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### RESEARCH ARTICLE

### Evaluation of Hygienic Status of Some Meat Products Retailed in Zagazig City, Egypt

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### **ABSTRACT**

This study aimed to assess the underexplored hygienic status of marketed meat products (minced meat, sausage, and luncheon) in Zagazig city, Sharkia Governorate, Egypt. Sixty meat product samples from different hypermarkets were analyzed for aerobic plate count (APC), Psychrotrophic bacteria count (PBC), coliforms, S. aureus and mold counts. The prevalence was 100% for APC, and PBC in all examined meat product samples, 100% for coliforms in minced meat, and sausage and 50% in luncheon samples, 80%, 90%, and 60% respectively for S. aureus in the examined meat product samples, meanwhile 100% for mold in all meat product samples. The mean counts of APC (4.73±0.21, 5.25±0.21, and 4.06±0.17 log10 CFU/g), PBC (4.35±0.20, 4.90±0.21, and 3.57±0.10 log10 CFU/g), and S. aureus (2.74±0.11, 3.01±0.13, and 2.42±0.14 log10 CFU/g) were statistically different (P < 0.05) in all tested meat products. Similarly, the counts for coliform and mold showed significant differences between the three types of meat products. This suggests an increased risk of spoilage and pathogen contamination. In conclusion, the obtained data demonstrated that the hygienic precautions that were implemented during the manufacturing of such products were insufficient. Therefore, there is an immediate and pressing requirement for improved hygienic standards in meat processing factories and supermarkets in order to limit the amount of microbial contamination and protect the safety of consumers.

### Introduction

Meat serves an important source of high-quality protein and bioavailable vitamins, along essential minerals with like iron, zinc, and phosphorus [1]. Meat products are gaining popularity as a result of the fact that they can be easily and quickly prepared into meat meals, which can help in addressing the issue of a

shortage of fresh meat that is extremely expensive [2].

One of the most major problems that affects the quality of meat, and the health of the general public is the presence of microbiological contamination. is possible that the microbiological quality of beef products is impacted by a variety of factors, such as the lengthy chain of preparation, processing, distribution, storage, and retailing [3].

A number of different methods, including aerobic plate count (APC). total psychrotrophic counts. total Enterobacteriaceae counts, total coliforms, Staphylococcus, and mold, can be utilized to assess the level of bacterial contamination and sanitary measures that are implemented during the manufacture meat as well as negative storage conditions for meat products [4-6]. APC is a significant measure of sanitary quality of a product throughout entire process of processing distribution [7]. The APC level in meat is regulated or recommended by a number of countries, which allows them to control the meat hygiene. Within the year 1978, Commission International **Specifications** Microbiological Foods (ICMSF) the sole entity was that guideline established the APC which was set at  $1.0 \times 10^7$  CFU/g or cm<sup>2</sup> The permissible limits for total bacterial count in meat products vary depending on the type of product and its processing. Generally, raw meat products have higher acceptable limits than cooked or processed products. For example, raw poultry products may have a limit not exceeding 10<sup>5</sup> CFU/g, while heat-treated products might have a limit of CFU/g [9]. As a result of their capacity to thrive at low temperatures, psychrotrophic the primary bacteria are responsible for the deterioration of meat products that are stored at temperatures that are below the refrigeration threshold [10]. There is a correlation between high counts of psychrotrophic bacteria and an environment that is conducive development of illnesses [11].

Psychrotrophic bacteria are excellent markers of the hygienic state of meats at low temperatures and their potential for spoilage. Coliforms are types of bacteria that are found in the stomach, and the number of them can be used as a measure of hygienic conditions [12]. The permissible limits for total psychrotrophic

counts in meat products vary depending on the type of product and regulations, but generally, raw poultry products should not exceed 10<sup>5</sup> CFU/g, and heat-treated products should be below 10<sup>4</sup> CFU/g [13].

Escherichia coli (E. coli) is a Gramnegative, facultative, anaerobic bacteria that is regarded to be a commensal organism in the human body [14]. Consumers experience may poisoning if proper cooking procedures are not followed [15]. As of right now, E. isolates that cause diarrhea are coliprimary divided into six pathotypes. These pathotypes are distinguished from one another by their unique virulence pathogenic factors and characteristics [14]. The shiga toxin producing E. coli (STEC) group is considered to be one of the most significant pathotypes associated human infectious diseases. meat should also have acceptable levels of coliform bacteria, with some standards specifying limits of 10<sup>2</sup> CFU/g. Cooked or processed meats generally have stricter limits, often around 10<sup>4</sup> CFU/g for APC [15].

