

Egyptian Journal of Chemistry

http://ejchem.journals.ekb.eg/



Gallic Acid and its Derivatives: A Review of their Antioxidant Properties and Applications for Fossil Fuel Preservation



Rabab M. Nasser, Nora A. Elebiary*& Khadra B. Alomari

Department of Physical Science, Chemistry Division, College of Science, Jazan University, P.O. Box. 114, Jazan 45142, Kingdom of Saudi Arabia.

Abstract

For many industrial and home applications, fossil fuels serve as the primary energy source. They are, however, prone to oxidative breakdown when exposed to air, light, heat, and metal catalysts, which results in the development of dangerous compounds such gums, sediments, acids, and peroxides. These products have the potential to negatively impact fuel quality, performance, and storage stability. Effective antioxidants that can neutralize free radicals and prevent oxidation are therefore required. Because of its capacity to donate hydrogen atoms and create stable radicals, gallic acid (3,4,5-trihydroxybenzoic acid) is a naturally occurring phenolic molecule with considerable antioxidant activity. Gallic acid and its derivatives have received a lot of attention for their potential use as antioxidants in a variety of industries, including food, cosmetics, medicine, and biotechnology. Gallic acid and its derivatives' chemistry, biology, and production, as well as their antioxidant mechanisms and uses for preserving fossil fuels, are all covered in this review. We also go over the benefits and drawbacks of using these substances as fossil fuel antioxidants, and we highlight the difficulties that now face their development and optimization as well as the potential that lie ahead.

Key words: Gallic acid; Antioxidants; Fossil fuels; Oxidative degradation; Phenolic compounds

1. Introduction

When fossil fuels are burned, oxygen reacts with carbon and hydrogen to produce water, carbon dioxide, heat, and light. However, sulfur and nitrogen in the fuel and air also create pollutants like sulfur dioxide (SO_2) and nitrogen oxides (NO_x). These pollutants can lead to acid rain, which damages structures, harms vegetation, and disrupts aquatic ecosystems. [1-8].

This review paper on gallic acid and its derivatives explores their antioxidant properties and applications, particularly in fossil fuel preservation. While focusing on these areas, the paper also highlights the broader relevance of the United Nations Sustainable Development Goals (SDGs). By investigating the role of gallic acid in various industries, the review supports SDG 12 on responsible consumption and production, SDG 9 on sustainable innovation and industry, and SDG 13 on environmental protection and climate action. Additionally, it emphasizes the importance of scientific research and technological advancements for sustainable development, aligning with the SDGs' call to foster innovation for a more sustainable future.

1.1. Effect of oxidation and degradation of fossil fuels:

Oxidation and degradation have an impact on the traits and issues of fossil fuels. The elderly ones emit greenhouse gases. Performance and stability provide evidence of how well fossil fuels can produce and sustain energy. Degradation and oxidation reduce their efficacy, dependability, longevity, compatibility, and safety [9, 10].

Factors influencing the oxidation of fossil fuels:

Free radicals and reactive oxygen species (ROS) are molecules or atoms that can oxidize fossil fuels by taking their hydrogen atoms or electrons. They can be formed by oxygen, heat, light, microorganisms, or catalysts. Catalysts are substances that speed up the oxidation process by activating oxygen or other molecules, or by generating more radicals from water or hydrogen peroxide [11, 12].

1.2. Gallic acid and its derivatives

Benzene rings with one or more hydroxyl groups attached are chemical compounds that make up the family of "phenolic acids," which includes the naturally occurring substance gallic acid. Gallic acid's chemical formula, "C₆H₂(OH)₃COOH", reflects the presence of three hydroxyl groups and one carboxylic acid group. Gallic acid and its derivatives exhibit a variety of "biological activities", including anti-inflammatory, anti-cancer, anti-microbial, and antioxidant properties. Various industrial applications, including those in food additives, preservatives, cosmetics, colors, and pharmaceuticals, are also possible for them [13-20]. Table 1 showed the various industrial applications of gallic acid and its derivatives. Figure 1 displays the percentage of applications for various gallic acid derivatives. Esters of gallic acid (green color) are used most frequently and have 7 applications (23.3%) in a variety of industries, including the food preservative, pharmaceutical, cosmetic, and chemical industries. The second most frequent use is for gallic acid itself (red color), which has six applications (20%) in a variety of industries including antioxidant, anticancer, antibacterial, antiulcer, anti-cholesterol, and drug metabolizing enzyme inhibitor. Ionic gallate (blue color) is used in two (6.7%) applications, including one as a carbonic anhydrase inhibitor and one as a

*Corresponding author e-mail: names@mail.com.; (Nora A. Elebiary).

Receive Date: 04 February 2025, Revise Date: 12 March 2025, Accept Date: 19 March 2025

DOI: 10.21608/ejchem.2025.357652.11253

©2025 National Information and Documentation Center (NIDOC)

278

chondro-protective action. There is just one application (3.3%) for each of the remaining gallic acid derivatives across all industries (Grey for Gallic acid and its catechin derivatives have one application as drug metabolizing enzyme inhibitor; Brown for Gallic acid and its ester derivatives have one application as anti-carcinogenic.; Yellow for Gallic acid and its ester derivatives have one application as anti-mutagenic.; Magenta for Gallic acid and its ester derivatives have one application as antiangiogenic. Silver color for gallic acid and its ester derivatives have one application as fuel additive. Cyan color for gallic acid and its ester derivatives have one application as anti-inflammatory.

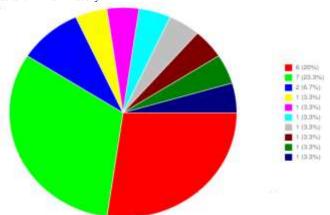


Figure 1. Applications of gallic acid and its derivatives.

Table 1. Applications of gallic acid and its derivatives.

#	Application	Gallic acid derivative	Source
1	Antioxidant	Gallic acid &Esters of gallic acid	[21-24]
2	Anticancer	Gallic acid	[21]
3	Antifungal	Gallic acid	[21]
4	Antibacterial	Gallic acid	[21]
5	Antiviral	Gallic acid	[21]
6	Antiulcer	Gallic acid	[21]
7	Anti-cholesterol	Gallic acid	[21]
8	Anticarcinogenic	Gallic acid Esters of gallic acid	[25, 26]
9	Antimutagenic	Gallic acid & Esters of gallic acid	[25, 26]
10	Antiangiogenic	Gallic acid & Esters of gallic acid	[25, 26]
11	Antiinflammatory	Gallic acid & Esters of gallic acid	[25, 26]
12	Drug metabolizing enzyme inhibitor	Gallic acid and its catechin derivatives	[22]
13	Food preservative	Esters of gallic acid	[22]
14	Cosmetic ingredient	Esters of gallic acid	[22]
15	Pharmaceutical agent	Esters of gallic acid	[22]
16	Chondro-protective effect	Ionic gallate	[23]
17	Carbonic anhydrase inhibitor	Ionic gallate	[23]
18	Antidiabetic activity	Ionic gallate	[23]
19	Cathepsin D inhibitor	Ionic gallate	[23]
20	Antimicrobial	Gallic acid and its ester derivatives	[25, 26]
21	Antifungal	Gallic acid and its ester derivatives	[25, 26]
22	Biofuel production	Gallic acid	[27]
23	Fuel additive	Esters of gallic acid	[21,23]
24	Lubricity enhancer	Gallic acid and its esters	[24]

Ordere New York The Destriction of Them And The Property and The Destriction of The Property o

1.2.1. Extraction Techniques for gallic acid:

There are numerous ways to extract gallic acid from natural sources, including maceration, percolation, soxhlet extraction, ultrasound-assisted extraction (UAE), microwave-assisted extraction (MAE), supercritical fluid extraction (SFE), enzyme-assisted extraction (EAE), etc. Additionally, pyrogallol and phloroglucinol can be used as starting materials for the chemical or enzymatic synthesis of gallic acid from other molecules. Utilizing a variety of processes including esterification, polymerization, condensation, complexation, etc., gallic acid derivatives can be made from gallic acid or other precursors. Recrystallization, column chromatography, fractional distillation, and other processes can be used to purify gallic acid and its derivatives. Methods like spectroscopy (UV-Vis, IR, NMR) can be used to characterize gallic acid and its derivatives [28-31].

1.3. Gallic acid occurrence, chemical, physical and biological activity

Occurrence:

Gallic acid is a naturally occurring substance that can be found in a variety of fruits, plants, and nuts. Gallnuts, sumac, witch hazel, tea, oak bark, grapes, berries, pomegranate, and mango are a few typical sources of gallic acid. Microbes like fungus and bacteria are also capable of producing gallic acid. Gallic acid derivatives can be made synthetically or from natural sources, such as gallic acid or other precursors [14, 32, and 33].

Gallic acid's chemical formula is $C_6H_2(OH)_3COOH$, and its molecular weight is 170.12 g/mol. The benzene ring has a carboxylic acid group at position 1, three hydroxyl groups at positions 3, 4, and 5, and three hydroxyl groups at positions 3, 4, and 5. Its four tautomeric forms are keto-enol (KE), enol (E), quinone-hydroquinone (QH), and semiquinone (SQ). The kind and location of the substituents on the carboxylic acid group or the benzene ring determine the structure of the derivative of gallic acid. Typical types of gallic acid derivatives include gallates (esters), gallotannins (polymers), ellagitannins (dimers), and metal complexes [34].

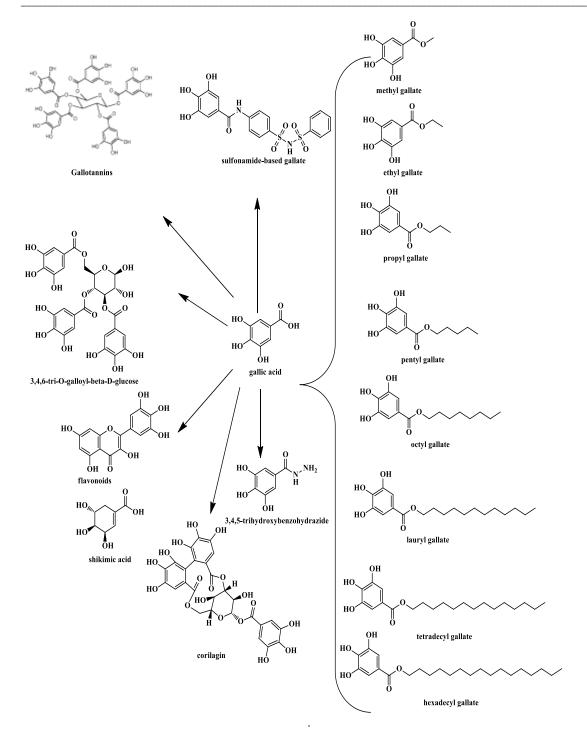
Chemical properties: Gallic acid is an acidic, insoluble, white or yellowish substance. It has a strong potential as an antioxidant and can create many compounds with various properties. Examples of derivatives of gallic acid include gallates, ellagitannins, metal complexes, and galllotannins [35]. Scheme 1 indicates the gallic acid and its derivatives.

The biological activity of gallic acid and its derivatives include anti-inflammatory, anti-microbial, antiviral, anti-cancer, and neuroprotective properties. They are capable of modifying several signaling pathways, removing free radicals, chelating metals, and controlling gene expression. Additionally, they may find use in the culinary, cosmetics, and pharmaceutical industries [23, 31, 36-38].

