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Staphylococcus Methicillin-Resistant aureus and Vancomycin-Resistant Staphylococcus aureus in Ready-toeat Meat Products



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Abstract

taphylococcus aureus is an emerging opportunistic threat to both animal and human health. It is Classified as a high-priority pathogen owing to its widespread nature, making it easy to enter the human food chain. This bacterium is highly versatile and can lead to diverse infections and several foodborne outbreaks all over the world, as it produces approximately 28 types of potent superantigens enterotoxins, the most common types of these toxins implicated in foodborne outbreaks are staphylococcal enterotoxins A, B, C, and D. Additionally, this pathogen can release another toxins which are responsible for another infections including: toxic shock syndrome, scale skin syndrome, folliculitis, and pneumonia, in addition to spreading factors facilitating its pathogenicity in the infected host. Moreover, S. aureus can cause various illnesses, including suppurative dermatitis, osteomyelitis, MRSA Heart Infection (Endocarditis), and septicemia. With the discovery of antimicrobial-resistant bacteria, S. aureus is one of the firstly pathogens developing multiple antimicrobial resistant to the antimicrobials used for its eradication and treatment in human through various mechanisms. Interestingly, the prevalence of S. aureus and its resistant strains varies across the different geographical regions in Egypt and worldwide, mainly influenced by the hygienic status, personal practices, and cultural traditions in the surveyed countries. This review provides concise and diverse information about the microbial characteristics, virulence factors, prevalence, and antibiotic resistance of S. aureus, MRSA, and VRSA, in ready-to-eat meat products, in Egypt and worldwide.

Keywords: S. aureus, Ready-to-eat meat products, Foodborne pathogens, MRSA, VRSA.

Introduction

Ready-to-eat (RTE) food is defined as food that is prepared and ready for consumption without further processing or requires only minimal steps, such as rewarming. However, despite its delicious taste, RTE food can be a silent source of foodborne pathogens. The preparation of RTE food involves various critical points through which it can be contaminated with such pathogens, including the quality of ingredients, the microbial status of the water used for washing food and utensils, the type of packaging material, the hygienic practices of all individuals involved in food processing, as well as the post-processing cross contamination of the prepared meals [1].

S. aureus is one of the most extensively studied foodborne pathogens related to RTE food after Salmonella and E. coli [2]. S. aureus is a versatile

pathogen that can infect various types of food during processing, as it is an asymptomatic colonizer found in all warm-blooded mammals, including human, inhabiting their noses and skin, which facilitates its entry into the food chain. Moreover, many surveys have confirmed that red meat and its products, particularly in African countries, are considered an important vehicle for this bacterium [3, 4, 5]. This potential food vehicle; meat is rich in proteins that, upon breaking down into amino acids and low molecular weight peptides, may supply the nutrients necessary to support the survival of S. aureus.

The main cause of Staphylococcal foodborne outbreaks over the world is Staphylococcal enterotoxins as reported by EFSA and ECDC [6] when these powerful enterotoxins were the primary cause of 207 outbreaks involving 2268 patients, 113 hospitalizations and 1 death in 2023. Moreover, these

secreted enterotoxins recorded 577 outbreaks involving 9092 cases during the period between 2010 and 2020 as mentioned by [7].

The adaptability of *S. aureus* to various stressful conditions enables it to be a major pathogenic agent in diverse food products, leading to life-threatening food poisoning outbreaks. *S. aureus* is a facultative anaerobic microbe has a strong salt and sugar tolerance, marked ability to survive at water activity levels as low as 0.83, allowing its persistence in even semi-dried food products. *S. aureus* can thrive in NaCl concentrations up to 15%, temperatures ranging from 7°C to 48.5°C, and a wide pH range of 4.2 to 9.3 [8].

Dissemination of *S. aureus* in the food chain is primarily attributed to the poor hygienic conditions of infected food workers, unclean utensils and machines used in meat plants, and inadequate food preparation environment [9]. Interestingly, *S. aureus* can invade and colonize various body tissues, including mammary glands, skin, mucous and serous membranes of the front nares, inguinal region, rectum, as well as internal organs of both humans and different animals, mainly cattle and chickens, leading to serious illnesses that are difficult to treat [10].

Material and Methods

Sources of RTE food contamination

Many studies suggest that the leading cause of food contamination with *S. aureus* is poor hygienic handling of food, poor personal hygiene before or after heat processing, followed by improper storage conditions that enable the growth and multiplication of the bacteria with production of staphylococcal enterotoxins within contaminated food, resulting in foodborne outbreaks [11, 12, 13].

Ray et al. [14] enumerated various pathways for the invasion of RTE food by microorganisms, including human mishandling, insufficient heat treatment, slow cooling, contaminated equipment, leaving food aside for extended periods before packaging, inappropriate packaging systems, and unhygienic distribution.

However, food handlers are the most common source of *S. aureus* during food preparation in unhygienic environment using contaminated cooking utensils, particularly when workers have hand or arm lesions or cough, as this bacterium is a common commensal of the skin and mucosal membranes of the nose and throat, with estimates indicating that 80% of healthy individuals carry it [15].

S. aureus as a serious foodborne pathogen

S. aureus is a problematic pathogen with various virulence factors essential for its survival and host infection. The organism's ability to

produce multiple toxins, enzymes, and biofilms plays a significant role in its pathogenicity, including thermonuclease, hemolysins, coagulase, Staphylococcal hyaluronidase, Panton-Valentine leukocidin (PVL), staphylokinase, toxic shock syndrome toxin (TSST), adhesins, and surface proteins like protein A [16, 17].

Furthermore, this pathogen produces several pyrogenic exotoxins known as enterotoxins, also some strains of S. aureus show multidrug resistance to the most common clinically used antibiotics [18]. Several foodborne poisoning outbreaks have occurred worldwide due to consuming food containing staphylococcal enterotoxins produced in contaminated food with S. aureus in response to various environmental conditions [19]. Malachowa and DeLeo [20] explained that S. aureus is highly adaptable to different nutritional and environmental stress factors, which sustains its ability to remain viable and infective across various settings by acquiring newly developed genes encoded on different mobile genetic elements.

Staphylococcus food poisoning is one of the major causes of hospitalization and foodborne outbreaks in many countries including USA, Europe, and France as mentioned by Fetsch and Johler [19]. Likewise, S. aureus reported as serious threaten as it can remain viable for long time on the initial contaminated surfaces and utensils with small dose of infective dose of one or more of staphylococcal enterotoxins (SEs), resulting in symptoms such as nausea, vomiting, abdominal pain, watery diarrhea, and chill, however, these cases are self-limiting, they may more sever in both elderly and children individuals. Symptoms generally appear nearly 3 hours after consumption of such contaminated food [21]. There are various types of SEs secreted by enterotoxigenic S. aureus strains, nearly about 28 different SEs identified in foodborne outbreaks. The 5 classical enterotoxins (SEA, SEB, SEC, SED, and SEE) are the most common toxins implicated in food poisoning outbreaks [22, 23].

SEs are superantigens proteins that can activate large numbers of T-cells of the host's immunity. Moreover, they exhibit significant resistance to heat, digestive enzymes, and acids such as pepsin and trypsin, facilitating outcome of human digestive immunity and withstanding conditions that could eliminate their secreting bacteria. A sufficient amount of the toxin is approximately 1 μ g; however, 0.5 μ g is enough to induce symptoms of poisoning [24].

Secretion of SEs can be stimulated by various stressed environmental factors in contaminated food, such as suitable temperature, NaCl concentration, and pH, which affect both the growth and secretion of these enterotoxins. The ability of producing bacteria to endure a wide range of osmotic sugar and

salt concentrations could explain the involvement of different types of RTE food in Staphylococcus food poisoning outbreaks [25].

