

Egyptian Journal of Veterinary Sciences

https://ejvs.journals.ekb.eg/



Influence of Supplemented Levels of *Acacia nilotica* Leaf Meal on Growth Performance, Faecal Egg Count and Haematological Parameters of West African Dwarf Bucks



Ayobami J. Owolabi¹, Azeez O. Yusuf¹*, Adekayode O. Sonibare², Olusiji S. Sowande¹ and Taiwo A. Olufemi¹

Abstract

THIS study was carried out to determine the performance of West African dwarf goats fed Acacia ■ nilotica leaf meal. Thirty (30) artificially infected West African Dwarf goats were randomly divided into five (5) groups of six (6) animals per group. Each group was randomly allotted to five (5) experimental diets containing varying levels (0. 5, 10, 15 and 20 %) of A. nilotica leaf meal (ANLM). Faecal samples were collected and analysed weekly from each animal to determine the faecal egg count. Weight changes of the animals were taken once a week throughout the experimental period and blood samples were collected on the 1st and 56th day of the experiment. Data collected were subjected to one-way analysis of variance. Results showed significant (p<0.05) impacts of the dietary treatments on most parameters considered. Goats fed 10% and 20% inclusion diets exhibited higher final weight gain and weight gains compared to other groups. Feed conversion ratios varied across the different dietary groups. Faecal egg counts decreased significantly (p<0.05) in goats on supplemented diets compared to the control group. Packed cell volume, haemoglobin concentration and red blood cell count were elevated (p<0.05) in the supplemented groups., Mean corpuscular volume, mean corpuscular haemoglobin, and mean corpuscular haemoglobin concentration were elevated in some of the treatment with no particular trend. White blood cell counts, and its differentials were also influenced (p<0.05) by the dietary treatments, it increased in animals offered 5% ANLM and least in those fed 10% ANLM. The study concluded that ANLM enhanced infected goats' performance, reduced faecal egg counts and maintained optimal blood levels demonstrating potential pharmaceutical and nutritional benefits for production.

Keywords: Acacia nilotica, faecal egg count, goats, growth performance, leaf meal.

Introduction

One of the significant hinderances to ruminant production other than nutrition is endoparasites. Endoparasites have grown to be a serious concern in large-scale sheep and goat production systems, as evidenced by sales data from several nations [1]. Animal health can be negatively impacted by gastrointestinal nematode infections [2], with

these infections causing clinical and subclinical disorders, which can lead to financial loss and generally lower animal productivity [3].

Helminthiasis in particular of the intestinal parasites is more severe in tropical and subtropical regions, because of the favourable environmental circumstances that lead to a greater abundance of this class of parasites [4]. The presence of these parasites

*Corresponding authors: Yusuf, Azeez Olanrewaju, E-mail: yusufao@funaab.edu.ng, Tel.: +2348036250504 (Received 24 March 2025, accepted 20 August 2025)

DOI: 10.21608/ejvs.2025.370905.2728

¹Department of Animal Production and Health, Federal University of Agriculture, Abeokuta, Nigeria

²Department of Veterinary and surgery, College of Veterinary Medicine, Federal University of Agriculture, Abeokuta, Nigeria

combined with the poor feed quality in these regions has resulted in poor productivity with increased cost of production.

The two-pronged approach to combat both nutritional and health challenges in ruminant production has resulted in a number of studies particularly the use of alternative feed resources that can boost the nutritional quality and improve health status of ruminant livestock [2]. The use of multipurpose tree species has an alternative feed source has been studied a lot. Tree species like *Moringa oleifera*, *Leucaena leucocephala*, *Newbouldia laevis*,... etc have been experimented with mostly for their nutritional properties.

However, the recent studies into sustainable agricultural production and shift from heavy reliance on synthetic chemotherapy has resulted in a second look at these plants for their medicinal attributes [5]. In addition, these plants have a deep root in ethno medicine with various attributes ranging from antibiotic, antiparasitic and antimicrobial documented orally through generations. These oral documentations together with recent research into the constituents of these plants has revealed that these plants do possess medicinal properties due to the level of phytochemicals present in them, all of which act in various modes to improve health and productivity of livestock [5-7].

On the other hand, over-ingestion of these phytochemicals can result in toxicity which could have adverse effect on the well-being of livestock. Hence, it is important to monitor the blood levels of animals that are fed with these plants with abundant phytochemicals.

This present study therefore aims at investigating the impact of *Acacia nilotica* leaf meal on the faecal egg count, performance and haematological profile of West African dwarf goats artificially infected with *Haemonchus contortus*.

Material and Methods

Experimental Animals

For the experiment, thirty (30) West African Dwarf (WAD) yearling goats were used. They were bought from local communities in Ogun State's Odeda Local Government. The goats averaged 6.5 kg at the onset of the experiment. Morigad® disinfectant was used to thoroughly clean and sanitize the experimental pens. The goats were kept in individual pen spaces with corrugated aluminium iron roofing and a slatted floor. Following purchase,

the animals received injections of ivermectin to eradicate any ectoparasites and an oral solution of albendazole to eradicate any endoparasites. The animals received tissue culture rinderpest vaccinations to protect them from *Peste des Petit Ruminants* (PPR). *Panicum maximum* was fed to the goats *ad libitum* by cut and carry methods.