1.2. Reactions of gallic acid

By esterification, methylation, or other changes, gallic acid can produce a number of derivatives. Examples include:

- **Esterification:** Gallic acid can be esterified to create methyl gallate, ethyl gallate, propyl gallate, octyl gallate, lauryl gallate, and other esters by reacting with alcohols or other organic acids. These esters can be employed as antioxidants in food, cosmetics, medicines, and fossil fuels since they are more stable and soluble than gallic acid [39-42].
- -Methylation: Gallic acid can be methylated to create derivatives like 3-methoxygallic acid, 4-methoxygallic acid, 5-methoxygallic acid, 3,4-dimethoxygallic acid, 3,5-dimethoxygallic acid, 4,5-dimethoxygallic acid, etc. by reacting with methylating chemicals like dimethyl sulfate or methyl iodide. As intermediates in the synthesis of other chemicals, these derivatives can have diverse pharmacological and antioxidant properties from gallic acid [13, 43, and 44].
- **Oxidation:** Gallic acid can be oxidized by a variety of oxidizing substances, including oxygen, hydrogen peroxide, potassium permanganate, etc. to generate oxidized derivatives such gallic acid hydrazide, gallic aldehyde, pyrogallol, etc. These derivatives can function as intermediates in the synthesis of other substances and can have chemical and biological properties that are distinct from those of gallic acid [45, 46].
- **Reduction:** Gallic acid can be reduced by a variety of reducing agents, including sodium borohydride, sodium dithionite, etc. to generate reduced derivatives such 3-deoxygallic acid, 4-deoxygallic acid, 5-deoxygallic acid, etc. These derivatives can function as intermediates in the synthesis of other substances and can have chemical and biological properties that are distinct from those of gallic acid [45-48].
- **Hydrolysis:** Acids or water can hydrolyze gallic acid to create hydrolyzed derivatives such 3-hydroxybenzoic acid, 4-hydroxybenzoic acid, 5-hydroxybenzoic acid, etc. When manufacturing other chemicals, these derivatives can be utilized as intermediates because they can have different chemical and biological properties from gallic acid [49, 50].
- Condensation: Gallic acid can condense with other substances like formaldehyde, acetaldehyde, phenols, etc. to create condensed derivatives such ellagic acid, gallotannins, ellagitannins, etc. These derivatives can be employed as natural products or antioxidants and have molecular weights that are higher than gallic acid and antioxidant activity [50, 52].
- **Complexation:** Gallic acid can combination with metal ions like iron, copper, zinc, etc. to create gallic acid metal complexes. These complexes can be utilized as dyes or catalysts and can differ from gallic acid in terms of color and catalytic activity [44, 53-55].



Scheme 1. Gallic acid and its derivatives.

Gallic acid can form different types of derivatives by modifying its hydroxyl or carboxylic acid groups. Some common types of gallic acid derivatives are:

Gallates: are esters of gallic acid with phenols or alcohols. For instance, methyl gallate, propyl gallate, octyl gallate, and dodecyl gallate are aliphatic alcohol-based esters of gallic acid. Catechins are flavonoids that can be found in tea and other plants, and catechin gallates are esters of gallic acid with catechins [56-60].

Gallotannins: They are polymer component of gallic acid, which also contain glucose or other sugars. They fall under the category of hydrolyzable tannins, which can be hydrolyzed enzymatically or in an acidic environment to produce gallic acid and sugars. They are abundant in plants and possess astringent and antioxidant qualities [61-65].

Ellagitannins: A class of hydrolyzable tannins that can be converted into ellagic acid and sugars through acidic or enzymatic hydrolysis. Ellagic acid is a dimer of gallic acid, linked by a carbon-carbon bond and two lactone rings. Found widely in plants, ellagitannins have demonstrated antioxidant and anticancer activities. [65-70].

Metal complexes: They are substances created when gallic acid or its derivatives combine with metal ions like iron, copper, zinc, or silver. They are used as catalysts, antibacterial agents, dyes, and in photography [71-75].

1. Fossil fuels antioxidants

Antioxidants can stop or delay the oxidation of other molecules. During oxidation, free radicals are produced as a result of electron loss. Free radicals are highly reactive and unstable atoms or molecules. They can damage the chemical structure and function of lipids, proteins, DNA, and other biomolecules [76-79].

Fossil fuels are hydrocarbons formed from prehistoric organic materials and are commonly used as energy sources for heating, lighting, transportation, and industrial activities. However, when exposed to air, heat, light, or metal catalysts, fossil fuels undergo oxidation. This process can produce peroxides, aldehydes, ketones, acids, and other compounds that can reduce the fuel's quality and efficiency. Additionally, these byproducts may increase the emissions of harmful substances such as carbon monoxide, hydrocarbons, nitrogen oxides, particulate matter, and smoke [80-84].

Antioxidants are therefore crucial and pertinent for use in fossil fuels since they can:

- 1. Prevent the creation of hazardous chemicals that could damage the stability, storage, transport, and combustion of fossil fuels by inhibiting or delaying the oxidation of those fuels [85-89].
- 2. Enhance engine performance and fuel efficiency by lowering fuel use, friction, wear, corrosion, and deposits [90-94].
- 3. Lower the emissions of greenhouse gases and other harmful substances that might cause cancer, acid rain, pollution, respiratory illnesses, and global warming in order to lessen their negative effects on the environment and health concerns [95-97].

2.1. Applications of gallic acid and its derivatives antioxidants:

Gallic acid and its derivatives can act as antioxidants for industrial fluids, heating systems, and fossil fuels. They can prevent or slow down oxidation and the formation of harmful chemicals. They can also improve the quality, performance, efficiency, and safety of these items. They can also reduce their consumption and emissions, and protect the environment and human health [98].

2.1.1. Mechanism of using gallic acid as antioxidant.

Gallic acid and its derivatives can act as antioxidants for fossil fuels by scavenging free radicals produced during the burning of hydrocarbons, such as hydroxyl radicals (•OH), peroxyl radicals (ROO•), and alkoxyl radicals (RO•). They can also stop lipid peroxidation, a process that starts a chain reaction that produces additional free radicals and dangerous substances. Additionally, they have the ability to bind metal ions that catalyze oxidation reactions, such iron and copper. In order to prevent oxidative damage to the fuel, they can also regulate endogenous antioxidant enzymes like superoxide dismutase (SOD), catalase (CAT), and glutathione peroxidase (GPx) [13, 98].

Gallic acid and its derivatives can reduce the oxidation of fossil fuels and the emissions of pollutants by: Preventing or delaying the development of peroxides, aldehydes, ketones, acids, and other chemicals that can reduce the quality and efficiency of the fuel. By lowering fuel consumption, friction, wear, corrosion, and deposits, one can increase engine performance and fuel economy. Decreasing the emissions of harmful compounds such carbon monoxide (CO), hydrocarbons (HC), nitrogen oxides (NOx), sulfur dioxide (SO2), particulate matter (PM), and smoke, as well as greenhouse gases like carbon dioxide (CO₂) and methane (CH₄) [13].

2.1.2. Forms of antioxidants in fossil fuel:

Phenolic anti-oxidants: These are substances that include a benzene ring together with one or more hydroxyl groups. By giving hydrogen atoms or electrons, they are able to scavenge free radicals. The metal ions that can catalyze oxidation reactions can also be chelated by them. Tert-butylhydroquinone (TBHQ), propyl gallate (PG), butylated hydroxytoluene (BHT), butylated hydroxyanisole (BHA), gallic acid (GA), and others are phenolic antioxidants [99, and 100].

Amine antioxidants: Antioxidants with one or more amino groups connected to an aromatic or aliphatic chain are known as amines. By contributing hydrogen atoms or electrons, they can neutralize free radicals. They have the ability to bind metal ions and prevent them from catalyzing. Diphenylamine (DPA), phenylenediamine (PPD), di-tert-butyl-p-cresol (DTBC), and N,N'-di-sec-butyl-p-phenylenediamine (DBPC) are a few examples of amine antioxidants [101, and 102].

Vitamin antioxidants: These are substances with structures and actions resembling those of vitamins. By giving hydrogen atoms or electrons, they are able to scavenge free radicals. In addition, by lowering other antioxidants back to their active states, they can renew them. Vitamins that act as antioxidants include beta-carotene (provitamin A), ascorbyl palmitate (vitamin C ester), and alpha-tocopherol (vitamin E) [103; 104]

Gallic acid and its derivatives have been shown in several experimental experiments to have antioxidant properties in a variety of fossil fuels, including gasoline, diesel, biodiesel, jet fuel, etc. Some examples include:

Sutanto and Nasikin [105] investigated the solubility and antioxidant properties of a pyrogallol derivative as a potential additive for biodiesel. The researchers prepared the derivative by reacting pyrogallol with methyl linoleate in the presence of a radical, resulting in a molecule with increased solubility. The solubility test and various tests for antioxidant potential were conducted, including acid value determination and the Rancimat test. The results showed that the pyrogallol derivative exhibited higher solubility and acid value stability in palm oil biodiesel compared to pyrogallol, tert-butylhydroquinone (TBHQ), and gallic acid. Additionally, the derivative demonstrated a higher Rancimat induction time, indicating superior performance under accelerated

oxidation conditions. These findings suggest the potential of the pyrogallol derivative as an effective antioxidant additive for

Varatharajan & Pushparani [106] addresses the issue of maintaining the stability of biodiesel fuels over an extended period. Biodiesel is susceptible to oxidation, resulting in the formation of insoluble gums that can clog fuel filters. The article highlights the importance of incorporating appropriate antioxidants to improve the storage stability of biodiesel. The selection of antioxidants depends on factors such as the chemistry of antioxidants, the composition of biodiesel, the presence of transition metals, and storage temperature. The study emphasizes the need for further research to identify effective antioxidants for different types of biodiesel fuels.

Longanesi et al [107] discussed the factors promoting oxidation, including molecular composition, metal contamination, temperature, and light exposure. It also explores the benefits of hydrogenated vegetable oil (HVO) and the analytical techniques used to study oxidation. Additionally, they addressed the influence of higher pressure injection systems on deposit formation, emphasizing that unsaturated biodiesel components are not solely responsible.

2.2. Factors that influence the antioxidant efficiency of gallic acid and its derivatives in fossil fuels

Gallic acid and its derivatives are natural compounds that can prevent or delay the oxidation of fossil fuels by free radicals and ROS. Their antioxidant performance depends on many factors that influence their characteristics and behavior in the fuel systems. They have benefits and drawbacks as antioxidants for fossil fuels, and they need to be optimized and researched to enhance their effectiveness and sustainability. [21, 23, 98, 108-115].

3. Determination of the antioxidant activity of gallic acid and its derivatives in fossil fuels:

There are some methods for measuring the antioxidant activity of gallic acid and its derivatives in fossil fuels, such as:

- **Induction period:** The induction period is the amount of time needed for free radicals and ROS to start oxidizing fossil fuels. Different methods, including differential scanning calorimetry (DSC), pressure differential scanning calorimetry (PDSC), and the Rancimat method [116, 117]; can be used to determine the induction duration. By delaying or inhibiting the oxidation of fossil fuels, the induction duration may reflect the antioxidant activity of gallic acid and its derivatives.
- **Gum content:** The quantity of sludge or insoluble deposits created during the oxidation of fossil fuels by free radicals and ROS is known as the gum content. Several methods [118, 119] like ASTM D381 [120], ASTM D873 [121], or IP 131 [122] can be used to determine the gum content. By lowering or limiting the formation of deposits or sludge, the gum content can serve as a proxy for the antioxidant activity of gallic acid and its derivatives.
- Oxidation stability: The ability of fossil fuels to withstand oxidation by free radicals and reactive oxygen species (ROS). Different methods, such as ASTM D2274 [123], ASTM D525 [124], ASTM D2272 [125], or EN 14112 [126], can be used to determine the oxidation stability. By improving or maintaining the quality and performance of fossil fuels, the oxidation stability can represent the antioxidant activity of gallic acid and its derivatives.
- **Acid value:** The amount of acid produced as a result of free radicals and reactive oxygen species oxidizing fossil fuels is known as the acid value. There are numerous methods that can be used to determine the acid value, including ASTM D664 [127], ASTM D974 [128], and IP 139 [129]. Gallic acid and its derivatives have antioxidant properties that can be reflected in the acid value by reducing or preventing the production of acid.
- **Peroxide value:** The peroxide value is the quantity of peroxides produced when free radicals and ROS oxidize fossil fuels. Different methods, such as ASTM D3703 [130], ASTM D2344 [131] or ISO 3960 [132], can be used to determine the peroxide value. By reducing or preventing the generation of peroxides, the peroxide value can indicate the antioxidant activity of gallic acid and its derivatives.

Accelerated oxidation tests: are techniques that can quickly and practically measure the antioxidant effectiveness and the quality and performance of fossil fuels under real oxidation and degradation conditions. The disadvantage of these techniques is that they may not be consistent, reproducible, or comparable across different laboratories or systems. They may also be influenced or interfered by various factors such as temperature, pressure, oxygen concentration, catalysts, etc.

3.1. Techniques for describing composition, characterization, and behavior of gallic acid and its derivatives in fossil fuels:

3.1.1. Differential scanning calorimetry (DSC):

DSC can be used to quantify the oxidation and decomposition of fossil fuels by looking for exothermic peaks that signify the beginning and growth of oxidation reactions [133]. Thermodynamics, antioxidants, and the kinetics of fossil fuels can all be studied using DSC. Although DSC is sensitive, accurate, and illuminating, it is also pricey, difficult, and demands several samples as well as pricy equipment. Various elements that affect the outcomes may also have an impact.

3.1.2. Gas chromatography (GC):

GC is a technique that separates and analyzes a sample by its volatility and polarity. It can measure the oxidation and degradation of fossil fuels by identifying and quantifying their components and changes. GC is selective, accurate, and informative, but it is also time-consuming, requires sample preparation and suitable column and detector. It could also be affected by various factors that could cause peak problems [134-136].

