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# Radiographic evaluation to assess bone density upon administration of Low-Intensity 660 nm Laser Therapy alongside Nano-hydroxyapatite and antioxidant compounds (N-acetylcysteine) for Dental Socket Preservation Following Extraction Utilizing CBCT

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### ABSTRACT:

**Purpose:** To assess the preservation of bone density upon tooth extraction utilising low-level laser therapy together with nano-hydroxyapatite and antioxidant substance (N-acetylcysteine).

**Materials and Methods:** The study sample comprised 30 randomly selected patients, evenly distributed across three groups. Group I, the socket was sealed by an absorbable collagen membrane without graft materials following tooth extraction. In Group II, a combination of nano-hydroxyapatite and antioxidant agents (N-acetylcysteine) was introduced into the empty socket, followed by placing a bioabsorbable collagenous membrane over the grafting material, then sutured. Group III, the same mixture of nano-hydroxyapatite and antioxidant agents was applied to the extracted socket, followed by the adaptation of an absorbable collagen membrane over the graft and suturing, in addition to low-level 660 nm laser therapy (LLLT) administered three times weekly for two weeks, power of 25 mW, and duration of 120 seconds. Bone density at the extraction site was evaluated using cone beam computed tomography (CBCT). Bone density was measured immediately and at three and six months postoperatively.

**Results:** The NHA and NAC groups subjected to LLLT exhibited the most significant improvement in bone density measurements compared to the other groups.

**Conclusions:** a combination of NHA and antioxidant agents (N-acetylcysteine), along with the application of LLLT to alveolar socket post-extraction, minimized bone resorption after six months from grafting that can be safely utilized for socket maintenance.

**Keywords:** LLLT, bone healing, NHA, NAC

### I. INTRODUCTION

To prevent changes in hard and soft tissues caused by tooth loss, alveolar socket preservation involves inserting biomaterials into the extracted tooth's socket during extraction [1].

Alloplastic, allogeneic, xenogeneic, and Autogenous bone substitutes, along with membranous barriers due to their capacity to preserve space, facilitate rapid bone turnover, and exhibit biocompatibility, as well as additional compounds as bone morphogenetic protein (BMP), platelet-rich plasma, platelet-rich fibrin (PRF), and titanium-prepared platelet-rich fibrin (T-PRF), have all been concurrently documented with various other substances [2-8].

Hydroxyapatite (HA), the predominant component of the mineral composition of bone and teeth and exhibiting biocompatibility, has been extensively studied as a biomaterial within the medical field [9].

Its significant similarity to the natural bone composition largely explains the regenerative potential mechanism of NHA [10, 11]. Furthermore, bone tissue promptly adheres to hydroxyapatite, resulting in the deposition of newly formed bone [12]. The HA surface has demonstrated the ability to enhance osteoblastic cell proliferation, differentiation, and adhesion [13]. In addition to its antioxidant properties, N-acetylcysteine (NAC) demonstrates a range of pharmacological effects on

osteoblast lineage cells [14]. NAC proved to enhance bone regeneration on implanted biomaterial by improving biomaterial cytocompatibility and reducing wound infection [15-17]. NAC has been shown to enhance osteogenesis by promoting differentiation of osteoblast-like cells rather than inducing bone marrow-derived mesenchymal stem cells (BMSCs) to differentiate into osteoblast progenitor cells [18].

Diode lasers employed in Low-level laser therapy (LLLT) have gained popularity for promoting faster wound healing. The photobiomodulation theory, or biostimulation, underpins its mechanism of action. Due to its nonthermal influence, it exerts a biostimulatory effect that facilitates modifications in cellular behavior. It induces cellular modifications by influencing membrane calcium channels and the mitochondrial respiratory chain. This intervention promotes cellular metabolism and proliferation, thereby facilitating expedited wound healing. Furthermore, LLLT exerts an immediate analgesic effect [19].

Photobiomodulation therapy has been demonstrated to expedite and augment the formation of new bone tissue in animals in vivo, enhance epithelial cell proliferation, and foster angiogenesis [20-25]. Diode lasers emitting light at 904 and 660 nm influence bone healing and enhance osteoblastic activity [8, 26-27].