Experimental design

The thirty (30) WAD goats were split into five (5) groups, each consisting of six (6) animals. Each group was assigned at random to one of the five (5) experimental diets using completely randomized design (CRD).

Inoculation and Treatment

Third stage (L3) *Haemonchus contortus* larvae, grown from the faeces of surgically afflicted sheep, were the source of the infection in the goats. The sheep who had already contracted the infection provided faeces, which were then processed and cultivated in a lab. For the purpose of ensuring infection, the experimental animals were given the cultured samples orally on day seven. Before the study started, the goats' infections were verified by taking faecal samples from them, and the quantity of nematode eggs in those samples was counted using the flotation method (McMaster). The animals utilized were limited to those whose counts of faecal eggs above 750 eggs per gram of faeces (epg).

Experimental Diet Preparation

Acacia nilotica was used in leaf meal form to compound the experimental diets. In order to create concentration diets with different inclusion levels of the test material as mentioned above, five (5) experimental diets were created utilizing 0, 5, 10, 15, and 20% inclusion levels of *A. nilotica* leaf meal (ANLM) together with other feed ingredients (Table 1).

Data Collection

Weight changes and feed intake

Feed intake: known quantity of feed at 3% of body weight was offered to individual animals on daily basis throughout the 56 days of the experiment, while the left over the following day was recovered and weighed to determine daily feed intake. The average daily feed intake was calculated as follows:

Average daily feed intake (kg DM) $= \frac{\text{total feed offered} - \text{total feed left over}}{\text{number of days}} (kg DM/day)$

Weight gain: each goat was weighed at the onset of the experiment before morning feeding, and at 7-day interval throughout the experimental period. Feed conversion ratio was also calculated using the formula:

 $FCR = \frac{\text{feed intake (kg)}}{\text{weight gain (kg)}}$

Parasite egg count

At the beginning of the experiment, a sample of faeces was taken straight from the rectum of the experimental animals in order to calculate the faecal egg count (FEC), which is a measure of the first infestation of parasites. Throughout the experimental phase, daily faecal samples were taken to assess the FEC as a measure of the subjects' gastrointestinal load.. The postgraduate laboratory of the Department of Animal Production and Health, Federal University of Agriculture, Abeokuta, was where the faeces samples taken to in order to calculate the FEC (eggs per gram of faeces).

A modified Wisconsin salt flotation technique, as described by [8] was applied to the faecal samples that were obtained. Three grams of pulverized faeces were combined with forty-two millilitres of floatation fluid, which is a saturated solution. Following filtering, a subsample was moved into each of the two compartments of the McMaster counting chamber and left for five minutes. All of the eggs were in focus against the higher slide as they ascended to the top. The number of eggs per gram in the original sample is represented by multiplying the number of eggs inside each ruled region by 50.

Blood sample collection

Using a hypodermic needle and syringe, a sample of approximately 5 millilitres of blood was taken directly from each animal's jugular vein on the first and 56th day of the experiment. In order to prevent coagulation, the blood samples were collected and placed into sample bottles containing EDTA (Ethyl Dimethyl Tetra Acetic Acid) as an anticoagulant. The bottles were then gently rolled to ensure that the blood and EDTA were properly mixed. Blood samples were analysed for packed cell volume (PCV) and haemoglobin (Hb) concentration as soon as they were collected following a standard procedure [9]. After dilution, the Neubauer the proper haemocytometer was used to measure the amounts of red blood cells (RBC), white blood cells (WBC), and differential WBC counts [10].

Statistical Analysis

All data generated were subjected to One-way Analysis of Variance as found in SPSS (2006), while significant means (p<0.05) were separated using the Tukey's test as contained in the same statistical package.

Results

Growth performance of artificially infected West African Dwarf goats fed diets containing varying levels of Acacia nilotica leaf meal.

The growth performance of artificially infected West African Dwarf goats fed diets containing varying levels of Acacia nilotica leaf meal is presented in Table 2. All the parameters considered were significantly influenced by the treatments. Goats offered 10% and 20% inclusion diets had significantly (p<0.05) higher mean values (8.16 and 9.06 kg) respectively, for final weight, while animals receiving the control diet recorded the lower mean value (7.00 kg). Weight gain, average weight gain and metabolic weight followed a similar trend having higher mean values (1.89 kg, 33.79 g/day, 13.91 $kgBW^{0.75}$ and 2.40 kg, 42.88 g/day, 16.33 kgBW^{0.75}) in animals fed 5% and 20% dietary inclusion of A. nilotica respectively, however, the lower mean values (0.75 kg, 13.43 g/day 6.92 kgBW^{0.75} and 0.95 kg, 17.03 g/day, 7.83 kgBW^{0.75}) were recorded in animals offered 0% and 15% inclusion of A. nilotica in their diets, respectively.

Dry matter intake was highest (265.40 g/day) in goats fed 10% inclusion diets, while the lowest value (186.51 g/day) was recorded in goats offered 15% inclusion diets. Animals fed on 15% inclusion diets had the higher feed conversion ratio value (24.60), while goats offered 5% and 20% recorded mean values (5.87 and 6.41) Kg feed intake/Kg gain, respectively.