The growth of enterotoxigenic S. aureus strains until exceed 10⁶ cells per each gram of such contaminated food is the starting point of enterotoxins production with subsequent food poisoning symptoms, moreover, production of these toxins is mainly organized by two factors including: storage temperature of prepared food and the ingredients of the meals itself [26]. Temperature is greatly influenced on SEs synthesis more than its effect on the bacterial growth; as it appears that the ideal SEA production is in a range of temperature between 10 °C and 45°C with a positive correlation with increased temperature in protein-rich meals [27]. Additionally, the emission of SEs is affected by pH, water activity, and NaCl concentration of RTE meals; as the acidic pH achieved by addition of lactic acid in certain types of meat products appeared to be have an inhibitory effect on the production of SEA [28], furthermore, SEB and SEC are the most types of these toxins affected by the availability of water in the surrounding media; as 0.86 is the minimal water activity, which permit enterotoxins production in the contaminated food [29]. Sihto et al. [30] reported that mild stress on S. aureus in 2% NaCl containing food was sufficient to induce SEs production.

S. aureus secretes various types of hemolysin toxins (alpha, beta, gamma, and delta); their primary functions include host tissue invasion, facilitating spread, and nutrient uptake. α -Hemolysin is the most common, exerting a damaging effect on different types of host cells, including epithelial cells, endothelial cells, erythrocytes, and monocytes, hence it can induce cell death. B-Hemolysin significantly impacts the plasma membrane of human immune cells, resulting in its degradation into phosphorylcholine and ceramides. γ - and δ -hemolysins have a greatly diminished effect on human immunity [31].

One of the life-threatening illnesses is toxic shock syndrome, which is mediated by toxic shock syndrome toxin, a pyrogenic polypeptide toxin encoded by the (*tsst-1*) gene in some strains of *S. aureus*. This syndrome is characterized by fever, hypotension, and multiple organ failure due to excessive fluid and electrolyte loss from the infected patient. All of these actions can occur as a response to excessive non-conventional T-cell activation and cytokine release in the susceptible host [32].

The ability of *S. aureus* to adhere to any surface and form a biofilm is one of the most critical issues, as it facilitates the persistence of this pathogen on both biotic and abiotic surfaces. The biofilm is an extracellular matrix generated by bacterial colony adhesion and extracellular polymeric substances

composed mainly of oligosaccharides, DNA, and proteins released during bacterial growth [8]. The biofilm enables the bacteria to sustain different environmental stressors, such as cleaning agents, disinfectants, and antimicrobials, through preventing chemicals or chemotherapeutics from reaching their target due to limited diffusion or repulsion caused by the biofilm matrix itself [33]. Moreover, the biofilm may enhance the virulence of this pathogen against host immunity, which raises the risk of such infection and hardens its treatment [34]. Hence, the ability of S. aureus to form biofilms serves as a serious weapon in its pathogenesis and control measures. The biofilm formation feature of MRSA adds an extra level of difficulty of the problem of antimicrobial resistance; as the slow-growing state of the bacteria encapsulated within the polymeric matrix of the biofilm can limit the action of βlactams antibiotics, which depends on the suppression of active cellular process of wall synthesis of the invading bacteria, hence S. aureus can resist methicillin through two mechanisms: the first is the limitation of antibiotic spread in the matrix of the biofilm, while the second is the inhibition of the antibiotic action itself [35]. The process of biofilm formation is a two-step course; the first is the bacterial attachment to the invading surface via capsular antigen, then production of multilayers of polysaccharide biofilm surrounding the bacteria increasing its resistance to be more than one thousand of the planktonic cells to the antibiotics and sanitizers [36]. In food industry, the ability of S. aureus to form a biofilm is a critical issues, as the stress factors encountered by the bacteria during food processing is a strong signal that triggers production of strong biofilm act a as a shield to protect the bacteria from any threat, hence the biofilm tend to raise the pathogenic tolerance to many used disinfectants, increasing the risk of food contamination [37].

Methods of detection of MRSA & VRSA in food

Detection of MRSA in food samples depends on the molecular identification of mecA gene, which carried on staphylococcal cassette chromosome mec (SCCmec), a DNA fragment ranging from 21 to 67 kb in size present as mobile genetic element carried the different genes horizontally among the bacterial cells [38]. Many studies confirmed that mecA gene is the reliable genetic marker for MRSA [5, 17, and 34]. On the other hand, VRSA detection requires the genetic analysis of the bacterial DNA, and finding of one of the van genes including: vanA, vanB, or vanC genes as discussed by [39]. Other available methods for detection of MRSA include colony morphology on the specific media, Gram staining reaction, and biochemical testing of catalase, hyaluronidase, and coagulase, in addition to crossing priming amplification (CPA). CPA relies on isothermal nucleic acid amplification strategy which can amplify the DNA extracted from *S. aureus*, then it depends on the visual display of the colored result within 40 minutes, hence this method overcomes PCR method by the short time required to obtain the final results [40].

Results and Discussion

Prevalence of S. aureus in RTE meat products in Delta region

In Egypt, different studies recorded the prevalence of *S. aureus* in RTE food samples gathered from various restaurants and street vendors to evaluate different levels of hygienic safety of the sold meals in our local market. Saif et al. [41] found that 50.8% (61/120) samples of RTE meat products including: beef kofta, shawarma, burger, and luncheon collected from street vendors in Benha city were contaminated with *S. aureus*. Higher results recorded by Mahros et al. [17], who found that 83.1% (187/225) of hot dog and burger sandwiches purchased from public restaurants in Mansoura city were positive for *S. aureus*.

However, lower results detected by Zakaria et al. [42], who carried a study on RTE chicken burger, sausage, and luncheon reporting that 11.3% (17/150) were contaminated with *S. aureus*. In Zagazig city, Morshdy et al. [43] reported that 22% (33/150) of various RTE chicken burger, fillet, luncheon, nuggets, and pané samples were positive for *S. aureus*. Different prevalence of *S. aureus* in RTE meat samples discussed in (Table1).

In our review, we collected data on 1659 RTE meat products sold in different food outlets across various districts in Egypt. Of these samples, 743 (44.8%) were contaminated with S. aureus at different levels. This contamination may be attributed to the variety of meat processing methods and the unhygienic practices of the workers involved in preparing and packaging of these products. The hygienic practices are critical points for the introduction of pathogens, as S. aureus is a commensal human flora inhabitant of the skin and mucous membrane of the nose, mouth, and throat. Additionally, the ambient temperature in Egypt ranges between 35 and 37°C, which encourages the survival and growth of S. aureus in initially contaminated meat products [55].

Prevalence of S. aureus in RTE meat products around the developing countries

S. aureus is a worldwide serious pathogen as it considers the main cause of several food poisoning outbreaks; hence many authors analyzed different types of RTE meat products to confirm its existence, prevalence, and characterization. Wu et al. [56 analyzed 645 RTE meat products selected from the favorable Chinese food to reported that 79 (12.2%) of the tested samples were positive for S. aureus.

Lower results detected by Lin et al. [57], who examined 1209 RTE meat products collected from different restaurants in Sichuan province, China and recording that 31 (2.6%) were contaminated with *S. aureus*.

In a Nigerian study, Omoruyi and Ibegbulam [58] made a survey on the microbiological quality of most common consumed RTE food including 20 meat pies recording that all of the tested samples (100%) were hygienically unsatisfactory for human consumption due to contamination with *S. aureus*. Various rates of *S. aureus* in RTE meat products in previous studies recorded in different countries illustrated in (Table 2).

Interestingly, we observed significant variations in the prevalence of *S. aureus* contamination rates among different countries worldwide. Our review included about 6626 RTE meat samples, within which only 441 (6.7%) were contaminated with S. aureus. Notably, Nigeria and Turkey showed the highest contamination rate, while RTE meat-based food sold in Algeria, Thailand, South Africa, Ghana, and Brazil showed a rate lower than 50% of the tested samples, indicating they were hygienically unacceptable. On the contrary, lower contamination rates were recorded by researchers in Libya, Afghanistan, Cameron, and Jordan. This variation in the incidence rates of S. aureus in RTE food influenced by several factors, including washing raw ingredients with contaminated water, the use of dirty equipment and utensils such as cutting tools, initial microbial contamination of meat with inadequate heat treatment, cross contamination between cooked and raw foods, prolonged food storage, and unhygienic personal practice during preparation, processing, and packaging [70, 71].