## II. MATERIALS AND INSTRUMENTS

### A- METHODOLOGY

Following the execution of an informed consent form detailing the procedure and expected outcomes post-treatment, a careful selection of thirty patients from the Oral and Maxillofacial Surgery Department's Outpatient Clinic Department of Kobri el Kobbah Military Hospital were divided into three groups based on treatment methods for participation in this study.

#### Inclusion criteria:

- Medically healthy adult patients aged 20–40 years.
- A hopeless, decayed root or teeth where extraction is the recommended treatment.
- posterior mandibular teeth.

#### Exclusion criteria:

- Systemically diseased participants.
- Existence of acute infections.
- Heavily smoking patients.
- Pregnancy or nursing.

#### Grouping:

To enhance the reliability of our findings. Thirty patients were randomized into three treatment groups, & each group was made up of ten individuals:

1. Group I: Following tooth extraction, the socket was covered with an absorbable collagen membrane (COLLAFLEX®, UNI CARE BIOMEDICAL, USA) and sutured with Vicryl 4-0 (Ethicon, USA).

2. Group II: After the tooth was extracted, the Socket was filled using a combination of Nano-hydroxyapatite (IngeniOs HA, USA) plus antioxidant agents (N-acetylcysteine) (Acetylcysteine, Pharmazell GmbH, Rosenheimer Str. 4383064 Raubling, Germany). Then, the socket was sutured using a resorbable membrane.
3. Group III: After the tooth was extracted, the Socket was filled using a combination of Nano-hydroxyapatite plus antioxidant agents (N-acetylcysteine). Then, the socket was sutured using a resorbable membrane, and low-level laser therapy (LLLT) was administered.

#### Surgical Procedures:

L.A. (Scandonest 3%, Canada) was achieved based on group diversity. The severely decayed tooth was extracted as a traumatically as possible to prevent soft tissue damage and bone loss. Following the dental extraction in the first group, a membrane was applied over the socket, and Vicryl 4-0 sutures were utilized.

In the second group, an empty alveolus received a Nano-hydroxyapatite filling plus antioxidant (N-acetylcysteine), after which the graft was covered with a collagen membrane that was then sutured [Figure 1].



**Figure 1:** alveolar socket after grafting with NHA & NAC & suturing

In the third group, the socket was preserved with a combination of Nano-hydroxyapatite and antioxidant agents (N-acetylcysteine). A bioabsorbable collagenous membrane was positioned, covering the graft and sutured in place. 660 nm LLLT was conducted three times weekly for two weeks using the Sirolaser advance plus (Dentsply Sirona, GMBH, Germany) at a power of 25 mW and for 120 seconds on the buccal, lingual, and occlusal surfaces [Figure 2].

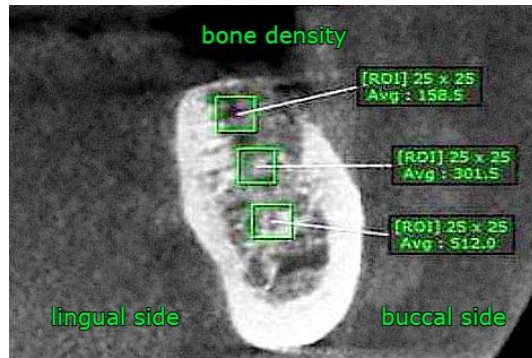


**Figure 2:** Sirolaser advance plus

## Radiographic Evaluation:

CBCT was conducted immediately post-surgery (baseline), at 3 months, and at 6 months to assess bone density as follows:

Bone density measurements were conducted precisely at the socket's Centre, allowing for a thorough comparison across multiple time intervals. This approach ensures a comprehensive understanding of changes over time. [Figure 3].



**Figure 3:** Coronal view Cone Beam CT showing bone density measurement

## III. RESULTS

### Statistical analysis:

IBM SPSS (version 20.0) was used for data analysis.

