



Menoufia Veterinary Medical  
Journal  
(MVMJ)

## Menoufia Veterinary Medical Journal

<https://vmmj.journals.ekb.eg>

MVMJ. Vol. 1 (1)



### Fungal Contamination of Milk and Some Cheese Varieties Retailed in Markets of Menoufia Governorate

<sup>1</sup> Abd Elrahman Elbagoury.<sup>2</sup> Marwa Mohammed Awad.<sup>3</sup> Fatma Hussein

<sup>1</sup> Elbagoury, Abd Elrahman Professor of Milk Hygiene and Control Department, Faculty of Veterinary Medicine .Menuofia University <sup>2</sup>Mohammed, Marwa Awad . Professor of milk Hygiene and Control Department Faculty of Veterinary Medicine shebien Elkome; EL Monofia University <sup>3</sup> Hussein, Fatma Department of Dairy and Dairy Products Health Control, Faculty of Veterinary Medicine, Menoufia University,

#### ABSTRACT

##### Key words:

Tallaga cheese, processed cheese, Ras cheese, Yeast, Moulds

[\\*Correspondence to](#)

Vet.fatma95@gmail.com

##### Article History

Received: 26 Nov 2024

Accepted: 10 Feb 2025

The goal of this research was isolation and identification of yeast and moulds some types of cheese, in raw milk. One hundred and twenty samples have been gathered from Menoufia Governorate, Egypt involving; 30 raw milk, 30 processed cheese 30 tallaga cheese and 30 Ras cheese. The outcomes stated which, total frequency probabilities of moulds within raw milk, tallaga, processed, and ras cheese were; 24/30 (80%), 12/30 (40%), 25/30 (83.3%) and 28/30 (93.3%) respectively with mean mould counts  $2.7 \times 10^3 \pm 1.6 \times 10^3$ ,  $3.4 \times 10^2 \pm 2.6 \times 10^2$ ,  $5.9 \times 10^3 \pm 1.3 \times 10^3$  and  $5.4 \times 10^3 \pm 1.6 \times 10^3$  cfu/g, respectively. Penicillium spp. and aspergillus spp. were the mould species that were isolated from these samples. While, the yeast contamination has been identified in Raw milk, processed, Ras and Tallaga cheese have been contaminated with yeast at percent of 66.6, 60, 33.3 and 80% respectively with mean count values of  $45.6 \times 10^2 \pm 8.4 \times 10^2$ ,  $7.6 \times 10^2 \pm 2.3 \times 10^2$ ,  $5.6 \times 10^2 \pm 1.2 \times 10^2$  and  $13.8 \times 10^3 \pm 3.4 \times 10^3$ , respectively. Sacharomyces cervisie and Candida tropicalis were the most frequently isolated yeast species from the samples. In conclusion, the presence of high frequency of fungi in cheese and raw milk samples could poses potential public health risks and may result in economically detrimental outcomes because of product spoilage consequently, strict hygienic measures are required to be applied through the chain of production of such products.

#### 1. Introduction

Milk and its products are being nutritious food for human and widely regarded as optimal environments for growth of various fungal species, as they contain favorable conditions and essential nutrients necessary for growth of fungi [1]. The presence of fungi in raw milk is affected by factors such as the physiological condition of the animal, its breeding circumstances, and the environmental conditions throughout storage, milking, and other pre-processing stages Additionally, the sanitary quality of both the cheese and milk was identified by the fungal count. [2, 3]. Growth of fungi on cheese is a commonly occurred during the ripening process and on surface of hard cheese when it is stored in the

refrigerator by both retailers and consumers [4]. The impact of these microorganisms on the biochemical characteristics of dairy products and milk results in their spoilage, which is characterized by a variety of metabolic by-products. These by-products cause undesirable flavors, odors, and distinct changes in color, texture, and appearance, making them commercially not desired and frequently leading to a decrease in the evaluating of the dairy product. The proliferation of certain yeasts in cheese produced yeasty flavors through the production of alcohol and carbon dioxide, while some yeasts generate sulfides that produce an egg-like odor. Conditions such as the presence of low pH and oxygen levels create an environment conducive to mold growth,

particularly in cheese; however, certain molds can thrive even in low-oxygen conditions [5]. Identification and characterization of fungi found in cheese and raw milk was carried out by analyzing their phenotypic features, which were determined by their gross cultural and microscopic characteristics, as well as their genotypic characters [6]. The health of animals and humans can be negatively impacted by certain molds, that are capable of producing mycotoxins, which are fungal 2<sup>ry</sup> metabolites. These mycotoxins may go in animal and human food chain through either indirect or direct contamination [7].