3.1.3. Mass spectrometry (MS):

- MS is a method that ionizes a sample and analyzes each of its constituent parts according to their mass-to-charge ratio. By identifying and quantifying their chemical fragments and isotopes, it can gauge the oxidation and breakdown of fossil fuels.

Although MS is sensitive, accurate, and illuminating, it is also costly, difficult, and labor-intensive and necessitates sample

3.1.4. The Molecular Docking of Gallic Acid and Its Derivatives

preparation and appropriate ionization and detection techniques [136-138].

Gallic acid and its derivatives exhibit various biological functions, including anti-inflammatory, anti-cancer, antioxidant, and antibacterial effects. Molecular docking, a computational technique, was used to predict the binding affinity of ligands to target proteins. In this study, molecular docking was performed using AutoDock Vina, with crystal structures of target proteins obtained from the Protein Data Bank (PDB). Energy minimization was carried out using Gaussian 09, and ligands were prepared with ChemDraw. Binding affinity was calculated using the inhibition constant (Ki) and binding energy (Δ G).

The outcomes demonstrated that gallic acid and its derivatives had strong binding affinity for the intended proteins. With a ΔG value of -8.1 kcal/mol and a Ki value of 0.13 μ M, gallic acid had the highest binding affinity towards cyclooxygenase-2 (COX-2). The derivatives likewise demonstrated strong binding affinity for COX-2, with G values between -7.5 and -8.0 kcal/mol. Gallic acid is a type of phenolic acid that is present in a wide variety of plants, including gallnuts, sumac, witch hazel, tea leaves, oak bark, etc. It contains antioxidant qualities and might also be analgesic, anti-inflammatory, antibacterial, anti-cancer, and have other health advantages.

Gallic acid and its derivatives can be evaluated using a variety of computational techniques, including as molecular docking, molecular dynamics simulations, quantum mechanics calculations, and density functional theory (DFT) studies.

3.1.4.1. Molecular docking:

Arsianti et al [139] presents the design and screening of gallic acid derivatives as potential inhibitors of malarial dihydrofolate reductase (DHFR) using in silico docking. The researchers designed fourteen gallic acid derivatives and modeled their three-dimensional structures. Through docking simulations and amino acid analysis, they identified three top-ranked compounds, with compound 12 (octyl gallate) showing the strongest interaction and greatest inhibitory activity against malarial DHFR. The study suggests that compound 12 holds promise as a potential candidate for the development of new antimalarial agents, Figures 2, 3.

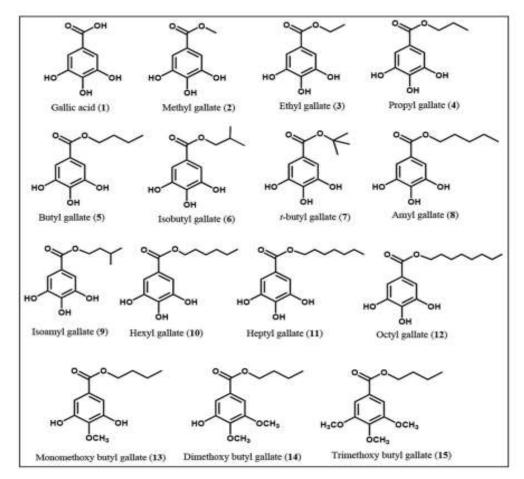


Figure 2 Chemical structure of gallic acid (1) and its derivatives (2-15) [139]

284 R. M. Nasser et.al.

Humaedi et al [140] investigates the potential of gallic acid and its derivatives as inhibitors of BRAF colon cancer through in silico molecular docking. The study examines the stability, affinity, and interaction of gallic acid and five derivatives as ligands with the BRAF protein. The results indicate that the derivatives have lower Gibbs energy values compared to gallic acid, suggesting higher stability. Additionally, the derivatives exhibit greater affinity and stronger interaction with the catalytic site of BRAF colon cancer compared to gallic acid. Among the derivatives, (2-hydroxy)-benzylgallate shows the highest stability and strongest interaction with BRAF, making it a promising candidate for colon cancer drug development, Figure 4.

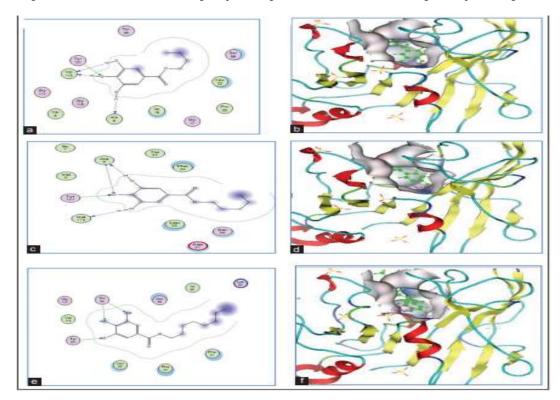


Figure 3 Two-dimensional and three-dimensional complex interaction of derivative 5 (a, b), 8 (c, d), and 12 (e, f) with dihydrofolate reductase [139]

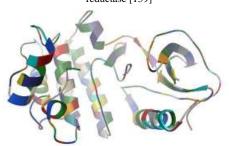


Figure 4 Crystal structure BRAF [140].

Kokila et al [141], explores the antihyperglycemic properties of gallic acid derived from the methanol extract of Thunbergia mysorensis flowers. The study demonstrates that the extract effectively inhibits α -glucosidase, α -amylase, and aldose reductase enzymes, which play a crucial role in postprandial hyperglycemia management. Gallic acid is identified as the primary active component of the extract, and it exhibits significant inhibition of the target enzymes. Molecular simulations reveal stable interactions between gallic acid and the active sites of the enzymes. These findings suggest that gallic acid derived from Thunbergia mysorensis could serve as a potential natural antidiabetic drug, warranting further in vivo and clinical studies.

Raghi et al. [142] explores the inhibitory activity of Gallic acid derivatives fused with 1,3, 4-Oxadiazole moieties against the ABL receptor, which is implicated in Chronic Myeloid Leukemia (CML). The study employs molecular electrostatic potential maps (MESP), molecular docking, and molecular dynamics (MD) simulations to evaluate the structural features and drug likeness of these derivatives. The compounds exhibit promising ABL kinase inhibitory activity and acceptable pharmacokinetic properties, making them potential candidates for the development of new CML drugs.

Umar et al. [143] presents molecular docking studies aimed at identifying potential therapeutic compounds against the non-structural proteins (nsp3, nsp5, nsp12, nsp13, and nsp14) of the novel coronavirus (SARS-CoV-2). The researchers docked 16 derivatives of gallic acid against these proteins and evaluated their binding energies. One derivative, 4-O-(6-galloylglucoside), displayed lower binding energy values than control drugs against multiple target proteins. Pharmacokinetics

as molecular dynamic simulation, Figure 5, in vivo, and in vitro experiments are needed to validate these findings.

screening revealed that this derivative could be metabolized by the liver and has high plasma protein binding. The study suggests that 4-O-(6-galloylglucoside) could be a promising inhibitor against the studied SARS-CoV-2 proteins, but further studies such

Figure 5 Molecular docking studies of gallic acid derivatives against main protease (Mpro) of SARS-Cov-2. a Protocol validation of molecular docking experiment using AutoDock Vina, PyMOL, and LigPlot+ . (a) Comparison of binding modes for re-docked ligand (red) vs. co-crystallized ligand (green) shown as stick representation. Amino acid residues interaction with (b) co-crystalized and (c) re-docked ligand accomplished in LigPlot+ . b Binding mode and molecular interaction of hit ligands with Mpro. (a) Surface representation of Mpro (PDB: 6 LU7) show the binding mode of docked 3-O-(6-galloylglucoside) (yellow), epicatechin gallate (green), and remdesivir (pink). 2D interaction of (b) 3-O-(6-galloylglucoside), (c) epicatechin gallate, and (d) remdesivir [143].

3.1.4.2. Molecular dynamics simulations:

Zhan et al [144] investigates the binding interaction between gallic acid (GA) and lysozyme (LYS) using a combination of molecular dynamics (MD) simulation and spectroscopic techniques. The results indicate that GA forms a stable complex with LYS, with hydrogen bonding and hydrophobic interactions being the main driving forces. The calculated binding energies suggest that van der Waals forces and electrostatic interactions play a significant role in the interaction, Figure 6. The study provides valuable insights into the pharmacological mechanism of GA and offers a reference for future research in this

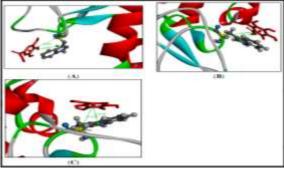


Figure 6 Distance map of GA to Trp62 of LYS, only picked Trp62 as a example. (A) The interaction mode between GA and LYS before MD simulation; (B) The interaction mode between GA and LYS after 298 K MD simulation; (C) The interaction mode between GA and LYS after 310 K MD simulations. (The ligand structure is represented using red stick model, and the red ball is the centroid of GA. The distances are remarked using green lines. The residue Trp62 is represented using a ball and stick model, the green ball represents the centroid of Trp and the yellow ball represents the $C\alpha$ of Trp) [144].

Nie et al [145] investigates the inhibitory effects of gallate moiety on the formation of A β amyloid aggregates, which are associated with Alzheimer's disease. Through molecular dynamics simulations, the study reveals that gallic acid (GA), containing the gallate moiety, effectively prevents the conformational changes of $A\beta 1-40$ monomers and inhibits the formation of β-sheet structures. The binding between GA molecules and Aβ1–40 monomers involves both hydrophilic and hydrophobic amino acid residues. Furthermore, the study demonstrates that polar interactions play a stronger role in the binding than nonpolar interactions. These findings provide molecular-level insights into the mechanisms by which gallate moiety contributes to the anti-amyloidogenic effects of polyphenols, offering potential implications for the development of therapeutic strategies for Alzheimer's disease.

Zhang et al [146] identified natural thrombin inhibitors from traditional Chinese medicine (TCM) and evaluate their biological activity and binding characteristics. Through a combination of molecular docking, thrombin inhibition assay, surface plasmon resonance (SPR), and molecular dynamics simulation, gallic acid was identified as a direct thrombin inhibitor with significant inhibitory effects on thrombin-induced platelet aggregation. The binding studies revealed that gallic acid interacted

with thrombin with a KD value of 8.29 lmol/L. Molecular dynamics and binding free energy analysis provided insights into the mechanism of thrombin inhibition by gallic acid, Figure 7. The study concluded that gallic acid could serve as a potential natural thrombin inhibitor, providing a basis for further research and development in this area.

Cappelli et al. [147], explores the solvation of gallic acid in water and acetonitrile through spectroscopic analysis. The study utilizes classical and quantum mechanical approaches to predict IR, UV, and NMR spectra and compares them with experimental data to validate the solvation models. The effects of hydrogen bonding and bulk solvent on the spectroscopic properties are evaluated. The authors employed a combination of continuum-only, discrete, and mixed continuum/discrete solvation models based on quantum-mechanical and classical molecular dynamics solute-solvent clusters. The study provides insights into the structural characteristics and spectroscopic properties of gallic acid, which can contribute to a better understanding of its biochemical structure-activity relationships and its potential applications in various fields.

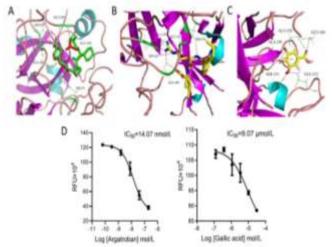


Figure 7. Binding mode of ligand MEL (A), argatroban (B) and gallic acid (C) in the inhibitor-binding site of thrombin. The green and red sticks represent the re-docked and cocrystallized conformations of ligand MEL, respectively. The red dotted lines represent the hydrogen bond interactions between the ligand and thrombin. Key residues are shown and the red dotted lines represent the hydrogen bond interactions between the inhibitors and thrombin. Yellow, white, red and blue atoms represent carbon, hydrogen, oxygen and nitrogen atoms, respectively. (D) Dose-response curves of thrombin inhibition for gallic acid and argatroban, respectively [146].

4. Quantum mechanics calculations:

Badhani and Kakkar [148] explored the structural and chemical properties of gallic acid using density functional theory (DFT) calculations. The study investigates the intramolecular interactions, determines the pKa values of gallic acid anions, and examines the influence of external factors such as pH and dielectric of the medium on the molecular orbitals and electronic spectra. Additionally, the paper simulates and validates the IR and NMR spectra of gallic acid. The researchers also analyze the global and local reactivities of gallic acid, assessing its susceptibility to nucleophilic, electrophilic, and radical attacks. The study provides insights into the structural and electronic characteristics of gallic acid, which are important for developing pharmacologically potent molecules.