### Demographic data:

The age range of the thirty patients was as follows: Study Group I ranged from 21.0 to 39.0 years, with a mean age of  $31.20 \pm 5.75$  years; Research Group II also ranged from 21.0 to 39.0 years, with a mean age of  $31.20 \pm 5.75$  years; and Study Group III ranged from 21.0 to 39.0 years, with a mean age of  $29.61 \pm 5.53$  years. The average age of the groups did not exhibit a statistically significant difference. Control Group I comprised five males and five females, Study Group II comprised five males and five females, and Study Group III included four males and six females. The sex distributions of the groups did not exhibit statistically significant differences.

### Radiographic evaluation (bone density):

The average bone density remained the same in all groups for six months. Upon comparing the bone density among the three groups at each follow-up interval, it was noted that group III exhibited the highest values, except at baseline, where group II held the highest value, while group I consistently recorded the lowest values at all follow-up times. The bone density increased in group III after the six-month interval, but the increase was nonsignificant.

*Table 1:* Overall mean bone density immediately, at 3 months, and 6 months post-extraction, along with the significance levels of the differences observed.

Group	Group I (control) N=10	Group II (study) N=10	Group III (study) N=10	H	P
Bone density	Mean $\pm$ SD	Mean $\pm$ SD	Mean $\pm$ SD		
Base line	386.95 $\pm$ 130.35	418.64 $\pm$ 165.67	405.24 $\pm$ 180.07	0.173	0.917 (ns)
3 months	427.12 $\pm$ 128.91a	480.54 $\pm$ 171.36	489.47 $\pm$ 179.16	1.18	0.555 (ns)
6 months	482.06 $\pm$ 136.36a	544.40 $\pm$ 150.81	606.61 $\pm$ 178.04	1.75	0.416 (ns)

H: test of Kruskal–Wallis.

P: p-value for comparison between the studied groups.

\*: "significance level  $p \leq 0.05$ ; ns=non-significant".

## IV. DISCUSSION

To optimize outcomes, surgeons must ensure high-quality bone structure and appropriate dimensions for implant insertion by preserving the alveolar ridge [28].

Various grafting materials have been utilized for socket preservation, including autogenous bone grafts, allografts, xenografts, and alloplasts [29].

Takata et al. [30] assert that incorporating collagen and HA may Encourage the formation of new bone. during the healing process & establish a more conducive medium for osteoblastic cell proliferation after adhesion and differentiation.

Supplementary HA markedly enhanced the mineralization of newly formed bone in vivo, as reported by Rajzer et al. [31]

A study on mouse calvarial cell cultures suggests that NAC may enhance osteoblastic development.[32]

This study utilized a combination of NHA and NAC as a bone substitute, with and without LLLT at 660 nm, 25 mW for 120 seconds, applied to the buccal, lingual, and occlusal surfaces three times weekly for two weeks, to assess the improvement of bone formation and density following tooth extraction.

Meanwhile, the NHA and NAC groups exhibited more enhancement in bone density than the control group.but non significantly.

The application of lasers increased the quantity of viable osteocytes in the irradiated area, demonstrating the substantial potential for new bone formation.

Other studies indicate that LLLT positively influences osteoblast development and proliferation in cell cultures and animal models of bone formation. [36, 37]

A recent study employing Nano-hydroxyapatite bone grafts and NAC to maintain the alveolar ridge post-tooth extraction. The minimal width reduction was observed in nanohydroxyapatite combined with NAC and NAC alone. The most significant enhancement in bone density and the

minimal decrease in bone height were noted with Nano-hydroxyapatite combined with NAC and NAC independently [38].

Brawn et al. investigated the impact of LED phototherapy on a sinus augmented with a granular bovine bony substitute in a distinct clinical case study [39]. A 20 mW/cm<sup>2</sup> Light Emitting Diode (LED) phototherapy treatment was administered twice daily for ten minutes over two weeks. A histological examination of a biopsy conducted four weeks later indicated that LED phototherapy had enhanced recovery.

