

Egyptian Journal of Veterinary Sciences

https://ejvs.journals.ekb.eg/



Enhancing the Antimicrobial Effect of Ciprofloxacin by Loading with Silica Nanoparticles Derived from Rice Straw: Spectral and Morphological Studies



Rania M. Alwakeel^{1,2}, Mohamed Moustafa Abdelaal³ and Ola M. El-Borady⁴*

- ¹ Chemistry department, School of Science & Engineering, The American University in Cairo (AUC), Egypt. E-mail: Raniaalwakeel@aucegypt.edu.
- ² Faculty of Biotechnology, October University for Modern Sciences and Arts (MSA University), 6th of October City, Egypt.

Abstract

NTIMICROBIAL resistance is a significant threat to both human and animal health, demanding a One Health approach to therapeutic strategies. The growing resistance of pathogens such as Salmonella, Brucella, E. coli and S. aureus complicates treatment and raises healthcare costs, highlighting the urgent need for alternative therapeutic strategies. Therefore, this study aims to prepare silica nanoparticles (SiO2-NPs) from rice straw through acid leaching method, then ciprofloxacin, one of the most abundant broad-spectrum antibiotics used worldwide, was loaded to SiO₂-NPs. Comprehensive characterization of TEM, SEM, EDX, and FTIR confirmed successful SiO₂-NPs synthesis and effective drug incorporation. TEM analysis for unmodified SiO₂-NPs showed spherical particles with a size of 130–170 nm, while ciprofloxacin-loaded SiO₂-NPs (CIP@SiO₂-NPs) expanded to 340-640 nm, indicating successful encapsulation and surface modification. These nanoparticles' antibacterial efficacy was tested against Gram-positive and Gram-negative bacteria frequently involved with zoonotic illnesses. The findings reveal that the ciprofloxacin-loaded SiO₂-NPs exhibit superior antibacterial activity to free ciprofloxacin especially for Gram (+ve) bacterial strains. Furthermore, the study proves the efficacy of rice straw-derived SiO₂-NPs as an eco-friendly nanocarrier approach for improving antibiotic delivery that enhances the management of bacterial infections in veterinary medicine, minimizing the risk of zoonotic disease transmission to humans.

Keywords: Antimicrobial resistance, One health approach, Silica Nanoparticles, Rice-Straw, Ciprofloxacin, Zoonotic diseases.

Introduction

Antimicrobial resistance (AMR) has caused a serious worldwide health concern, affecting the efficiency of antibiotics in both human and veterinary medicine [1, 2]. The misuse of antibiotics in clinical, agricultural, and animal care settings is a major contributor to the rising prevalence of AMR [3]. The extreme usage of antibiotics, generally in veterinary and aquaculture medicine, has had an essential influence on establishing and spreading resistant bacterial strains [4, 5]. Accordingly, AMR may cause the spread of

many diseases transmitted to humans by consuming contaminated animal products or bacterial strains that affect humans by direct contact or environmental exposure, leading to severe health consequences [6, 7]. The essential need for novel therapeutic strategies to combat these resistant pathogens is more evident than ever [8, 9].

It was detected that Zoonotic diseases, transmitted from animals to humans, are a significant part of emerging infectious diseases. The World Health Organization (WHO) has recorded over 200

*Corresponding authors: Ola M. El-Borady, E-mail: olachem_elborady@yahoo.com, Tel.: 01004725266 (Received 22 May 2025, accepted 19 August 2025)

DOI: 10.21608/ejvs.2025.387882.2864

³Department of Cardiothoracic Surgery, Faculty of Medicine, Kafrelsheikh University, Egypt.

⁴ Institute of Nanoscience and Nanotechnology, Kafr-Elsheikh University, Kafr-Elsheikh, Egypt.

zoonotic diseases, many associated with antimicrobial-resistant pathogens [10]. The overuse of antibiotics in veterinary practices, especially for disease prevention and growth promotion in veterinary medicine, has been a key factor in the development of AMR [11, 12].

Furthermore, antibiotic-resistant bacteria can spread from animals to humans through various means, including direct contact, consumption of contaminated animal products, and environmental exposure [13]. For instance, urban wildlife like ducks and crows have been found to carry antibiotic-resistant bacteria, posing a risk to public health by contaminating the food chain [7]. To address the escalating AMR crisis, a One Health approach is crucial, emphasizing the interconnectedness of human, animal, and environmental health [14, 10].

Infections induced by resistant bacteria could lead to prolonged illness, increased veterinary costs, and, in severe cases, animal mortality [4]. Resistant bacteria in animals also act as reservoirs that can infect humans, complicating treatment [6]. As a result, implementing antimicrobial stewardship programs in veterinary settings is critical to encouraging ethical antibiotic use and preventing resistance development [5]. Alternative therapeutic strategies, with nanotechnology gaining attention for their potential in combating resistant pathogens [15, 16]. The unique physicochemical features of nanomaterial's (such as their high surface area and small size) offer multiple mechanisms to inhibit growth. [17]. Amorphous bacterial nanoparticles (SiO2-NPs) have shown particular promise in antimicrobial applications due to their high surface area compared to other nanoparticles [19, 20]. SiO₂-NPs can encapsulate various therapeutic agents, including antibiotics, ensuring controlled release and targeted delivery to infection sites [19, 20]. These leading SiO2-NPs enable them to interact with bacterial cell membranes, inducing disruption and bacterial cell death. SiO2-NPs also generate reactive oxygen species (ROS), further enhancing their antimicrobial activity [16]. The antimicrobial properties and drug delivery capabilities of SiO2-NPs offer a dual-functional approach to combating AMR [17].

