Journal of Applied Veterinary Sciences, 10 (3): 118-128 (July, 2025).

ISSN: Online: 2090-3308, Print: 1687-4072

Journal homepage: https://javs.journals.ekb.eg



Potential Application of Saccharomyces cerevisiae as an Antioxidant Agent for Broiler Chickens – A Brief Overview

Sugiharto Sugiharto*, Ikania Agusetyaningsih, Endang Widiastuti and Turini Yudiarti Faculty of Animal and Agricultural Sciences, Universitas Diponegoro, Tembalang Campus, Semarang 50275, Central Java, Indonesia

*Corresponding author: Sugiharto Sugiharto, E-mail: sgh undip@yahoo.co.id

ABSTRACT

Broiler chicken strains and the intensive production system are typically attributed to the stress conditions in broiler production. The use of synthetic antioxidants is commonly practiced by the farmers to alleviate the adverse effects of stress on broiler chickens. Yet, excessive use of synthetic antioxidants can leave residues in broiler meats, which can be harmful to consumer health. As a probiotic, yeast Saccharomyces cerevisiae has been widely used as a feed additive by broiler producers. Beyond its probiotic properties, S. cerevisiae has also been demonstrated to have antioxidant properties and may therefore be exploited as an antioxidant agent for broiler chickens. However, the in-depth understanding about the potential use of S. cerevisiae as an antioxidant agent for broiler chickens is still limited. Hence, the overview regarding the application of S. cerevisiae as an alternative antioxidant agent for broiler chickens is of importance. The current review article elucidates the antioxidant potential of S. cerevisiae, the use of S. cerevisiae to increase the antioxidant activity of fermentation products, as well as the application of S. cerevisiae and its derivative products as an antioxidant agent in broiler production. Moreover, the potential application of S. cerevisiae as an antiinflammatory agent for broiler chickens is also briefly discussed in this current review.

Review Article:

DOI:https://dx.doi.org/10.21608/j avs.2025.381713.1604

Received: 04 May, 2025. Accepted: 05 June, 2025. Published in July, 2025.

This is an open access article under the term of the Creative Commons Attribution 4.0 (CC-BY) International License . To view a copy of this license, visit:

http://creativecommons.org/licenses/by/4

Keywords: Broiler, Gut health, Inflammation, Natural antioxidant, Stress, β-glucans.

J. Appl. Vet. Sci., 10(3): 118-128.

INTRODUCTION

Broiler production is one of the sectors that plays an important role in meeting society's animal protein needs while also contributing positively to the economy of various countries (Sumanu et al., 2022). Farmers are currently raising broiler chickens intensively and in large numbers in order to achieve economic efficiency (Sugiharto, 2022). According to Gržinić et al. (2023), the intensive rearing system for broiler chickens (indoor open floor housing) can cause stress due to uncomfortable conditions. These stress conditions can be exacerbated when broiler chickens are overcrowded (Sugiharto, 2022).

Other factors that may pose a threat to the comfort include climate change and global warming, which can result in heat stress in broiler chickens, particularly those housed in open houses (Gržinić et al., 2023). The use of environmentally controlled broiler houses is an alternative that can protect broiler chickens from environmental stressors. However, using this type of broiler house is very expensive, so not all farmers can afford it.

Oxidative stress in broilers is a significant concern due to its detrimental impact on productivity and overall health. It can result from various environmental and physiological stressors (Sugiharto, 2019; Gržinić et al., 2023). Heat stress is one of the primary contributors to oxidative stress in broilers. One of the consequences of heat stress is the increase in reactive oxygen species (ROS), which are by-products of normal cellular respiration. When the production of ROS surpasses the capacity of antioxidant defenses, oxidative stress occurs, resulting in cellular damage, such as lipid peroxidation, protein oxidation, and DNA damage (Shakeri et al., 2022; Apalowo et al., 2024). Dietary intervention with synthetic antioxidants (e.g., butylated hydroxytoluene [BHT], butylated hydroxyanisole [BHA], ethoxyquin, etc.) is commonly applied to reduce the negative impact of stress on poultry. However, excessive use of synthetic antioxidants can leave residues in broiler meats, which can be harmful to consumer health (Sugiharto, 2019; Xu et al., 2021).

Farmers have been using yeast Saccharomyces cerevisiae as a feed additive following the prohibition of antibiotic growth promoters in broiler production. Numerous studies have demonstrated that S. cerevisiae is an effective probiotic that can enhance the health and productivity of broiler chickens (Sugiharto et al., 2019; Sumanu et al., 2023). Apart from its probiotic properties, S. cerevisiae has also been shown to have antioxidant properties that can ameliorate the negative effects of stress on broiler chickens (Het et al., 2022; Sumanu et al., 2023). In this respect, Aluwong et al., (2013) and Abo-Sriea et al., (2024) confirmed that selenium, β-glucans, and mannan oligosaccharides (MOS) found in S. cerevisiae have the capacity to scavenge excess free radicals or ROS in the bodies of broiler chickens under stress.

Currently, the published reviews discussing the potential use of *S. cerevisiae* as an antioxidant agent for broiler chickens are still lacking. Therefore, this article provides a short overview regarding the application of *S. cerevisiae* as an alternative antioxidant agent for broiler chickens.

Stress in broiler chicken production

Broiler chicken strains are types or strains of chicken that have the genetic potential to grow very quickly. However, despite their very high growth potential, broiler strains have several weaknesses, including poor cardiovascular health, a high prevalence of mortality, and being highly susceptible to ascites. Broiler strains are also highly susceptible to stress (e.g., heat stress and high-density stress) and have low welfare (Riber and Wurtz, 2024). In addition to stress due to high stocking density, intensive broiler chicken production is often negatively affected by excessive ammonia production in the broiler house. Guo et al. (2023) reported that excessive ammonia inhaled by chickens can cause an imbalance of antioxidants and oxidants in the broiler chicken's body, which can cause oxidative stress and immunosuppression. In fact, ammonia is the result of the decomposition of uric acid and undigested protein by broiler chickens involving special microorganisms (Sugiharto et al., 2023). Ammonia production in the broiler house can be higher along with stress on the chicken, high stocking density, excessive protein content in the feed, high temperature, and poor litter management (Sugiharto et al., 2023).

The imbalance between growth potential and immune capacity to face immune challenges or incoming pathogenic microorganisms makes broiler strains very susceptible to immune stress and disease during the rearing period (Aylward et al., 2024). The term immune stress refers to several external factors that can affect chickens during the rearing period, such as pathogenic microorganisms, vaccination, and excessive

use of drugs. These factors can affect the body's immune system and disrupt immune homeostasis in the body of broiler chickens (Wang et al., 2023). Like stress due to heat or high stocking density, immune stress also has a negative effect on the productivity and health of broiler chickens.