Development of Antimicrobial-Resistant S. aureus

The increase in the emergence of antimicrobialresistant bacteria has become a significant health challenge, leading to a rise in prolonged hospital stays, treatment failure, extensive treatment costs, and eventually, deaths. S. aureus is one of the most commonly occurring bacteria, developing resistance to most of the antimicrobials used to treat it. S. aureus first developed resistance to penicillin and methicillin in 1961 within American hospitals; subsequently, it was isolated from cattle with mastitis in Belgium, as reported by Krumova-Valcheva [72]. Nowadays, these bacteria continue to develop resistance to many available antimicrobial drugs, including vancomycin, linezolid, and daptomycin, which are used as a final line treatments for humans and animals [73].

The misuse and overuse of antimicrobials in the veterinary field, either as growth promoters or as treatments of infections, as well as in human medicine, is the main cause of the development of antimicrobial drug resistance genes. These genes have entered the population food chain not only

through direct contact with animals but also via food of animal origin, particularly from certain species known to be common reservoirs for multidrugresistant *S. aureus*, including pigs, cattle, and poultry [74].

Mecithillin (β-lactams antimicrobial) was the drug of choice for the treatment of S. aureus infections for only two years, while some strains of S. aureus could produce a modified penicillinbinding protein known as PBP2a. The PBP2a blocks the activity of most of β-lactams as the inhibitory control unit of transpeptidation reactions that link peptidoglycan chains, the key element of bacterial cell walls. Hence, the bacteria can cross-link a stable peptidoglycan layer in their cell wall, preserving their cytoplasmic contents [35]. Mecithillin-resistant S. aureus (MRSA) not only deactivates the action of also mecithillin, but renders penicillins, cephalosporins, and carbapenems inactive against bacterial strains expressing PBP2a.

An additional pathway for resistance to β -lactams is the synthesis of β -lactamases, which is a group of enzymes encoded by the *blaZ* gene carried by certain plasmids [75]. However, these enzymes have restricted substrates, including natural penicillins and aminopenicillins.

The resistance mechanism to β-lactams is encoded by the *mecA* gene located on staphylococcal chromosomal cassette mec (SCC*mec*) cassettes, which are classified as mobile genetic elements present within the genomes of methicillin-resistant *S. aureus* strains, but not in susceptible strains, and can be transmitted either vertically or horizontally within bacterial generations. [75]. The *mecA* gene is not only confined to *S. aureus*, but it can also be detected in 64 species belonging to the genus Staphylococcus. The *mecA* gene is the most commonly identified one in MRSA; moreover, the *mecC* gene is also detected in many human and animal cases [76].

Overall, due to the widespread concern over MRSA contamination of food, a detailed investigation of the prevalence of this serious pathogen will be discussed, including the type and number of examined samples and the percentage of *mecA*-positive isolates among the recovered *S. aureus* isolates (Table 3-4).

In the last two decades, the global increase in vancomycin-resistant *S. aureus* (VRSA) prevalence has become an emerging health issue, as vancomycin has been described as the final resort for treating difficult cases of *S. aureus* infections. Until 2020, the highest prevalence of VRSA among *S. aureus* isolates was recorded in Africa at 16%, followed by 5%,4%,3%, and 1% in Asia, North America, South America, and Europe, respectively [86].

Vancomycin is one of the oldest glycopeptide antibiotics, first used to treat MRSA infections at the

end of the 1980s. In 2002, the first isolate of *S. aureus* showing complete resistance to vancomycin was recorded in the USA [87]. Vancomycin resistance is mediated by *van* gene clusters, which originated from *Enterococcus faecalis* and were then transmitted to MRSA in a mixed infection in mice [88]. Although several gene clusters are responsible for the resistance of bacteria to vancomycin, including *vanA*, *vanB*, *vanD*, *vanF*, *vanI*, *vanM*, *vanC*, *vanE*, *vanG*, *vanL*, and *vanN*, which have been confirmed until now, the *vanA* gene is the most common one identified in food and clinical samples [88].

few studies reported Unfortunately, incidences of VRSA in RTE meat products as most studies depend on the disc diffusion method which is not accurate method for detection of vancomycin resistant S. aureus strains. In Egypt, Saber et al. [53] recorded a high prevalence of VRSA by 26.7% among the S. aureus isolates recovered from shawarma and burger sandwiches collected from different restaurants in Al-Sharkia government. In contrary, El-Elshebrawy et al. [34] recorded that only 3.9% of S. aureus isolates in tested chicken sandwiches were VRSA. Further studies in Egypt and worldwide discussed in (Table 5-6)The continuously emerged antimicrobial resistance of S. aureus can be attributed to the high level of chemical pollution by heavy metals, polyfluoroalkyl compounds (PAFS), disinfectants, and pesticides which used in cleaning, sanitation and disinfection process in animal farms; hence subjecting of such pathogen to these chemicals result in appearance of what is called co-resistance, and co-selection make the bacteria resistant to multiple antimicrobial [95].

Challenges on food industries regarding MRSA & VRSA

Food industry faces significant challenges due to the emergence of MRSA in the food chain. MRSA can contaminate raw meat, vegetables, and processed meat sandwiches [96]. The existence of MRSA foodborne pathogen throughout the human food chain is a critical public health issue, aside from the food poisoning outbreaks caused by the bacteria and their enterotoxins, which are responsible for many human diseases, MRSA can transmit the resistance genes to the consumers, reducing treatment options and increasing the morbidity and mortality [97]. Furthermore, the ability of MRSA to produce an ideal biofilm reducing the efficacy of most of detergents and sanitizers used in the hygienic practices in environment of food plants and slaughterhouses [98]. Moreover, controlling MRSA and VRSA requires strict hygienic measures, regular screening in food production environments, and effective antibiotic stewardship in animal farming. These efforts increase operational costs and demand enhanced monitoring systems, making MRSA a complex and persistent threat to food safety and consumer health.

Trails considering Control and prevention of MRSA & VRSA in RTE meat products

Several recent trails have explored effective strategies for the prevention and control of MRSA and VRSA in processed meat products. Firstly, various natural antimicrobials, such as clove essential oils, has demonstrated strong inhibitory effects against MRSA owing to the active compounds eugenol, which suggested having inhibitory effect exceeding 98% in a period of 8 hours on the growing S. aureus [99]. Secondly, Hu et al.,[100] reported that the essential oils extracted from Litsea cubeba have a destructive effect on the cell membrane of MRSA with subsequent leakage of the intracellular components. Moreover, Chinese cinnamon and red thyme recorded a significant suppressive action of the most pathogens threat the food industry including MRSA in a study conducted by [101]. All of these natural preservatives overcome the synthetic ones, which preserve the product at the expense of the consumer health.

Furthermore, the golden key to prevent the existence of MRSA and VRSA in RTE meals is improving hygienic handling and personal practices in food processing facilities, which is shown significant reduction in the contamination rate of these foodborne pathogens [102]. Gamma irradiation is another intervention, where doses around 3 KGy have successfully eliminated MRSA in poultry meat without any deleterious effect on the meat quality [103]. Innovative packaging approaches, including antimicrobial films containing plant extracts or silver nanoparticles, have also contributed in the reduction

of MRSA growth curve during storage period [104]. These trails collectively underscore the importance of integrated control measures combining natural antimicrobials, advanced processing techniques, strict hygiene protocols, as well as the periodical surveillance studies to ensure food safety and limit the spread of MRSA and VRSA through meat products.

Conclusion

This review provides inclusive data on *S. aureus* and MRSA infections in ready-to-eat meat products. It discusses the virulence factors, prevalence, antibiotic resistance of *S. aureus*, and the different mechanisms of MRSA development and its prevalence. The data indicate that *S. aureus* and its antibiotic-resistant strains pose a significant public health risk. Therefore, critical measures should be implemented for regulating antibiotic use in veterinary medicine to prevent the transmission of these resistant bacteria to humans via the food chain, and also require a great wake-up call for public health authorities to establish high hygienic standards in all restaurants and food outlets to safeguard human health globally.

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Declaration of Conflict of Interest

The authors declare that there is no conflict of interest.