Vahedi et al, [149] investigated the interaction mechanism between gallic acid (GA) and α -Chymotrypsin (α -CT) using spectroscopic methods, computational docking, and molecular dynamics (MD) simulation. The results show the formation of a stable complex between GA and α -CT, with fluorescence spectra analysis indicating a static quenching mechanism. The binding constants suggest moderate affinity between GA and α -CT. CD findings reveal alterations in the protein's secondary structure upon interaction with GA. Enzyme activity assays demonstrate a significant decrease in α -CT's activity in the presence of GA, indicating its role as an effective inhibitor. Molecular docking and MD simulations provide insights into the optimal binding site and stability of the α -CT-GA complex.

Pardeshi et al, [150] presents a comprehensive investigation on the molecular interactions and spectroscopic properties of Gallic acid (GA) and acrylic acid (AA) in Gallic acid imprinted polymers (MIPs). The study utilizes density functional theory (DFT) calculations to optimize the structures of GA, AA, and their complex, and examines the effect of the porogen acetonitrile (ACN) using the polarizable continuum model (PCM). The results reveal the formation of a stable GA-AA complex through intermolecular hydrogen bonding. The vibrational spectra simulations are compared with experimental FT-IR spectra, demonstrating the accuracy of the computational approach. Moreover, the study determines the optimal GA-AA mole ratio for the synthesis of MIPs for GA.

Guendouzi, et al, [151] investigated the formation of an inclusion complex between gallic acid (GA) and β -cyclodextrin (β -CD) at a 1:1 stoichiometry ratio. β -CD is known for its ability to encapsulate bioactive compounds, providing protection and improved solubility. The study aims to understand the complexation mechanism and the capability of β -CD to encapsulate GA in both gas and solution phases. Different quantum mechanical methods, including HF/6-31G* and density functional theory (DFT) with dispersion correction, were compared to evaluate the importance of dispersion forces and hydrogen bonding in the interaction. The stability of the optimized complex geometries was assessed using the super molecule method. Various techniques such as frontier molecular orbital (FMO) theory, global indices of reactivity, condensed natural bond orbital

(NBO) analysis, and molecular docking were employed to confirm the inclusion complex and examine the nature of hydrophobic interactions during the complexation process.

Ghouari, et al [152] investigated a new polymorph of gallic acid monohydrate (GAM-VII) for its potential application in the pharmaceutical industry. The synthesis, crystal structure, and computational quantum investigations of GAM-VII were presented. The structural analysis reveals that the presence of water molecules and moderate intermolecular interactions involving carboxyl groups contribute to the formation and stability of this polymorph. A comprehensive comparison is made with the other six known GAM polymorphs, focusing on molecular conformations, hydrogen bonding, and intermolecular interactions. The Hirshfeld surface method is employed to analyze the supramolecular assemblies and quantify the contributions of different intermolecular interactions. The study also provides insights into the reactivity process of GAM-VII through theoretical calculations, including dipole moment, ionization, chemical potential, electronegativity, and electrophilicity index.

Zhan et al. [153] conducted molecular dynamics simulations on the GA-LYS complex to study its binding mechanism. They found that temperature affects GA

binding to LYS, with less protein flexibility at lower temperatures. Molecular docking confirmed the interaction was driven by hydrogen bonding, van der Waals forces, and hydrophobic interactions, consistent with spectroscopy results. Using the MM-PBSA method, they showed the complex remained stable during the simulations.

Additionally, they observed that the binding strength of GA to LYS weakened at higher temperatures (310K) compared to 298K.

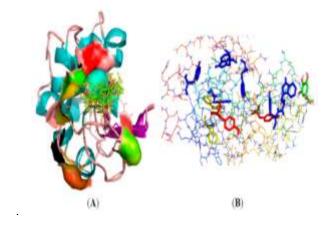


Figure 8 Interaction of GA with LYS. (A) Electron density and hydrophobic surface map between GA- LYS, only residues around 20.0 Å of the ligand are displayed; (B) Interaction mode between GA and LYS. The ligand structures are represented using a stick model, the blue sticks represented Trp residues, the red sticks represented Typ residues, the yellow sticks represented Phe residues [153].

5. Advantages and disadvantages of using gallic acid and its derivatives as antioxidants in fossil fuels

It is possible to examine and contrast the following benefits and drawbacks of employing gallic acid and its derivatives as antioxidants in fossil fuels:

The benefits are Free radicals and reactive oxygen species (ROS) can oxidize and degrade fossil fuels, whereas gallic acid and its derivatives are organic, biodegradable, and renewable sources of antioxidants. In addition to chelating metal ions and modulating cellular signaling pathways, they can scavenge free radicals, prevent lipid peroxidation, and limit combustion of lipids. By extending the induction period, oxidation stability, viscosity, density, etc. of fossil fuels, they can also enhance their quality and efficiency. They may also possess a variety of biological and pharmacological properties, including those that are anticancer, antifungal, antibacterial, antiviral, antiulcer, and anticholesterol, among others.

Gallic acid's and its derivatives' low solubility, stability, and availability in fossil fuels are disadvantages. In the interfacial region of the emulsions, they may be quickly oxidized or destroyed by oxygen, light, heat, metal ions, or microorganisms. Depending on their dosage, structure, and surrounding circumstances, they may also exhibit a dual function as an antioxidant and prooxidant. Additionally, they might have negative consequences like cytotoxicity, genotoxicity, allergenicity, or disruption of other molecules or enzymes.

6. Potential uses and effects of gallic acid and its derivatives' as antioxidants in numerous fields: Advantages of using gallic acid

Gallic acid and its derivatives are antioxidants that can improve the quality, performance, and longevity of fossil fuels and the equipment that uses them. They can prevent or reduce the harmful effects of oxidation and other chemical reactions in fossil fuels, as indicated in the following paragraphs:

- **Transportation:** By enhancing the fluidity, flow-ability, lubricity, etc. of fossil fuels, gallic acid and its derivatives might enhance their transportation. Additionally, they can guard against the corrosion, fouling, clogging, etc. that may happen as a result of the oxidation and degradation of fossil fuels in transportation pipelines and tanks. They can also prevent the production of hazardous substances like aldehydes, ketones, esters, ethers, and other gases from occurring when fossil fuels are burned.
- Environment: By lowering the environmental effect and reliance on synthetic antioxidants that may be hazardous, non-biodegradable, or non-renewable, gallic acid and its derivatives can enhance the environment. Additionally, they can lessen the

harm that fossil fuel oxidation and degradation may do to the environment by releasing noxious substances into the air, water, or soil, such as acids, peroxides, hydroxyl radicals, and other hazardous substances. Additionally, they may encourage the use of antioxidants derived from various plant sources or manufactured using sustainable and renewable methods.

- **Health:** By lowering the risks for illnesses and conditions brought on by contact with or consumption of oxidized and degraded fossil fuels, which may have cytotoxic, genotoxic, allergic, or carcinogenic effects, gallic acid and its derivatives can improve health. Additionally, they can have a number of biological and pharmacological advantages, including, but not limited to, anticancer, antifungal, antibacterial, antiviral, antiulcer, and anti-cholesterol properties.

Limitations and potential drawbacks of using gallic acid

Limitations and potential drawbacks of using gallic acid and its derivatives as antioxidants may vary depending on the context and application, here are some general considerations:

Stability and Shelf Life: Gallic acid and its derivatives may exhibit limitations in terms of stability, especially under certain environmental conditions such as exposure to light, heat, or pH variations. This can affect their efficacy as antioxidants and their shelf life in practical applications.

Compatibility and Interactions: Compatibility issues may arise when incorporating gallic acid and its derivatives into different products or systems. They may interact with other

components or ingredients, leading to changes in taste, color, or stability. Understanding the compatibility and potential interactions is crucial for successful application.

Extraction and Production Challenges: Obtaining gallic acid and its derivatives from natural sources or synthesizing them can present challenges. Extraction methods may require specific solvents or conditions, and large-scale production might be costly or complex. These factors can impact the availability and cost-effectiveness of these antioxidants.

Sensitivity to Processing Conditions: Gallic acid and its derivatives may be sensitive to processing conditions, such as high temperatures or certain chemical reactions. This sensitivity can result in the degradation or alteration of their antioxidant properties, limiting their effectiveness in specific applications or processing techniques.

Regulatory Considerations: Depending on the industry and application, there may be regulatory considerations or restrictions on the use of gallic acid and its derivatives as antioxidants. Compliance with regulations related to safety, labeling, dosage, and maximum allowable concentrations may be necessary.

Potential Interference with Other Processes: While gallic acid and its derivatives exhibit antioxidant properties, they may also interfere with other processes or reactions. For example, in certain industrial processes or biological systems, they might impact enzymatic reactions or interfere with desired chemical transformations.

7. Gallic acid and its derivatives' potential as antioxidants in fossil fuels: Opportunities and challenges

Fossil fuels, such as gasoline and diesel, are prone to oxidative degradation, leading to the formation of harmful byproducts and reduced fuel quality. Gallic acid, a naturally occurring phenolic compound, possesses strong antioxidant properties due to its ability to scavenge free radicals and inhibit oxidation reactions. Its derivatives, including esters and polymers, can be tailored to enhance stability and solubility in fuel matrices. These antioxidants have demonstrated promising results in mitigating fuel degradation and improving fuel performance, such as reducing deposit formation, minimizing corrosion, and extending fuel shelf life. However, challenges remain, including the optimization of antioxidant dosage, compatibility with different fuel compositions, and potential side effects on combustion efficiency and emissions. Additionally, the interactions between gallic acid derivatives and other fuel additives need to be thoroughly investigated. Despite these challenges, gallic acid and its derivatives offer a promising avenue for developing effective antioxidant strategies in fossil fuels, contributing to the overall sustainability and longevity of fuel systems.

8. New Directions for further research of gallic acid and its derivatives as antioxidants in fossil fuels

More research is needed on gallic acid and its derivatives' antioxidant properties in fossil fuels in:

Improving their characteristics: improving their solubility, stability, and availability by using different formulations and optimizing the pH and the ratios of the components.

Characterizing them: determining their chemical composition and behavior and studying their role in preventing oxidation and degradation by using advanced methods.

Evaluating them: measuring their antioxidant activity and effectiveness based on various factors and comparing the results of different systems and methods.

Optimizing them: developing models for their distribution, concentration, and structure by using computer-based methods and predicting and controlling their antioxidant and pro-oxidant effects when they interact with other molecules or enzymes.

9. List of Abbreviations:

Abbreviation Full Form

•OH Hydroxyl Radicals

ASTM American Society for Testing and Materials

BHA Butylated Hydroxyanisole BHT Butylated Hydroxytoluene

CAT Catalase
CD Cyclodextrin
CH4 Methane

CO Carbon Monoxide
CO₂ Carbon Dioxide

DBPC N,N'-Di-sec-butyl-p-phenylenediamine

DFT Density Functional Theory

DG Dodecyl Gallate
DPA Diphenylamine

DSC Differential Scanning Calorimetry

DTBC Di-tert-butyl-p-cresol

E Enol

EAE Enzyme-assisted Extraction

EG Ethyl Gallate

EN European Patent Specification

GA Gallic Acid

GADs Gallic Acid Derivatives
GC Gas Chromatography
GPx Glutathione Peroxidase

HC Hydrocarbons

hCA IX Human Carbonic Anhydrase IX HAS Human Serum Albumin

IP Institute of Petroleum Test Methods

IR Infrared

ISO International Organization for Standardization

KE Keto-enol

Ki Inhibition Constant

MAE Microwave-assisted Extraction

MG Methyl Gallate
MS Mass Spectrometry

NMR Nuclear Magnetic Resonance

NOx Nitrogen Oxides
OG Octyl Gallate
PDB Protein Data Bank

PDSC Pressure Differential Scanning Calorimetry

PG Propyl Gallate

PM Particulate Matter

PPD Phenylenediamine

QH Quinone-hydroquinone

RMSD Root Mean Square Deviation

RMSF Root Mean Square Fluctuation

RO• Alkoxyl Radicals

10. Conclusions:

Gallic acid and its derivatives, with their potent antioxidant properties and ability to prevent the oxidative degradation of fossil fuels, present promising candidates for use as antioxidants in fossil fuel preservation. While their synthesis, formulation, characterization, assessment, and optimization can still be improved, there are significant opportunities for advancement in this field. This study aims to stimulate further interest and innovation by providing a comprehensive review of the current knowledge and identifying research gaps regarding the use of gallic acid and its derivatives as antioxidants in fossil fuels.