Ciprofloxacin is a broad-spectrum bactericidal second-generation fluoroquinolone that inhibits crucial bacterial growth enzymes [21, 22]. Despite its efficacy, ciprofloxacin has potential side effects, and many efforts are now being made to develop resistant strains, emphasizing the need for responsible usage to maintain the effectiveness of ciprofloxacin [22]. This research aims to investigate aspects like encapsulation efficacy and the antimicrobial effect of ciprofloxacin-loaded SiO₂-NPs against different bacterial strains. The study findings seek to provide insights into enhancing the therapeutic efficacy of ciprofloxacin and offer solutions for combating

AMR, particularly through advanced drug delivery systems like amorphous silica nanoparticles.

Material and Methods

Synthesis of Silica Nanoparticles

SiO₂-NPs were synthesized via acid leaching of grinded rice straw, a method known for producing high-purity amorphous silica with substantial surface area [19]. Initially, 25 g of finely ground rice straw immersed in 100 mL of concentrated hydrochloric acid (HCl) and stirred continuously for 3 hours at ambient temperature to remove metallic Post-leaching, impurities. the mixture neutralized using sodium hydroxide (NaOH) until a neutral pH was achieved, followed by filtration to separate the solid residue. The obtained residue was oven-dried at 100 °C and subsequently subjected to calcination in a muffle furnace at 750 °C for 5 hours. This thermal treatment yielded a white, amorphous silica powder, which was stored at room temperature for further analysis and applications.

Surface Functionalization of Silica Nanoparticles

The synthesized SiO₂-NPs were functionalized using the crosslinker. (3-aminopropyl) trimethoxysilane (APTMS) to introduce amine groups onto their surfaces, enhancing their reactivity for subsequent drug loading. The functionalization process involved dispersing the SiO2-NPs in an ethanol solution containing APTMS, followed by stirring for a specified duration to allow silanization. Post-reaction, the modified nanoparticles were washed thrice with distilled water and once with 70% ethanol to remove unreacted silane and by-products. The final product was dried in a petri dish, resulting in a white powder of amine-functionalized SiO₂-NPs.

Loading of Ciprofloxacin onto Functionalized Silica Nanoparticles

To ciprofloxacin (CIP) load onto functionalized SiO₂-NPs, 0.16 g of CIP dissolved in 100 mL of distilled water using magnetic stirring to ensure complete dissolution. Subsequently, 0.3 g of the amine-functionalized SiO₂-NPs was added to the solution, and the mixture was stirred at 80 °C for 3 hours to facilitate adsorption of CIP onto the nanoparticle surfaces. The CIP@SiO2-NPs then separated were centrifugation, washed twice with distilled water and once with 70% ethanol to remove unbound drug molecules, and dried in a petri dish. The supernatants from the washing steps were collected to determine drug loading efficiency.

Characterization of Silica Nanoparticles

The synthesized and functionalized SiO₂-NPs were comprehensively characterized using various analytical techniques. Transmission Electron Microscopy (TEM), EOL JEM-2010 (Japan) operated at 200 kV, was employed to investigate the

morphology and size distribution of three types of samples: unmodified SiO₂-NPs, amine-functionalized SiO₂-NPs, and CIP@SiO₂-NPs. The Energy-Dispersive X-ray Spectroscopy (EDX) was employed to analyze their elemental composition. Additionally, Fourier-Transform Infrared Spectroscopy (FTIR), JASCO spectrometer (Japan), was conducted to identify characteristic functional groups and to confirm the successful surface modification with amine groups as well as the subsequent loading of ciprofloxacin onto the nanoparticles. characterizations were conducted at the institute of nanoscience and Nanotechnology, Kafr El-Sheikh University, Egypt.

Antibacterial Activity Assessment

The antibacterial efficacy of the CIP@SiO2-NPs was assessed using the agar well-diffusion approach against four bacterial strains: Staphylococcus aureus (Gram-positive), Pseudomonas aeruginosa (Gramnegative), Bacillus cereus (Gram-positive), and Escherichia coli (Gram-negative). These strains were obtained from the Microbial Genetics Laboratory, Genetic Engineering and Biotechnology Division, National Research Centre, Egypt. Each bacterial strain was cultured in Mueller-Hinton broth and incubated at 35 °C with shaking at 120 rpm for 24 hours. The cultures were then adjusted to an optical density of 0.5 at 570 nm, corresponding to approximately 1.5×10^8 CFU/mL. Sterile cotton swabs spread the bacterial suspensions uniformly onto Mueller-Hinton agar plates. After that, wells (6 mm) were punched into the agar, and 100 μL of the CIP-loaded SiNP suspension was introduced into each well. The plates included tested samples were incubated for 24 hrs at 37 °C, after which the inhibition zones were measured to antibacterial activity.