Free radicals, especially ROS, play a very important role in the body's immune response to incoming pathogens. Typically, ROS are naturally produced in broiler chickens and can be exacerbated by stress (Shakeri et al., 2022; Apalowo et al., 2024). Indeed, excessive ROS production can pose a threat to the body because ROS has the potential to damage proteins and nucleic acids, resulting in tissue damage and causing negative effects on the physiological conditions, health and growth of broiler chickens (Wang et al., 2021). To prevent the body from oxidative damage due to stress, the animal body has an enzymatic (superoxide dismutase [SOD], glutathione peroxidase [GPx], catalase [CAT]) and non-enzymatic (glutathione [GSH]) antioxidant system in cells. SOD can convert superoxide anions into oxygen and hydrogen peroxide, which are then broken down into water by antioxidant enzymes including GPx and CAT. GPx plays a role in protecting the structure and function of cell membranes from disruption and oxide damage (Luo et al., 2003). However, when the activity of antioxidant enzymes decreases, polyunsaturated fatty acids will be attacked by ROS. The latter condition will lead to lipid peroxidation, resulting in malondialdehyde (MDA) production (Cheng et al., 2020).

Antioxidant potential of S. cerevisiae

Farmers have frequently employed synthetic antioxidants to mitigate the detrimental effects of stress on the health, productivity, and physiological conditions of broiler chickens. In addition to being applied as antistress agents for broiler chickens, synthetic antioxidants are also often applied to feed (mixed into feed) to inhibit or prevent nutrient oxidation in feed so that feed quality is maintained (Sithole *et al.*, 2023). However, Sugiharto (2019) and Xu *et al.* (2021) have confirmed that the use of synthetic antioxidants in excess or improperly can result in residue on meats, which could pose a health risk to consumers. In this respect, excessive use of butylated hydroxytoluene (BHT) can induce apoptosis and encourage tumorigenesis.

Likewise, butylated hydroxyanisole (BHA) metabolites are also involved in the carcinogenicity process (**Xu** et al., 2021). Based on these conditions and to ensure the sustainability of the broiler chicken industry, it is necessary to find alternative natural antioxidants to replace synthetic antioxidants for broiler chickens. Several strategies have been implemented, including using natural ingredients as a source of

antioxidants for broiler chickens. The use of natural antioxidant sources is expected to be effective in alleviating the negative effects of stress on broiler chickens, as well as providing proof that broiler chicken products are safe for consumers who are becoming more health conscious. Among these natural antioxidants, broiler farmers often use plant extracts or herbal ingredients as a source of natural antioxidants for broiler chickens (**Zdanowska-Sąsiadek** *et al.*, **2019**).

Another studies also report the potential of probiotic microorganisms and postbiotics (metabolites that are produced by probiotic microorganisms and exhibit biological activity to the host) as alternative sources of natural antioxidants for broiler chickens (Rakngam et al., 2024). Probiotics are defined as live microorganisms that, when administered in adequate amounts, confer a health benefit on the host (Sugiharto, 2016; 2022). According to Sugiharto (2016), probiotics can improve the health and productivity of broiler chickens by boosting the immune system, colonization resistance against infections, and the host intestine microbial balance. Lactic acid bacteria, Bacillus species, and specific yeast strains are some of the most commonly employed microorganisms as probiotics for broiler chickens (Sugiharto, 2016; 2019).

S. cerevisiae is a yeast that has been widely known by farmers to have the potential as a probiotic for broiler chickens. S. cerevisiae is also reported to have the potential as a prebiotic for broiler chickens so that it can replace the role of antibiotic growth promoters (Ahiwe et al., 2021). In addition, several studies have reported the potential of S. cerevisiae as a source of natural antioxidants that are very potential to be applied to broiler chickens (Attia et al., 2022; Mohamed et al., 2022). Makky et al., (2021) reported that the antioxidant potential of S. cerevisiae is closely related to the secondary metabolites produced by the yeast, namely tryptophol. Tryptophol is a metabolic derivative of tryptophan that has the ability to scavenge free radicals in the body. Fakruddin et al., (2017) further reported that S. cerevisiae exhibits very strong metal chelating activity so that it can function as a very potential source of antioxidants. In this case, the ability of metal chelating activity is very important because it can reduce the concentration of metals that have a catalytic effect on lipid peroxidation.

In addition, metal chelating agents are considered secondary antioxidants because they can reduce the oxidation-reduction (redox) potential and thus stabilize oxidized metal ions (Fakruddin et al., 2017). S. cerevisiae is also a source of GSH, which is an endogenous antioxidant compound found in all eukaryotic cells (Santos et al., 2022). In this case, GSH can function as an antioxidant in conditions where

broiler chickens experience stress due to its low reduction potential. GSH can also provide a protective effect against free radicals, either by reacting directly with free radicals or acting as a cofactor for redox enzymes such as glutathione reductase, GPx, glutaredoxin (thioltransferase: serves a crucial role in cellular redox regulation), and glutathione S-transferase (GST) (Santos et al., 2022; Abd El-Hamid, 2024).

In addition to GSH, the antioxidant activity of S. cerevisiae is largely determined by the content of sulphur-containing amino acids and Maillard reaction products in the yeast (Hassan, 2011). Sulphurcontaining amino acids are involved in the synthesis of intracellular antioxidants such as glutathione and Nacetyl cysteine (Atmaca et al., 2004; Colovic et al., 2018). Maillard reaction products are natural antioxidants that are formed mainly during the fermentation process of S. cerevisiae (Hassan, 2011). S. cerevisiae is one of the sources of MOS extracted from its cell walls. According to Zhao et al. (2022), MOS extracted from S. cerevisiae shows excellent free radical neutralization activity, so that it can be used as a source of natural antioxidants for broiler chickens. It is further explained that the free radical scavenging potential of MOS can be caused by the presence of carboxyl groups in MOS. However, it should be noted that the antioxidant activity of MOS from S. cerevisiae is greatly influenced by the molecular weight, glycosidic bonds, and sulfation level of MOS (Zhao et al., 2022).

The use of S. cerevisiae has been reported to improve antioxidant status in broiler chickens both under commercial conditions and stress conditions (Abo-Sriea et al., 2024; Al-Abdullatif et al., 2024). In general, the use of S. cerevisiae can increase SOD and CAT activity and decrease MDA levels in serum, liver, and other tissues or organs such as intestines (He et al., 2021; Wang et al., 2021). The use of S. cerevisiae has also been reported to reduce MDA levels in breast meat and decrease thiobarbituric acid reactive substances (TBARS) in meat during storage (Mohamed et al., 2022; Sumanu et al., 2023). The use of S. cerevisiae can also decrease the heterophil-to-lymphocyte (H/L) ratio, which indicates a decrease in physiological stress in broiler chickens (Attia et al., 2022). Al-Abdullatif et al., (2024) reported that hydrolyzed S. cerevisiae can maintain the balance of antioxidants and prooxidants in the body of broiler chickens under heat stress conditions so that the nuclear factor kappa-light-chain enhancer of activated B cells (NF-kB) and nuclear factor erythroid 2-related factor 2 (Nrf2) can be balanced.

In line with this study, **Wang** *et al.*, **(2021)** reported that from a molecular perspective, *S. cerevisiae* supplementation resulted in beneficial changes in the relative gene expression of Nrf2-related mRNA. Nrf2 is

a transcription factor that can regulate the expression of antioxidant-related genes such as heme oxygenase-1 (HO-1), GPx, and SOD. The Nrf2 and NF-κB pathways essentially regulate cellular responses to oxidative stress, with Nrf2 protecting against oxidative stress and NF-κB regulating inflammatory responses in the body. The Nrf2 pathway inhibits NF-κB pathway activation by increasing antioxidant defenses and HO-1 expression, which efficiently neutralizes ROS, thereby reducing ROS-mediated NF-κB activation (Yerra et al., **2013).** Recent reports also confirm the ability of S. cerevisiae to affect the hypothalamic-pituitary-adrenal (HPA) axis in broiler chickens under stress conditions (Sumanu et al., 2024). The latter investigators further confirmed that the use of S. cerevisiae can help neutralize ROS, thereby reducing oxidative stress and supporting better thermal regulation. Such a condition may therefore control the HPA axis and corticosterone production during heat stress (Sumanu et al., 2024).