Mycotoxins, particularly aflatoxins might be contaminated milk and its products s either directly by accidental growth of fungi from unhygienic media of processing or the usage of polluted fungal starter cultures that used in manufacturing and then toxins production or indirectly by ingestion of contaminated feed by AFB1 which excreted into milk as AFM1 [8].

There are approximately 20 varieties of aflatoxins, which belong to a broad category of toxic compounds. Among these, only four types—AFB1, AFG1, AFB2, and AFG2—are known to cause natural contamination of food products. Additionally, AFM1 can be present in milk and its derivatives as hydroxyl metabolites, resulting from the consumption of feed contaminated with these toxins by dairy animals [10].

Therefore, objective of this investigation was to detect and isolate fungal contamination within some types of cheese and raw milk samples.

## 2. Material and methods:

### 2.1. Collection of samples:

A total of 100 and twenty samples of raw milk, processed, Tallaga, and Ras cheese have been gathered from supermarkets and dairy shops in the Menoufia Governorate. The samples have been transported to the laboratory in sterile, airtight jars with minimal delay to be exposed and prepared to a mycological investigation.

### 2.2. Preparation of cheese samples:

Ten grams of each cheese sample and ten milliliters of raw milk were homogenized in a stomacher bag (Lab-blender 400, Seward, UAC House Friars Road, London SE19UG. Model No. 6021) with ninety milliliters of sterile 0.2 percent sodium citrate solution. To achieve a dilution of 102, one milliliter of the original sample homogenate was added to a test tube that contained nine milliliters of 0.1% sterile peptone water. The same procedure was followed for preparing a ten-fold serial dilution [11].

### 2.3. Enumeration, Isolation and determination of fungi:

The characterization of different yeast and mould colonies was applied by microscopical examination according to [12, 13].

### 2.4 Statistical analysis:

The SPSS program was utilized to conduct a statistical analysis. The outcomes were presented as the mean of 3 readings with the standard error, regarding [14].

## 3. Results:

**Table (1): Statistical analytical results of molds count of examined samples (Total mold count per gram). Number=30**

| Samples Types    | Total number of studied samples | Positive sample number | %    | Mean ± SE (colony-forming units per milliliter or gram) |
|------------------|---------------------------------|------------------------|------|---------------------------------------------------------|
| Raw milk         | 30                              | 24                     | 80   | $2.7 \times 10^3 \pm 1.6 \times 10^3$                   |
| Tallaga cheese   | 30                              | 12                     | 40   | $3.4 \times 10^2 \pm 2.6 \times 10^2$                   |
| Processed cheese | 30                              | 25                     | 83.3 | $5.9 \times 10^3 \pm 1.3 \times 10^3$                   |
| Ras cheese       | 30                              | 28                     | 93.3 | $5.4 \times 10^3 \pm 1.6 \times 10^3$                   |

**Table (2): Incidence and statistical analytical outcomes of total yeast count per gram for examined samples (Number=30)**

| Samples Types    | Total number of studied samples | Positive sample number | %    | Mean $\pm$ SE (colony-forming units per milliliter or gram) |
|------------------|---------------------------------|------------------------|------|-------------------------------------------------------------|
| Raw milk         | 30                              | 20                     | 66.6 | $45.6 \times 10^2 \pm 8.4 \times 10^2$                      |
| Tallaga cheese   | 30                              | 18                     | 60   | $7.6 \times 10^2 \pm 2.3 \times 10^2$                       |
| Processed cheese | 30                              | 10                     | 33.3 | $5.6 \times 10^2 \pm 1.2 \times 10^2$                       |
| Ras cheese       | 30                              | 24                     | 80   | $13.8 \times 10^3 \pm 3.4 \times 10^3$                      |