TABLE 1. Prevalence of S. aureus in different RTE meat products sold in Delta region

| Type of product | Region | Number of tested samples | Number (%) of S. aureus +ve | Reference |
|--|----------------------------|-----------------------------|-----------------------------|-----------|
| Beef burger, luncheon, shawerma, and sausage | Dakahlia Governorate | 80 (20 each of) | 15 (18.75%) | [33] |
| Beef luncheon and chicken luncheon | Cairo Governorate | 120 (60 each of) | 8 (6.7%) | [34] |
| Beef shawerma, hawawshi, and kofta | El-Qalyobia governorate | 105 (35 each of) | 28 (26.7%) | [35] |
| Beef Kofta (40), burger (40), hawawshi (35), beef shawerma (35), and beef sausage (25) | Sohag governorate | 175 | 14 (8%) | [36] |
| Beef kofta shawerma, sausage, and liver | El-Qalyobia governorate | 100 (25 each of) | 58 (58%) | [37] |
| Beef kofta and luncheon | El-Gharbia governorate | 50 (25 each of) | 15 (30%) | [38] |
| Beef kofta, luncheon, burger, shawarma, hawawshi, liver, and sausage | Al-Sharqia Governorate | 140 (20 each of) | 95 (67.9%) | [39] |
| Beef kofta and liver | El-Qalyobia governorate | 60 (30 each of) | 20 (33.3%) | [40] |
| Beef kofta shawarma, burger, and luncheon | El-Qalyobia governorate | 120 (30 each of) | 61 (50.8%) | [30] |
| Chicken burger, sausage, and luncheon | El-Qalyobia governorate | 150 (50 each of) | 17 (11.3%) | [31] |

| Type of product | Region | Number of tested samples | Number (%) of S. aureus +ve | Reference |
|---|----------------------------|--------------------------|-----------------------------|-----------|
| beef burger and hot dog | Al-Dakahlia Governorate | 225 | 187 (83.1%) | [17] |
| Beef liver, beef burger, sausage, shawarma, and kofta | Al-Sharkia Governorate | 240 | 168 (70%) | [41] |
| Beef shawarma and burger | Al-Sharkia Governorate | 64 | 15 (23.4%) | [42] |
| Chicken burger, fillet, luncheon, nuggets, and panne | Al-Sharkia Governorate | 150 (30 each of) | 33 (22%) | [32] |
| Chicken crispy sandwiches | Al-Dakahlia Governorate | 90 | 38 (42.2%) | [5] |
| grilled and fried beef, grilled and fried chicken | Al-Menoufia Government | 120 (30 each of) | 31 (25.8%) | [43] |

TABLE 2. Prevalence of S. aureus in different RTE meat products sold in different countries all over the world

| Type of product | Country | Number of tested samples | Number (%) of S. aureus +ve | Reference |
|---|--------------|------------------------------|-----------------------------|-----------|
| sausage sandwiches | Turkey | 270 | 191 (70.7%) | [48] |
| Shawarma sandwiches | Jordon | 100 (80 chicken, 20 beef) | 8 (8%) | [49] |
| Beef sandwiches | Brazil | 25 | 5 (20%) | [50] |
| chicken meals | Thailand | 45 | 12 (26.7%) | [51] |
| Sausage, dried meat, meat with sauce, fried meat, smoked meat | China | 4047 | 32 (0.79%) | [52] |
| Roasted and spiced RTE meat | Nigeria | 105 | 17 (16.2%) | [53] |
| Beef products | China | 645 | 79 (12.2%) | [45] |
| RTE chicken, beef head meat, beef intestine | South Africa | 115 | 25% | [54] |
| beef products | China | 1209 | 31 (2.6%) | [46] |
| Grilled beef | Ghana | 36 | 9 (25%) | [55] |
| Beef burger, ground meat, beef sausage | Libya | 44 | 8 (18%) | [56] |
| RTE meat-based meals | Algeria | 55 | 21 (38.2%) | [22] |
| RTE hot meat pot | Cameron | 70 | 7(10%) | [57] |
| RTE meat pie | Nigeria | 20 | 20 (100%) | [47] |
| Chapli kebab, Shami steak, Liver kebab, beef burger, sausage | Afghanistan | 100 (20 each of) | 13 (13%) | [58] |

TABLE 3. Prevalence of MRSA in different RTE meat products in Egypt $\,$

| Type of product | Region | Number of tested samples | Percent (%) of MRSA among S. aureus isolates | Reference |
|--|---------------------------|--------------------------|--|-----------|
| Beef products (minced meat, luncheon, burger, sausage, and beef steak) | El-Gharbia Province | 100 (20 each of) | 35% | [67] |
| Beef luncheon and chicken nuggets | Al-Sharqia Governorate | 80 | 8.8% | [68] |
| Beef smoked sausage | Al-Sharqia Governorate | 50 | 8% | [39] |
| Beef burger and hot dog | Dakahlia Governorate | 225 | 22.6% | [17] |

| Type of product | Region | Number of tested samples | Percent (%) of MRSA among S. aureus isolates | Reference |
|--|-------------------------|--------------------------|--|-----------|
| Chicken luncheon, beef burger, and | Al-Qalubiya | 150 | 2.6% | [31] |
| sausage | Governorate | (50 each of) | 2.070 | [31] |
| Beef sausage, burger, luncheon and | Alexandria | 100 | 12% | [69] |
| pasterami | Governorate | (25 each of) | 1270 | [09] |
| Doof hungan and hatdage conduciones | Dakahlia | 100 | 44.3% | [70] |
| Beef burger and hotdogs sandwiches | Governorate | (50 each of) | 44.5% | |
| Beef burger, sausage, and chicken | Alexandria | 60 | 15% | [71] |
| pane | Governorate | (20 each of) | 13% | [71] |
| Beef shawarma and burger | Sharkia Governorate | 64 | 23.4% | [42] |
| Chicken burger, fillet, luncheon, nuggets, and panne | Sharkia Governorate | 150 (30 each of) | 33 % | [32] |
| Chicken sandwiches | Dakahlia Governorate | 90 | 23.7% | [29] |

 $TABLE\ 4.\ Prevalence\ of\ MRSA\ in\ different\ RTE\ meat\ products\ sold\ in\ different\ countries\ all\ over\ the\ world$

| Type of product | Region | Number of tested samples | Prevalence (%) of MRSA among S. aureus isolates | Reference |
|---|-------------|--------------------------|---|-----------|
| Cooked chicken products | China | 31 | 9.7% | [72] |
| RTE chicken and beef meals | Turkey | 150 | 38.6% | [73] |
| Cooked chicken meat | China | 153 | 10.1% | [52] |
| Meat and chicken barbecue | Iran | 113 | 10.6% | [74] |
| Roast chicken, salt-baked chicken, and stewed meat sausage | China | 645 | 7.4% | [45] |
| Street RTE meat products | Bangladesh | 112 | 6.3% | [75] |
| RTE meat products | China | 1209 | 3.2% | [46] |
| RTE hot meat pot | Cameron | 70 | 2(28.6%) | [57] |
| Chapli kebab, Shami steak, Liver kebab, beef burger, sausage | Afghanistan | 100 (20 each of) | 31.25% | [58] |

TABLE 5. Prevalence of VRSA in different RTE meat products sold in Egypt

| Type of product | Region | Number of tested samples | Prevalence(%) of VRSA among S. aureus isolates | Reference |
|--|-------------------------------|--------------------------|--|-----------|
| Chicken carcasses | Dakahlia Governorate | 200 | 17(5.9%) | [5] |
| beef burger and hot dog | Dakahlia Governorate | 225 | 4(2.1%) | [17] |
| Beef shawarma and burger | Sharkia Governorate | 64 | 4(26.7%) | [42] |
| Beef and chicken products | Kafr El-Sheikh governorate | 49 | 5 (10.2%) | [79] |
| Chicken crispy sandwiches | Dakahlia Governorate | 90 | 3(3.9%) | [29] |
| grilled and fried beef , grilled fried chicken | Menoufia Government | 120 (30 each of) | 1(3.2%) | [43] |