Results

Characterization of Silica Nanoparticles

Transmission Electron Microscopy (TEM)

TEM images Fig.1(a,b) revealed that the synthesized silica nanoparticles (SiO2-NPs) were spherical with predominantly uniform distribution. The average particle size was estimated to be approximately 130 to 170 nm, showing slight agglomeration. Post-functionalization with APTMS, Fig.1(c,d) showed images maintained their morphology, with a minor increase in apparent particle size due to surface coating, indicating successful surface modification without significant aggregation. On the other hand, the HR-TEM images Fig.1(e,f) for the CIP@SiO2-NPs also retained their spherical shape, with a slight increase in size to approximately 340-640 nm, suggesting effective drug loading.

Scanning Electron Microscopy (SEM)

Morphological examination using SEM confirmed the structural changes throughout different stages. The original silica nanoparticles, in Fig.2(a) revealed characteristic non-uniform flake shape structure with a relatively rough surface topology. Fig.2(b) showed surface topology alteration upon modification with APTMS. Drug delivery system formation has been confirmed in Fig.2(c) where particles exhibit surface heterogeneity and enhanced roughness, that serves as morphological evidence indicating high capacity drug incorporation within the silica structure.

Energy-Dispersive X-ray Spectroscopy (EDX)

EDX pattern Fig.2(d) confirmed the elemental composition of the SiO₂-NPs, primarily consisting of silicon and oxygen, and minor Ca and P impurities may be due to the residual of rice straw. Fig.2(e), showed additional signals due to the presence of nitrogen in the functionalized SiO₂-NPs, indicating successful amine group incorporation, while the EDX pattern in Fig.2(f) for the *ciprofloxacin* loaded SiO₂-NPs detect fluorine atoms, confirmed drug loading.

Fourier-Transform Infrared Spectroscopy (FTIR)

FTIR spectra in Fig.3 of unmodified SiO₂-NPs shows characteristic peaks at ~3400 cm⁻¹ (O-H stretching), $\sim 1630 \text{ cm}^{-1}$ (H-O-Hbending), ~1080 cm⁻¹ (Si–O–Si asymmetric stretching), and $\sim 800 \text{ cm}^{-1}$ (Si-O-Si symmetric stretching), confirming the formation of amorphous silica. After surface modification with APTMS, new peaks at 2900-2950 cm⁻¹ (C-H stretching) and ~1550 cm⁻¹ (N-H bending) appear, indicating successful functionalization with amine groups. Following ciprofloxacin loading, additional bands at ~1700-1725 cm⁻¹ (C=O stretching) and ~1450-1380 cm⁻¹ (C-N and CH₃ bending) emerge, confirming the successful incorporation of ciprofloxacin onto the functionalized silica nanoparticles.

Drug Loading Efficiency

The loading efficiency of ciprofloxacin onto the functionalized SiO₂-NPs was determined by measuring the concentration of unbound drug in the supernatant post-loading. Using UV-Vis spectrophotometry at 276 nm, the loading efficiency was calculated to be 87%, indicating a substantial amount of drug was successfully adsorbed onto the nanoparticles.

Antibacterial Activity

The antimicrobial activity of CIP@SiO₂-NPs was evaluated using the agar well diffusion method against a panel of Gram-positive and Gram-negative bacterial strains. The results demonstrated, in Fig.4, that the CIP@SiO₂-NPs exhibited enhanced antibacterial activity compared to free ciprofloxacin

against most tested strains. For Gram-positive bacteria, B. subtilis and B. licheniformis showed increased inhibition zones with CIP@SiO2-NPs (16.29 mm and 19.21 mm, respectively) compared to ciprofloxacin alone (15.33 mm and 18.09 mm, respectively) as shown in Fig.5. Similarly, a slight improvement was observed against B.cereus and S.aureus. Among Gram-negative strains, a notable enhancement was detected for E.coli, where the inhibition zone increased from 11.55 mm for free ciprofloxacin to 13.07 mm for the nanoparticles. P.fluorescens also exhibited a slight increase in the inhibition zone with CIP@SiO2 NPs, while P.aeruginosa showed a minor decrease. No significant difference was observed for E.ludwigii. Inhibition zone diameters were expressed as mean values. To assess the effect of treatment, a Wilcoxon signed-rank test was performed comparing ciprofloxacin and SiO₂-NPs-APTMS ciprofloxacin within each bacterial group. The analysis was conducted separately for Gram-positive (B.subtilis, B. licheniformis, B. cereus, S.aureus) and Gram-negative (E.coli, P.aeruginosa, P. fluorescens, E.ludwigii) strains. A statistically significant increase in inhibition zone diameter was observed in the Gram-positive group following nanoparticle encapsulation (p = 0.038), whereas the difference in Gram-negative strains was not significant (p =0.875). These findings suggest that loading ciprofloxacin onto silica nanoparticles can improve its antibacterial efficacy compared to free ciprofloxacin, particularly against certain Grampositive bacteria and specific Gram-negative strains such as E. coli and P. fluorescens.