11. Conflicts of interest

The authors have no conflict of interest.

12. Formatting of funding sources

This research did not receive any specific grant from funding agencies in the public, commercial, or not-for-profit sectors.

13. References

- [1] Yıldız, İ. (2018). 1.12 Fossil Fuels. In Comprehensive Energy Systems (pp. 521–567). Elsevier. https://doi.org/10.1016/B978-0-12-809597-3.00111-5
- [2] Arku, P., Regmi, B., & Dutta, A. (2018). A review of catalytic partial oxidation of fossil fuels and biofuels: Recent advances in catalyst development and kinetic modelling. Chemical Engineering Research & Design: Transactions of the Institution of Chemical Engineers, 136, 385–402. https://doi.org/10.1016/j.cherd.2018.05.044
- [3] Nie, X., Zhao, T., & Su, Y. (2021). Fossil fuel carbon contamination impacts soil organic carbon estimation in cropland. Catena, 196(104889), 104889. https://doi.org/10.1016/j.catena.2020.104889
- [4] Luo, H., Barrio, J., Sunny, N., Li, A., Steier, L., Shah, N., Stephens, I. E. L., & Titirici, M.-M. (2021). Progress and perspectives in photo- and electrochemical-oxidation of biomass for sustainable chemicals and hydrogen production. Advanced Energy Materials, 11(43). https://doi.org/10.1002/aenm.202101180
- [5] Zhang, X., & Caldeira, K. (2015). Time scales and ratios of climate forcing due to thermal versus carbon dioxide emissions from fossil fuels. Geophysical Research

Letters, 42(11), 4548-4555. https://doi.org/10.1002/2015gl063514

- [6] Blasing, T. J., Broniak, C. T., & Marland, G. (2005). The annual cycle of fossil-fuel carbon dioxide emissions in the United States. Tellus. Series B, Chemical and Physical Meteorology, 57(2), 107–115. https://doi.org/10.1111/j.1600-0889.2005.00136.x
- [7] Hao, F., & Van Brown, B. L. (2019). An analysis of environmental and economic impacts of fossil fuel production in the U.s. from 2001 to 2015. Society & Natural Resources, 32(6), 693–708. https://doi.org/10.1080/08941920.2019.1574044
- [8] Hasheminasab, H., Hashemkhani Zolfani, S., Kazimieras Zavadskas, E., Kharrazi, M., & Skare, M. (2022). A circular economy model for fossil fuel sustainable decisions based on MADM techniques. Economic Research-Ekonomska Istraživanja, 35(1), 564–582. https://doi.org/10.1080/1331677x.2021.1926305
- [9] Pirou, S., Bermudez, J. M., Na, B. T., Ovtar, S., Yu, J. H., Hendriksen, P. V., Kaiser, A., Reina, T. R., Millan, M., & Kiebach, R. (2018). Performance and stability of (ZrO 2) 0.89 (Y 2 O 3) 0.01 (Sc 2 O 3) 0.10 -LaCr 0.85 Cu 0.10 Ni 0.05 O 3-8 oxygen transport membranes under conditions relevant for oxy-fuel combustion. Journal of Membrane Science, 552, 115–123. https://doi.org/10.1016/j.memsci.2018.01.067
- [10] Gani, A. (2021). Fossil fuel energy and environmental performance in an extended STIRPAT model. Journal of Cleaner Production, 297(126526), 126526. https://doi.org/10.1016/j.jclepro.2021.126526
- [11] Xu, L., Wang, Y., & Liu, D. (2022). Effects of oxygenated biofuel additives on soot formation: A comprehensive review of laboratory-scale studies. Fuel (London, England), 313(122635), 122635. https://doi.org/10.1016/j.fuel.2021.122635
- [12] Lushchak, V. I. (2014). Free radicals, reactive oxygen species, oxidative stress and its classification. Chemico-Biological Interactions, 224, 164–175. https://doi.org/10.1016/j.cbi.2014.10.016
- [13] Huang, Y., Richardson, S. J., Brennan, C. S., & Kasapis, S. (2024). Mechanistic insights into α-amylase inhibition, binding affinity and structural changes upon interaction with gallic acid. Food Hydrocolloids, 148(109467), 109467. https://doi.org/10.1016/j.foodhyd.2023.109467
- [14] Weng, C.-J., & Yen, G.-C. (2012). Chemopreventive effects of dietary phytochemicals against cancer invasion and metastasis: Phenolic acids, monophenol, polyphenol, and their derivatives. Cancer Treatment Reviews, 38(1), 76–87.

https://doi.org/10.1016/j.ctrv.2011.03.001

- [15] Wianowska, D., & Olszowy-Tomczyk, M. (2023). A concise profile of Gallic acid—from its natural sources through biological properties and chemical methods of determination. Molecules (Basel, Switzerland), 28(3), 1186. https://doi.org/10.3390/molecules28031186
- [16] Pandurangan, A. K., Norhaizan, M., Mohebali, N., & Yeng, L. C. (2015). Gallic acid attenuates dextran sulfate sodium-induced experimental colitis in BALB/c mice. Drug Design, Development and Therapy, 3923. https://doi.org/10.2147/dddt.s86345
- [17] Jiang, Y., Pei, J., Zheng, Y., Miao, Y.-J., Duan, B.-Z., & Huang, L.-F. (2022). Gallic acid: A potential anti-cancer agent. Chinese Journal of Integrative Medicine, 28(7), 661–671. https://doi.org/10.1007/s11655-021-3345-2
- [18] Uddin, S. J., Afroz, M., Zihad, S. M. N. K., Rahman, M. S., Akter, S., Khan, I. N., Al-Rabbi, S. M. S., Rouf, R., Islam, M. T., Shilpi, J. A., Nahar, L., Tiralongo, E., & Sarker, S. D. (2022). A systematic review on anti-diabetic and cardioprotective potential of Gallic acid: A widespread dietary phytoconstituent. Food Reviews International, 38(4), 420–439. https://doi.org/10.1080/87559129.2020.1734609
- $[19] \ Qu, Y., Wang, L., \& \ Mao, Y. \ (2022). \ Gallic \ acid \ attenuates \ cerebral \ is chemia/re-perfusion-induced \ blood-brain \ barrier \ injury \ by \ modifying \ polarization \ of \ microglia.$

- Journal of Immunotoxicology, 19(1), 17–26. https://doi.org/10.1080/1547691x.2022.2043494
- [20] Shabani, S., Rabiei, Z., & Amini-Khoei, H. (2020). Exploring the multifaceted neuroprotective actions of gallic acid: a review. International Journal of Food Properties, 23(1), 736-752. https://doi.org/10.1080/10942912.2020.1753769
- [21] Badhani, B., Sharma, N., & Kakkar, R. (2015). Gallic acid: a versatile antioxidant with promising therapeutic and industrial applications. RSC Advances, 5(35), 27540–27557. https://doi.org/10.1039/c5ra01911g
- [22] Williamson, G., & Manach, C. (2005). Bioavailability and bioefficacy of polyphenols in humans. II. Review of 93 intervention studies. The American Journal of Clinical Nutrition, 81(1), 243S-255S. https://doi.org/10.1093/ajcn/81.1.243s
- [23] AL Zahrani, N. A., El-Shishtawy, R. M., & Asiri, A. M. (2020). Recent developments of gallic acid derivatives and their hybrids in medicinal chemistry: A review. European Journal of Medicinal Chemistry, 204(112609), 112609. https://doi.org/10.1016/j.ejmech.2020.112609
- [24] Freiría-Gándara, J., Losada-Barreiro, S., Paiva-Martins, F., & Bravo-Díaz, C. (2018). Enhancement of the antioxidant efficiency of gallic acid derivatives in intact fish oil-in-water emulsions through optimization of their interfacial concentrations. Food & Function, 9(8), 4429–4442. https://doi.org/10.1039/c8fo00977e
- [25] Bai, J., Zhang, Y., Tang, C., Hou, Y., Ai, X., Chen, X., Zhang, Y., Wang, X., & Meng, X. (2021). Gallic acid: Pharmacological activities and molecular mechanisms involved in inflammation-related diseases. Biomedecine & Pharmacotherapie [Biomedicine & Pharmacotherapy], 133(110985), 110985. https://doi.org/10.1016/j.biopha.2020.110985
- [26] Choubey, S., Varughese, L. R., Kumar, V., & Beniwal, V. (2015). Medicinal importance of gallic acid and its ester
- derivatives: a patent review. Pharmaceutical Patent Analyst, 4(4), 305-315. https://doi.org/10.4155/ppa.15.14
- [27] Dhiman, S., & Mukherjee, G. (2021). Gallic acid (GA): A multifaceted biomolecule transmuting the biotechnology era. Recent Developments in Microbial Technologies (pp. 163–202). Springer Nature https://DOI:10.4155/ppa.15.14
- [28] Masoud, M. S., Hagagg, S. S., Ali, A. E., & Nasr, N. M. (2012). Synthesis and spectroscopic characterization of gallic complexes. Journal of Molecular Structure, of its azo https://doi.org/10.1016/j.molstruc.2012.01.041
- [29] Li, C., Wu, H., Masisi, K., Malunga, L. N., & Song, Y. (2020). Strawberries. In Nutritional Composition and Antioxidant Properties of Fruits and Vegetables (pp. 423-435). Elsevier. https://doi.org/10.1016/B978-0-12-812780-3.00026-X
- [30] Guo, L., Qiang, T., Ma, Y., Ren, L., & Dai, T. (2021). Purification and characterization of hydrolysable tannins extracted from Coriaria nepalensis bark using macroporous resin and their application in gallic acid production. Industrial Crops and Products, 162(113302), 113302. https://doi.org/10.1016/j.indcrop.2021.113302
- [31] Khatkar, A., Nanda, A., Kumar, P., & Narasimhan, B. (2017). Synthesis, antimicrobial evaluation and QSAR studies of gallic acid derivatives. Arabian Journal of Chemistry, 10, S2870-S2880. https://doi.org/10.1016/j.arabjc.2013.11.014
- [32] Forester, S. C., & Waterhouse, A. L. (2009). Metabolites are key to understanding health effects of wine polyphenolics. The Journal of Nutrition, 139(9), 1824S-1831S. https://doi.org/10.3945/jn.109.107664
- [33] Punithavathi, V. R., Prince, P. S. M., Kumar, R., & Selvakumari, J. (2011). Antihyperglycaemic, antilipid peroxidative and antioxidant effects of gallic acid on streptozotocin induced diabetic Wistar rats. European Journal of Pharmacology, 650(1), 465–471. https://doi.org/10.1016/j.ejphar.2010.08.059
- [34] Govea-Salas, M., Rivas-Estilla, A. M., Ascacio-Valdés, J., Zugasti-Cruz, A., Rodríguez-Herrera, R., Belmares-Cerda, R., & Morlett-Chávez, J. (2018). Gallic acid as a putative antioxidant in usage against liver disease. In The Liver (pp. 317-322). Elsevier. https://doi.org/10.1016/B978-0-12-803951-9.00026-4
- [35] Shen, Y., Guo, C., Lu, T., Ding, X.-Y., Zhao, M.-T., Zhang, M., Liu, H.-L., Song, L., & Zhou, D.-Y. (2021). Effects of gallic acid alkyl esters and their combinations with other antioxidants on oxidative stability of DHA algae oil. Food Research International (Ottawa, Ont.), 143(110280), 110280. https://doi.org/10.1016/j.foodres.2021.110280
- [36] Ojeaburu, S. I., & Oriakhi, K. (2021). Hepatoprotective, antioxidant and, anti-inflammatory potentials of gallic acid in tetrachloride-induced hepatic damage in Wistar rats. Toxicology Reports, 8, https://doi.org/10.1016/j.toxrep.2021.01.001
- [37] Chanwitheesuk, A., Teerawutgulrag, A., & Rakariyatham, N. (2005). Screening of antioxidant activity and antioxidant plants compounds some edible of Thailand. Food Chemistry, 92(3), https://doi.org/10.1016/j.foodchem.2004.07.035
- [38] Ghasemzadeh, A. (2011). Flavonoids and phenolic acids: Role and biochemical activity in plants and human. Journal of Medicinal Plant Research, 5(31). https://doi.org/10.5897/jmpr11.1404
- [39] Wen, W., Jia, X., Zhang, W., Jiang, X., & Fernie, A. R. (2022). Understanding carotenoid biosynthetic pathway control points using metabolomic analysis and natural genetic variation. In Carotenoids: Carotenoid and apocarotenoid biosynthesis metabolic engineering and synthetic biology (pp. 127-151). Elsevier. https://doi.org/10.1016/bs.mie.2022.03.015
- [40] Khan, Z., Javed, F., Shamair, Z., Hafeez, A., Fazal, T., Aslam, A., Zimmerman, W. B., & Rehman, F. (2021). Current developments in esterification reaction: A review on process and parameters. Journal of Industrial and Engineering Chemistry, 103, 80–101. https://doi.org/10.1016/j.jiec.2021.07.018
- [41] Delfanian, M., Sahari, M. A., Barzegar, M., Ahmadi Gavlighi, H., & Barba, F. J. (2023). Interfacial behavior of gallic acid and its alkyl esters in stripped soybean oil in combination with monoacylglycerol and phospholipid. Food Chemistry, 413(135618), 135618. https://doi.org/10.1016/j.foodchem.2023.135618
- [42] Otero-Pareja, M., Casas, L., Fernández-Ponce, M., Mantell, C., & Ossa, E. (2015). Green extraction of antioxidants from of varieties red grape pomace. Molecules (Basel, Switzerland), https://doi.org/10.3390/molecules20069686