References

- Makinde, O. M., Ayeni, K. I., Sulyok, M., Krska, R., Adeleke, R. A. and Ezekiel, C. N., Microbiological safety of ready-to-eat foods in low-and middle-income countries: A comprehensive 10-year (2009 to 2018)- A Review, Comprehensive Reviews in Food Science and Food Safety, 19(2), 703-732 (2020).
- Wang, S. K., Fu, L. M., Chen, G. W., Xiao, H. M., Pan, D., Shi, R. F. and Sun, G. J., Multisite survey of bacterial contamination in ready-to-eat meat products throughout the cooking and selling processes in urban supermarket, Nanjing, China, Food Sci. & Nut., 8(5), 2427-2435 (2020).
- Aydın, A., Sudağıdan, M. and Muratoğlu, K., Prevalence of staphylococcal enterotoxins, toxin genes and genetic-relatedness of foodborne Staphylococcus aureus strains isolated in the Marmara Region of Turkey, Int. J. of Food Microbiol., 148(2), 99-106 (2011)
- Hennekinne, J. A., De Buyser, M. L. and Dragacci, S., Staphylococcus aureus and its food poisoning toxins: characterization and outbreak investigation, *FEMS Microbiol. Reviews*, 36(4), 815-836 (2012).
- Sallam, K. I., Abd-Elghany, S. M., Elhadidy, M., and Tamura, T., Molecular Characterization and Antimicrobial Resistance Profile of Methicillin-Resistant Staphylococcus aureus in Retail Chicken, *J.* of Food Prot., 78(10), 1879-1884 (2015).
- EFSA and ECDC (European Food Safety Authority and European Centre for Disease Prevention and Control), The European Union One Health 2023 Zoonoses report. EFSA J., 22(12), e9106. (2024).
- Lu, D., Liu, J., Liu, H., Guo, Y., Dai, Y., Liang, J., ...
 and Li, N., Epidemiological features of foodborne
 disease outbreaks in catering service facilities—China,
 2010–2020, China CDC Weekly, 5(22), 479 (2023).
- 8. Saad, T., Naguib, D., Gwida, M., Elgohary, A. H., and Mohamed, A., Current Perspectives on Methicillin-Resistant Staphylococcus aureus in Small Ruminants, *Egypt. J. of Vet. Sci.*, 1-12 (2024).
- Algammal, A. M., Hetta, H. F., Elkelish, A., Alkhalifah, D. H. H., Hozzein, W. N., Batiha, G. E. S., ... and Mabrok, M. A., Methicillin-Resistant Staphylococcus aureus (MRSA): one health perspective approach to the bacterium epidemiology, virulence factors, antibiotic-resistance, and zoonotic impact, *Infect. and drug resist.*, 3255-3265 (2020).
- Persoons, D., Van Hoorebeke, S., Hermans, K., Butaye, P., De Kruif, A., Haesebrouck, F., and Dewulf, J., Methicillin-resistant Staphylococcus aureus in poultry. *Emerg. infect. Dis.*, **15**(3), 452 (2009).
- Argudín, M. Á., Mendoza, M. C., and Rodicio, M. R., Food poisoning and Staphylococcus aureus enterotoxins, *Toxins*, 2(7), 1751-1773 (2010).
- 12. Umeda, K., Nakamura, H., Yamamoto, K., Nishina, N., Yasufuku, K., Hirai, Y., ... and Ogasawara, J., Molecular and epidemiological characterization of staphylococcal foodborne outbreak of Staphylococcus aureus harboring seg, sei, sem, sen, seo, and selu genes

- without production of classical enterotoxins, *Int. J. of food microbiol.*, **256**, 30-35 (2017).
- Thwala, T., Madoroba, E., Basson, A., and Butaye, P., Prevalence and characteristics of Staphylococcus aureus associated with meat and meat products in African countries-A Review, *Antibiotics*, 10(9), 1108 (2021).
- 14. Ray, S., Das, J., Pande, R., and Nithya, A., Foodborne Pathogens cum Contamination, Hygiene Practices, and Other Associated Issues in Ready-to-Eat Food, *In Rec. Adv. in Ready-to-Eat Food Tech.*, 195-222 (2024).
- Marriott, N. G., Schilling, M. W., Gravani, R. B., Marriott, N. G., Schilling, M. W., and Gravani, R. B., Food contamination sources, *Prin. of food sanit.*, 83-91 (2018)
- 16. Missiakas, D., and Schneewind, O., Staphylococcus aureus vaccines: Deviating from the carol, *J. of Exp. Med.*, **213**(9), 1645-1653 (2016).
- Mahros, M. A., Abd-Elghany, S. M., and Sallam, K. I.,. Multidrug-, methicillin-, and vancomycin-resistant Staphylococcus aureus isolated from ready-to-eat meat sandwiches: An ongoing food and public health concern, *Int. J. of Food Microbiol.*, 346, 109165 (2021).
- 18. Lin, Y. C. and Peterson, M. L., New insights into the prevention of staphylococcal infections and toxic shock syndrome, *Expert review of Clin. Pharm.*, **3**(6), 753-767 (2010).
- 19. Fetsch, A. and Johler, S., Staphylococcus aureus as a foodborne pathogen, *Curr. Clin. Microbiol. Reports*, **5**, 88-96. (2018).
- 20. Malachowa, N. and DeLeo, F. R., Mobile genetic elements of Staphylococcus aureus, *Cell. and Mol. Life Sci.*, **67**, 3057-3071. (2010).
- Tarekgne, E. K., Skjerdal, T., Skeie, S., Rudi, K., Porcellato, D., Felix, B. and Narvhus, J. A., Enterotoxin gene profile and molecular characterization of Staphylococcus aureus isolates from bovine bulk milk and milk products of Tigray region, northern Ethiopia, *J. of Food Prot.*, 79(8), 1387-1395. (2016).
- 22. Mekhloufi, O. A., Chieffi, D., Hammoudi, A., Bensefia, S. A., Fanelli, F. and Fusco, V., Prevalence, enterotoxigenic potential and antimicrobial resistance of Staphylococcus aureus and methicillin-resistant Staphylococcus aureus (MRSA) isolated from Algerian ready to eat foods, *Toxins*, 13(12), 835 (2021).
- Pérez-Boto, D., D'Arrigo, M., García-Lafuente, A., Bravo, D., Pérez-Baltar, A., Gaya, P. and Arqués, J. L., Staphylococcus aureus in the processing environment of cured meat products, *Foods*, 12, 2161 (2023).
- Pinchuk, I. V., Beswick, E. J. and Reyes, V. E., Staphylococcal enterotoxins, *Toxins*, 2(8), 2177-2197 (2010).
- Al-Bahry, S. N., Mahmoud, I. Y., Al-Musharafi, S. K. and Sivakumar, N., Staphylococcus aureus contamination during food preparation, processing and handling, *Int. J. of Chem. Eng. and Appl.*, 5(5), 388 (2014).