Faecal egg count of artificially infected West African Dwarf goats fed diets containing varying levels Acacia nilotica leaf meal.

Figure 1 shows the faecal egg count of infected West African Dwarf goats fed diets containing varying levels *A. nilotica* leaf meal at the onset and culmination, and on weekly basis throughout the experiment, respectively. All goats fed the varying diets had steeper reductions in their faecal egg counts except for the control which had the least reduction.

Red blood cell composition of artificially infected West African Dwarf goats fed diets containing varying levels of A. nilotica leaf meal

Table 3 shows the red blood cell composition of artificially infected WAD goats fed diets containing varying levels of A. nilotica leaf meal. All parameters considered at the beginning of the experiment were not significantly differed, however, at the end of the experiment significant differences were observed in the parameters considered across the treatments. Packed cell volume had mean values (33.63 and 30.38 %) in goats fed 10 and 20 % inclusion diets respectively, while the 0% and 15% inclusion groups recorded the lower values (24.13 24.75 %), respectively. Haemoglobin concentration was lower in goats fed the control diet compared to the other supplemented groups. Red blood cell count was lower in goats fed 5 % ANLM inclusion diet, compared to other supplemented groups and the control. Mean corpuscular volume recorded the lower mean value (16.10 fl) in goats fed 10% inclusion of A. nilotica leaf meal diet compared to the control and other supplemented groups. The higher mean values (7.79 and 7.28 pg) were recorded in goats fed 5 and 10 % ANLM inclusion diet for mean corpuscular haemoglobin. Mean corpuscular haemoglobin concentration had higher mean values (37.02, 38.91 and 37.98 g/dL) in the 0, 5 and 15 % treatment groups, respectively, while the lower mean values (32.69 and 32.86 g/dL) were recorded in the 10 and 20 % groups, respectively.

White blood cell counts, and its differentials of artificially infected West African Dwarf goats fed varying levels of A. nilotica leaf meal.

Table 4 shows the white blood cell count and its differentials of West African dwarf goats artificially infected goats fed varying levels of *A. nilotica* leaf meal. At the inception of the experiment, only the eosinophil count was significantly differed across the treatment groups, having significantly (p<0.05) mean value (0.00 %) in the 15 % treatment group, compared to the other groups.

However, at the culmination of the experiment all the parameters considered were significantly (p<0.05) influenced by the treatments, except for eosinophil count. The white blood cell count had significantly (p<0.05) highest value $(11.00\times10^{9/L})$ in goats offered 5 % inclusion of *A. nilotica*, while the least value $(7.25\times10^{9/L})$ was recorded in the 10% treatment group. Neutrophil count recorded value (33.75%) in the control group compared to the other supplemented groups. Goats in the control and 10 % group had the highest lymphocyte count (65.88 and 64.88 %), while goats offered 15% dietary inclusion

of *A. nilotica* had the lowest value (59.88%). Basophil count was lower in goats fed 5 and 10 % ANLM diets compared to the control and supplemented groups. Monocyte count was higher (1.00 and 0.63 %) in goats offered 15 and 20 % *A. nilotica* leaf meal inclusions compared to the other supplemented groups and control.

Discussion

The potentials of shrubs and multipurpose trees are not limited to the medicinal attributes they possess, but rather all physiological processes for optimum performance. Several reports of various plants which have been used in feeding trials have been documented. Plants like Moringa oleifera, Newbouldia laevis [11], and aquatic plants like Eichhornia crassipes [12, 13] have not been left out in the search for alternative feed resources. Plants rich in secondary metabolites have been credited by researchers [14, 15] to have various nutritional benefits such as improving fatty acid profile and reducing enteric methane production and are gaining recognition as being suitable for potential increase in ruminant production. The abundance of plant secondary metabolites (PSM) in multipurpose tree species like Acacia nilotica used in this study, their nutritional profile containing significant level of nutrients important for substantial increase in production parameters, coupled with their all-yearround availability further makes these tree species economically important.

The growth response (final weight and weight gain) documented in this study corresponds with that of other authors [16] who fed different species of Acacia (Acacia seyal) to sheep and observed an increasing weight gain with increase in inclusion levels. In addition, Yusuf et al. (2016) reported a significant increase in weight gain with increase in Moringa oleifera diets fed to WAD sheep. A similar observation was documented in another experiment [17] where an increase weight gain in Red Sokoto goats fed A. nilotica pods was documented. The result observed in this study could be as a function of higher crude protein (12.80 %) levels, as well as the PSM which has antibiotic and antioxidative capabilities to promote growth and ensures better immune response for optimal production [18] with increasing inclusion of A. nilotica leaf meal in the diets. This could possibly be responsible for the higher growth performance exhibited by the animals in the 20% ANLM treatment group. The findings of this study indicate that, because ANLM includes a

significant amount of important nutrients, adding up to 20% of it to the diet of goats may increase livestock performance in terms of body weight changes and high yields of high-quality products. The dual action of tannin components in the fed diets serving both nutritional and medicinal purposes may be responsible for higher performance results recorded in this study. It was documented by different researchers [19, 20] that the competition for nutrients between gastrointestinal nematodes (GIN) and their hosts often leads to reduced growth performance. Since the results indicated reduced FEC, this depicts that feeding *A. nilotica* to animals could enhance inhibition of both worm eggs and larvae development.