- ______
- [43] Giglio, R. V., Patti, A. M., Cicero, A. F. G., Lippi, G., Rizzo, M., Toth, P. P., & Banach, M. (2018). Polyphenols: Potential use in the prevention and treatment of cardiovascular diseases. Current Pharmaceutical Design, 24(2), 239–258. https://doi.org/10.2174/1381612824666180130112652
- [44] Rosas, E. C., Correa, L. B., & das Graças Henriques, M. (2019). Antiinflammatory properties of Schinus terebinthifolius and its use in arthritic conditions. In Bioactive Food as Dietary Interventions for Arthritis and Related Inflammatory Diseases (pp. 489–505). Elsevier. https://doi.org/10.1016/B978-0-12-813820-5.00028-3
- [45] Das, A. B., Goud, V. V., & Das, C. (2019). Phenolic compounds as functional ingredients in beverages. In Value-Added Ingredients and Enrichments of Beverages (pp. 285–323). Elsevier. https://doi.org/10.1016/B978-0-12-816687-1.00009-6
- [46] Du, Y., Chen, H., Zhang, Y., & Chang, Y. (2014). Photodegradation of gallic acid under UV irradiation: Insights regarding the pH effect on direct photolysis and the ROS oxidation-sensitized process of DOM. Chemosphere, 99,254–260 https://doi.org/10.1016/j.chemosphere.2013.10.093
- [47] Mystrioti, C., Koursari, S., Xenidis, A., Papassiopi, N., 2021. Hexavalent chromium reduction by gallic aci Chemosphere. 273, 129737. https://doi.org/10.1016/j.chemosphere.2021.12973
- [48] Rocha, J. E., Guedes, T. T. A. M., Bezerra, C. F., Costa, M. do S., Campina, F. F., de Freitas, T. S., Souza, A. K., Sobral Souza, C. E., de Matos, Y. M. L. S., Pereira-Junior, F. N., da Silva, J. H., Menezes, I. R. A., Teixeira, R. N. P., Colares, A. V., & Coutinho, H. D. M. (2019). Identification of the gallic acid mechanism of action on mercury chloride toxicity reduction using infrared spectroscopy and antioxidant assays. International Biodeterioration & Biodegradation, 141, 24–29. https://doi.org/10.1016/j.ibiod.2018.07.002
- [49] Chang RM, Zhang YY, Zhang GB, Zhang XX, Chen AJ, Zhang W, Li Y. (2022) Application of thermal alkaline hydrolysis technology to improve the loading and in-vitro release of gallic acid in UiO-66. Food Chem. 15; 391:133238. https://doi:10.1016/j.foodchem.2022.133238. Epub 2022 May 18.PMID:35598392.
- [50] Crestini, C., & Lange, H. (2015). A novel and efficient immobilised tannase coated by the layer-by-layer technique in the hydrolysis of gallotannins and ellagitannins. Microchemical Journal, Devoted to the Application of Microtechniques in All Branches of Science, 123, 139–147. https://doi.org/10.1016/j.microc.2015.05.025
- [51] Garro^Galvez, J. M., Fechtal, M., & Riedl, B. (1996). Gallic acid as a model of tannins in condensation with formaldehyde. Thermochimica Acta, 274, 149–163. https://doi.org/10.1016/0040-6031(95)02630-4
- [52] Chen, L.-Y., Cheng, C.-W., & Liang, J.-Y. (2015). Effect of esterification condensation on the Folin–Ciocalteu method for the quantitative measurement of total phenols. Food Chemistry, 170, 10–15. https://doi.org/10.1016/j.foodchem.2014.08.038
- [53] Zhan, F., Hu, J., He, C., Sun, J., Li, J., & Li, B. (2020). Complexation between sodium caseinate and gallic acid: Effects on foam properties and interfacial properties of foam. Food Hydrocolloids, 99(105365), 105365. https://doi.org/10.1016/j.foodhyd.2019.105365
- [54] Masoud, M. S., Hagagg, S. S., Ali, A. E., & Nasr, N. M. (2012). Synthesis and spectroscopic characterization of gallic acid and some of its azo complexes. Journal of Molecular Structure, 1014, 17–25. https://doi.org/10.1016/j.molstruc.2012.01.041
- [55] Lu, L.-L., Li, Y.-H., & Lu, X.-Y. (2009). Kinetic study of the complexation of gallic acid with Fe(II). Spectrochimica Acta. Part A, Molecular and Biomolecular Spectroscopy, 74(3), 829–834. https://doi.org/10.1016/j.saa.2009.08.025
- [56] Assunção, M., Pinheiro, J., Cruz, S., Brazão, J., Queiroz, J., Eiras Dias, J. E., & Canas, S. (2019). Gallic acid, sinapic acid and catechin as potential chemical markers of Vitis graft success. Scientia Horticulturae, 246, 129–135. https://doi.org/10.1016/j.scienta.2018.10.056
- [57] Díaz-Gómez, R., López-Solís, R., Obreque-Slier, E., & Toledo-Araya, H. (2013). Comparative antibacterial effect of gallic acid and catechin against Helicobacter pylori. Lebensmittel-Wissenschaft Und Technologie [Food Science and Technology], 54(2), 331–335. https://doi.org/10.1016/j.lwt.2013.07.012
- [58] Díaz-Gómez, R., Toledo-Araya, H., López-Solís, R., & Obreque-Slier, E. (2014). Combined effect of gallic acid and catechin against Escherichia coli. Lebensmittel-Wissenschaft Und Technologie [Food Science and Technology], 59(2), 896–900. https://doi.org/10.1016/j.lwt.2014.06.049
- [59] Li, Y., Zhang, S., & Sun, Y. (2020). Measurement of catechin and gallic acid in tea wine with HPLC. Saudi Journal of Biological Sciences, 27(1), 214-221. $\underline{\text{https://doi.org/}10.1016/j.sjbs.2019.08.011}$
- [60] Oriakhi, K., & Orumwensodia, K. O. (2021). Combinatorial effect of Gallic acid and Catechin on some biochemical ampro-inflammatory markers in CCl4-mediated hepatic damage in rats. Phytomedicine plus: International Journal of Phytotherap and Phytopharmacology, 1(1), 100017 . https://doi.org/10.1016/j.phyplu.2020.100017
- [61] Bharathi, D., Dhanasekaran, S., Varshini, R., Bhuvaneswari, S., Periyasami, G., Pandiaraj, S., Lee, J., & Ranjithkumar, R. (2023). Preparation of gallotannin loaded chitosan/zinc oxide nanocomposite for photocatalytic degradation of organic dye and antibacterial applications. International Journal of Biological Macromolecules, 243(125052), 125052 https://doi.org/10.1016/j.ijbiomac.2023.125052
- [62] Campos, D., Chirinos, R., Huaraca-Espinoza, P., Aguilar-Galvez, A., García-Ríos, D., Pedreschi, F., & Pedreschi, R. (2024). Atmospheric immersion and vacuum impregnation of gallotannins and hydrolysed gallotannins from tara pods (Caesalpinia spinosa) mitigate acrylamide and enhances the antioxidant power in potato chips. Food Chemistry, 436(137675), 137675. https://doi.org/10.1016/j.foodchem.2023.137675
- [63] Guo, P., Tong, Y., Yang, R., Zhang, M., Lin, Q., Lin, S., & Wang, C. (2023). Effects of hydrolyzed gallotannin on intestinal physical barrier, immune function, and microbiota structure of yellow-feather broilers. Poultry Science, 102(11), 103010. https://doi.org/10.1016/j.psj.2023.103010

- [64] Huang, C., Li, K., Zhang, H., Zhang, W., Sun, Y., Xing, S., Shao, Y., Zhu, J., Chen, Y., & Chen, L. (2023). Chitosan immobilized Chinese gallotannin: A potential adsorbent to enrich the rare metals Ge (IV), Ga (III), and In (III). Journal of Environmental Chemical Engineering, 11(5), 110816. https://doi.org/10.1016/j.jece.2023.110816
- [65] Liu, L., Jia, W., Jiang, S., Zhang, G., Zhao, J., Xu, J., Wang, L., Wu, D., Tao, J., Yue, H., & Zhao, X. (2023). Inhibitory activities and rules of plant gallotannins with different numbers of galloyl moieties on sucrase, maltase and α-amylase in vitro and in vivo. Phytomedicine: International Journal of Phytotherapy and Phytopharmacology, 120(155063), 155063. https://doi.org/10.1016/j.phymed.2023.155063
- [66]Landete, J.M., 2011. Ellagitannins, ellagic acid and their derived metabolites: a review about source, metabolism, Food functions and health. Res Int [Internet]. 44 1150-60. https://www.sciencedirect.com/science/article/pii/S0963996911002572
- [67] Lorenzo, J.M., Barba, F.J., 2019. Sources, chemistry, and biological potential of ellagitannins and ellagic acid derivatives. Natural Products Chemistry. Studies in Elsevier; https://www.sciencedirect.com/science/article/pii/B9780444642418000064.
- [68] Alfei, S., Marengo, B., Dall'Asta, M., Pajno, R., Robino, A., Rossi, R., Arlorio, M., Ellagitannins: Bioavailability and biotransformation by human gut microbiota. J Funct Foods [Internet]. 2020 Jan [cited 2023 Apr 17]; 64:103658. Available from: https://www.sciencedirect.com/science/article/pii/S1756464619306107
- [69] Gao, X., Liu, Y., Liu, Y., Wang, Y., Liu, C-F., 2019. Ellagitannins in cancer prevention and therapy: chemistry, pharmacology and clinical applications. Pharmacol Res. 104505. https://www.sciencedirect.com/science/article/pii/S104366181931489X
- [70] Kowalska, K., Olejnik, A., 2020. Ellagitannins as natural modulators of the gut microbiota: mechanisms of action and health benefits. Crit Rev Food Sci Nutr. 60, 3438-51. https://www.tandfonline.com/doi/full/10.1080/10408398.2019.1682326 [71] Asghari-Varzaneh, E., Sharifian-Mobarakeh, S., & Shekarchizadeh, H. (2024). Enhancing hamburger shelf life and quality using gallic acid encapsulated in gelatin/tragacanth gum complex coacervate. Heliyon, 10(2), e24917. https://doi.org/10.1016/j.heliyon.2024.e24917
- [72] Azhar, B., Avian, C., & Tiwikrama, A. H. (2023). Green synthesis optimization with artificial intelligence studies of copper-gallic acid metal-organic framework and its application in dye removal from wastewater. Journal of Molecular Liquids, 389(122844), 122844. https://doi.org/10.1016/j.molliq.2023.122844
- [73] El-Megharbel, S. M., & Hamza, R. Z. (2022). Synthesis, spectroscopic characterizations, conductometric titration and investigation of potent antioxidant activities of gallic acid complexes with Ca (II), Cu (II), Zn(III), Cr(III) and Se (IV) metal ions. Journal of Molecular Liquids, 358(119196), 119196. https://doi.org/10.1016/j.molliq.2022.119196
- [74] Izadi, S., Tashkhourian, J., & Alireza Hosseini Hafshejani, S. (2024). Ecofriendly ratiometric colorimetric determination of mercury(II) ion in environmental watersamples using gallic acid-capped gold nanoparticles. Spectrochimica Acta. Part A, Molecular and Biomolecular Spectroscopy, 308(123778), 123778. https://doi.org/10.1016/j.saa.2023.123778
- [75] Yang, Y., Ma, Y., Wu, M., Wang, X., Zhao, Y., Zhong, S., Gao, Y., & Cui, X. (2024). Fe3+-induced coordination crosslinking gallic acid-carboxymethyl cellulose self-healing hydrogel. International Journal of Biological Macromolecules, 267(131626), 131626. https://doi.org/10.1016/j.ijbiomac.2024.131626
- [76] Liang, C., Wang, S., Hu, R., Huang, G., Xie, J., Zhao, B., Li, Y., Zhu, W., Guo, S., Jiang, J., & Hao, J. (2023). Molecular tracers, mass spectral tracers and oxidation of organic aerosols emitted from cooking and fossil fuel burning sources. The
- Science of the Total Environment, 868(161635), 161635. https://doi.org/10.1016/j.scitotenv.2023.161635 [77] Ou, B., Huang, D., Hampsch-Woodill, M., Flanagan, J. A., & Deemer, E. K. (2002). Analysis of antioxidant activities of common vegetables employing oxygen radical absorbance capacity (ORAC) and ferric reducing antioxidant power (FRAP) A comparative study. Journal of Agricultural and Food Chemistry, 50(11), 3122-3128. https://doi.org/10.1021/jf0116606
- [78] Sailaja Rao, P., Kalva, S., Yerramilli, A., & Mamidi, S. (2011). Free radicals and tissue damage: Role of antioxidants. Free Radicals and Antioxidants, 1(4), 2-7. https://doi.org/10.5530/ax.2011.4.2
- [79] Wang, J., Jiang, H., Jiang, H., Mo, Y., Geng, X., Li, J., Mao, S., Bualert, S., Ma, S., Li, J., & Zhang, G. (2020). Source apportionment of water-soluble oxidative potential in ambient total suspended particulate from Bangkok: Biomass burning versus fossil fuel combustion. Atmospheric Environment (Oxford, England: 1994), 235(117624), 117624 https://doi.org/10.1016/j.atmosenv.2020.117624
- [80] Blondeel, M., Bradshaw, M. J., Bridge, G., & Kuzemko, C. (2021). The geopolitics of energy system transformation: A review. Geography Compass, 15(7). https://doi.org/10.1111/gec3.12580
- [81] Lam, V., Li, G., Song, C., Chen, J., Fairbridge, C., Hui, R., & Zhang, J. (2012). A review of electrochemical desulfurization technologies for fossil fuels. Fuel Processing Technology, https://doi.org/10.1016/j.fuproc.2012.01.022
- [82] Nie, X., Zhao, T., & Su, Y. (2021). Fossil fuel carbon contamination impacts soil organic carbon estimation in cropland. Catena, 196(104889), 104889. https://doi.org/10.1016/j.catena.2020.104889
- [83] Ranzi, E., Frassoldati, A., Stagni, A., Pelucchi, M., Cuoci, A., & Faravelli, T. (2014). Reduced kinetic schemes of complex reaction systems: Fossil and biomass-derived transportation fuels. International Journal of Chemical Kinetics, 46(9), 512-542. https://doi.org/10.1002/kin.20867
- [84] Tashie-Lewis, B. C., & Nnabuife, S. G. (2021). Hydrogen production, distribution, storage and power conversion in a hydrogen economy - A technology review. Chemical Engineering Journal Advances, 8(100172), 100172. https://doi.org/10.1016/j.ceja.2021.100172