- 26. Min, K. J., Jung, Y. J., Kwon, K. Y., Kim, J. H., Hwang, I. G. and Yoon, K. S., Effect of Temperature on the Production of Staphylococcal Enterotoxin and Thermal Inactivation Kinetics of Staphylococcus aureus in Selected Ready-to-Eat (RTE) Foods in Korea, J. of Food Safety, 33(1), 17-24 (2013).
- Tsutsuura, S., Shimamura, Y. and Murata, M., Temperature dependence of the production of staphylococcal enterotoxin A by Staphylococcus aureus, *Biosci. Biotech. and Biochem.*, 77(1), 30-37 (2013).
- 28. Zeaki, N., Cao, R., Skandamis, P. N., Rådström, P. and Schelin, J., Assessment of high and low enterotoxin A producing Staphylococcus aureus strains on pork sausage, *Int. J. of Food Microbiol.*, **182**, 44-50 (2014).
- 29. Schelin, J., Wallin-Carlquist, N., Thorup Cohn, M., Lindqvist, R. and Barker, G. C., The formation of Staphylococcus aureus enterotoxin in food environments and advances in risk assessment, *Virulence*, **2**(6), 580-592 (2011).
- 30. Sihto, H. M., Stephan, R., Engl, C., Chen, J. and Johler, S., Effect of food-related stress conditions and loss of agr and sigB on seb promoter activity in S. aureus, *Food Microbiol.*, **65**, 205-212 (2017).
- 31. Divyakolu, S., Chikkala, R., Ratnakar, K. S. and Sritharan, V., Hemolysins of Staphylococcus aureus—an update on their biology, role in pathogenesis and as targets for anti-virulence therapy, *Adv. in Infect. Dis.*, **9**(2), 80-104 (2019).
- 32. Gottlieb, M., Long, B. and Koyfman, A., The evaluation and management of toxic shock syndrome in the emergency department: a review of the literature, *The J. of Emerg. Med.*, **54**(6), 807-814 (2018).
- 33. Léguillier, V., Pinamonti, D., Chang, C. M., Mukherjee, R., Kumar, H., Cossetini, A. and Vidic, J., A review and meta-analysis of Staphylococcus aureus prevalence in foods, *The Microbe*, 100131 (2024).
- 34. Elshebrawy, H. A., Kasem, N. G. and Sallam, K. I., Methicillin-and vancomycin-resistant Staphylococcus aureus in chicken carcasses, ready-to-eat chicken meat sandwiches, and buffalo milk, *Int. J. of Food Microbiol.*, 427, 110968 (2025).
- Craft, K. M., Nguyen, J. M., Berg, L. J. and Townsend,
 D., Methicillin-resistant Staphylococcus aureus (MRSA): antibiotic-resistance and the biofilm phenotype, *Med. Chem. Comm.*, 10, 1231–1241(2019).
- Guo, Y., Song, G., Sun, M., Wang, J. and Wang, Y., Prevalence and therapies of antibiotic-resistance in Staphylococcus aureus, Front. in cellular and inf. *Microbiol.*, 10, 107 (2020).
- Silva-de-Jesus, A. C., Ferrari, R. G., Panzenhagen, P. and Conte-Junior, C. A., Staphylococcus aureus biofilm: The role in disseminating antimicrobial resistance over the meat chain, *Microbiol.*, 168(10), 001245 (2022).
- 38. Katayama, Y., Ito, T. and Hiramatsu, K., A new class of genetic element, staphylococcus cassette chromosome mec, encodes methicillin resistance in

- Staphylococcus aureus. Antimicrob. Agents And Chemo., 44(6), 1549-1555 (2000).
- Shahid, A. H., Nazir, K. N. H., El Zowalaty, M. E., Kabir, A., Sarker, S. A., Siddique, M. P. and Ashour, H. M., Molecular detection of vancomycin and methicillin resistance in Staphylococcus aureus isolated from food processing environments, *One Health*, 13, 100276 (2021).
- 40. Ou, A., Wang, K., Mao, Y., Yuan, L., Ye, Y., Chen, L. and Huang, T., First report on the rapid detection and identification of methicillin-resistant Staphylococcus aureus (MRSA) in viable but non-culturable (VBNC) under food storage conditions, *Front. in Microbiol.*, 11, 615875. (2021).
- 41. Saif, M., Saad, S., Hassanin, F. S., Shaltout, F. and Zaghloul, M., Prevalence of methicillin-resistant Staphylococcus aureus in some ready-to-eat meat products, *Benha Vet. Med. J.*, **37**(1), 12-15 (2019).
- 42. Zakaria, I. M., Elsayed, S. N. and Sehim, A. E., Prevalence of multidrug-resistant Staphylococcus aureus in some processed chicken meat products, Egypt. Acad. J. of Biol. Sci., G. Microbiol., 13(2), 49-58 (2021).
- 43. Morshdy, A. E. M. A., Mahmoud, A. F. A., Khalifa, S. M., El-Dien, W. M. S., Darwish, W. S. and El Bayomi, R., Prevalence of Staphylococcus aureus and Salmonella species in chicken meat products retailed in Egypt, *Slov. Vet. Res.*, 60(25), 425-32 (2023).
- 44. HFA, E. D., and WEAM, M., Detection of Spoilage and Food Poisoning Bacteria in Some Ready to Eat Meat Products in Dakahlia Governorate, *Assiut Vet. Med. J.*, **59**(138), 55-59 (2013).
- 45. Awadallah, M. A., Ahmed, H. A. and Merwad, A. M., Prevalence of non-O157 shiga toxin-producing Escherichia coli and Enterotoxigenic staphylococci in ready-to-eat meat products, handlers and consumers in Cairo, Egypt, *Glob. Vet.*, 12(5), 692-699. (2014).
- 46. Ibrahim, M. A. and Nassif, M. Z., Incidence of Staphylococcus aureus in meat products with special reference to enterotoxins, *Benha Vet. Med. J.*, **30**(2), 23-27 (2016).
- 47. Hassanien, A. A. and Abdel-Aziz, N. M., Staphylococcus aureus and their enterotoxin genes in fast food and food handlers from different food premises in Sohag city, Egypt, *Int. J. Agric. Sci. Vet. Med.*, 5(1), 107-113 (2017).
- 48. Shaltout, F. A. E., Farouk, M., Ibrahim, H. A. and Afifi, M. E., Incidence of Coliform and Staphylococcus aureus in ready to eat fast foods, *Benha Vet. Med. J.*, **32**(1), 13-17 (2017).
- Saad, S. M., F Khater, D. and Zaki, S. M., Food poisoning bacteria in ready to eat meat and chicken meat products, *Benha Vet. Med. J.*, 35(1), 301-310 (2018).
- 50. Morshdy, A. E. M., Darwish, W. S., El-Dien, W. M. S. and Khalif, S. M., Prevalence of multidrug-resistant Staphylococcus aureus and Salmonella enteritidis in meat products retailed in Zagazig city, Egypt, *Slov. Vet. Res.*, 55, 295–301 (2018).

- 51. Gaafar, R., Hassanin, F. S., Shaltout, F. and Zaghloul, M., Molecular detection of enterotoxigenic Staphylococcus aureus in some ready to eat meat based sandwiches, *Benha Vet. Med. J.*, 37(1), 22-26 (2019).
- 52. Hussein, M. A., Samar, H. M., Tharwat, A. E., Hafez, A. E. and Gomaa, R. A., Effect of ingredients on the bacteriological profile of ready to eat meat sandwiches, *J. Anim. Health Prod.* 9(s1), 69-75 (2021).
- 53. Saber, T., Samir, M., El-Mekkawy, R. M., Ariny, E., El-Sayed, S. R., Enan, G. and Tartor, Y. H., Methicillin-and vancomycin-resistant Staphylococcus aureus from humans and ready-to-eat meat: characterization of antimicrobial resistance and biofilm formation ability, Front. in Microbiol., 12, 735494 (2022).
- 54. Ghanem, M. A., Idris, A. B. M., Abu El Roos, N. and Darwish, W. S., Prevalence of Multidrug-Resistant Staphylococcus aureus in Meals Served at Hospitals, *Egypt. J. of Vet. Sci.*, **56**(7), 1479-1488 (2025).
- 55. Oyeyi, T. I. and Lum-Nwi, M. E. F., Bacteriological quality of some street vended foods in Bayero University campuses, Kano, Nigeria, *Biol. and Enviro. Sci. J. for the Trop.*, **5**(4), 239-243 (2008).
- Wu, S., Huang, J., Wu, Q., Zhang, J., Zhang, F., Yang, X. and Xue, L., Staphylococcus aureus isolated from retail meat and meat products in China: incidence, antibiotic resistance and genetic diversity, *Front. in Microbiol.*, 9, 2767 (2018).
- 57. Lin, Q., Sun, H., Yao, K., Cai, J., Ren, Y. and Chi, Y., The prevalence, antibiotic resistance and biofilm formation of Staphylococcus aureus in bulk ready-to-eat foods, *Biomolecules*, **9**(10), 524 (2019).
- 58. Omoruyi, I. M. and Ibegbulam, O. S., Ready-to-eat food sold in healthcare facilities could contribute to the increased disease burden of multidrug resistant Staphylococcus aureus, *Bio-Res.*, 21(2), 1923-1934 (2023).
- 59. Büyükyörük, S., Beyaz, D., Göksoy, E. Ö., Kök, F. and Koçak, P., Microbiological evaluation of ready-to-eat sandwiches served near hospitals and schools, *Ankara Üni. Vet. Fakültesi Dergisi*, **61**(3), 193-198 (2014).
- Nimri, L., Al-Dahab, F. A., and Batchoun, R., Foodborne bacterial pathogens recovered from contaminated shawarma meat in northern Jordan, *The J. of Infect. in Devel. Count.*, 8(11), 1407-1414. (2014).
- 61. Contreras, C. P. Á., da Silva, L. N. N., Ferreira, D. C. G., dos Santos Ferreira, J. and de Castro Almeida, R. C., Prevalence of methicillin-resistant Staphylococcus aureus in raw hamburgers and ready-to-eat sandwiches commercialized in supermarkets and fast food outlets in Brazil, Food and Nut. Sci., 6(14), 1324 (2015).
- Ananchaipattana, C., Bari, M. L. and Inatsu, Y., Bacterial contamination into ready-to-eat foods sold in middle Thailand, *Biocontrol. sci.*, 21(4), 225-230 (2016).
- 63. Yang, S., Pei, X., Wang, G., Yan, L., Hu, J., Li, Y. and Yang, D., Prevalence of food-borne pathogens in ready-to-eat meat products in seven different Chinese regions, *Food Control*, **65**, 92-98 (2016).