F. Mafrá de Lima et al. assert that LLLT and NAC synergistically augment MIP-2 (Macrophage Inflammatory Protein 2) mRNA expression in alveolar macrophages under the influence of lipopolysaccharide or H<sub>2</sub>O<sub>2</sub>. They propose that intracellular reactive oxygen species generation and NF-κB signaling may be implicated [40].

### Limitations

The duration of observation in this study may be limited.

## IV-CONCLUSION

Under the constraints of the present study, LLLT combined with a mixture of nanohydroxyapatite (NHA) and antioxidant agents (N-acetylcysteine) as a graft in the alveolar socket can be effectively employed for ridge preservation following tooth extraction.

### Recommendations

Future studies may necessitate histological assessments. Additional research is required to examine the effects of NHA and NAC over extended observation periods (12 to 24 months).

### Declarations

### Funding:

The authors financed this research independently.

### Data availability:

All data pertinent to this research were gathered from the participants utilizing CBCT, which facilitated recording their bone density measurements post-treatment.

### Ethics approval:

Ethical approval reference: NILES-EC-CU 24/10/16. Additionally, consent was documented in a written format that complied with the institutional ethics committee's approval and was comprehended by each participant.

### Competing interests:

There are no conflicts of interest.

## REFERENCES

- Vignoletti F, Matesanz P, Rodrigo D, Figuero E, Martín C, Sanz M. Surgical protocols for ridge preservation after tooth extraction. A systematic review. *Clin Oral Implants Res* 2012;23(5):22–38.
- Histological comparison of healing following tooth extraction with ridge preservation using enamel matrix derivatives versus Bio-Oss Collagen: a pilot study. *Int J Oral Maxillofac Surg* 2013;42(12):1522–8.
- Eskow AJ, Mealey BL Evaluation of healing following tooth extraction with ridge preservation using cortical versus cancellous freeze-dried bone allograft. *J Periodontol* 2014;85(4):514–24.
- Calasans-Maia M, Resende R, Fernandes G, Calasans-Maia J, Alves AT, Granjeiro JM. A randomised controlled clinical trial to evaluate a new xenograft for alveolar socket preservation. *Clin Oral Implants Res* 2014;25(10):1125–30.
- Suttapreyasri S, Leepong N. Influence of platelet-rich fibrin on alveolar ridge preservation. *J Craniofac Surg* 2013;24(4):1088–94.
- Kakar A, Rao BHS, Hegde S, Deshpande N, Lindner A, Nagursky H, et al. Ridge preservation using an in situ hardening biphasic calcium phosphate (beta TCP/HA) bone graft substitute- a clinical, radiological, and histological study. *Int J Implant Dent* 2017; 3(1):25.
- Kim YJ, Lee JY, Kim JE, Park JC, Shin SW, Cho KS. Ridge preservation using demineralised bone matrix gel with recombinant human bone morphogenetic protein-2 after tooth extraction: a randomised controlled clinical trial. *J Oral Maxillofac Surg* 2014;72(7):1281–90.
- S. Hemaid, A. Saafan, M. Hosny, and G. Wimmer, 'Enhancement of Healing of Periodontal Intrabony Defects Using 810 nm Diode Laser and Different Advanced Treatment Modalities: A Blind Experimental Study'. *Open Access Maced J Med Sci*. 2019; 7(11):1847-53.
- Erlind, P., Lait, K.B., Gaspare, P., Gianluca, T. and Guido, M. Nanohydroxyapatite and its applications in preventive, restorative and regenerative dentistry: a review of literature. *Ann Stomatol (Roma)* 2014;5(3):108–14.
- Chaves, M.D., DeSouza, L.S., DeOliveira, R.V., Holgado, L.A., Filho, H.N., Matsumoto, M.A. and Ribeiro, D.A. Bovine hydroxyapatite (Bio-Oss ((R))) induces osteocalcin, RANK-L and osteoprotegerin expression in sinus lift of rabbits. *J Craniomaxillofac Surg* 2012; 40:315-20.
- Meimandi, P.A., Oryan, A.Z., Shafiei, S., Bigham, A.S. Effectiveness of synthetic hydroxyapatite versus Persian Gulf coral in an animal model of long bone defect reconstruction. *J. OrthopTraumatol*. 2013; 14:259-68.