**Table (3) Frequency of fungal isolates in the examined samples (Number =30)**

| Type of moulds                 | Raw milk |       | Tallaga cheese |       | Processed cheese |       | Ras cheese |       |
|--------------------------------|----------|-------|----------------|-------|------------------|-------|------------|-------|
|                                | No.      | %     | No.            | %     | No.              | %     | No.        | %     |
| <i>Aspergillus flavus</i>      | 8        | 26.6  | 5              | 16.6  | 4                | 13.3  | 15         | 50    |
| <i>Aspergillus niger</i>       | 2        | 6.6   | -----          | ----- | 1                | 3.3   | 4          | 13.3  |
| <i>Aspergillus fumigatus</i>   | -----    | ----- | 2              | 6.6   | 2                | 6.6   | 1          | 3.3   |
| <i>Geotrichum candidum</i>     | 3        | 10    | -----          | ----- | -----            | ----- | 1          | 3.3   |
| <i>Mucor spp</i>               | -----    | ----- | -----          | ----- | -----            | ----- | 2          | 6.6   |
| <i>Penicillium chrysogenum</i> | 2        | 6.6   | -----          | ----- | -----            | ----- | 1          | 3.3   |
| <i>Penicillium hemtrachum</i>  | -----    | ----- | 2              | 6.6   | 4                | 13.3  | -----      | ----- |
| <i>Penicillium caseifulvum</i> | 2        | 6.6   | -----          | ----- | 2                | 6.6   | 1          | 3.3   |
| <b>Type of yeast</b>           |          |       |                |       |                  |       |            |       |
| <i>Candida albicans</i>        | -----    | ----- | 2              | 6.6   | -----            | ----- | 4          | 13.3  |
| <i>Rhodotorula spp.</i>        | 5        | 16.6  | -----          | ----- | -----            | ----- | -----      | ----- |
| <i>Sacharomyces cervisieoe</i> | 2        | 6.6   | 1              | 3.3   | 2                | 6.6   | 3          | 10    |
| <i>Candida tropicalis</i>      | 2        | 6.6   | 1              | 3.3   | 1                | 3.3   | 1          | 3.3   |

**Table (4): Quality of studied samples regarding EOSQC (2005)**

| Examined samples | Standards according to EOSQC (2005) |      |              |      |                                                |      |              |      |
|------------------|-------------------------------------|------|--------------|------|------------------------------------------------|------|--------------|------|
|                  | Moulds $\leq 10$ cfu/g              |      |              |      | Yeast $\leq 400$ colony-forming units per gram |      |              |      |
|                  | Acceptable                          |      | Unacceptable |      | Acceptable                                     |      | Unacceptable |      |
|                  | No./30                              | %    | No./30       | %    | No./30                                         | %    | No./30       | %    |
| Raw milk         | 6                                   | 20   | 24           | 80   | 10                                             | 33.4 | 20           | 66.6 |
| Tallaga cheese   | 18                                  | 60   | 12           | 40   | 12                                             | 40   | 18           | 60   |
| Processed cheese | 5                                   | 16.7 | 25           | 83.3 | 20                                             | 66.7 | 10           | 33.3 |
| Ras cheese       | 2                                   | 6.7  | 28           | 93.3 | 6                                              | 20   | 24           | 80   |

#### 4. Discussion:

The contamination of cheese and raw milk by fungi is a consequence of the unhygienic conditions that happen throughout their manufacturing, transportation, production, and storage periods. Additionally, fungal contamination can be the consequence of contaminated air, unhygienic measures in certain factories, or storage. [15]. The provision of all essential nutrients and conditions for development of fungi is the reason why cheese and raw milk are generally regarded as a perfect growth medium for numerous fungal species [16].

Within the present research a total of 120 samples (30 raw milk, 30 processed cheese, 30

tallaga cheese, and 30 ras cheese) obtained from Menoufia Governorate, Egypt. Have been analyzed mycological. The outcomes showed in **Table (1)** discovered which, total frequency rate of the mould in tallaga, raw milk, processed and ras cheese samples were; 24/30 (80%), 12/30 (40%), 25/30 (83.3 %) and 28/30 (93.3%) respectively with mean mould counts  $2.7 \times 10^3 \pm 1.6 \times 10^3$ ,  $3.4 \times 10^2 \pm 2.6 \times 10^2$ ,  $5.9 \times 10^3 \pm 1.3 \times 10^3$  and  $5.4 \times 10^3 \pm 1.6 \times 10^3$  cfu/g. An insignificant variance in the mean mold counts between processed and ras cheeses.