The possibilities of S. cerevisiae being able to improve antioxidant status in broiler chickens have also been confirmed by several researchers. Aluwong et al., (2013) and Abo-Sriea et al., (2024) reported that βglucans and MOS in S. cerevisiae have the ability to scavenge free radicals, whose presence increases due to stress conditions in broiler chickens. In line with this study, Wang et al., (2021) reported that β-glucans and MOS in the cell walls of S. cerevisiae play a significant role in increasing antioxidant capacity in the body of broiler chickens. Furthermore, Aluwong et al., (2013) reported that the sodium selenite fraction of S. cerevisiae also plays a role in increasing antioxidative activity in broiler chickens under stress conditions. In line with the study above, Wang et al., (2021) reported that the cell wall components of S. cerevisiae containing β-glucans and MOS, in addition to increasing the activity of antioxidant-related enzymes, can also reduce ROS production and lipid peroxidation levels, resulting in a decrease in MDA levels in blood and tissues in broiler chickens.

S. cerevisiae has been studied for the modulation of the heat shock protein 70 (HSP70) in broiler chickens (Al-Abdullatif et al., 2024). HSP70 is a vital protein that plays a crucial role in stress response by helping to maintain protein integrity during stress conditions. Environmental factors such as high temperature and other stress factors significantly impact HSP70 expression in broilers. For instance, exposure to high temperatures can increase HSP70 expression, which helps the birds cope with heat stress (Najafi et al., 2015). The work by Al-Abdullatif et al., (2024) showed that supplementing with hydrolyzed yeast S. cerevisiae reduced the duodenal mRNA expression of HSP70. According to this, giving broiler chickens exposed to heat stress a supplement of hydrolyzed yeast

may enhance intestinal redox equilibrium, suggesting the potential benefits of yeast in enhancing stress resilience.

The use of *S. cerevisiae* to increase the antioxidant activity of fermentation products

S. cerevisiae is a type of yeast that is widely applied in the production of fermented products that can be used in the food industry or as animal feed. Typically, fermented products of S. cerevisiae experience increased nutritional value, especially increased amino acid and vitamin B content, and decreased anti-nutritional factors compared to products before fermentation (Prabhu et al., 2016; Qiu et al., 2025). Furthermore, the use of S. cerevisiae as a fermenter has been reported to increase antioxidant activity in fermented products. In this Setivoningrum et al. (2022) reported an increase in antioxidant capacity, total polyphenols, alkaloids, and total flavonoids in garlic fermented with S. cerevisiae. In line with that, Ejuama et al. (2021) also reported an increase in the quantity of phenolic compounds and antioxidant activity with fermentation using S. cerevisiae in roselle calyx aqueous extract. Darwesh et al. (2023) also reported an increase in antioxidant activity, total phenols and flavonoids in cinnamon following the fermentation using S. cerevisiae. In the latter case, the S. cerevisiae fermentation process increased p-hydroxybenzoic acid, gentisic acid, catechin and chlorogenic acid so that it had an impact on increasing antioxidant activity in cinnamon (Darwesh et al., 2023).

Another example was reported by Chandra et al. (2024) who reported an increase in phenols, flavonoids and antioxidant activity in Orthosiphon stamineus Benth leaves after fermentation using S. cerevisiae. Another example was presented by Jelena and Yustiantara (2021), where there was an increase in antioxidant activity in green coffee beans after fermentation using S. cerevisiae. Fermentation using S. cerevisiae was also reported to increase alkaloids, saponins, flavonoids, phenolics, and antioxidant activity of flaxseed extracts (Ghosal et al., 2021). An increase in polyphenols, flavonoids and total antioxidant capacity (TAC) in wheat bran after fermentation using S. cerevisiae was also reported by Prabhu et al. (2016). Moreover, Afriliana et al. (2023) documented that fermentation using S. cerevisiae was able to enhance antioxidant activity, phenolic compounds, flavonoids, and tannins of Robusta coffee beans.

The literature shows that most yeast have the ability to produce antioxidants (Sugiharto, 2019). Liang et al. (2023) reported that glycosidic bonds in phenolic compounds are hydrolysed during fermentation, resulting in the formation of additional

phenolic metabolites such as gallic acid, protocatechuic acid, and quercetin, thereby increasing antioxidant activity in the fermented material. During the fermentation process, yeast is able to produce enzymes that can break down glycosidic bonds in several phenol hydroxyl groups bound to complex compounds and this degradation in turn increases the amount of free phenol (Ejuama et al., 2021). According to Setiyoningrum et al., (2022), increasing the flavonoid content in fermented products using S. cerevisiae is also possible because there is an increase in acid during fermentation which frees the bound flavonoid component. A more recent study by Anggraeni et al., (2025) further showed that fermentation using S. cerevisiae can increase the selenium content in infertile egg powder, thereby increasing antioxidant activity in the fermentation product. Moreover, Wang et al., (2025) reported that S. cerevisiae fermentation was capable of reducing the molecular weight and modifying the structure of plant polysaccharides and thereby augmenting antioxidant protective action of plant polysaccharides. Indeed, the latter feature is beneficial in protecting cells from oxidative damage induced by free radicals (Wang et al., 2025).

Several *in vivo* studies have evaluated the *S*. cerevisiae fermentation products to improve the antioxidant status of broiler chickens. Sobotik et al. (2022) and Heinsohn et al. (2024) reported a decrease in plasma corticosterone and H/L ratio in broiler chickens given S. cerevisiae-fermentation products under heat stress conditions. It was most likely that S. cerevisiae fermentation products are able to modulate the gut microbiome of broiler chickens. Considering that gut microbiota has a critical role in regulating broiler thermogenesis through the gut-brain axis, the ability of yeast-fermentation products to modulate gut therefore microbiome may be beneficial thermoregulation in broiler chickens (Heinsohn et al., 2024).

Another study by **Aristides** *et al.* (2018) demonstrated the efficacy of *S. cerevisiae* fermentation products in improving the meat quality of broiler chickens. In such cases, flavonoids and vitamin E present in the *S. cerevisiae*-fermentation products are responsible for reducing lipid oxidation in broiler chicken meats and therefore increasing the shelf life of meats (**Aristides** *et al.*, 2018).

Moreover, the use of *S. cerevisiae*-derived yeast fermentate was able to improve the antioxidant capacity in broiler chickens exposed to acute and chronic stress during the rearing period in the study of **Nelson** *et al.* **(2020)**. They confirmed that the use of *S. cerevisiae*-derived yeast fermentate reduced melanocortin-2-receptor (MC2R: the adrenocorticotropic hormone

receptor on adrenocortical cells, serving a critical role in regulating cortisol production in the adrenal glands) gene expression. The reduced MC2R gene expression may consequently reduce corticosterone production and alleviate stress conditions in broiler chickens (Nelson *et al.*, 2020).