Although, optimal production assessment is hinged on the inverse relationship between feed intake and weight gain, as it is more beneficial in production terms to eat less and gain more. Thus, it can be opined that PSM levels present in the fed diets was able to combat the *H. contortus* present in the goats, while ensuring better utilization of the consumed feed as evident in most of the supplemented groups.

It was opined that there is need for more in vivo testing of plants purported to have anthelminthic properties, since in vitro assays puts the parasites in direct contact with the metabolites which are in concentrated forms [21]. Faecal egg count is a fast, simple, and easy method of estimating the worm burden in infected animals. The results of this study revealed significant decrease in faecal egg count with increasing level of A. nilotica incorporation in the diets of experimental animals. A similar study [22] on sheep infected with Trichostrongylus nematodes fed dried A. nilotica seeds showed a significant decrease in faecal egg count with increasing level of inclusion. However, another study [23] on Boer goats infected with H. contortus and fed dried Acacia karoo and Acacia nilotica documented A. nilotica as having no effect on faecal egg count of infected animals. Lower worm burden and faecal egg count may imply that the PSMs present in ANLM suppressed the viability of already established nematodes. This result may be attributed to the action of the many PSMs (most especially tannin) on the prolificacy of nematodes. Tannins have been reported severally as one of the secondary metabolites responsible for anthelmintic properties ascribed to various plants [6, 24 - 26]. Its mode of action was reported to include impairing crucial activities such as reproduction and feeding, and

damage to the parasites' carapace's integrity [27]. These include the possibility of ion channel complexion development, with proteins, polysaccharides, and enzyme inhibition. These actions may disrupt helminths' normal biochemical and physiological functions resulting in nutrient deprivation, structural modifications, neuromuscular disruptions, and other impacts [5]. It has been documented that most of these modes of actions are known targets for routinely used anthelmintics [7, 28]. It can be deduced from the results of this study that inclusion of ANLM in WAD goat diets is capable of mitigating H. contortus population in infected animals, thereby reducing faecal egg count and subsequently the deleterious effect of H. contortus on the performance of the animals.

The detrimental effects of gastrointestinal nematode infections especially H. contortus cannot be overemphasized. Disease conditions caused by nematode infections are usually most observed in performance, faecal egg count and blood profile of animals as Haemonchus majorly feeds on the blood of their hosts, thereby predisposing them to secondary infection and other diseases [29]. While faecal egg count is the easiest way to assess the presence of worm burden on the animal, haematological assay actually shows the indirect effect of these parasites on animals. Goats' haematological characteristics provide insight into the quality and quantity of nutrients they receive from their diet. Since it is used to diagnose illness and/or parasite infection in animals, it is also a crucial metric when it comes to infected goats [30, 31]. Red blood cells (RBCs) deliver oxygen throughout the body through haemoglobin (Hb), although packed cell volume (PCV) indicates the percentage of RBCs in the blood. Certain disorders may be indicated by an excess or deficiency of red blood cells.

Often, there is concern about feeding PSM-rich plants to animals due to poisoning. It is therefore imperative to monitor blood levels of animals to ensure that the test samples do not adversely affect the organ and general health status of the animals. In this study, the level of infection was low to express subclinical signs of infection as evident in the nonsignificantly different values recorded at the beginning of the study. Results obtained at the end of the experiment, showed that all the haematological parameters determined were within recommended range for healthy goats as reported by several authors [32, 34]. A similar result was

observed when *A. nilotica* pod meal was fed to growing male goats [35]. This could be attributed to the rich nutritional profile of the fed diets in the supplemented groups, the action of PSMs (especially tannin) in improving protein bypass in the rumen to make more protein available for absorption and utilisation, and the potential anthelmintic effect of the PSMs on adult *H. contortus* which would naturally compete for nutrients by feeding on the blood from the abomasal walls. These results further affirm the significance of ANLM in goat diets through improved performance, reduced faecal egg count with attendance improved in health status by mitigating the negative effects of the worm and subsequently aiding normal RBC production.

The primary function of white blood cell (WBC/leucocytes) is to provide mobile system of protection for the body and is affected by factors such as diseases, age, pregnancy, parturition, lactation, nutritional status, stress and/or excitement [36]. Often, parasitic infection triggers automatic immune response from the host leading to elevated WBC over red blood cells to combat the infection, which may result in anaemia. The results obtained in this study at the termination of the experiment revealed that WBC counts and its differentials were within the recommended range. The action of the fed diets both nutritionally and therapeutically could be responsible for the balanced level of WBC observed in the supplemented groups, by mitigating the negative actions of the nematodes on the animals and nutritionally supporting the animal while combating the parasites itself.

The reduced, but significantly different WBC values at the end of the experiment were within the normal reference range by Merck (2012) for healthy goats. In relation to this present study, a related study documented a decrease in WBC of Red Sokoto goats fed tannin-rich *Pterocarpus erinaceus* forage diets [37]. This could be due to the actions of PSMs in combating various forms of infections thus, reducing the natural response of the body in combating these infections in the animals if PSMs were absent in their diets.