Mold contamination in dairy products serves as an indicator of sanitary quality conditions [17]. The distribution of isolated mold species from the positive samples is shown in **Table (2)**

In raw milk *Aspergillus flavus* is the most isolate (8/30, 26.6%), *Geotrichum candidum* (3/30, 10%) and *Penicillium* spp. (4/30, 10.2%) while In Tallaga cheese, the most prevalent isolates were *Aspergillus flavus* (5/30, 16.6%), *Aspergillus fumigatus* 2/30, 6.6%), and *Penicillium hemtrachum* (2/30, 6.6%). In processed cheese, the most common mold isolates were *Penicillium* spp. (6/30, 19.9%) and *Aspergillus* spp. (7/30, 23.2%). In ras cheese, the most abundant isolates were *Aspergillus* spp. (20/30, 66.6%), *Penicillium* spp. (1/30, 3.3%), *Mucor* spp (2/30, 6.6 %) and *Geotrichum candidum* (1/30, 3.3%). Similar findings were reported by [18], he identified *Aspergillus* spp., *Geotrichum* spp., *Penicillium* spp., and *Fusarium* spp. as the primary fungi responsible for mycotoxin contamination in cheese and milk. [19] found that the most prevalent genera among the four types of cheese samples examined were *Aspergillus* and *Penicillium* species. Additionally, [20] noted that *Penicillium* was the most common mold genus found in cheese samples, after that *Aspergillus*, *Geotrichum*, *Cladosporium*, and *Mucor* species.

The outcomes documented within **Table (3)** was noted that the studied samples of Tallaga, Raw milk, Ras and processed cheese have been contaminated with yeast at percentage of 60, 66.6, 80 and 33.3% respectively with mean count values of  $7.6 \times 10^2 \pm 2.3 \times 10^2$ ,  $45.6 \times 10^2 \pm 8.4 \times 10^2$ ,  $13.8 \times 10^3 \pm 3.4 \times 10^3$ , and  $5.6 \times 10^2 \pm 1.2 \times 10^2$  respectively. [21] noticed a nearly identical frequency of yeast in Tallaga cheese samples. They analyzed fifty soft white cheese samples mycologically and determined which frequency of yeasts was sixty. [22] and [23] discovered greater outcomes, whereas [24] and [25] noticed fewer frequency and counts. [26] and [27] reported a relatively fewer frequency and counts in processed cheese. However, [28] achieved a more favorable result. [29] reported fewer frequency and counts of Ras cheese, while [30] and [19] reported a greater frequency and counts. Yeasts and mold count in cheese and raw milk are utilized as an indicator of the quality of sanitation. These counts could be the outcome of contaminated air, a change in season, a poor environment in certain factories, unregulated sterilization, the utilization of milk of inferior

quality, equipment, and storage, unsensitized instruments, and other factors.

The most predominant yeast species as shown in **Table 2** were isolated from raw milk were *Rhodotorula* 5 (16.6%), *Sacharomyces* 2 (6.6%) and *Candida tropicalis* 2 (6.6%). While in Tallaga cheese were *Candida albicans* 2 (6.6%) *Sacharomyces* 1(3.3%) and *Candida tropicalis* 1 (3.3%). In processed cheese only *Sacharomyces* 2(6.6%) and *Candida tropicalis* 1 (3.3%). [31] achieved outcomes that were nearly identical. In the processed cheese samples that were investigated. *Candida albicans* is the most type of yeast in Ras cheese 4(13.3%), followed by *Sacharomyces* 3 (10%) and *Candida tropicalis* 1 (3.3%) lower than result recorded by [32].

Regarding the outcomes in **Table (4)**, 80, 83.3, 40, and 93.3% of the studied raw milk, processed, Tallaga, and Ras cheese samples, respectively had molds more than the acceptable limit (undesirable) and in comparison, to EOSQC (2005), the yeast counts were 66.6, 60, 33.3, and 80 percent higher. Such numbers are considered undesirable. Certain food borne yeasts & molds may be hazardous due to their potential to allergic reactions [33]. Additionally, the proliferation of fungi on cheese can lead to discoloration and undesirable flavors, as well as the production of mycotoxins, which have been linked to numerous instances of mycotoxicosis [34].