Discussion

The results of this study provide important insights into the evolving application of nanoparticlebased drug delivery systems in veterinary antimicrobial therapy, particularly targeting zoonotic bacterial pathogens [14, 8]. The observed increase in particle size following ciprofloxacin encapsulation, as demonstrated by transmission electron microscopy (TEM), aligns with previous findings indicating that drug loading onto silica nanoparticles induces particle expansion. This size enlargement is associated with improved sustained drug release, a critical attribute for the effective management of chronic and recurrent infections in veterinary medicine [23,24]. The successful surface modification and encapsulation were further supported by scanning electron microscopy (SEM) and elemental analysis measured by (EDX) analysis, which detected elemental signatures critical for confirming functionalization [25]. In veterinary contexts, such modifications are particularly valuable for improving drug stability and enhancing bioavailability in complex biological environments such as infected tissues or abscesses in companion animals and livestock [26].

The spectral analysis including the Fourier-transform infrared spectroscopy (FTIR) data revealed the presence of Si–O–Si networks, amine groups, and ciprofloxacin functional bands, suggesting strong electrostatic interactions between the drug and the nanoparticle matrix. These findings align with previous studies highlighting that amine-functionalized SiO₂-NPs improve antibiotic loading and release kinetics, enhancing therapeutic outcomes in animal models [27].

The improved antimicrobial efficacy of CIP@SiO₂-NPs against Gram-positive and Gram-negative veterinary pathogens such as *Bacillus subtilis*, *Bacillus licheniformis*, and *Escherichia coli* demonstrates the potential of this platform in managing common animal infections. Therefore, by improved local drug concentration and sustained release the need for frequent dosing reduced, thereby minimizing stress in treated animals and decreasing the risk of antibiotic overuse, a major driver of antimicrobial resistance [28, 29].

The slightly reduced efficacy observed against *Pseudomonas aeruginosa*, a known multidrugresistant organism in both human and veterinary medicine, underlines the importance of developing combination therapies or nanoparticle-based synergistic strategies to overcome intrinsic bacterial defense mechanisms [30, 31].

In addition to acting as carriers, SiO₂-NPs may exhibit intrinsic antimicrobial properties, such as membrane disruption and ROS generation [32, 16]. Such synergistic influences could be particularly beneficial in veterinary wound management, mastitis treatment in dairy cattle, and surgical site infection control, particularly in small animals.

Utilizing rice straw biomass as a precursor for silica nanoparticles provides an environmentally sustainable and cost-effective approach. This strategy is highly relevant for veterinary applications in low and middle-income countries such as Egypt, where restricted access to therapeutics can be limited [33].

In summary, this study presents a promising, ecofriendly approach for enhancing the antimicrobial efficacy of conventional antibiotics through silica nanoparticle-based delivery systems. This strategy holds potential for addressing the urgent need for effective antimicrobial therapies in veterinary practice, particularly in the face of growing concerns about zoonotic disease transmission and antibiotic stewardship [14, 8].

Limitations for the present work: There are some challenges to apply this solution such as rigorous toxicity study should be done to assess the effect of this nano-drug delivery system on animal and human health. Also, for future work, another preparation method parameter will be tested to get smaller SiO₂-NPs with enhanced antimicrobial activity.

Conclusion

In conclusion, SiO₂-NPs were successfully synthesized from rice straw through acid extraction processes, resulting in high-purity amorphous SiO2-NPs with desirable surface properties. Surface functionalization with APTMS was achieved, providing amine groups that facilitated the efficient loading of ciprofloxacin onto the nanoparticles. Comprehensive characterization using TEM, SEM-EDX, and FTIR confirmed the successful synthesis, processes. modification. and drug loading Antibacterial assays demonstrated that ciprofloxacinloaded SiO₂-NPs exhibited enhanced or comparable antimicrobial activity against a range bacterial strains, compared to free ciprofloxacin, particularly improving efficacy against E.coli, B.subtilis, and B.licheniformis. These determinations emphasize the possibility of SiO2-NPs being utilized as drug systems for improving delivery antibiotic performance and comprising new alternative antibiotics, including nano-sized materials.

Acknowledgments

The authors gratefully express their gratitude to Prof. Dr. Ayman Diab and Prof. Dr. Gehan Safwat (Faculty of Biotechnology, October University for Modern Sciences & Arts (MSA), Egypt) for their encouragement and support.

Funding statement

This study didn't receive any funding support.

Declaration of Conflict of Interest

None of the authors have any conflicting interests

Authors' contributions

Rania M. Al wakeel, Ola M. El-Borady the experimental work of the characterization part, investigation, data analysis, and writing of the original draft. Mohamed Abdelaal; supervision, validation, and reviewing of the final manuscript.