The reduced and significantly different neutrophil values observed at the end of the experiment were within the normal reference value for healthy goats [31, 33], which maybe as a direct result of the variations observed for WBC. The lymphocyte count

observed in this study was also within the reference range for clinically healthy goats [33].

Eosinophil, basophil and monocyte values reported in this study were within recommended range for clinically healthy goats [31, 33]. However, these values are lower than those reported [34] in a study that focused on feeding *A. nilotica* pod meal to growing male goats. Eosinophil functions include but not limited to trapping substances and antiparasitic activities. The stable WBC, basophil, monocytes, and other differential count can be hinged on the curtailing action of the supplemented Acacia as well as the embedded PSM which helped the animals combat the infection. This shows that inclusion of ANLM in goat diets had no deleterious effect on the health status of the animals.

Conclusion

The efficacy of *Acacia nilotica* in leaf meal form as a suitable antihelminth and potential nutritive alternative *in vivo* especially for diseased ruminants has been reflected in the results obtained in this study. The reduced faecal egg count, improved performance and optimal blood parameters under diseased conditions affirms the potentials of *A. nilotica* as a suitable alternative in feeding and treating nematode infections in ruminants.

Acknowledgements

The authors wish to acknowledge DAAD, Germany for the award of postdoctoral fellowship and funding this research under the ClimapAfrica postdoctoral fellowship.

Authors contribution

Yusuf, Azeez Olanrewaju, Sonibare Adekayode Olanrewaju and Sowande Olusiji Sunday conceptualize the project idea and monitor the research throughout the project duration of the project. Owolabi Ayobami John is the PhD student on the research. He carried out the field and laboratory research while Taiwo Ajayi Olufemi was in charge of most of the laboratory activities. Yusuf, Azeez Olanrewaju proofread the first and second draft of the manuscript before being forwarded to Sowande Olusiji Sunday for final draft reading.

Conflict of Interest

The authors declared that there is no contrasting interest in the course of this research.

TABLE 1. Gross Composition of the Experimental Diet (%)

Ingredients	Inclusion Levels (%) of ANLM							
nigredients	0 ANLM	5 ANLM	10 ANLM	15 ANLM	20 ANLM			
Palm Kernel Cake	15	14	13	12	11			
Cassava Peels	20	19	18	17	16			
Rice Bran	30	29	28	27	26			
Wheat Offal	17	16	15	14	13			
Maize	15	14	13	12	11			
Acacia nilotica Leaf Meal	0	5	10	15	20			
Bone Meal	1	1	1	1	1			
Salt	2	2	2	2	2			
Total	100	100	100	100	100			
Determined Analysis (%) on air dried basis								
Dry Matter	89.51	87.47	91.39	90.06	88.26			
Crude Protein	15.75	16.18	16.35	16.59	16.85			
Crude Fibre	10.45	7.62	8.12	7.79	7.20			
Ash	7.17	6.52	8.28	6.45	8.69			
Crude Fat	6.93	4.09	5.29	5.84	6.86			
Nitrogen Free Extract	49.21	53.06	53.35	53.39	48.66			
Neutral Detergent Fibre	36.7	40.40	33.60	34.80	41.10			
Acid Detergent Fibre	21.60	19.30	22.40	18.90	23.80			
Acid Detergent Lignin	5.50	5.90	5.90	6.20	5.40			

ANLM: Acacia nilotica leaf meal.

TABLE 2. Growth Performance of artificially infected West African Dwarf Goats fed Varying Levels of *Acacia nilotica* Leaf Meal

Parameters	0% ANLM	5% ANLM	10% ANLM	15% ANLM	20% ANLM	SEM
Initial Weight (kg)	6.25	6.68	6.56	6.37	6.66	0.19
Final Weight (kg)	7.00^{b}	8.58 ^{ab}	8.16 ^{ab}	7.32 ^{ab}	9.06 ^a	0.31
Weight Gain (kg)	0.75^{b}	1.90^{a}	1.60 ^{ab}	0.95^{b}	2.40^{a}	0.17
Average Weight Gain (g/day)	13.39 ^b	33.93 ^a	28.57 ^{ab}	16.96 ^b	42.86 ^a	3.00
Concentrate mixture Feed Intake	189.60 ^{bc}	197.63 ^{abc}	241.52 ^a	169.73 ^c	224.58 ^{ab}	8.38
(g/day)						
Total Dry Matter Intake (g/day)	208.35 ^{bc}	217.17 ^{abc}	265.40 ^a	186.51 ^c	246.79 ^{ab}	9.21
Feed Conversion Ratio	15.56 ^a	6.40^{b}	9.29^{b}	11.00.60 ^{ab}	5.76 ^b	2.17

^{a, b, c} Means across rows with different superscripts are significantly (p<0.05) different.