## Conclusion

This study has clearly demonstrated the presence of fungal contamination in cheese samples and raw milk obtained from markets in the Menoufia Governorate, Egypt. This finding suggests inadequate hygienic practices during the processes of milk production, cheese & handling manufacturing, storage and ripening process. The predominant mold species identified have been *Penicillium* and *Aspergillus* spp. The proliferation of these fungi poses potential health risks to customers. Consequently, it is imperative to implement stringent hygienic measures throughout production of milk, cheese processing, handling, and storage, ensuring that final products adhere to Egyptian standar.

## 5. References

1. **Gulbe, G., Valdovska, A. (2014):** Diversity of Microscopic Fungi in the Raw Milk from Latvian Organic farm. *Proc. Latv. Univ. Agr.*, 31(1), 46-53.
2. **Callon, C., Duthoit, F., Delbes, C., Ferrand, M., Le Frileux, Y., De Cremoux, R., Montel, M. C. (2007):** Stability of microbial communities in goat milk during a lactation year: approaches. *Systematic and Applied Microbiology*, 30, 547-560.
3. **Moubasher, A. A. H., Abdel-Sater, M. A., Soliman, Z. S. M. (2018):** Yeasts and filamentous fungi associated with some dairy products in Egypt. *Journal de Mycologie Medicale* 28(1),76-86.
4. **Gandomi, H., Misaghi A., Basti A. A., Bokaei S., Khosravi A., Abbasifar A. and Javan A. J. (2009):** Effect of Zataria multiflora Boiss. essential oil on growth and aflatoxin formation by *Aspergillus flavus* in culture media and cheese., *Food and Chemical Toxicology* 47: 2397-2400.
5. **Ledenbach, L. H., Marshall, R. T. (2009):** Microbiological Spoilage of Dairy Products. *Compendium of the microbiological spoilage of food and beverages*. Pp.41-67.
6. **El-Fadaly H. M., El-Kadi S. M., Hamad M. N. and Habib A. A. (2015)** Isolation and Identification of Egyptian Ras Cheese (Romy) Contaminating Fungi during Ripening Period. *Journal of Microbiology Research*, 5 (1): 1-10.
7. **Bohra, N. K. and Purohit, D. K. (2003):** Fungal toxicity with special reference mycotoxins. *Journal of Environmental biology*, 24(3):213- 221.
8. **Sengun, I., Yaman, D., Gonul, S. (2008):** Mycotoxins and mould contamination in cheese. *World Mycotox. J.* 3, 291-298
9. **Marin, S., Ramos, A. J., Cano-Sancho, G., Sanchis, V. (2013):** Mycotoxins, occurrence, toxicology, and exposure assessment. *Food and Chemical Toxicology* 60, 218-237.
10. **Dohnal, V., Wu, Q. and Kuca, K. (2014):** Metabolism of aflatoxins, Key enzymes and interindividual as well as interspecies differences. *Arch. Toxicol.* 88, 1635-1644.
11. **APHA (American Public Health association) 2003:** Compendium of methods for the microbiological examination of foods. Third Ed. (Vanderzant, C and SPlittoesser, D. Eds) Washington DC, USA, 675-8000
12. **Pitt JI, Hocking AD. (1999):** Fungi and food spoilage. Second ed. Gaithersburg: Aspen Publishers
13. **Manga M., Bansa S. and Sharma M. (2014):** Macro and micro morphological characterization of different *Aspergillus* isolates. *Legume Res.*, 37 (4): 372-378.
14. **Verma, J.P., (2012):** Data analysis in management with SPSS software. Springer Science & Business Media.
15. **Younis, G., Ibrahim, D., Awad, A., El Bardisy, M.M. (2016):** Determination of aflatoxin M1 and ochratoxin A in milk and dairy products in supermarkets located in Mansoura City, Egypt. *Adv. Anim. Vet. Sci.* 4, 114-121.
16. **Gulbe, G., Valdovska, A. (2014):** Diversity of Microscopic Fungi in the Raw Milk from Latvian Organic farm. *Proc. Latv. Univ. Agr.*, 31(1), 46-53
17. **Varga, L. (2007):** Microbiological quality of commercial dairy products, pp. 487- 494. In *Communicating current research and educational topics and trends in applied microbiology*. Formatex microbiology Series, Formatex Badajoz, 2007. ISBN 978-84-611-9422-3.
18. **Silva, J. L.; Aparecido, C. C.; Hansen, D.; Pereira, T. A. M.; Felicio, J. D. and Goncalez, E. (2015):** Identification of toxigenic *Aspergillus* species from diet dairy goat using a polyphasic approach. *Ciência Rural*, Santa Maria,45(8): 1466-1471.
19. **EL-Bagory, A. M., Amal, E. M., Hammad, A. M., Salwa, A. D. (2014):** prevalence of fungi in locally produced cheese and molecular characterization of isolated toxigenic molds. *BenhaVet. Med. Journal* 27(2), 9-20
20. **Finne Kure, C.; Skaar, I. and Brendehaug, J. (2004):** Mould contamination in production of semi-hard cheese. *International Journal of Food Microbiology*, 93: 41-49.
21. **El Sayed, M. A., Hosny I. M., El Kholy W.I., El-Dairouty, R.K. and Sahar, H.S. (2011):** Microbiological evaluation of