- 64. Okoli, C. E., Njoga, E. O., Enem, S. I., Godwin, E. E., Nwanta, J. A. and Chah, K. F., Prevalence, toxigenic potential and antimicrobial susceptibility profile of Staphylococcus isolated from ready-to-eat meats, *Vet. World*, 11(9), 1214 (2018).
- 65. Tshipamba, M. E., Lubanza, N., Adetunji, M. C. and Mwanza, M., Molecular characterization and antibiotic resistance of foodborne pathogens in street-vended ready-to-eat meat sold in South Africa. *J. of Food Prot.*, 81(12), 1963-1972 (2018).
- 66. Adzitey, F., Ekli, R. and Abu, A., Prevalence and antibiotic susceptibility of *Staphylococcus aureus* isolated from raw and grilled beef in Nyankpala community in the Northern Region of Ghana, *Cogent Food & Agri.*, **5**(1), 1671115 (2019).
- 67. Naas, H. T., Edarhoby, R. A., Garbaj, A. M., Azwai, S. M., Abolghait, S. K., Gammoudi, F. T. and Eldaghayes, I. M., Occurrence, characterization, and antibiogram of Staphylococcus aureus in meat, meat products, and some seafood from Libyan retail markets, *Vet. World*, 12(6), 925 (2019).
- 68. Esemu, S. N., Njoh, S. T., Ndip, L. M., Keneh, N. K., Kfusi, J. A. and Njukeng, A. P., Ready-to-Eat Foods: A Potential Vehicle for the Spread of Coagulase-Positive Staphylococci and Antimicrobial-Resistant Staphylococcus aureus in Buea Municipality, South West Cameroon, Canadian J. of Inf. Dis. and Med. Microbiol., 2023(1), 9735319 (2023).
- 69. Ahmadi, S. A., Aziz, A., Afshar, M. F. and Abi, A. J., Exploring Staphylococcus aureus prevalence and antimicrobial resistance in ready-to-eat meat products in Kabul city, *Egypt. J. of Vet. Sci.*, (In press) (2025).
- Lehto, M., Kuisma, R., Määttä, J., Kymäläinen, H. R. and Mäki, M., Hygienic level and surface contamination in fresh-cut vegetable production plants, Food Control, 22(3-4), 469-475 (2011).
- González-Machado, C., Alonso-Calleja, C. and Capita, R., Methicillin-Resistant Staphylococcus aureus (MRSA) in Different Food Groups and Drinking Water. Foods, 13(17), 2686 (2024).
- Krumova-Valcheva, G. L., The role of food in the spread of the Methicillin–resistant Staphylococcus aureus (MRSA), Food Sci. and Appl. Biotech., 6(1), 87-94 (2023).
- 73. Markwart, R., Willrich, N., Eckmanns, T., Werner, G. and Ayobami, O., Low proportion of linezolid and daptomycin resistance among bloodborne vancomycin-resistant Enterococcus faecium and methicillin-resistant Staphylococcus aureus infections in Europe, Front. in Microbiol., 12, 664199 (2021).
- 74. De Neeling, A. J., Van den Broek, M. J. M., Spalburg, E. C., van Santen-Verheuvel, M. G., Dam-Deisz, W. D. C., Boshuizen, H. C. and Huijsdens, X. W., High prevalence of methicillin resistant Staphylococcus aureus in pigs, *Vet. Microbiol.*, 122(3-4), 366-372 (2007).
- Mlynarczyk-Bonikowska, B., Kowalewski, C., Krolak-Ulinska, A. and Marusza, W., Molecular mechanisms of drug resistance in Staphylococcus aureus, *Int. J. of Mol. Sci.*, 23(15), 8088 (2022).

- Butaye, P., Argudín, M. A. and Smith, T. C., Livestock-associated MRSA and its current evolution, *Curr. Clin. Microbiol. Reports*, 3, 19-31 (2016).
- 77. Eldaly, E. A., El-Shopary, N. F. and Gamal, R. H. E., Prevalence of methicillin-resistant Staph. aureus (MRSA) in retail meat products in El-Gharbia Province, Egypt, Global J. of Agri. and Food Safety Sci., 1(2), 270-282 (2014).
- 78. El Bayomi, R. M., Ahmed, H. A., Awadallah, M. A., Mohsen, R. A., Abd El-Ghafar, A. E. and Abdelrahman, M. A., Occurrence, virulence factors, antimicrobial resistance, and genotyping of Staphylococcus aureus strains isolated from chicken products and humans, *Vector-Borne and Zoo. Dis.*, 16(3), 157-164 (2016).
- Khamis, M. A., Mousa, M. M. and Helmy, N. M., Methicillin-Resistant Staphylococcus aureus (MRSA) in some meat products, *Alexandria J. of Vet. Sci.*, 70(1) (2021).
- 80. Mohamed, E., Ramadan, H., Abd-Elghany, S. and Mahros, M., Isolation and characterization of enterotoxigenic coagulase-positive and methicillinresistant Staphylococcus aureus contaminating beef burger and hot dog sandwiches retailed in Mansoura city, *Mansoura Vet. Med. J.*, 22(1), 20-24 (2021).
- 81. Nossair, M. A., Elaadli, H., Mansour, A. M., Shaaban, S. I., Khatab, S. A. and Severin, M., Prevalence and antimicrobial resistance profile of methicillin resistant Staphylococcus aureus strains isolated from food products and food handlers in Egypt, *Alexandria J. of Vet. Sci.*, 75(2) (2022).
- Tang, J., Zhang, R., Chen, J., Zhao, Y., Tang, C., Yue, H. and Shi, H., Incidence and characterization of Staphylococcus aureus strains isolated from food markets. *Annals of Microbiol.*, 65, 279-286 (2015).
- 83. Özdemir, H. and Keyvan, E., Isolation and characterisation of Staphylococcus aureus strains isolated from beef, sheep and chicken meat, *Ankara Üni. Vet. Fakültesi Dergisi*, **63**(4), 333-338 (2016).
- 84. Safarpoor Dehkordi, F., Gandomi, H., Basti, A. A., Misaghi, A. and Rahimi, E., Phenotypic and genotypic characterization of antibiotic resistance of methicillinresistant Staphylococcus aureus isolated from hospital food, Antimicrob. Resis. & Infect. Control, 6, 1-11(2017).
- 85. Islam, M. A., Parveen, S., Rahman, M., Huq, M., Nabi, A., Khan, Z. U. M. and Wagenaar, J. A., Occurrence and characterization of methicillin resistant Staphylococcus aureus in processed raw foods and ready-to-eat foods in an urban setting of a developing country, Front. in Microbiol., 10, 503 (2019).
- 86. Wu, Q., Sabokroo, N., Wang, Y., Hashemian, M., Karamollahi, S. and Kouhsari, E., Systematic review and meta-analysis of the epidemiology of vancomycinresistance Staphylococcus aureus isolates, *Antimicrob. Resis. & Infec. Control*, 10, 1-13 (2021).
- 87. Chang, S., Sievert, D. M., Hageman, J. C., Boulton, M. L., Tenover, F. C., Downes, F. P. and Fridkin, S. K., Infection with vancomycin-resistant Staphylococcus