TABLE 3. Haematological Indices of West African Dwarf Goats Artificially Infected with *Haemonchus contortus* and fed Varying Levels of *Acacia nilotica* Leaf Meal (ANLM)

Parameters		0%	5%	10%	15%	20%	SEM	Reference
		ANLM	ANLM	ANLM	ANLM	ANLM		Range
Packed Cell Volume (%)	Initial	25	25.67	24.67	24.67	26.33	0.95	22-38+
	Final	24.13^{b}	28.38^{ab}	33.63 ^a	24.75 ^b	30.38^{a}	1.02	
	Difference	-0.87	2.71	8.96	0.08	4.05	0.07	
Haemoglobin (g/dL)	Initial	11.97	12.33	11.8	11.73	12.53	0.46	7-15*
0 10 1	Final	8.81^{b}	10.76^{a}	10.95^{a}	9.14 ^b	9.98^{ab}	0.24	
	Difference	-3.16	-1.57	-0.85	-2.59	-2.55	-0.22	
Red Blood Cell (×10 ^{12/L})	Initial	1.57	1.27	1.99	4.85	1.46	0.61	8-18+
	Final	14.55 ^{ab}	14.22 ^b	15.48 ^{ab}	15.65 ^{ab}	17.10^{a}	0.39	
	Difference	12.98	12.95	13.49	10.8	15.64	-0.22	
Mean Corpuscular	Initial	15.97	15.63	17.3	17.9	16.1	0.77	16-25
Volume (fl)								
	Final	16.77 ^{ab}	20.74^{ab}	22.48^{a}	16.10^{b}	18.04^{ab}	0.91	
	Difference	0.8	5.11	5.18	-1.8	1.94	0.14	
Mean Corpuscular	Initial	7.6	7.53	8.27	8.47	7.63	0.36	5.2-8
Haemoglobin (pg)								
3 43	Final	6.13 ^b	7.79^{a}	7.28^{ab}	$5.90^{\rm b}$	5.95 ^b	0.26	
	Difference	-1.47	0.26	-0.99	-2.57	-1.68	-0.1	
Mean Corpuscular	Initial	47.87	48.1	47.87	47.5	47.53	0.16	30-36
Haemoglobin								
Concentration (g/dL)								
,	Final	37.02^{ab}	38.91 ^a	32.69^{b}	37.98^{a}	32.86^{b}	0.81	
	Difference	-10.85	-9.19	-15.18	-9.52	-14.67	0.65	

^{a, b} Means across rows with different superscripts are significantly (p<0.05) different.

^{*}Daramola et al. (2005)

⁺Merck Manual (2012)

TABLE 4. White Blood Cell and Differentials Count of West African Dwarf Goats Artificially Infected with Haemonchus contortus and fed Varying Levels of Acacia nilotica Leaf Meal

Parameters		0%	5%	10%	15%	20%	SE	Referenc
		ANLM	ANLM	ANLM	ANLM	ANLM	\mathbf{M}	e Range
White Blood Cell (×10 ^{9/L})	Initial	13.27	14.20	11.93	11.17	14.60	0.98	4-13 ⁺
	Final	8.05 ^{bc}	11.00^{a}	7.25°	8.69 ^{bc}	9.24 ^b	0.35	
	Difference	-5.22	-3.20	-4.68	-2.48	-5.36	-0.63	
Neutrophils (%)	Initial	44.33	40.00	37.33	39.33	40.67	5.51	30-48+
•	Final	33.75 ^b	37.25 ^{ab}	34.38^{ab}	39.13 ^a	36.38^{ab}	0.7	
	Difference	-10.58	-2.75	-2.95	-0.20	-4.29	-4.81	
Lymphocytes (%)	Initial	53.33	59.00	61.67	59.67	58.00	1.53	47-82*
	Final	65.88^{a}	61.63 ^{bc}	64.88^{ab}	59.88 ^c	62.00^{abc}	0.70	
	Difference	12.55	2.63	3.21	0.21	4	-0.83	
Eosinophils (%)	Initial	1.00^{a}	0.67^{ab}	0.67^{ab}	0.00^{b}	0.67^{ab}	0.13	$0.05 \text{-} 0.65^{+}$
_	Final	0.00	0.38	0.38	0.00	0.38	0.07	
	Difference	-1.00	-0.29	-0.29	0.00	-0.29	-0.06	
Basophils (%)	Initial	0.33	0.33	0.00	0.33	0.00	0.11	0-1+
	Final	0.25^{ab}	0.00^{b}	0.00^{b}	0.25^{ab}	0.63^{a}	0.08	
	Difference	-0.08	-0.33	0.00	-0.08	0.63	-0.03	
Monocytes (%)	Initial	1.00	0.00	0.33	0.67	0.67	0.17	0-1*
	Final	0.38^{b}	0.38^{b}	0.38^{b}	1.00^{a}	0.63^{ab}	0.08	
	Difference	-0.62	0.38	0.05	0.33	-0.04	-0.09	

 $^{^{\}text{a, b, c}}$ Means across rows with different superscripts are significantly (p<0.05) different.