- Egyptian white soft cheeses style. Journal of American Science, 7(5):517- 526
22. **Hegazy, M.I. and Mahgoub, S.A. (2013):** Microbiological characterization of the Egyptian soft white cheese and identification of its dominant yeasts. African journal of microbiology research, 7(20): 2205-2212.
23. **Salih, A. Zakaria, Sulieman, E. Abdel Moneim, Elkhalfa, A. Elamin and Ali, O., Ali, (2012):** Chemical and Microbiological Characteristics of White Cheese (Jibna-beida) Produced in Sudan. Food and Public Health 2012, 2(6): 259-264.
24. **Aly, A. Salwa, Farag, D. E. and Galal, E. (2012):** Effect of Gamma Irradiation on the Quality and Safety of Egyptian Karish Cheese. Journal of American Science, 8(10):761-766.
25. **Hakim, A.S., Abuelnaga, A.S.M., and Sayed El Ahl, R.M.H. (2013):** Isolation, Biochemical Identification and Molecular Detection of Yeasts from Kareish Cheese. International Journal of Microbiological Research, 4(1):95-100
26. **Hussein, F.S.E.; ElZubeir, I.M., Fadelel and Moula, A.A. (2011):** Quality evaluation of imported and locally produced processed cheese in Sudan. Jordan journal of biological science, 4:231-236.
27. **EL-Shibiny, Safinaz.; Abd El-Gawad, A., M., Mona; Assem, M., Fayza; Seleet, L. Faten; Abou Dawood, A.; Shireen and Elaaser, M. (2013):** Preparation, composition and microbiological and rheological properties of functional processed cheese supplemented with rice bran. Journal of Applied Sciences Research, 9(8): 4927-4934.
28. **Nour El Diam, M. S. A. and El Zubeir, I. E. M. (2006):** Comparison of microbiological quality of processed and non-processed Sudanese white cheese. Res. J. Microbiol. 1(3):273-279.
29. **Torkar, G. K. and Teger, S., G. (2006):** The presence of some pathogenic microorganisms, yeast and mould in cheese samples produced at small dairy processing. Acta agriculture Slovenica 88(1):37-51
30. **Sadek, I., Zeinab; Hosny, I. M.; El-Kholy, W. I.; and El-Dairouty, R. K. (2009):** Comparative investigations for detection of food born microorganisms in Egyptian hard cheese "Ras" using conventional and fast biochemical tests. Global veterinaria3 (3):189-195
31. **EL-Shafei, Kawther; Abd EL-Gawad, A.M. Mona; Dabiza, Nadia; Sharaf, O.M.; Effat, B.A. (2008):** A mixed culture of propioni bacterium Thoenii p-127, *Lactobacillus Rhamnosus* and *Lactobacillus plantarum* as protective cultures in Kareish cheese. Pol. J. Food Nutri. Sci., 58(4): 433-441
32. **Spanamberg, A., Fraga, C.F., Ferreiro, L., Aguinisky, M.S., Sanches, E.M.C., Roehe, C., Lautert, C., Santurio, J. M. (2014):** Yeasts in the raw ewe's milk. Acta Scientiae Veterinariae42,1236-1240
33. **Mislivec, P.B., Beuchat, L.R., Cousin, M. A. (1992):** Yeasts and molds. Chapter 16. In: Compendium of methods for microbiological examination of foods. 3rd ed. APHA, Washington DC.
34. **Neal, G.E., Eaton, D.L., Judah, D.J., Verma, A. (1998):** Metabolism and toxicity of aflatoxin M1 and B1 in human-derived in vitro systems. *Toxicol. Appl. Pharmacol.* 151:152-158