Application of *S. cerevisiae* and its derivative products as antioxidant agents in broiler production

In commercial broiler production, S. cerevisiae and its derivatives have gained significant attention for their potential as alternative antioxidant agents. The utilization of these yeast products is primarily driven by the pressing need to find sustainable alternatives to synthetic antioxidants, which have been traditionally used to maintain the antioxidative status of broiler chickens but now pose risks of synthetic antioxidant residues in broiler meat products (Bilal et al., 2021). Antioxidants are crucial in poultry production as they help mitigate oxidative stress, a significant challenge in enhancing growth performance and health in broilers. In addition to improving gut health and immune competency, yeast products and their derivatives can increase broiler antioxidant capacity, making them a possible substitute for synthetic antioxidants and antibiotics for broiler production (Obianwuna et al., 2024).

A number of studies have been carried out to evaluate the efficacy of S. cerevisiae and its derivatives as natural antioxidants for broiler chickens (**Table 1**). In addition to the improved antioxidant status and physiological response to stressors, using S. cerevisiae and its derivatives as antioxidant agents enhances broiler chicken production performance and health. Under commercial conditions, **Abo-Sriea** et al. (2024) used NutriFix® (consisting of S. cerevisiae, MOS and β -glucan) as a feed additive for broilers. They found that such dietary treatment improved growth performance, feed conversion ratio, intestinal health, immune responses and antioxidant capacity of broiler chickens.

Al-Abdullatif et al. (2024) also revealed that hydrolyzed S. cerevisiae supplementation improved duodenal redox homeostasis and the immune system, and hence improved growth performance and feed efficiency, as well as decreased the mortality rate of broiler chickens during heat stress. During stress conditions, dietary administration of S. cerevisiae also improved the meat quality of broiler chickens, as indicated by the reduced MDA and TBARS contents in meat (He et al., 2022; Mohamed et al., 2022). The latter condition may therefore increase the shelf life of broiler meats during storage (Aristides et al., 2018; Mohamed et al., 2022).

Potential Application of Saccharomyces cerevisiae as

Table 1: Application of *S. cerevisiae* and its derivative products as an antioxidant agent in broiler production.

S. cerevisiae products	Levels in feed/water	Conditions during rearing	Effects on broiler chickens	References
S. cerevisiae- fermented infertile egg flour	0.7% feed	Challenged with high-stocking density	Decreased serum SOD level with no effect on serum MDA level	(Anggraeni <i>et al.</i> , 2025)
Activated <i>S.</i> cerevisiae (1 10 ¹⁰ cells/g)	0.75 g/kg feed	Infected with Clostridium perfringens	Increased TAC and reduced MDA levels in serum	(Al-Baadani et al., 2025)
Inactivated <i>S.</i> cerevisiae (1 10 ¹⁰ cells/g)	0.75 g/kg feed	Infected with C. perfringens	Increased TAC and reduced MDA levels in serum	(Al-Baadani et al., 2025)
NutriFix® (S. cerevisiae, MOS and β-glucan)	250 g/ton of feed	Reared under commercial conditions	Increased serum TAC	(Abo-Sriea <i>et al.</i> , 2024)
Hydrolysed yeast derived from <i>S. cerevisiae</i>	400, 800, or 1,200 mg/kg feed	Exposed to heat stress	Decreased duodenal mRNA expression of heat shock protein 70 (HSP70)	(Al-Abdullatif et al., 2024)
Concentrated S. cerevisiae fermentation product	0.625 kg/metric ton feed	Challenged with overcrowding condition and heat stress for 12 hours	Reduced corticosterone levels and H/L ratio	(Heinsohn <i>et al.</i> , 2024)
Enzymatically treated yeast (<i>S. cerevisiae</i>)	1 and 2 g/kg feed	Coccidia challenge condition	Increased serum CAT activities	(Alagbe <i>et al.</i> , 2023)
Commercial S. cerevisiae Commercial S. cerevisiae (12×10 ⁹ active yeast per gram)	1 g/kg feed 0.02% or 0.04% of feed	Exposed to heat stress Reared under commercial conditions	Reduced MDA level in breast meat Decreased H/L ratio, serum MDA level, and TAC	(Sumanu et al., 2023) (Attia et al., 2022)
Live yeast (S. cerevisiae; 1×10 ¹⁰ cfu/g)	500 and 1000 mg/kg feed	Transport stress condition	At 1000 mg/kg feed enhanced TAC and reduced MDA in serum and muscles	(He <i>et al.</i> , 2022)
Commercial S. cerevisiae	5% of feed	Exposed to heat stress	Increased CAT, GPx and SOD activities, and decreased MDA level in serum and TBARS in meat at days 1, 7 and 30 of storage	(Mohamed <i>et al.</i> , 2022)
S. cerevisiae and phytase cofermented wheat bran	5 and 10%	Reared under commercial conditions	Increased mRNA expression of Nrf2 on chicken peripheral blood mononuclear cells	(Chuang <i>et al.</i> , 2021)

Live yeast (<i>S. cerevisiae</i> ; 47,1 × 10 ¹⁰ cfu/g)	500 and 1000 mg/kg feed	Reared under commercial conditions	Increased SOD and CAT activities and decreased MDA level in serum	(He <i>et al.</i> , 2021)
Yeast (S. cerevisiae)-derived product	50, 100 and 150 mg/kg	Reared under commercial conditions	At 50 to 100 mg/kg improved jejunal antioxidant capacity (increased GSH, SOD, GPx activities and decreased MDA level)	(Wang <i>et al.</i> , 2021)
Selenium- enriched <i>S.</i> cerevisiae (10 ⁸ cfu/g)	1 g/kg feed	Reared under commercial conditions	Increased GPx and thioredoxin reductase activities and TAC, and decreased MDA level in meats	(Hou <i>et al.</i> , 2020)
Se-enriched <i>S. cerevisiae</i>	0.15, 0.5 and 1.5 mg per kg of diet	Reared under commercial conditions	Improved antioxidant status (increased GPx, SOD and TAC and lowered MDA in plasma and liver)	(Chen <i>et al.</i> , 2017)
S. cerevisiae culture	0.5 mL/L of drinking water daily	Reared under commercial conditions	Enhanced serum antioxidant enzyme (GPx) activities	(Aluwong <i>et al.</i> , 2013)

CAT= catalase, cfu= colony forming units, GPx= glutathione peroxidase, GSH= glutathione, H/L ratio= heterophils to lymphocytes ratio, MDA= malondialdehyde, MOS= mannan oligosaccharides, Nrf2= nuclear factor erythroid 2-related factor 2, SOD= superoxide dismutase, TAC= total antioxidant capacity, TBARS= thiobarbituric acid reactive substances.