- aureus containing the vanA resistance gene, New England J. of Med., **348**(14), 1342-1347 (2003).
- 88. Cong, Y., Yang, S. and Rao, X., Vancomycin resistant Staphylococcus aureus infections: A review of case updating and clinical features, *J. of adv. Res.*, **21**, 169-176 (2020).
- Naeim, D. E., Eldesoukey, I. E., Moawad, A. A. and Ahmed, A. M., Molecular detection of methicillinresistant Staphylococcus aureus isolated from different foodstuffs in Egypt, In Vet. Res. Forum, 14(5), 243 (2023).
- 90. Yahya, A. M., Abdalbagi, A. H., Mohammed, T. G. B., Omer, A. S., Saif, A. A. E., Hamza M. M. and Altayb, H. N., Molecular detection of Staphylococcal enterotoxins and mecA genes products in selected food samples collected from different areas in Khartoum State. *Int. J. of Microbiol.*, 2021(1), 5520573 (2021).
- 91. Liu, Y., Zheng, X., Xu, L., Tong, P., Zhu, M., Peng, B. and Yao, G., Prevalence, antimicrobial resistance, and molecular characterization of Staphylococcus aureus isolated from animals, meats, and market environments in Xinjiang, China, *Foodborne Path. and Dis.*, **18**(10), 718-726 (2021).
- 92. Afshari, A., Taheri, S., Hashemi, M., Norouzy, A., Nematy, M. and Mohamadi, S., Methicillin-and vancomycin-resistant Staphylococcus aureus and vancomycin-resistant enterococci isolated from hospital foods: Prevalence and antimicrobial resistance patterns, *Curr. Microbiol.*, **79**(11), 326 (2022).
- 93. Ain, Q. T., Arif, M., Ema, F. A., Shathi, M. S. A., Islam, M. A. and Khatun, M. M., Multidrug-resistant Staphylococcus aureus isolated from chicken nuggets sold at superstores in Mymensingh, Bangladesh, *J. of Adv. Vet. and Ani. Res.*, **9**(4), 601 (2022).
- 94. Adeyemi, F. M., Oyedara, O. O., Yusuf-Omoloye, N. A., Ajigbewu, O. H., Ndaji, O. L., Adegbite-Badmus, M. K. and Oluokun, T. E., Guardians of resistance and virulence: detection of mec, femA, Van, pvl, hlg and spa genes in methicillin and vancomycin-resistant Staphylococcus aureus from clinical and food samples in Southwestern Nigeria, *BMC Microbiol.*, 24(1), 498 (2024).
- 95. Huo, M., Xu, X., Mi, K., Ma, W., Zhou, Q., Lin, X. and Huang, L., Co-selection mechanism for bacterial resistance to major chemical pollutants in the environment, *Sci.of The Total Envi.*, **912**, 169223 (2024).
- 96. Alonso, V. P., Queiroz, M. M., Gualberto, M. L. and Nascimento, M. S., Klebsiella pneumonia carbapenemase (KPC), methicillin-resistant Staphylococcus aureus (MRSA), and vancomycinresistant Enterococcus spp.(VRE) in the food production chain and biofilm formation on abiotic surfaces, *Curr.Opin. in Food Sci.*, 26, 79-86 (2019).
- 97. Miao, J., Liang, Y., Chen, L., Wang, W., Wang, J., Li, B. and Xu, Z., Formation and development of Staphylococcus biofilm: with focus on food safety. *J. of Food Safety*, 37(4), e12358 (2017).

- Vergara, A., Normanno, G., Di Ciccio, P., Pedonese, F., Nuvoloni, R., Parisi, A. and Ianieri, A., Biofilm formation and its relationship with the molecular characteristics of food-related methicillin-resistant Staphylococcus aureus (MRSA). *J. of Food Sci.*, 82(10), 2364-2370 (2017).
- Li, J., Li, C., Shi, C., Aliakbarlu, J., Cui, H. and Lin, L., Antibacterial mechanisms of clove essential oil against Staphylococcus aureus and its application in pork, *Int. Journal of Food Microbiol.*, 380, 109864 (2022).
- 100. Hu, W., Li, C., Dai, J., Cui, H. and Lin, L., Antibacterial activity and mechanism of Litsea cubeba essential oil against methicillin-resistant Staphylococcus aureus (MRSA). *Indust. Crops and Prod.*, 130, 34-41 (2019)
- 101. Ghabraie, M., Vu, K. D., Tata, L., Salmieri, S., and Lacroix, M., Antimicrobial effect of essential oils in

- combinations against five bacteria and their effect on sensorial quality of ground meat. *LWT-Food Sci. and Tech.*, **66**, 332-339 (2016).
- 102. Fetsch, A. and Johler, S., Staphylococcus aureus as a foodborne pathogen. *Curr. Clin. Microbiol. Reports*, **5**(2), 88-96 (2018).
- 103. Hwang, H. J., Lee, E. J., Yu, H. H. and Lee, J. H., Studies on the radiation sensitivity of food microorganism by high dose irradiation . KAERI/CM-1304/2009 (2010).
- 104. Sánchez-Ortega, I., García-Almendárez, B. E., Santos-López, E. M., Amaro-Reyes, A., Barboza-Corona, J. E. and Regalado, C., Antimicrobial edible films and coatings for meat and meat products preservation, The Sci. World J., 2014(1), 248935 (2014).

الميكروب العنقودي الذهبي المقاوم للميثيسيلين و الميكروب العنقودي الذهبي المقاوم للفائكوميسين في منتجات اللحوم

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الملخص

تعد المكورات العنقودية الذهبية تهديدا خطيرا متزايدا لصحة الحيوان و الإنسان حيث تصنف هذة الجراثيم كمسبب مرضي أولوية بالغة و ذلك طبقا لتصنيف منظمة الصحة العالمية نظرا لإنتشارها الواسع إلي جانب أنها تملك عوامل ضراوة متعددة و خطيرة ، كما ان كونها من الجراثيم المتعايشة علي الجلد و الأغشية المخاطية للإنسان و الحيوان كذلك في الجهاز التنفسي له مما يسهل دخولها إلي سلسلة الغذاء البشري و الذي بدوره يؤدي إلي العديد من حالات التسمم الغذائي في مصر و العديد من دول العالم. تسبب العدوي بالميكروب العنقودي الذهبي الكثير من الاعراض الاكلينيكية للحيوان و التي يصاحبها خسائر رمالية كبيرة، كما أنها تسبب أمراض عديدة و خطيرة للجنس البشري و تشمل: التسمم الغذائي، الإلتهاب الرئوي، التهاب العظام، التهاب الغشاء المبطن القلب، و التسمم الدموي. يعرف الميكروب العنقودي الذهبي بأنه واحد من الرئوي، التهاب العظام، التهاب الغشاء المبطن القلب، و التسمم الامريكية القولونية و بكتيريا السالمونيلا حيث انه يفرز مما يزيد عن ثماني و عشرين نوعا من السموم المعوية التي تستثير أعداد هائلة من الخلايا و الأجسام المناعية في جسم العائل المصاب و التي قد تنتهي بالوفاة. هذا البحث المرجعي يعرض بإيجاز طرق تلوث منتجات اللحوم و الوجبات الجاهزة المصابة، مقاومتها للمضادات الحيوية، مدي انتشارها في منتجات اللحوم الجاهزة للأكل في مصر و بعض البلاد حول العالم. العالم.

الكلمات الدالة: الميكروب العنقودي الذهبي، الأمراض المنقولة بالغذاء، مقاومة للميثيسيلين، مقاومة للفانكوميسين.