Ethical of approval

This study does not need ethical regulations.

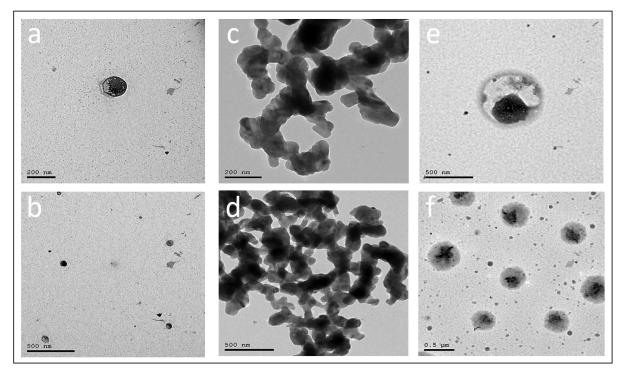


Fig. 1. (a,b) TEM images of prepared SiO₂-NPs from different spots with different magnification scale; (c, d).) APTMS-functionalized SiO₂-NPs, (e,f) Ciprofloxacin @ SiO₂-NPs.

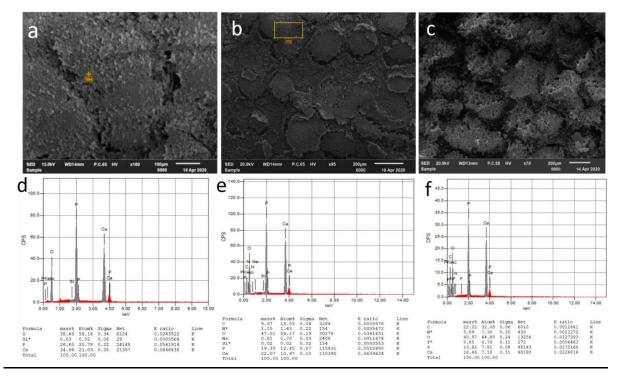


Fig. 2. SEM-EDX patterns of prepared (a,d) SiO₂-NPs, (b,e) APTMS-functionalized SiO₂-NPs SEM image & EDX results confirming successful SiO₂-NPs surface modification and (c,f) Ciprofloxacin @ SiO₂-NPs, showing successful drug loading on SiO₂-NPs.

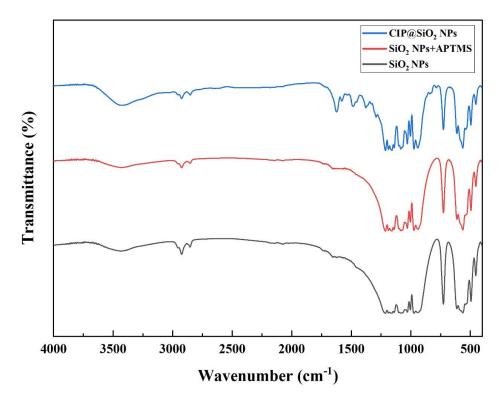


Fig. 3. FTIR spectra of prepared SiO₂ NPs (black line), SiO₂ NPs + APTMS (Red line) and CIP@ SiO₂ NPs (blue line).

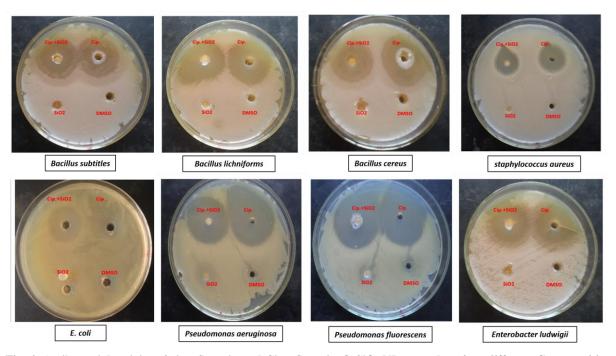


Fig. 4. Antibacterial activity of ciprofloxacin and Ciprofloxacin @ SiO₂ NPs, tested on four different Gram-positive and four different Gram-negative bacterial strains.

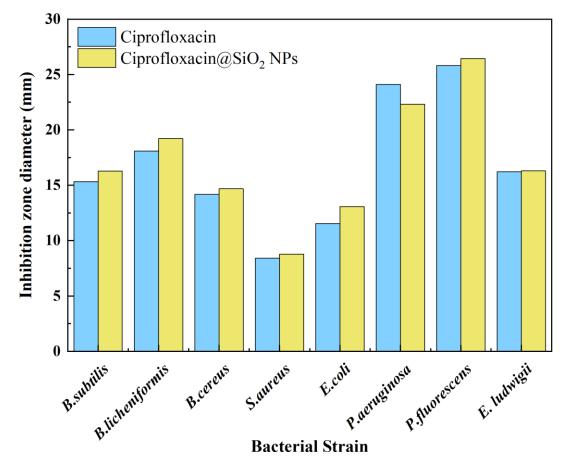


Fig. 5: Bar chart illustrating the inhibition zone diameters (in mm) of ciprofloxacin (blue bars) and Ciprofloxacin @ SiO₂ (yellow bars) against eight bacterial strains, including both Gram-positive (Bacillus subtilis, Bacillus licheniformis, Bacillus cereus, Staphylococcus aureus) and Gram-negative (Escherichia coli, Pseudomonas aeruginosa, Pseudomonas fluorescens, Enterobacter ludwigii) bacteria.