⁺Merck Manual (2012)

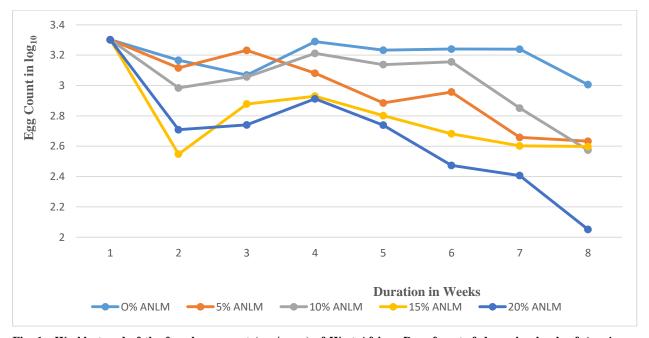


Fig. 1. Weekly trend of the faecal egg count (egg/gram) of West African Dwarf goats fed varying levels of *Acacia nilotica* leaf meal.

^{*}Daramola et al. (2005)

References

- Coles, G.C. Anthelmintic resistance looking to the future: a UK perspective. Research in Veterinary Science, 78(2), 99-108.(2005).
- Lüscher, A., Häring, D.A., Heckendorn, F., Scharenberg, A., Dohme, F., Maurer, V. and Hertzberg, H. Use of tanniferous plants against gastrointestinal nematodes in ruminants. In Researching Sustainable Systems-International Scientific Conference on Organic Agriculture, Adelaide, Australia (2005).
- 3. Rahmann, G., Koopmann, R. and Hertzberg, H. Gesundheit erhalten statt Krankheit Kurieren. [Maintain health instead of disease curing-Animal health in organic farming] FORSCHUNGS Report, Verbraucherschutz, Ernährung, Landwirtschaft. Forschungs, 1(25), 4-7 (2002).
- 4. Waller, P.J. Sustainable helminth control in developing countries. *Veterinary Parasitology*, **71**, 195-207(1997).
- Badar, N., Iqbal, Z., Khan, M.N. and Akhtar, M.S. 2011. In vitro and in vivo anthelmintic activity of Acacia nilotica (L.) Willd. ex Delile bark and leaves. Pakistan Veterinary Journal, 31(3).
- Min, B.R. and Hart, S.P. Tannins for suppression of internal parasites. *Journal of Animal Science*, 81(14_suppl_2): E102-E109(2003)..
- 7. Mottier L, L Alvarez, L Ceballos and C Lanusse,. Drug transport mechanisms in helminth parasites: Passive diffusion of benzimidazole anthelmintics. *Experimental Parasitology*, **113**, 49-57(2006).
- 8. Jordan, H.I., Philips, W.A., Morrison, R.D., Deyle, J. and Mckenzic, K. A three-year study of continuous mixed grazing of cattle and sheep: parasitism of offspring, *International Journal for Parasitology*, **18**, 779-784(1988)..
- Jain, N.N. Essentials of veterinary haematology. Lea & Febieger, Philadelphia, PA. J. Clin. Microbiol., 35, 823-829(1993).
- 10. Lamb, G.N., Manual of veterinary laboratory technique. *CIBA-Geigy, Kenya*, 96-107(1981).
- Yusuf, A.O., Iposu, S.O., Sanwo, K.A., Akinde, K.T., Owolabi, A.J., Omotosho, O.O.A., Adedayo, O.A. and Okwelum, N., Influence of Moringa leaf meal (MOLM) supplementation on Growth performance, Haematological parameters and faecal egg count of West African dwarf sheep. *Nigerian Journal of Animal Science*, 18(2), 492-506(2016).
- Tham, H. T. Water Hyacinth (Eichhornia crassipes) Biomass Production, Ensilability and Feeding Value to Growing Cattle. Faculty of Veterinary Medicine and Animal Science Department of Animal Nutrition and Management Uppsala. Acta Universitatis Agriculturae Sueciae, 1652-6880 (2012). 90 ISBN 978-91-576-7737-2 [Doctoral Thesis].
- 13. Yusuf, A.O., Owolabi, A.J., Adebayo, K.O., Aina, A.B.J., Sowande, O.S. and Ajayi, T.O. Growth performance of goats fed diets containing varying levels of water hyacinth (*Eichhornia crassipes*). *Agricultura Tropica et Subtropica*, **54**(1), 228-237 (2021).

