed pasteurized and buttermilk samples were free from bacterial contamination. This is in agreement with the findings of Garedew *et al.* [23], who found that all pasteurized and packed milk samples taken from various supermarkets and restaurants at different shelf life were culture negative for Gram-negative staining bacteria. This may be explained by the effectiveness of pasteurization at the processing plant that minimized the chance of postpasteurization contamination.

The results of this study indicated that camel's milk had antibacterial activity against *E. coli* and *K. pneumoniae*. Different concentrations were used, and it was found that 40 and 70% were the highly effective concentrations on both organisms, respectively. *E. coli* was the common organism affected by the four types of milk tested. It was found that pasteurized cow's milk was the highly effective milk type against this organism.

The concentration and the activity of lactoferrins and lysozyme (antibacterial agents) differ in cow's and camel's milk [4].

Lactoferrin works as an antimicrobial compound through chelating the iron ion, making this essential ion unavailable to the invading pathogens. There have been many reports on the antibacterial effects of lactoferrin of different origins [24–26].

The mean concentrations of lactoferrin in normal and mastitic cow milk have been reported previously and

- diabetes: a 1 year randomized controlled trial. *Diabetes Res Clin Pract* 2005; 68:176–177.
- 8 Bartowska J, Szwajkowska M, Litwińcuk Z, Król J. Nutritional value and technological suitability of milk from various animal species used for dairy production. *Tal J Food Sci* 2011; 10:291–302.
 - 9 Walstra P, Wouters JTM, Geurts TJ. *Dairy science and technology*. 2nd ed. Boca Raton, FL: CRC Press Taylor & Francis Group; 2006.
 - 10 Barbour EK, Nabbut NH, Frechs WM, AL-Nakhli HM. Inhibition of pathogenic bacteria by camels milk relation to whey lysozyme and stage of lactation. *J Food Prod* 1984; 47:838–840.
 - 11 Hahn G. *Pathogenic bacteria in raw milk – situation and significance of pathogenic microorganisms in raw milk*. Brussels, Belgium: International Dairy Federation, IDF; 1996; 68–77.
 - 12 Quinn PJ, Carter ME, Markey B, Carter GR. *Clinical veterinary microbiology*. 2nd ed. Grafos: Mosby International; 2002; 6–346.
 - 13 Barrow GI, Feltham RKA. *Cowan and Steel's manual for the identification of medical bacteria*. UK: Cambridge University Press; 2004; 39–725.
 - 14 Cappuccino JG, Sherman N. *Microbiology lab manual*. USA: Benjamin-Cummings Publishing Company; 1995; 477.
 - 15 National Committee for Clinical Laboratory Standards (NCCLS). *Performance standards for antimicrobial susceptibility testing: fifteen information supplement. CLSI/NCCLS document M100-S15*. Wayne, PA: CLSI; 2005.
 - 16 Magiorakos AP, Srinivasan A, Carey RB, Carmeli Y, Falagas ME, Giske CG, et al. Multidrug resistant, extensively drug-resistant and pandrug-resistant bacteria: an international expert proposal for interim standard definitions for acquired resistance. *Clin Microbiol Infect* 2012; 18:268–281.
 - 17 Jones RN, Barry AL, Gavan TL, Washington JA. Microdilution and macrodilution broth procedures. In: Lennette EH, Balows A, Hausler WJ Jr, Shadomy HJ, editors. *Manual of clinical Microbiology*. 4th ed. Washington DC: American Society for Microbiology; 1985; 972–977.
 - 18 Barry AL, Craig WA, Nadler H, Reller B, Christine CS, Jana MS. National Committee for Clinical Laboratory Standards (NCCLS). *M26-A: Methods for determining bactericidal activity of antimicrobial agents, approved guideline*; 1999; 19–18.
 - 19 Hammer MU, Brauser A, Olak C, Brezesinski G, Goldmann T, Gutschmann T, Andrä J. Lipopolysaccharide interaction is decisive for the activity of the antimicrobial peptide NK-2 against *Escherichia coli* and *Proteus mirabilis*. *Biochem J* 2010; 427:477–488.
 - 20 Torrent M, Navarro S, Moussaoui M, Nogués MV, Boix E. Eosinophil cationic protein high-affinity binding to bacteria-wall lipopolysaccharides and peptidoglycans. *Biochemistry* 2008; 47:3544–3555.
 - 21 Kris-Etherton PM, Hecker KD, Bonanome A, Coval SM, Binkoski AE, Hilpert KF, et al. Bioactive compounds in foods: their role in the prevention of cardiovascular disease and cancer. *Am J Med* 2002; 113:71S–88S.
 - 22 Sboui A, Khorchani T, Agrebi A, Djegham M, Mokni M, Belhadj O. Antidiabetic effect of camel milk on alloxan-induced diabetic dogs. *Afr J Microbiol Res* 2012; 6:4023–4029.
 - 23 Garedew L, Berhanu A, Mengesha D, Tsegay G. Identification of Gram-negative bacteria from critical control points of raw and pasteurized cow milk consumed at Gondar town and its suburbs, Ethiopia. *BMC Public Health* 2012; 12:950.
 - 24 Bethell DR, Huang J. Recombinant human lactoferrin treatment for global health issues: iron deficiency and acute diarrhea. *Biomaterials* 2004; 17:337–342.
 - 25 Arnold RR, Russell JE, Champion WJ, Brewer M, Gauthier JJ. Bactericidal activity of human lactoferrin: differentiation from the stasis of iron deprivation. *Infect Immun* 1982; 35:792–799.
 - 26 Farnaud S, Patel A, Odell EW, Evans RW. Variation in antimicrobial activity of lactoferricin-derived peptides explained by structure modeling. *FEMS Microbiol Lett* 2004; 238:221–226.
 - 27 Lee NY, Kawai K, Nakamura I, Tanaka T, Kumura H, Shimazaki K. Susceptibilities against bovine lactoferrin with microorganisms isolated from mastitic milk. *J Vet Med Sci* 2004; 66:1267–1269.
 - 28 Hagiwara S, Kawai K, Anari A, Nagahata H. Lactoferrin concentrations in milk from normal and subclinical mastitic cows. *J Vet Med Sci* 2003; 65:319–323.
 - 29 Al-Majali AM, Ismail ZB, Al-Hami Y, Nour AY. Lactoferrin concentration in milk from camels (*Camelus dromedarius*) with and without subclinical mastitis. *Intern J Appl Res Vet Med* 2007; 5:120–124.
 - 30 Elagamy EI. Effect of heat treatment on camel milk proteins with respect to antimicrobial factors: a comparison with cows' and buffalo milk proteins. *Food Chem* 2000; 68:227–232.
 - 31 Cardoso RR, Ponte M, Leite V. Protective action of camel milk in mice inoculated with *Salmonella enterica*. *Isr Med Assoc J* 2013; 15:5–8.
 - 32 Dhaiman AS. Purification of camel milk lysozyme and its lytic effect on *Escherichia coli* and *Micrococcus lysodeikticus*. *Comp Biochem Physiol B* 1988; 91:793–796.
 - 33 Mwambete KD. The in vitro antimicrobial activity of fruit and leaf crude extracts of *Momordica charantia*: a Tanzania medicinal plant. *Afr Health Sci* 2009; 9:34–39.
 - 34 Arnold RR, Brewer M, Gauthier JJ. Bactericidal activity of human lactoferrin: sensitivity of a variety of microorganisms. *Infect Immun* 1980; 28:893–898.