References

- Horvat, O., & Kovačević, Z. Human and Veterinary Medicine Collaboration: Synergistic Approach to Address Antimicrobial Resistance Through the Lens of Planetary Health. Antibiotics (Basel, Switzerland), 14(1),38(2025). https://doi.org/10.3390/antibiotics14010038
- World Health Organization. Antimicrobial resistance. (2023). https://www.who.int/news-room/fact-sheets/detail/antimicrobial-resistance
- Ahmed, S. K., Hussein, S., Qurbani, K., Ibrahim, R. H., Fareeq, A., Mahmood, K. A. and Mohamed, M. G. Antimicrobial resistance: Impacts, challenges, and future prospects. *Journal of Medicine, Surgery, and Public Health*, 2, 100081. (2024). https://doi.org/10.1016/j.glmedi.2024.100081
- Kaur, K., Singh, S. and Kaur, RImpact of antibiotic usage in food-producing animals on food safety and possible antibiotic alternatives. *The Microbe.*, 100097. (2024). https://doi.org/10.1016/j.bsheal.2020.09.004
- Hoelzer, K., Wong, N., Thomas, J., Talkington, K., Jungman, E. and Coukell, A. Antimicrobial drug use in food-producing animals and associated human health risks: What, and how strong, is the evidence? *BMC Veterinary Research*, 13, 211. (2017). https://doi.org/10.1186/s12917-017-1131-3
- Collineau, L., Carson, C. A. and Moreno, M. A. antimicrobial usage in companion and food animals: methods, surveys and relationships with antimicrobial resistance in animals and humans, volume II. *Frontiers in Veterinary Science*, 8, 728267. (2021). https://doi.org/10.3389/fvets.2021.728267
- Vittecoq, M., Godreuil, S., Prugnolle, F., Durand, P., Brazier, L., Renaud, N. and Renaud, F. Antimicrobial resistance in wildlife. *Journal of Applied Ecology*, 53(2), 519–529. (2016). https://doi.org/10.1111/1365-2664.12596
- 8. Naylor, N. R., Atun, R., Zhu, N., Kulasabanathan, K., Silva, S., Chatterjee, A. and & Knight, G. M. Estimating the burden of antimicrobial resistance: A systematic literature review. *Antimicrobial Resistance & Infection Control*, **7**,58(2018). https://doi.org/10.1186/s13756-018-0336-y
- Hernando-Amado, S., Coque, T. M., Baquero, F. and Martínez, J. L. Defining and combating antibiotic resistance from One Health and Global Health perspectives. *Nature Microbiology*, 4(9), 1432–1442. (2019). https://doi.org/10.1038/s41564-019-0503-9
- World Health Organization, Food and Agriculture Organization of the United Nations, United Nations Environment Programme, & World Organisation for Animal Health. A One Health priority research agenda for antimicrobial resistance. World Health Organization.(2023). https://www.who.int/publications/i/item/978924007592
- 11. European Medicines Agency, & European Food Safety Authority. Joint scientific opinion on measures to reduce the need to use antimicrobial agents in animal husbandry in the European Union, and the resulting