Potential application of *S. cerevisiae* as an antiinflammatory agent for broiler chickens

It is generally known that stress not only causes oxidative stress but also causes inflammation in broiler chickens. In this case, stress may induce the secretion of hormones such as corticosterone, which can cause oxidative stress and can also initiate the inflammatory pathways. Indeed, oxidative stress and inflammation are interrelated, as each can exacerbate the other (Ramos-González et al., 2024). In addition to being used as antistress agents, antioxidants are also often used by farmers to alleviate inflammation in broiler chickens under stress conditions. With regard particularly to S. cerevisiae, Wang et al., (2016) reported that the use of live S. cerevisiae alleviated intestinal inflammation in broiler chickens infected with Escherichia coli by reducing NF-κB and interleukin (IL)-1β expressions in the ileum. Furthermore, Lin et al., (2023) reported that the use of S. cerevisiae hydrolysate improves antioxidant status, reduces the expression levels of intestinal inflammatory factors, and alleviates the intestinal inflammatory response of broilers. In the above case, the S. cerevisiae hydrolysate may exert an anti-inflammatory effect by inhibiting the excessive expressions of IL-1β, IL-6, and tumor necrosis factor (TNF)-α. In addition, S. cerevisiae hydrolysate has the potential to improve the microbial population in the

intestine, thereby positively influencing the inflammatory response in the intestine of broiler chickens (Lin et al., 2023). In line with the above study, Yang et al. (2019) also reported that the use of S. cerevisiae-fermented product reduced inflammation-related mRNA as indicated by the reduced IL-1ß, inducible nitric oxide synthase (iNOS), interferon (IFN)-γ, and NFκB mRNA expressions in the peripheral blood mononuclear cells of broiler chickens. In agreement with Lin et al. (2023), Yang et al. (2019) also confirmed that the decrease in the population of pathogenic bacteria in the intestine (e.g., Clostridium perfringens) due to treatment using S. cerevisiaefermented product had an impact on the decrease in the amount of pro-inflammatory cytokines in the intestine of broiler chickens.

It is further explained that β -glucans contained in *S. cerevisiae* play a significant role in inhibiting IL-1 β production and thereby reducing NF- κ B-mediated inflammatory responses (Yang *et al.*, 2019). Another study by Chuang *et al.*, (2021) also demonstrated the anti-inflammatory effect of *S. cerevisiae* in broiler chickens. They reported that the use of *S. cerevisiae* and phytase co-fermented wheat bran in feed could inhibit the expression of IL-1 β , which is one of the main regulators of inflammation in broiler chickens. Again, components of the *S. cerevisiae* cell wall, especially β -

glucans and MOS, have a very critical role in reducing inflammatory-related cytokine release so that the negative effect of stress on inflammation can be reduced (Chuang et al., 2021). Another study further reported that S. cerevisiae hydrolysate showed anti-inflammatory effects in broiler chickens (Wang et al., 2022). The latter authors revealed that S. cerevisiae hydrolysate reduced TNF- α concentration, IL-6, IL-1 β , TNF- α and NF κ B, while increasing IL-10 expression levels in the jejunum of broiler chickens. In line with other studies, the anti-inflammatory effect of S. cerevisiae hydrolysate is closely related to β -glucans and MOS in S. cerevisiae, which can serve as anti-inflammatory agents.

CONCLUSIONS

The potential of S. cerevisiae as a source of natural antioxidants for broiler chickens is largely related to the content of several active ingredients in the yeast, such as tryptophol, GSH, sulfur-containing amino acids and Maillard reaction products. The antioxidant potential of S. cerevisiae is also closely related to βglucans and MOS, which are the main components of the cell wall of the yeast. Dietary supplementation with S. cerevisiae has been shown to enhance antioxidant enzyme activity and reduce oxidative markers in broilers. In such cases, the active components in yeast, as mentioned above, play a major role in neutralizing ROS and preventing lipid peroxidation, especially under stress conditions. S. cerevisiae also plays a very important role in improving the response of broiler chickens to stress so that the negative consequences of stress can be alleviated. The sustainability of broiler production will greatly benefit from a field commercial study investigating the efficacy of S. cerevisiae and its derivatives as antioxidant agents and as substitute antibiotic growth promoters. Further research on the dose-dependent effects of S. cerevisiae and its derivatives as antioxidants and substitute antibiotics is also crucial.

Conflict of interests

We declare having no conflict of interest

Acknowledgement

We thank to Universitas Diponegoro for supporting this review article through "Riset Artikel Review (RAR)" scheme with contract nr. 222-038/UN7.D2/PP/IV/2025.

REFERENCES

- **ABD El-HAMID, I.S., 2024.** Influence of Spirulina platensis supplementation alone or mixed with live yeast on blood constituents and oxidative status of Damascus goats and their new born. Journal of Applied Veterinary. Sciences, 9 (1), 1-12. https://doi.org/10.21608/javs.2023.231998.1266
- ABO-SRIEA, T. M., ISMAEL, E., SOBHI, B. M., HASSAN, N. H., ELLEITHY, E. M., OMAR, S. A.,

- and RAMADAN, A., 2024. Impact of dietary-nucleotides and Saccharomyces cerevisiae-derivatives on growth-performance, antioxidant-capacity, immuneresponse, small-intestine histomorphometry, caecal-Clostridia, and litter-hygiene of broiler-chickens treated with florfenicol. International Journal of Veterinary Science and Medicine, 12(1), 11-24. https://doi.org/10.1080/23144599.2024.2324411
- AFRILIANA, A., PURNOMO, B. L., and WITONO, Y., 2023. Antioxidant activity of fermented inferior Jember Robusta coffee beans using *Saccharomyces cerevisiae* as a starter in the semi-carbonic maceration technique. Asian Journal of Food Research and Nutrition, 2(4), 451-461.

https://journalajfrn.com/index.php/AJFRN/article/view/69

- AHIWE, E. U., DOS SANTOS, T. T., GRAHAM, H., and IJI, P. A., 2021. Can probiotic or prebiotic yeast (*Saccharomyces cerevisiae*) serve as alternatives to infeed antibiotics for healthy or disease-challenged broiler chickens?. a review. Journal of Applied Poultry Research, 30(3), 100164. https://doi.org/10.1016/j.japr.2021.100164
- AL-ABDULLATIF, A. A., ALHOTAN, R. A., ALBADWI, M. A., DONG, X., KETTUNEN, H., VUORENMAA, J., and AZZAM, M. M., 2024. Effects of hydrolyzed yeast on growth performance, intestinal redox homeostasis, and woody breast myopathy in heat-stressed broilers. Frontiers in Veterinary Science, 11, 1484150. https://doi.org/10.3389/fvets.2024.1484150
- ALAGBE, E. O., SCHULZE, H., and ADEOLA, O., 2023. Growth performance, nutrient digestibility, intestinal morphology, cecal mucosal cytokines and serum antioxidant responses of broiler chickens to dietary enzymatically treated yeast and coccidia challenge. Journal of Animal Science and Biotechnology, 14(1), 57. https://doi.org/10.1186/s40104-023-00846-z
- AL-BAADANI, H. H., ALHARTHI, A. S., ABBAS, N. I., QASEM, A. A., SALEH, A., and IBRAHEEM, M. A., 2025. Effect of activated and inactivated Saccharomyces cerevisiae as alternative to antibiotic growth promoter on the performance and health of broilers infected with *Clostridium perfringens*. Italian Journal of Animal Science, 24(1), 43-52. https://doi.org/10.1080/1828051X.2024.2441349
- ALUWONG, T., KAWU, M., RAJI, M., DZENDA, T., GOVWANG, F., SINKALU, V., and AYO, J., 2013. Effect of yeast probiotic on growth, antioxidant enzyme activities and malondialdehyde concentration of broiler chickens. Antioxidants, 2(4), 326-339. https://doi.org/10.3390/antiox2040326
- ANGGRAENI, D., SUNARTI, D., and SUGIHARTO, S., 2025. Effects of *Saccharomyces cerevisiae*-fermented infertile egg flour on growth performance, antioxidant status and carcass of broilers raised in high density pens. Pakistan Journal of Agricultural Science, 62(1), 131-139. https://doi.org/10.21162/PAKJAS/25.375
- APALOWO, O. O., EKUNSEITAN, D. A., and FASINA, Y. O., 2024. Impact of heat stress on broiler chicken production. Poultry, 3(2), 107-128. https://doi.org/10.3390/poultry3020010
- ARISTIDES, L. G. A., VENANCIO, E. J., ALFIERI, A. A., OTONEL, R. A. A., FRANK, W. J., and OBA, A.