- ______
- [85] Abbasi, K. R., Zhang, Q., Ozturk, I., Alvarado, R., & Musa, M. (2024). Energy transition, fossil fuels, and green innovations: Paving the way to achieving sustainable development goals in the United States. Gondwana Research: International Geoscience Journal, 130, 326–341. https://doi.org/10.1016/j.gr.2024.02.005
- [86] Ali, M. A., Islam, M. A., Othman, N. H., Noor, A. M., Hossen, J., & Ibrahim, M. (2019). Effect of heating on compositional characteristics and oxidative stability of crude and refined rice bran oil. Journal of Oleo Science, 68(11), 1085–1097. https://doi.org/10.5650/jos.ess19140
- [87] Knothe, G., & Steidley, K. R. (2005). Kinematic viscosity of biodiesel fuel components and related compounds. Influence of compound structure and comparison to petrodiesel fuel components. Fuel, 84(9), 1059–1065. https://doi.org/10.1016/j.fuel.2005.01.016
- [88] Mohamed Shameer, P., & Ramesh, K. (2017). FTIR assessment and investigation of synthetic antioxidant on the fuel stability of Calophyllum inophyllum biodiesel. Fuel, 209, 411–416. https://doi.org/10.1016/j.fuel.2017.08.006
- [89] Srivastava, Y., Singh, B., Kaur, B., Ubaid, M., & Semwal, A. D. (2024). Kinetic study of thermal degradation of flaxseed oil and moringa oil blends with physico-chemical, oxidative stability index (OSI) and shelf-life prediction. Journal of Food Science and Technology, 61(4), 675–687. https://doi.org/10.1007/s13197-023-05868-z
- [90] Khorsheed Zangana, L. M., Yaseen, A. H., Hassan, Q. H., Mohammed, M. M., Mohammed, M. F., & Alalwan, H. A. (2023). Investigated kerosene-diesel fuel performance in internal combustion engine. Cleaner Engineering and Technology, 12(100591), 100591. https://doi.org/10.1016/j.clet.2022.100591
- [91] Manimaran, S., Athimoolam, A., Albeshr, M. F., Praveenkumar, T. R., & Beata, G. (2024). Effects of nano-additives and ammonia on performance and emission characteristics of diesel engine fueled with Aleurites moluccanus. Fuel, 363(130787), 130787. https://doi.org/10.1016/j.fuel.2023.130787
- [92] Mofijur, M., Ahmed, S. F., Ahmed, B., Mehnaz, T., Mehejabin, F., Shome, S., Almomani, F., Chowdhury, A. A., Kalam, M. A., Badruddin, I. A., & Kamangar, S. (2024). Impact of nanoparticle-based fuel additives on biodiesel combustion: An analysis of fuel properties, engine performance, emissions, and combustion characteristics. Energy Conversion and Management: X, 21(100515), 100515. https://doi.org/10.1016/j.ecmx.2023.100515
- [93] Mohite, A., Jyoti Bora, B., Sharma, P., Medhi, B. J., Barik, D., Balasubramanian, D., Nguyen, V. G., Js, F. J., Cuong Le, H., Kamalakannan, J., Varuvel, E. G., & Cao, D. N. (2024). Maximizing efficiency and environmental benefits of an algae biodiesel-hydrogen dual fuel engine through operational parameter optimization using response surface methodology. International Journal of Hydrogen Energy, 52, 1395–1407. https://doi.org/10.1016/j.ijhydene.2023.10.134
- [94] Youssef, A., & Ibrahim, A. (2024). An experimental evaluation for the performance of a single cylinder CI engine fueled by a Diesel-Biodiesel blend with alcohols and Zinc-Aluminate nanoparticles as additives. Materials Today: Proceedings. https://doi.org/10.1016/j.matpr.2024.04.015
- [95] Chai, J., Yang, Y., Wang, S., & Lai, K. K. (2016). Fuel efficiency and emission in China's road transport sector: Induced effect and rebound effect. Technological Forecasting and Social Change, 112, 188–197. https://doi.org/10.1016/j.techfore.2016.07.005
- [96] García-Martos, C., Rodríguez, J., & Sánchez, M. J. (2013). Modelling and forecasting fossil fuels, CO2 and electricity prices and their volatilities. Applied Energy, 101, 363–375. https://doi.org/10.1016/j.apenergy.2012.03.046
- [97] Meng, X., Sørensen, P., Møller, H. B., & Petersen, S. O. (2023). Greenhouse gas balances and yield-scaled emissions for storage and field application of organic fertilizers derived from cattle manure. Agriculture, Ecosystems & Environment, 345(108327), 108327. https://doi.org/10.1016/j.agee.2022.108327
- [98]AL Zahrani, N. A., El-Shishtawy, R. M., & Asiri, A. M. (2020). Recent developments of gallic acid derivatives and their hybrids in medicinal chemistry: A review. European Journal of Medicinal Chemistry, 204(112609), 112609. https://doi.org/10.1016/j.ejmech.2020.112609
- [99] Chen, X., Yang, Y., Ye, G., Liu, S., & Liu, J. (2023). Chiral ruthenium nanozymes with self-cascade reaction driven the NO generation induced macrophage M1 polarization realizing the lung cancer "cocktail therapy." Small, 19 (28). https://doi.org/10.1002/smll.202207823
- [100] Baite, T. N., Mandal, B., & Purkait, M. K. (2024). Exploring gallic acid-rich leaf extract: Formulation and characterization of antioxidant blends. Measurement: Food, 14(100162), 100162. https://doi.org/10.1016/j.meafoo.2024.100162
- [101] Liu, Q., Peng, D., Wei, P., Song, H., Cong, C., Meng, X., & Zhou, Q. (2023). Synthesis of macromolecular antioxidants containing thioether and aromatic secondary amine to improve the anti-oxidation properties of EPDM. Polymer Degradation and Stability, 218(110585), 110585. https://doi.org/10.1016/j.polymdegradstab.2023.110585
- [102] Zhang, S., Cheng, Z., Cao, Y., He, F., Zhao, L., Baqar, M., Zhu, H., Zhang, T., & Sun, H. (2024). Aromatic amine antioxidants (AAs) and p-phenylenediamines-quinones (PPD-Qs) in e-waste recycling industry park: Occupational exposure and liver X receptors (LXRs) disruption potential. Environment International, 186(108609), 108609. https://doi.org/10.1016/j.envint.2024.108609
- [103] Mehrabani, M., Lotfian sargazi, M., Amirkhosravi, A., Farhadi, S., Vasei, S., Raeiszadeh, M., & Mehrabani, M. (2023). The influence of harvest time on total phenolic and flavonoid contents, antioxidant, antibacterial and cytotoxicity of Rheum khorasanicum root extract. Annales Pharmaceutiques Francaises,81(3),475–483 . https://doi.org/10.1016/j.pharma.2022.11.010
- [104] Papas, A. M. (2019). Vitamin E: Tocopherols and Tocotrienols. In Antioxidant Status, Diet, Nutrition, and Health (pp. 188–210). CRC Press. https://doi.org/10.1201/9780367811099-10
- [105] Sutanto, Susanto, & Nasikin. (2019). Solubility and antioxidant potential of a pyrogallol derivative for biodiesel additive. Molecules (Basel, Switzerland), 24(13), 2439. https://doi.org/10.3390/molecules24132439