- impacts on food safety. EFSA Journal, 15(1), e04666. (2017). https://doi.org/10.2903/j.efsa.2017.4666
- 12. Food and Agriculture Organization of the United Nations. The FAO action plan on antimicrobial resistance 2021–2025. (2021). https://doi.org/10.4060/cb5545en
- 13. Godijk, N.G., Bootsma, M.C.J. and Bonten, M.J.M. Transmission routes of antibiotic resistant bacteria: a systematic review. *BMC Infect. Dis.*, **22**, 482 (2022). https://doi.org/10.1186/s12879-022-07360-z
- Adisasmito, W., Almuhairi, S., Behravesh, C. B., Bilivogui, P., Bukachi, S., Casas, N. and Rüegg, S. R. One Health: A new definition for a sustainable future. *PLOS Pathogens*, 18(6), e1010537. (2022). https://doi.org/10.1371/journal.ppat.1010537
- 15. Bharti, S. Harnessing the potential of bimetallic nanoparticles: Exploring a novel approach to address antimicrobial resistance. *World J. Microbiol. Biotechnol.*, **40**, 89 (2024). https://doi.org/10.1007/s11274-024-03923-1
- Murugaiyan, J., Kumar, P.A., Rao, G.S., Iskandar, K., Hawser, S., Hays, J.P., Mohsen, Y., Adukkadukkam, S., Awuah, W.A., Jose, R.A.M., et al. Progress in Alternative Strategies to Combat Antimicrobial Resistance: Focus on Antibiotics. *Antibiotics*. 11(2),200. (2022) https://doi.org/10.3390/antibiotics11020200
- Xie, M., Gao, M., Yun, Y., Malmsten, M., Rotello, V. M., Zboril, R. and Li, R. Antibacterial nanomaterials: mechanisms, impacts on antimicrobial resistance and design principles. *Angewandte Chemie International Edition*, 62(17), e202217345. (2023). https://doi.org/10.1002/anie.202217345
- Croissant, J.G., Butler, K.S. and Zink, J.I. Synthetic amorphous silica nanoparticles: toxicity, biomedical and environmental implications. *Nat Rev. Mater.*, 5, 886–909 (2020). https://doi.org/10.1038/s41578-020-0230-0
- Seghir, B. B., Hemmami, H., Hocine, B. M. E., Soumeia, Z., Sharifi-Rad, M., Awuchi, C. G. and Messaoudi, M. Methods for the preparation of silica and its nanoparticles from different natural sources. *Biological Trace Element Research*, 201(12), 5871-5883 (2023). https://doi.org/10.1007/s12011-023-03628-w
- Almutairi, B. A. A. ., Almotawa, A. A. ., Almutairi, M. A. M., Baabbad, R. S. ., Abdullah S, A. A. ., Alagedi, H. S. K. ., Aljaizani, E. A. H. ., Alahmar, M. A. ., Shabi, F. A. A. ., Alharbi, S. A. ., Alaboudi, I. S. A., Kaabi, A. A. and Saleh A, A. A. . Ciprofloxacin: An Overview of Uses, Mechanism of Action, and Adverse Effects. *Journal of Ecohumanism*, 3(8), 9328 (2024). https://doi.org/10.62754/joe.v3i8.5546
- 21. Shariati, A., Arshadi, M., Khosrojerdi, M. A., Abedinzadeh, M., Ganjalishahi, M., Maleki, A. and Khoshnood, S. The resistance mechanisms of bacteria against ciprofloxacin and new approaches for enhancing the efficacy of this antibiotic. *Frontiers in Public Health*, **10**, 1025633. (2022). https://doi.org/10.3389/fpubh.2022.1025633

- Alandiyjany, M. N., Abdelaziz, A. S., Abdelfattah-Hassan, A., Hegazy, W. A., Hassan, A. A., Elazab, S. T. and Ibrahim, D. Novel in vivo assessment of antimicrobial efficacy of ciprofloxacin loaded mesoporous silica nanoparticles against salmonella typhimurium infection. *Pharmaceuticals*, 15(3), 357 (2022). https://doi.org/10.3390/ph15030357
- 23. Hetta, H. F., Ramadan, Y. N., Al-Harbi, A. I., A. Ahmed, E., Battah, B., Abd Ellah, N. H. and Donadu, M. G. Nanotechnology as a promising approach to combat multidrug resistant bacteria: a comprehensive review and future perspectives. Biomedicines, 11(2), 413 (2023). https://doi.org/10.3390/biomedicines11020413
- 24. Khan, I., Saeed, K. and Khan, I. Nanoparticles: Properties, applications and toxicities. *Arabian Journal of Chemistry*, 12(7), 908-931(2019). https://doi.org/10.1016/j.arabjc.2017.05.011
- 25. Caneschi, A., Bardhi, A., Barbarossa, A. and Zaghini, A. (2023). The use of antibiotics and antimicrobial resistance in veterinary medicine, a complex phenomenon: A narrative review. *Antibiotics*, 12(3), 487. (2023). https://doi.org/10.3390/antibiotics12030487
- 26. Ghobadi, M., Salehi, S., Ardestani, M. T. S., Mousavi-Khattat, M., Shakeran, Z., Khosravi, A. and Zarrabi, A. Amine-functionalized mesoporous silica nanoparticles decorated by silver nanoparticles for delivery of doxorubicin in breast and cervical cancer cells. *European Journal of Pharmaceutics and Biopharmaceutics*, 114349. (2024). https://doi.org/10.1016/j.ejpb.2024.114349
- Salam, M. A., Al-Amin, M. Y., Salam, M. T., Pawar, J. S., Akhter, N., Rabaan, A. A. and Alqumber, M. A. Antimicrobial resistance: a growing serious threat for