12. Kattimani, V., Lingamaneni, K.P., Chakravarthi, P.S., Kumar, T.S. and Siddharthan, A. Eggshell Derived Hydroxyapatite: A New Era in Bone Regeneration. *J Craniofac Surg*. 2016; 27:112-17.
13. Abdel-Fattah, W.I. and Elkhooly, T.A. Nano-beta-tricalcium phosphates synthesis and Biodegradation: 2. Biodegradation and apatite layer formation on nano-beta-TCP synthesised via microwave treatment. *Biomed Mater* 2010; 5:35015.
14. Almeida, M.; Han, L.; Martin-Millan, M.; O'Brien, C.A.; Manolagas, S.C. Oxidative stress Antagonizes Wnt signalling in osteoblast precursors by diverting beta-catenin from T cell factor to forkhead box O-mediated transcription. *J. Biol. Chem*. 2007;282(37):27298-305.
15. Yamada, M.; Ishihara, K.; Ogawa, T.; Sakurai, K. The inhibition of infection by wound pathogens on scaffold in tissue-forming process using N-acetyl cysteine. *Biomaterials* 2011; 32:8474-85.
16. Tsukimura, N.; Yamada, M.; Aita, H.; Hori, N.; Yoshino, F.; Chang-Il Lee, M.; Kimoto, K.; Jewett, A.; Ogawa, T. N-acetyl cysteine (NAC)-mediated detoxification and functionalisation of poly(methyl methacrylate) bone cement. *Biomaterials* 2009; 30:3378-89.
17. Yamada, M.; Ueno, T.; Minamikawa, H.; Sato, N.; Iwasa, F.; Hori, N.; Ogawa, T. N-acetyl cysteine alleviates cytotoxicity of bone substitute. *J. Dent. Res*. 2010; 89:411-6.
18. Yamada, M.; Tsukimura, N.; Ikeda, T.; Sugita, Y.; Att, W.; Kojima, N.; Kubo, K.; Ueno, T.; Sakurai, K.; Ogawa, T. N-acetyl cysteine as an osteogenesis-enhancing molecule for bone regeneration. *Biomaterials*. 2013; 34:6147-56.
19. Bhagyashree RK, Amit AA, Chetan PR. Effect of low-level laser therapy on wound healing and patients' response after scalpel gingivectomy: A randomized clinical split-mouth study. *J Indian Soc Periodontol*. 2018; 22(5):419-26.
20. Kawasaki K, Shimizu N Effects of low-energy laser irradiation on bone remodeling during Experimental tooth movement in rats. *Lasers Surg Med* 2000; 26(3):282-91
21. Shirazi M, Akhoundi MSA, Javadi E, Kamali A, Motahhari P, Rashidpour M, Chiniforush N The effects of diode laser (660 nm) on the rate of tooth movements: an animal study. *Lasers Med. Sci*. 2015; 30(2):713-8.
22. Kesler G, Romanos G, Koren R Use of Er: YAG laser to improve osseointegration of titanium alloy implants—a comparison of bone healing. *Int. J. Oral Maxillofac Implants*. 2006; 21(3):375-9.
23. Jonasson TH, Zancan R, de Oliveira-Azevedo L, Fonseca AC, Silva MCD, Giovanini AF, Zielak JC, Araujo MR Effects of low-level laser therapy and platelet concentrate on bone repair: histological, histomorphometric, immunohistochemical, and radiographic study. *J. Craniomaxillofac Surg*. 2017; 45(11):1846-53.
24. Angeletti P, Pereira MD, Gomes HC, Hino CT, Ferreira LM. Effect of low-level laser therapy (GaAlAs) on bone regeneration in mid palatal anterior suture after surgically assisted rapid maxillary expansion. *Oral Surg Oral Med Oral Pathol Oral Radiol Endod*. 2010;109(3): e38-e46.
25. Avila-Ortiz G, Elangovan S, Kramer KW, Blanchette D, Dawson DV. Effect of alveolar ridge Preservation after tooth extraction: a systematic review and meta-analysis. *J. Dent. Res*. 2014; 93(10):950-8.
26. Nissan J, Assif D, Gross MD, Yaffe A, Binderman I. Effect of low-intensity laser irradiation on surgically created bony defects in rats. *J. Oral Rehabil* 2006; 33(8):619-24.
27. Nicolau RA, Jorgetti V, Rigau J, Pacheco MT, dos Reis LM, Zangaro RA. Effect of low-power GaAlAs laser (660nm) on bone structure and cell activity: An experimental animal study. *Lasers Med Sci* 2003;18(2):89-94.
28. Álvarez-Camino JC, Valmaseda-Castellón E, Gay-Escoda C. Immediate implants placed in fresh sockets associated to periapical infectious processes. a systematic review. *Med. Oral Patol. Oral Cir. Bucal* 2013;18(5): e780-5.
29. Balli G, Ioannou A, Powell CA, Angelov N, Romanos GE, Soldatos N. Ridge Preservation Procedures after Tooth Extractions: A Systematic Review. *Int J Dent*. 2018; 3:8546568.
30. Takata T, Miyauchi M and Wang HL. Migration of osteoblastic cells on various guided bone Regeneration membranes. *Clin Oral Implant Res* 2001;12(4):332-8.
31. Rajzer I, Menaszek E, Kwiatkowski R, Chrzanowski W. Bioactive nanocomposite PLDL/nanohydroxyapatite electrospun membranes for bone tissue engineering. *J Mater Sci. Mater Med*. 2014; 25(5):1239-47.
32. Jun JH, Lee SH, Kwak HB, Lee ZH, Seo SB, Woo KM, et al. N-acetylcysteine stimulates osteoblastic differentiation of mouse calvarial cells. *J Cell Biochem* 2008; 103:1246-55.
33. Wheeler SL. Sinus augmentation for dental implants: The use of alloplastic materials. *J Oral Maxillofac Surg*. 1997; 55(11):1287-93.
34. Cunha MJ, Esper LA, Sbrana MC, de Oliveira, do Valle, de Almeida. Effect of low-level laser on bone defects treated with bovine or autogenous bone grafts: in vivo study in rat calvaria. *Biomed Res Int*. 2014; 5:104-230.
35. de Almeida AL, Medeiros IL, Cunha MJ, Sbrana MC, de Oliveira PG, Esper LA, et al. The effect of low-level