- **2018.** Carcass characteristics and meat quality of broilers fed with different levels of *Saccharomyces cerevisiae* fermentation product. Poultry science, 97(9), 3337-3342. https://doi.org/10.3382/ps/pey174
- ATMACA, G., 2004. Antioxidant effects of sulfur-containing amino acids. Yonsei Medical Journal, 45(5), 776-788. https://doi.org/10.3349/ymj.2004.45.5.776
- ATTIA, Y. A., AL-KHALAIFAH, H., EL-HAMID, A., HATEM, S., AL-HARTHI, M. A., ALYILEILI, S. R., and EL-SHAFEY, A. A., 2022. Antioxidant status, blood constituents and immune response of broiler chickens fed two types of diets with or without different concentrations of active yeast. Animals, 12(4), 453. https://doi.org/10.3390/ani12040453
- AYLWARD, B. A., JOHNSON, C. N., PERRY, F., WHELAN, R., and ARSENAULT, R. J., 2024. Modern broiler chickens exhibit a differential gastrointestinal immune and metabolic response to repeated CpG injection relative to a 1950s heritage broiler breed. Frontiers in Physiology, 15, 1473202. https://doi.org/10.3389/fphys.2024.1473202
- BAI, K., HUANG, Q., ZHANG, J., HE, J., ZHANG, L., and WANG, T., 2017. Supplemental effects of probiotic *Bacillus subtilis* fmbJ on growth performance, antioxidant capacity, and meat quality of broiler chickens. Poultry Science, 96(1), 74-82. https://doi.org/10.3382/ps/pew246
- BILAL, R. M., HASSAN, F. U., SAEED, M., RAFEEQ, M., ZAHRA, N., FRAZ, A., SAEED, S., KHAN, M. A., MAHGOUB, H. A. M., FARAG, M. R., and ALAGAWANY, M., 2021. Role of yeast and yeast-derived products as feed additives in broiler nutrition. Animal Biotechnology, 34(2), 392-401. https://doi.org/10.1080/10495398.2021.1942028
- CHANDRA, M. A., SYAMSU, K., AMBARSARI, L., FATIMAH, N., and NURCHOLIS, W., 2024. Increasing polyphenol antioxidant in *Orthosiphon stamineus* Benth leaves with fermentation extraction by *Saccharomyces cerevisiae* ATCC-9763. Journal of Applied Biology and Biotechnology, 12(1), 111-116. https://doi.org/10.7324/JABB.2024.149026
- CHEN, F., ZHU, L., QIU, H., and QIN, S., 2017. Selenium-enriched Saccharomyces cerevisiae improves growth, antioxidant status and selenoprotein gene expression in Arbor Acres broilers. Journal of Animal Physiology and Animal Nutrition, 101(2), 259-266. https://doi.org/10.1111/jpn.12571
- CHUANG, W. Y., LIN, L. J., HSIEH, Y. C., CHANG, S. C., and LEE, T. T., 2021. Effects of *Saccharomyces cerevisiae* and phytase co-fermentation of wheat bran on growth, antioxidation, immunity and intestinal morphology in broilers. Animal Bioscience, 34(7), 1157. https://doi.org/10.5713/ajas.20.0399
- CHUANG, W. Y., LIN, W. C., HSIEH, Y. C., HUANG, C. M., CHANG, S. C., and LEE, T. T., 2019. Evaluation of the combined use of *Saccharomyces cerevisiae* and *Aspergillus oryzae* with phytase fermentation products on growth, inflammatory, and intestinal morphology in broilers. Animals, 9(12), 1051. https://doi.org/10.3390/ani9121051
- COLOVIC, M. B., VASIC, V. M., DJURIC, D. M., and KRSTIC, D. Z., 2018. Sulphur-containing amino acids: protective role against free radicals and heavy

- metals. Current Medicinal Chemistry, 25(3), 324-335. https://doi.org/10.2174/0929867324666170609075434
- DARWESH, O. M., EWEYS, A. S., ZHAO, Y. S., and MATTER, I. A., 2023. Application of environmental-safe fermentation with *Saccharomyces cerevisiae* for increasing the cinnamon biological activities. Bioresources and Bioprocessing, 10(1), 12. https://doi.org/10.1186/s40643-023-00632-9
- EJUAMA, C. K., ONUSIRIUKA, B. C., BAKARE, V., NDIBE, T. O., YAKUBU, M., and ADEMU, E. G., 2021. Effect of *Saccharomyces cerevisiae*-induced fermentation on the antioxidant property of *Roselle Calyx* aqueous extract. European Journal of Biology and Biotechnology, 2(3), 33-38. https://doi.org/10.24018/ejbio.2021.2.3.201
- FAKRUDDIN, M. D., HOSSAIN, M. N., AND AHMED, M. M., 2017. Antimicrobial and antioxidant activities of *Saccharomyces cerevisiae* IFST062013, a potential probiotic. BMC Complementary and Alternative Medicine, 17, 1-11. https://doi.org/10.1186/s12906-017-1591-9
- GHOSAL, S., BHATTACHARYYA, D. K., and BHOWAL, J., 2021. Effect of Saccharomyces cerevisiae fermentation process on the phenolic content, flavonoid content and antioxidant properties of flaxseeds. In Advances in Bioprocess Engineering and Technology: Select Proceedings ICABET 2020 (pp. 119-131). Springer Singapore. https://doi.org/10.1007/978-981-15-7409-2 12
- GRŽINIĆ, G., PIOTROWICZ-CIEŚLAK, A., KLIMKOWICZ-PAWLAS, A., GÓRNY, R. L., ŁAWNICZEK-WAŁCZYK, A., PIECHOWICZ, L., and WOLSKA, L., 2023. Intensive poultry farming: A review of the impact on the environment and human health. Science of the Total Environment, 858, 160014. https://doi.org/10.1016/j.scitotenv.2022.160014
- GUO, Y., ZHANG, J., LI, X., WU, J., HAN, J., YANG, G., and ZHANG, L., 2023. Oxidative stress mediated immunosuppression caused by ammonia gas via antioxidant/oxidant imbalance in broilers. British Poultry Science, 64(1), 36-46. https://doi.org/10.1080/00071668.2022.2122025
- HASHEMITABAR, S. H., and HOSSEINIAN, S. A., 2024. The comparative effects of probiotics on growth, antioxidant indices and intestinal histomorphology of broilers under heat stress condition. Scientific Reports, 14(1), 23471. https://doi.org/10.1038/s41598-024-66301-9
- HASSAN, H. M., 2011. Antioxidant and immunostimulating activities of yeast (*Saccharomyces cerevisiae*) autolysates. World Applied Science Journal, 15(8), 1110-9. https://www.idosi.org/wasj/wasj15(8)11/11.pdf
- HE, T., MA, J., MAHFUZ, S., ZHENG, Y., LONG, S., WANG, J., and PIAO, X., 2022. Dietary live yeast supplementation alleviates transport-stress-impaired meat quality of broilers through maintaining muscle energy metabolism and antioxidant status. Journal of the Science of Food and Agriculture, 102(10), 4086-4096. https://doi.org/10.1002/jsfa.11758
- HE, T., MAHFUZ, S., PIAO, X., WU, D., WANG, W., YAN, H., and LIU, Y., 2021. Effects of live yeast (Saccharomyces cerevisiae) as a substitute to antibiotic on growth performance, immune function, serum