- Durmic, Z., Black, J.L., Martin, G.B. and Vercoe, P.E. Harnessing plant bioactivity for enteric methane mitigation in Australia. *Animal Production Science*, 62(12) 1160-1172 (2021).
- 15. Gemeda, B.S. and Hassen, A. The potential of tropical tannin rich browses in reduction of enteric methane. *Approaches in Poultry, Dairy and Veterinary Sciences*, **2**, 154-162(2018).
- 16. Yahaya, B. Evaluation of nutritive value of Acacia (*Acacia seyal del*.) fruits on performance of Yankasa sheep (Doctoral dissertation, M. Sc Thesis submitted to animal Sci. Dept. Ahmadu Bello University, Zaria, Nigeria) (2011)..
- 17. Uguru, C., Lakpini, C.A.M., Akpa, G.N. and Bawa, G.S. Nutritional potential of acacia (Acacia nilotica (l.) del.) pods for growing Red Sokoto goats. *IOSR Journal of Agriculture and Veterinary Science*, **7**(6), 43-49 (2014).
- 18. Yang, C., Chowdhury, M.K., Hou, Y. and Gong, J. Phytogenic compounds as alternatives to in-feed antibiotics: potentials and challenges in application. *Pathogens*, **4**(1),137-156(2015).
- 19. Ferket, P.R., Van Heugten, E., Van Kempen, T.A.T.G. and Angel, R., Nutritional strategies to reduce environmental emissions from nonruminants. *Journal of Animal Science*, **80**(2), 168-182(2002).
- 20. Charlier, J., Höglund, J., Morgan, E.R., Geldhof, P., Vercruysse, J. and Claerebout, E., Biology and epidemiology of gastrointestinal nematodes in cattle. *Veterinary Clinics: Food Animal Practice*, **36**(1), 1-15(2020).
- 21. Athanasiadou, S., Kyriazakis, I., Jackson, F. and Coop, R. L. Direct anthelmintic effects of condensed tannins towards different gastrointestinal nematodes of sheep: in vitro and in vivo studies. *Veterinary Parasitology*, **99**, 205-219(2001)..
- 22. Bachaya, H.A., Iqbal, Z., Khan, M.N. and Jabbar, A. Anthelmintic activity of *Ziziphus nummularia* (bark) and *Acacia nilotica* (fruit) against *Trichostrongylid nematodes* of sheep, *Journal of Ethnopharmacology*, 123, 325–329(2009).
- 23. Kahiya, C., Mukaratirwa, S. and Thamsborg, S.M. Effects of *Acacia nilotica* and *Acacia karoo* diets on *Haemonchus contortus* infection in goats. *Veterinary Parasitology*, **115**(3), 265-274(2003).
- 24. Hoste, H., Jackson, F., Athanasiadou, S., Thamsborg, S.M. and Hoskin, S.O. The effects of tannin-rich plants on parasitic nematodes in ruminants. *Trends in Parasitology*, **22**(6),253-261(2006).
- 25. Athanasiadou, S., Githiori, J. and Kyriazakis, I., Medicinal plants for helminth parasite control: facts and fiction. *Animal*, **1**(9), 1392-1400(2007).
- 26. Brunet, S., Aufrere, J., El Babili, F., Fouraste, I. and Hoste, H., The kinetics of exsheathment of infective nematode larvae is disturbed in the presence of a tannin-rich plant extract (sainfoin) both in vitro and in vivo. *Parasitology*, **134**(9), 1253-1262(2007).

- Niezen, J.H., Waghorn, T.S., Charleston, W.A.G. and Waghorn, G.C. Growth and gastrointestinal nematode parasitism in lambs grazing either lucerne (*Medicago* sativa) or sulla (*Hedysarum coronarium*) which contains condensed tannins. The Journal of Agricultural Science, 125(2), 281-289(1995).
- Kohler, P., The biochemical basis of anthelmintic action and resistance. *International Journal of Parasitology*, 31, 336-345(2001).
- Awais, M., Abbas, K., Qureshi, A.S., Awais, M., Bayer, M., Awais, M., ul Hassan, M., Nargis, S., Qureshi, A.S., Masood, A. and Ali, M.Z., November. LIST OF ABSTRACTS. In *Proc 3rd Int Workshop Dairy Science Park* (p. 1). (2015).
- Karesh, W.B. and Cook, R.A. Applications of veterinary medicine to in situ conservation efforts. *Oryx*, 29(4), 244-252(1995).
- Yusuf, A.O., Oyebanji, O.A., Yusuf, D.A., Ekunseitan, K.A., Adeleye, O.S., Sowande, O.S. and Fasae, O.A. Blood profile of West African dwarf goats fed Panicum maximum supplemented with Newbouldia laevis leaves. *Bulletin of Animal Health and Production in Africa*, 60(4), 481-490(2012).
- 32. Daramola, J. O., Adeloye, A. A., Fatoba, T. A. and Soladoye, A. O. Haematological and biochemical

- parameters of West African Dwarf goats. *Livestock Research for Rural Development*, **17**(8), 3(2005).
- Opara, M.N., Udevi, N. and Okoli, I.C. Haematological parameters and blood chemistry of apparently healthy West African Dwarf (Wad) goats in Owerri, South Eastern Nigeria. New York Science Journal, 3(8), 68-72(2010).
- 34. Merck, Manual, 2012. Haematological reference ranges. *Mareck Veterinary Manual*. Retrieved from http://www.merckmanuals.com
- 35. Paswan, J.K., Kumar, K., Kumar, S., Chandramoni, Kumar, A., Kumar, D. And Kumar, A., Effect of feeding *Acacia nilotica* pod meal on hematobiochemical profile and faecal egg count in goats. *Veterinary World*, **9**(12), 1400-1406(2016)...
- Tornquist, S.J. and Rigas, J. Interpretation of ruminant leukocyte responses In: Schalm,s Veterinary Haematology. Weiss. D.J and Wardrop, K.J. (Ed.) 6th ed.. A John Wiley and Sons, Ltd. USA, pp. 307-313(2010).
- 37. Olafadehan, O.A., Adewumi, M.K. and Okunade, S.A. Effects of feeding tannin-containing forage in varying proportion with concentrate on the voluntary intake, haematological and biochemical indices of goats. *Trakia Journal of Sciences*, **12**(1),73(2014).