- [106] Varatharajan, K., & Pushparani, D. S. (2018). Screening of antioxidant additives for biodiesel fuels. Renewable and Sustainable Energy Reviews, 82, 2017–2028. https://doi.org/10.1016/j.rser.2017.07.020
- [107] Longanesi, L., Pereira, A. P., Johnston, N., & Chuck, C. J. (2022). Oxidative stability of biodiesel: recent insights. Biofuels, Bioproducts & Biorefining: Biofpr, 16(1), 265–289. https://doi.org/10.1002/bbb.2306
- [108] Noubigh, A., Jeribi, C., Mgaidi, A., & Abderrabba, M. (2012). Solubility of gallic acid in liquid mixtures of (ethanol+water) from (293.15 to 318.15)K. The Journal of Chemical Thermodynamics, 55, 75-78. https://doi.org/10.1016/j.jct.2012.06.022
- [109] Yen, G.C., Duh, P.D., Tsai, H.L., 2002. Antioxidant and pro-oxidant properties of ascorbic acid and gallic acid. Food Chem. 79, 307-313. https://doi:10.1016/S0308-8146(02)00145-0
- [110] Delfanian, M., Sahari, M. A., Barzegar, M., & Ahmadi Gavlighi, H. (2021). Structure–antioxidant activity relationships of gallic acid and phloroglucinol. Journal of Food Measurement & Characterization, 15(6), 5036-5046. https://doi.org/10.1007/s11694-021-01045-y
- [11] Xia, T., Wang, Y., Wang, B., Yang, Z., Pan, G., Zhang, L., & Zhang, J. (2019). Natural compounds Gallic acid derivatives for long-life Li/Na organic batteries. ChemElectroChem, 6(18),https://doi.org/10.1002/celc.201901064
- [112] Belin, F., Barthélémy, P., Ruiz, K., Lacombe, J. M., & Pucci, B. (2003). Synthetic Gallic acid derivatives as models comprehensive study of antioxidant activity. HelveticaChimicaActa,86(2),247–265. https://doi.org/10.1002/hlca.200390053
- [113] Tomi, I. H. R., Ali, G. Q., Jawad, A. H., & Yousif, E. (2017). Synthesis and characterization of gallic acid derivatives and their utilized as organic photo-stabilizers for poly (vinyl chloride). Journal of Polymer Research, 24(8). https://doi.org/10.1007/s10965-017-1283-7
- [114] Xu, J., Liu, X., & Fu, S. (2022). Bio-based epoxy resin from gallic acid and its thermosets toughened with renewable tannic acid derivatives. Journal of Materials Science, 57(20), 9493-9507. https://doi.org/10.1007/s10853-022-07174-z
- [115] Freiría-Gándara, J., Losada-Barreiro, S., Paiva-Martins, F., & Bravo-Díaz, C. (2018). Enhancement of the antioxidant efficiency of gallic acid derivatives in intact fish oil-in-water emulsions through optimization of their interfacial concentrations. Food & Function, 9(8), 4429–4442. https://doi.org/10.1039/c8fo00977e
- [116] Liu, H., Wang, Q., Xing, L., Zhang, Y., Zhang, T., Ran, W., & Cao, J. (2021). Measurement report: quantifying source contribution of fossil fuels and biomass-burning black carbon aerosol in the southeastern margin of the Tibetan Plateau. Atmospheric Chemistry and Physics, 21(2), 973–987. https://doi.org/10.5194/acp-21-973-2021
- [117] Odziemkowska, M., Czarnocka, J., & Wawryniuk, K. (2018). Study of stability changes of model fuel blends. In Improvement Trends for Internal Combustion Engines. InTech. https://doi.org/10.5772/67056
- [118] Pradelle, F., Braga, S. L., Martins, A. R. F. A., Turkovics, F., & Pradelle, R. N. C. (2015). Gum formation in gasoline and its blends: A review. Energy & Fuels: An American Chemical Society Journal, 29(12), 7753-7770. https://doi.org/10.1021/acs.energyfuels.5b01894
- Jin, Q., Zhu, W., Jiang, D., Zhang, R., Kutyreff, C. J., Engle, J. W., Huang, P., Cai, W., Liu, Z., & Cheng, L. (2017). Ultra-small iron-gallic acid coordination polymer nanoparticles for chelator-free labeling of 64Cu and multimodal imagingguided photothermal therapy. Nanoscale, 9(34), 12609–12617. https://doi.org/10.1039/c7nr03086i
- ASTM D381-22. Standard Test Method for Gum Content in Fuels by Jet Evaporation. West Conshohocken, PA: ASTM International; 2022. https://www.astm.org/d0381-22.html
- ASTM D873-22. Standard Test Method for Oxidation Stability of Aviation Fuels (Potential Residue Method). West Conshohocken, PA: ASTM International; 2019. https://www.astm.org/d0873-22.html
- IP 131: Petroleum products Gum content of light and middle distillate fuels Jet evaporation method. IP 131: Petroleum products - Gum content of light and middle distillate fuels - Jet evaporation method | EI - Publishing (energyinst.org)
- ASTM D2274-14 (2019). Standard Test Method for Oxidation Stability of Distillate Fuel Oil (Accelerated Method). West Conshohocken, PA: ASTM International; 2019. https://www.astm.org/doi:10.1520/D2274-14R19
- ASTM D525-19. Standard Test Method for Oxidation Stability of Gasoline (Induction Period Method). West [124] Conshohocken, PA: ASTM International; 2019. https://www.astm.org/doi:10.1520/D0525-12AR19
- ASTM D2272-22. Standard Test Method for Oxidation Stability of Steam Turbine Oils by Rotating Pressure Vessel. West Conshohocken, PA: ASTM International;2014. https://www.astm.org/doi:10.1520/D2272-22
- [126] EN 14112:2020. Fat and oil derivatives Fatty Acid Methyl Esters (FAME) Determination of oxidation stability (accelerated oxidation test). Brussels: European Committee for Standardization; 2020. BS EN 14112:2020 Fat and oil derivatives. Fatty Acid Methyl Esters (FAME). Determination of oxidation stability (accelerated oxidation test) (enstandard.eu)
- [127] ASTM D664-18e02. Standard Test Method for Acid Number of Petroleum Products by Potentiometric Titration. West Conshohocken, PA: ASTM International; 2018. https://www.astm.org/d0664-18e02.html
- ASTM D974-21. Standard Test Method for Acid and Base Number by Color-Indicator Titration. West Conshohocken, PA: ASTM International; 2020. https://www.astm.org/d0974-21.html
- [129] IP 139/18. Determination of acid number Colour-indicator titration method. London: Energy Institute; 2018. IP 139: Petroleum products and lubricants - Determination of acid or base number - Colour-indicator titration method | EI - Publishing (energyinst.org)
- [130] ASTM D3703-18. Standard Test Method for Hydroperoxide Number of Aviation Turbine Fuels, Gasoline and Diesel Fuels. West Conshohocken, PA: ASTM International; 2018. https://www.astm.org/d3703-18.html

16.html

- [131] ASTM D2344/D2344M-16. Standard Test Method for Short-Beam Strength of Polymer Matrix Composite Materials and Their Laminates. West Conshohocken, PA: ASTM International; 2016. https://www.astm.org/d2344_d2344m-
- [132] ISO 3960:2020. Animal and vegetable fats and oils Determination of peroxide value Iodometric (visual) endpoint determination. Geneva: International Organization for Standardization; 2020. https://www.iso.org/obp/ui/#iso:std:iso:3960:ed-5:v1:en
- [133] Leroy, V., Cancellieri, D., & Leoni, E. (2006). Thermal degradation of ligno-cellulosic fuels: DSC and TGA studies. Thermochimica Acta, 451(1–2), 131–138. https://doi.org/10.1016/j.tca.2006.09.017
- [134] Olivella, M. A., Palacios, J. M., Vairavamurthy, A., del Río, J. C., & de las Heras, F. X. C. (2002). A study of sulfur functionalities in fossil fuels using destructive- (ASTM and Py–GC–MS) and non-destructive- (SEM–EDX, XANES and XPS) techniques. Fuel (London, England), 81(4), 405–411. https://doi.org/10.1016/s0016-2361(01)00198-3
- [135] He, Y.-F., Wang, R.-M., Liu, Y.-Y., Chang, Y., Wang, Y.-P., Xia, C.-G., & Suo, J.-S. (2000). Study on oxidation mechanism of cumene based on GC-MS analysis. Journal of Molecular Catalysis. A, Chemical, 159(1), 109–113. https://doi.org/10.1016/s1381-1169(00)00178-3
- [136] Rontani, J.-F. (2022). Use of gas chromatography-mass spectrometry techniques (GC-MS, GC-MS/MS and GC-QTOF) for the characterization of photooxidation and autoxidation products of lipids of autotrophic organisms in environmental samples. Molecules (Basel, Switzerland), 27(5), 1629. https://doi.org/10.3390/molecules27051629
- [137] Robbat, A., Jr, Considine, T., & Antle, P. M. (2010). Subsurface detection of fossil fuel pollutants by photoionization and gas chromatography/mass spectrometry. Chemosphere, 80(11), 1370–1376. https://doi.org/10.1016/j.chemosphere.2010.06.005
- [138] Zhan, D., & Fenn, J. B. (2000). Electrospray mass spectrometry of fossil fuels. International Journal of Mass Spectrometry, 194(2–3), 197–208. https://doi.org/10.1016/s1387-3806(99)00186-4
- [139] Arsianti, A. A., Astuty, H., Fadilah, F., Bahtiar, A., Tanimoto, H., & Kakiuchi, K. (2017). Design and screening of Gallic acid derivatives as inhibitors of malarial dihydrofolate reductase (dhfr) by in silico docking. Asian Journal of Pharmaceutical and Clinical Research, 10(2), 330. https://doi.org/10.22159/ajpcr.2017.v10i2.15712
- [140] Humaedi, A., Arsiyanti, A., & Radji, M. (2017). In silico molecular docking study of Gallic acid and its derivatives as inhibitor BRAF colon cancer. International Journal of Chemtech Research, 10(1), 310–315. http://repository.binawan.ac.id/id/eprint/848
- [141] Kokila, N. R., Mahesh, B., Ramu, R., Mruthunjaya, K., Bettadaiah, B. K., & Madhyastha, H. (2023). Inhibitory effect of gallic acid from Thunbergia mysorensis against α -glucosidase, α -amylase, aldose reductase and their interaction: Inhibition kinetics and molecular simulations. Journal of Biomolecular Structure & Dynamics, 41(20), 10642–10658. https://doi.org/10.1080/07391102.2022.2156923
- [142] Raghi, K. R., Sherin, D. R., Saumya, M. J., Arun, P. S., Sobha, V. N., & Manojkumar, T. K. (2018). Computational study of molecular electrostatic potential, docking and dynamics simulations of gallic acid derivatives as ABL inhibitors. Computational Biology and Chemistry, 74, 239–246. https://doi.org/10.1016/j.compbiolchem.2018.04.001
- [143]Umar, H. I., Siraj, B., Ajayi, A., Jimoh, T. O., & Chukwuemeka, P. O. (2021). Molecular docking studies of some selected gallic acid derivatives against five non-structural proteins of novel coronavirus. Journal, Genetic Engineering & Biotechnology, 19(1), 16. https://doi.org/10.1186/s43141-021-00120-7
- [144] Zhan, M., Guo, M., Jiang, Y., & Wang, X. (2015). Characterization of the interaction between Gallic acid and lysozyme by molecular dynamics simulation and optical spectroscopy. International Journal of Molecular Sciences, 16(12), 14786–14807. https://doi.org/10.3390/ijms160714786
- [145] Nie, R., Huo, Y.-Q., Yu, B., Liu, C.-J., Zhou, R., Bao, H.-H., & Tang, S.-W. (2020). Molecular insights into the inhibitory mechanisms of gallate moiety on the A β 1–40 amyloid aggregation: A molecular dynamics simulation study. International Journal of Biological Macromolecules, 156, 40–50. https://doi.org/10.1016/j.ijbiomac.2020.04.007
- [146] Zhang, Y., Wang, X., Lu, B., Gao, Y., Zhang, Y., Li, Y., Niu, H., Fan, L., Pang, Z., & Qiao, Y. (2022). Functional and binding studies of gallic acid showing platelet aggregation inhibitory effect as a thrombin inhibitor. Chinese Herbal Medicines, 14(2), 303–309. https://doi.org/10.1016/j.chmed.2021.09.001
- [147] Cappelli, C., Mennucci, B., & Monti, S. (2005). Environmental effects on the spectroscopic properties of Gallic acid: A combined classical and quantum mechanical study. The Journal of Physical Chemistry. A, 109(9), 1933–1943. https://doi.org/10.1021/jp044781s
- [148] Badhani, B., & Kakkar, R. (2017). DFT study of structural and electronic properties of gallic acid and its anions in gas phase and in aqueous solution. Structural Chemistry, 28(6), 1789–1802. https://doi.org/10.1007/s11224-017-0958-3
- [149] Vahedi, S. Z., Farhadian, S., Shareghi, B., & Asgharzadeh, S. (2024). Thermodynamic and functional changes of alphachymotrypsin after interaction with gallic acid. Spectrochimica Acta. Part A, Molecular and Biomolecular Spectroscopy, 313(124109), 124109. https://doi.org/10.1016/j.saa.2024.124109
- [150] Pardeshi, S., Dhodapkar, R., & Kumar, A. (2013). Quantum chemical density functional theory studies on the molecular structure and vibrational spectra of Gallic acid imprinted polymers. Spectrochimica Acta. Part A, Molecular and Biomolecular Spectroscopy, 116, 562–573. https://doi.org/10.1016/j.saa.2013.07.067
- [151] Guendouzi, O., Guendouzi, A., Ouici, H. B., Brahim, H., Boumediene, M., & Elkeurti, M. (2020). A quantum chemical study of encapsulation and stabilization of gallic acid in β-cyclodextrin as a drug delivery system. Canadian Journal of Chemistry, 98(4), 204–214. https://doi.org/10.1139/cjc-2019-0464
- [152] Ghouari, N., Benali-Cherif, R., Takouachet, R., Falek, W., Missaoui, D., Rahmouni, A., Bendeif, E.-E., & Benali-Cherif, N. (2023). Crystal engineering of a new pharmaceutical polymorph of gallic acid monohydrate: a structural

 $comparative \ study \ and \ chemical \ computational \ quantum \ investigations. \ CrystEngComm, \ 25(45), \ 6279-6290. \\ \underline{https://doi.org/10.1039/d3ce00766a}$

[153] Zhan M, Guo M, Jiang Y, Wang X. Characterization of the interaction between Gallic acid and lysozyme by molecular dynamics simulation and optical spectroscopy. Int J Mol Sci. 2015; 16(12):14786–807. http://dx.doi.org/10.3390/ijms160714786