- biochemical parameters and intestinal morphology of broilers. Journal of Applied Animal Research, 49(1), 15-22. https://doi.org/10.1080/09712119.2021.1876705
- HEINSOHN, Z., BROWN, A., SOBOTIK, E., HOUSE, G., STIEWERT, A., CHANEY, W. E., and ARCHER, G. S., 2024. Evaluating the effects of feeding a concentrated *Saccharomyces cerevisiae* fermentation product on the performance and stress susceptibility of broiler chickens. Poultry, 3(1), 57-65. https://doi.org/10.3390/poultry3010006
- HOU, L., QIU, H., SUN, P., ZHU, L., CHEN, F., and QIN, S., 2020. Selenium-enriched *Saccharomyces cerevisiae* improves the meat quality of broiler chickens via activation of the glutathione and thioredoxin systems. Poultry Science, 99(11), 6045-6054. https://doi.org/10.1016/j.psj.2020.07.043
- JELENA, J., and YUSTIANTARA, P. S., 2021.

 Antioxidant activity of fermented coffee beans. Pharmacy Reports, 1(2), 25-25.

 https://doi.org/10.51511/pr.25
- LIANG, Z., HUANG, Y., ZHANG, P., and FANG, Z., 2023. Impact of fermentation on the structure and antioxidant activity of selective phenolic compounds. Food Bioscience, 56, 103147. https://doi.org/10.1016/j.fbio.2023.103147
- LIN, J., COMI, M., VERA, P., ALESSANDRO, A., QIU, K., WANG, J., and ZHANG, H. J., 2023. Effects of *Saccharomyces cerevisiae* hydrolysate on growth performance, immunity function, and intestinal health in broilers. Poultry Science, 102(1), 102237. https://doi.org/10.1016/j.psj.2022.102237
- LUO, G. M., REN, X. J., LIU, J. Q., MU, Y., and SHEN, J. C., 2003. Towards more efficient glutathione peroxidase mimics: Substrate recognition and catalytic group assembly. Current Medicinal Chemistry, 10(13), 1151-83. https://doi.org/10.2174/0929867033457502
- MAKKY, E. A., ALMATAR, M., MAHMOOD, M. H., TING, O. W., and QI, W. Z., 2021. Evaluation of the antioxidant and antimicrobial activities of ethyl acetate extract of *Saccharomyces cerevisiae*. Food technology and biotechnology, 59(2), 127-136. https://doi.org/10.17113/ftb.59.02.21.6658
- MOHAMED, H. E., IBRAHIM, W. A., and GAAFAR, R. E., 2022. Impact of *Moringa olifera* leaves or *Saccharomyces* supplementation on carcass quality, mRNA of heat shock proteins and antioxidants in broilers exposed to heat stress. SVU-International Journal of Veterinary Sciences, 5(4), 193-227. https://doi.org/10.21608/svu.2022.167661.1232
- NAJAFI, P., RAMIAH, S. K., AMAT JAJULI, N., FARJAM, A. S., ZULKIFLI, I., O'REILY, E., ECKERSALL, D., and AMIR, A. A., 2015. Environmental temperature and stocking density effects on acute phase proteins, heat shock protein 70, circulating corticosterone and performance in broiler chickens. International Journal of Biometeorology, 59(11), 1577-1583. https://doi.org/10.1007/s00484-015-0964-3
- NELSON, J. R., SOBOTIK, E. B., ATHREY, G., and ARCHER, G. S., 2020. Effects of supplementing yeast fermentate in the feed or drinking water on stress susceptibility, plasma chemistry, cytokine levels, antioxidant status, and stress- and immune-related gene

- expression of broiler chickens. Poultry Science, 99, 3312-3318. https://doi.org/10.1016/j.psj.2020.03.037
- OBIANWUNA, U. E., CHANG, X., OLEFORUH-OKOLEH, V. U., ONU, P. N., ZHANG, H., QIU, K., and WU, S., 2024. Phytobiotics in poultry: revolutionizing broiler chicken nutrition with plant-derived gut health enhancers. Journal of Animal Science and Biotechnology, 15(1), 169. https://doi.org/10.1186/s40104-024-01101-9
- PRABHU, A., GARG, Y., CHITYALA, S., and VENKATA, D., V., 2016. Improvement of phytonutrients and antioxidant properties of wheat bran by yeast fermentation. Current Nutrition and Food Science, 12(4), 249-255. https://doi.org/10.2174/1573401312666160830155812
- QIU, S., YAO, K., SUN, J., LIU, S., and SONG, X., 2025. Impact of fermentation by *Saccharomyces cerevisiae* on the macronutrient and in vitro digestion characteristics of Chinese noodles. Food Chemistry, 462, 140967. https://doi.org/10.1016/j.foodchem.2024.140967
- RAKNGAM, S., ZHU, Y., OKRATHOK, S., PUKKUNG, C., and KHEMPAKA, S., 2024. Enhancing heat stress resilience in broiler chickens through the use of probiotics and postbiotics: a review. Tropical Animal Science Journal, 47(4), 538-548. https://doi.org/10.5398/tasj.2024.47.4.538
- RAMOS-GONZÁLEZ, E. J., BITZER-QUINTERO, O. K., ORTIZ, G., HERNÁNDEZ-CRUZ, J. J., and RAMÍREZ-JIRANO, L. J., 2024. Relationship between inflammation and oxidative stress and its effect on multiple sclerosis. Neurologia, 39(3), 292-301. https://doi.org/10.1016/j.nrleng.2021.10.010
- RIBER, A. B., and WURTZ, K. E., 2024. Impact of growth rate on the welfare of broilers. Animals, 14(22), 3330. https://doi.org/10.3390/ani14223330
- RYBARCZYK, A., HARAF, G., TOKARCZYK, G., TELESZKO, M., TOBOLSKA, I., and BIENKIEWICZ, G., 2024. Effect of distiller's yeast in feed on texture, fatty acid profile and antioxidant properties of breast muscle of broiler chickens. Agricultural and Food Science, 33, 2. https://doi.org/10.23986/afsci.145308
- SANTOS, L. O., SILVA, P. G. P., LEMOS JUNIOR, W. J. F., DE OLIVEIRA, V. S., and ANSCHAU, A., 2022. Glutathione production by *Saccharomyces cerevisiae*: current state and perspectives. Applied Microbiology and Biotechnology, 106(5), 1879-1894. https://doi.org/10.1007/s00253-022-11826-0
- SETIYONINGRUM, F., PRIADI, G., and AFIATI, F., 2022. Chemical properties of solo black garlic fermented by *Saccharomyces cerevisiae*. IOP Conference Series: Earth and Environmental Science, 976(1): 012044. https://doi.org/10.1088/1755-1315/976/1/012044
- SITHOLE, A. N., HLATINI, V. A., and CHIMONYO, M., 2022. Potential of combining natural-derived antioxidants for improving broiler meat shelf-life a review. Animal Bioscience, 36(9), 1305. https://doi.org/10.5713/ab.22.0188
- SHAKERI, M., LE, H., SHAKERI, M., and OSKOUEIAN, E. 2020. Strategies to combat heat stress in broiler chickens: Unveiling the roles of