- global public health. In *Healthcare*, **11**(13), 1946). (2023).. https://doi.org/10.3390/healthcare11131946
- 28. Aljeldah, M. M. Antimicrobial resistance and its spread is a global threat. *Antibiotics*, **11**(8), 1082 (2022). https://doi.org/10.3390/antibiotics11081082
- Wood, S. J., Kuzel, T. M. and Shafikhani, S. H. Pseudomonas aeruginosa: infections, animal modeling, and therapeutics. *Cells*, 12(1), 199 (2023). https://doi.org/10.3390/cells12010199
- Anand, U., Carpena, M., Kowalska-Góralska, M., Garcia-Perez, P., Sunita, K., Bontempi, E. and Simal-Gandara, J. Safer plant-based nanoparticles for combating antibiotic resistance in bacteria: A comprehensive review on its potential applications, recent advances, and future perspective. *Science of The Total Environment*, 821, 153472 2(2022). https://doi.org/10.1016/j.scitotenv.2022.153472
- AlQurashi, D. M., AlQurashi, T. F., Alam, R. I., Shaikh, S. and Tarkistani, M. A. M. Advanced Nanoparticles in Combating Antibiotic Resistance: Current Innovations and Future Directions. *Journal of Nanotheranostics*, 6(2), 9 (2022). https://doi.org/10.3390/jnt6020009
- 32. Oliveira, M., Antunes, W., Mota, S., Madureira-Carvalho, Á., Dinis-Oliveira, R. J. and da Silva, D. D. An overview of the recent advances in antimicrobial resistance. *Microorganisms*, **12**(9), 1920 (2024). https://doi.org/10.3390/microorganisms12091920
- 33. Abdel Hamid, E. M., Ismail, M., Moussa, M., Abdel Aziz, M., Sobhy, M., Mohamed, R. and Mohamed, Y. Optimization and characterization of bio-silica extraction from rice straw using RSM. *Egyptian Journal of Chemistry*, **66**(11), 373-383 (2023). https://doi.org/10.21608/ejchem.2023.161205.6968

تعزيز التأثير المضاد للميكروبات للسيبروفلوكساسين عن طريق التحميل بجسيمات السيليكا النانوية المشتقة من قش الأرز: دراسات طيفية ومورفولوجية

رانيا م. الوكيل 1 ، محمد مصطفى عبد العال 3 و علا محمد البردي 4

- أ قسم الكيمياء، كلية العلوم والهندسة، الجامعة الأمريكية بالقاهرة، مصر.
- 2 كلية التكنولوجيا الحيوية، جامعة أكتوبر للعلوم الحديثة والأداب، مدينة السادس من أكتوبر، مصر.
 - ³ قسم جراحة القلب والصدر ، كلية الطب، جامعة كفر الشيخ، مصر .
 - ⁴ معهد علوم وتكنولوجيا النانو، جامعة كفر الشيخ، كفر الشيخ، مصر

الملخص

تشكل مقاومة مضادات الميكروبات تهديدًا كبيرًا لصحة الإنسان والحيوان مما يستدعي اتباع نهج الصحة الواحدة في الاستراتيجيات العلاجية, تُعقد المقاومة المتزايدة لهذه المُمْرضات ، مثل عدوى السالمونيلا، وعدوى البروسيلا، وعدوى الإشريكية القولونية والمكورات العنقودية الذهبية المقاومة للعلاج وترفع تكاليف الرعاية الصحية، مما يُبرز الحاجة المُلِحة لاستراتيجيات علاجية القولونية والمكورات العنقودية الذهبية المقاومة للعلاج وترفع تكاليف الرعاية الصحية، مما يُبرز الحاجة المُلِحة لاستراتيجيات علاجية الالله المنتخدام طريقة الاستخلاص الحمضي، ثم تم تحميل السيبروفلوكساسين، أحد أكثر المضادات الحيوية واسعة الطيف انتشارًا واستخدامًا عالميًا، في جسيمات نانوية من السيليكا (SiO2-NPs) وقد أكّد التوصيف الشامل لتقنيات المجهر الإلكتروني النافذ المسلمة والمجهر الإلكتروني النافذ المسلمة والمجهر الإلكتروني النافذ الجسيمات النانو من السيليكا (SiO2-NPs) وفعالية دمج الدواء. أظهر تحليل المجهر الإلكتروني النافذ الجسيمات النانو من السيليكا (SiO2-NPs) عير المُحمّلة بالسيبروفلوكساسين (SiO2-NPs) المُحمّلة بالسيبروفلوكساسين المضادة المبكتيريا ضد البكتيريا موجبة وسالبة الجرام التي غالبًا ما ترتبط بأمراض حيوانية فعالية هذه الجسيمات النانوية المضادة للبكتيريا ضد البكتيريا الموجبة وسالبة الجرام التي غالبًا ما ترتبط بأمراض حيوانية المنشأ. وكشفت الناتائج أن جسيمات أكسيد السيليكون النانوية المحملة بالسيبروفلوكساسين أظهرت نشاطًا مضادًا البكتيريا يغوق السيبروفلوكساسين الحر، وخاصة ضد سلالات البكتيريا الموجبة الجرام. علاوة على ذلك، أثبتت الدراسة فعالية جسيمات أكسيد السيليكون النانوية المشتقة من قش الأرز كنهج ناقل نانوي صديق للبيئة لتحسين توصيل المضادات بعسيمات أكسيد السيليكون النانوية المشتقة من قش الأرز كنهج ناقل نانوي صديق للبئة التحسين توصيل المضادات المحيوية، مما يغزز إدارة العدوى البكتيرية في الطب البيطري، ويقلل من خطر انتقال الأمراض حيوانية المنشأ إلى البشر.

الكلمات الدالة: مقاومة المضادات الحيوية، نهج الصحة الواحدة، جسيمات السيليكا النانوية، قش الأرز، سيبروفلوكساسين الأمراض الحيوانية المنشأ.