- laser on bone healing in critical size defects treated with or without autogenous bone graft: An experimental study in rat calvaria. *Clin. Oral Implants Res.* 2014; 25(10):1131–6.
36. Medrado AR, Pugliese LS, Reis SR, Andrade ZA. Influence of low-level laser therapy on wound healing and its biological action upon myofibroblasts. *Lasers Surg. Med.* 2003; 32(3):239–44.
  37. Hopkins JT, McLoda TA, Seegmiller JG, David Baxter G. Low-level laser therapy facilitates Superficial wound healing in humans: a triple-blind, sham-controlled study. *J. Athl. Train.* 2004; 39(3):223–9.
  38. Elkhier, S.G., fakharany, A.M., & Hassanen, A.M. “Evaluation of N-acetyl Cysteine with NanoHydroxyapatite Bone Graft for Preservation of Alveolar Ridge After Teeth Extraction.” *Al-Azhar Journal of Dental Science* 2020; 23(3): 279-84.
  39. Brawn P, K won-Hong A. Phototherapy enhances bone regeneration in direct sinus lifts. In: BIOS Biomedical Optics Symposium and exhibition SPIE Photonics west. 2007. Scientific poster.
  40. De Lima FM, Villaverde AB, Albertini R, de Oliveira AP, Faria Neto HC, Aimbire F. Low-level laser therapy associated to N acetylcysteine lowers macrophage inflammatory protein-2 (MIP-2) mRNA expression and generation of intracellular reactive oxygen species in alveolar macrophages. *Photomed. Laser Surg.* 2010 ;28(6):763-71.

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