- selenium, vitamin E and vitamin C. Veterinary Sciences, 7(2), 71. https://doi.org/10.3390/vetsci7020071
- SOBOTIK, E. B., NELSON, J. R., PAVLIDIS, H. O., and ARCHER, G. S., 2022. Evaluating the effects of supplementing *Saccharomyces cerevisiae* in the feed or drinking water on stress susceptibility of broilers. Journal of Applied Poultry Research, 31(1), 100220. https://doi.org/10.1016/j.japr.2021.100220
- SUGIHARTO, S. 2016. Role of nutraceuticals in gut health and growth performance of poultry. Journal of the Saudi Society of Agricultural Sciences, 15, 99-111. http://dx.doi.org/10.1016/j.jssas.2014.06.001
- SUGIHARTO, S. 2019. A review of filamentous fungi in broiler production. Annals of Agricultural Sciences, 64(1), 1-8. https://doi.org/10.1016/j.aoas.2019.05.005
- SUGIHARTO, S. 2022. Dietary strategies to alleviate highstocking-density-induced stress in broiler chickens—a comprehensive review. Archives Animal Breeding, 65(1), 21-36. https://doi.org/10.5194/aab-65-21-2022
- SUGIHARTO, S., AGUSETYANINGSIH, I., WIDIASTUTI, E., WAHYUNI, H. I., YUDIARTI, T., and SARTONO, T. A., 2023. Dietary supplementation of enzymes: An approach to mitigate ammonia emission during broiler production. Iranian Journal of Applied Animal Science, 13(4), 615-625. https://journals.iau.ir/article 708962.html
- SUGIHARTO, S., YUDIARTI, T., ISROLI, I., WIDIASTUTI, E., WAHYUNI, H. I., and SARTONO, T. A., 2019. Effect of formic acid, Saccharomyces cerevisiae or their combination on the growth performance and serum indices of the Indonesian indigenous crossbred chickens. Annals of Agricultural Sciences, 64(2), 206-210. https://doi.org/10.1016/j.aoas.2019.12.004
- SUMANU, V. O., CHAMUNORWA, J. P., OOSTHUIZEN, M. C., and NAIDOO, V. 2022. Adverse effects of heat stress during summer on broiler chickens production and antioxidant mitigating effects. International Journal of Biometeorology, 66(12), 2379–2393. https://doi.org/10.1007/s00484-022-02372-5
- SUMANU, V. O., BYARUHANGA, C., BOSMAN, A. M., OCHAI, S. O., NAIDOO, V., OOSTHUIZEN, M. C., and CHAMUNORWA, J. P., 2023. Effects of probiotic (*Saccharomyces cerevisiae*) and ascorbic acid on oxidative gene damage biomarker, heat shock protein 70 and interleukin 10 in broiler chickens exposed to heat stress. Animal Gene, 28, 200150. https://doi.org/10.1016/j.angen.2023.200150
- SUMANU, V. O., NAIDOO, V., OOSTHUIZEN, M. C., and CHAMUNORWA, J. P., 2024. Evaluating the efficacy of probiotics and ascorbic acid as anti-stress agents against heat stress in broiler chickens. Frontiers in Veterinary Science, 11, 1482134. https://doi.org/10.3389/fvets.2024.1482134
- WANG, T., CHENG, K., LI, Q., and WANG, T., 2022. Effects of yeast hydrolysate supplementation on intestinal morphology, barrier, and anti-inflammatory

- functions of broilers. Animal Bioscience, 35(6), 858. https://doi.org/10.5713/ab.21.0374
- Wang, T., Cheng, K., Yu, C. Y., Li, Q. M., Tong, Y. C., Wang, C., and Wang, T., 2021. Effects of a yeast-derived product on growth performance, antioxidant capacity, and immune function of broilers. Poultry Science, 100(9), 101343. https://doi.org/10.1016/j.psj.2021.101343
- WANG, W., LI, Z., HAN, Q., GUO, Y., ZHANG, B., and D'INCA, R., 2016. Dietary live yeast and mannanoligosaccharide supplementation attenuate intestinal inflammation and barrier dysfunction induced by *Escherichia coli* in broilers. British Journal of Nutrition, 116(11), 1878-1888. https://doi.org/10.1017/S0007114516004116
- WANG, X., LIU, X., LIU, S., QU, J., YE, M., WANG, J., and LI, R., 2023. Effects of anti-stress agents on the growth performance and immune function in broiler chickens with vaccination-induced stress. Avian Pathology, 52(1), 12-24. https://doi.org/10.1080/03079457.2022.2114874
- WANG, Z., ZHENG, Y., LAI, Z., KONG, Z., HU, X., ZHANG, P., YANG, Y., and LI, N. 2025. Effect of Saccharomyces cerevisiae CICC 32883 fermentation on the structural features and antioxidant protection effect of Chinese yam polysaccharide. Foods, 14(4), 564. https://doi.org/10.3390/foods14040564
- XU, X., LIU, A., HU, S., ARES, I., MARTÍNEZ-LARRAÑAGA, M. R., WANG, X., and MARTÍNEZ, M. A., 2021. Synthetic phenolic antioxidants: Metabolism, hazards and mechanism of action. Food Chemistry, 353, 129488. https://doi.org/10.1016/j.foodchem.2021.129488
- YERRA, V. G., NEGI, G., SHARMA, S. S., and KUMAR, A., 2013. Potential therapeutic effects of the simultaneous targeting of the Nrf2 and NF-κB pathways in diabetic neuropathy. Redox Biology, 1(1), 394-397. https://doi.org/10.1016/j.redox.2013.07.005
- ZDANOWSKA-SĄSIADEK, Z., LIPIŃSKA-PALKA, P., DAMAZIAK, K., MICHALCZUK, M., GRZYBEK, W., KRUZIŃSKA, B., and MARCHEWKA, J., 2019. Antioxidant effects of phytogenic herbal-vegetable mixtures additives used in chicken feed on breast meat quality. Animal Science Papers and Report, 36(4), 393-408.
- ZHAO, Y., WANG, J., FU, Q., ZHANG, H., LIANG, J., XUE, W., and ODA, H., 2022. Characterization and antioxidant activity of mannans from *Saccharomyces cerevisiae* with different molecular weight. Molecules, 27(14), 4439. https://doi.org/10.3390/molecules27144439

How to cite this article:

Sugiharto Sugiharto, Ikania Agusetyaningsih, Endang Widiastuti and Turini Yudiarti, 2025. Potential Application of Saccharomyces cerevisiae as an Antioxidant Agent for Broiler Chickens – A Brief Overview. Journal of Applied Veterinary Sciences, 10 (3): 118-128.

DOI:https://dx.doi.org/10.21608/javs.2025.381713.1604