Staphylococcus aureus (S. aureus) is a opportunistic pathogen known spread through food and has been linked to multiple outbreaks in hospitals and communities worldwide Furthermore. S. frequently aureus colonizes human skin and can lead to opportunistic infections in the host, which can contaminate or re-contaminate cooked meals through the hands, tools, or utensils of workers [17]. The acceptable limit for aureus count in meat products generally less than 100 CFU/g according the International Microbiological to Criteria [18].

Mold is considered one of the causative agents of meat deterioration; it is responsible for the world's economic losses from food's production by 5-10% [19]. Meat products contaminated by mold were a sign of unhygienic and unsanitary handling, processing, and

storage conditions [4]. Heat-treated meat products should be free from molds [20].

In light of the previous facts, the current study was carried out with the purpose of assessing the microbiological condition of a selection of meat products including minced meat, sausage, and luncheon in Zagazig city. This was accomplished through the assessment of APC, mold, coliform, S. aureus and psycrotrophic bacterial counts.

#### Materials and methods

This conducted study was with Zagazig University's accordance and no living animals were procedures, used.

### Samples collection and preparation

A total of sixty samples of meat products meat, including minced sausage, luncheon, 20 each were obtained randomly from a variety of stores in Zagazig city, Sharkia Governorate, Egypt. For the purpose of conducting microbiological analysis, the samples were without delay and delivered to the Hygiene Laboratory, Faculty Veterinary Medicine, Zagazig University, Egypt.

### Microbiological analyses

accordance with In methodology outlined in ISO 6887-2 [21], samples of meat products and serial dilutions were created. In summary, twenty-five grams of each sample were meticulously blended using sterile homogenizer flask (Homogenizer type MPW-302, Poland) containing 225 milliliters of sterile peptone water (Hi media, India) diluted 0.1% to a dilution concentration. Α 1/10 produced homogenate was by homogenizing the 2.5 contents minutes at 5000 revolutions per minute. The homogenate was permitted to rest for roughly 5 minutes. Following the transfer of the homogenate into a sterile test tube containing nine milliliters of one percent tenfold serial dilutions peptone water,

were generated, achieving a concentration of 10<sup>6</sup> per milliliter.

The following analytical procedures were carried out on the samples that had been prepared: In accordance with ISO 4833-1 the APC was carried out [22],employing a sterile plate count agar and inoculating it at 37 °C for 24 hours. On plates that contained between 30 and 300 colonies, the APC computed per gram was determined.

For determination of the psychrotrophic counts [23], accurately 0.1 mL from each of the previously prepared serial dilution was spread over a dry surface of a plate count agar medium (Oxoid, UK) by a sterile bent glass spreader. The plates were incubated at 7 °C for 10 days with the control plates. together counted, the average colonies were number of colonies was recorded, and the psychrotrophic bacterial (CFU/g) was calculated.

After incubating the samples at 37 °C for 24-48 hours, the most probable number (MPN) of coliforms was determined by counting the number of coliforms that were incubated in three MacConkey broth tubes with inverted Durham's tube. It was determined that the generation of acid and gas in the inverted Durham's tubes was found to be positive and evaluated in accordance with the ICMSF [24].

The count of S. aureus was carried out using each of the serial dilutions that had been produced in advance. Using sterilized glass spreader, 0.1 mL from previously prepared serial dilutions was evenly distributed on Baired Parker agar plates. The plates were incubated at 37 °C for 48 hours. The shiny black colonies surrounded by a halo zone were counted per gram of sample [25].

After inoculating the agar at a temperature of 25 °C for five days, the total mold count was determined by using Dichloran Chloramphenicol Rose Bengal agar (Oxoid, UK) that contained chloramphenicol at a concentration 0.05 mg/mL [26]. A "star-shape" mold

growth was counted and recorded as the total mold count per gram of sample.

### Statistical analysis

Microbial counts were transformed into their logarithmic values (Log<sub>10</sub> CFU/g). One-way analysis was performed on the data that was acquired using the SPSS program (SPSS Inc., Chicago, 16.0 Illinois, United States of America). order to determine whether or not there were significant differences between the values, the Tukey's multiple mean comparison tests utilized. The were results were denoted as the mean plus the standard error (SE), and a significance level of P < 0.05was utilized determine the statistical significance [27].

### **Results**

In Table 1, the findings of the prevalence analysis APC, psychrotrophic, of coliform, S. and mold aureus, contamination in sixty samples are presented.

The results indicated that 100% of all examined meat products were positive for psychrotrophic bacteria, coliforms (except 50% for luncheon). It is important to note that both APC and PBC positive for all examined meat products, which indicated that aerobic and psychrotrophic bacteria were present rather high concentrations. Coliform detection was high in samples of minced beef and sausage (100%), but it was low in samples of luncheon (50%). Furthermore, the prevalence of S. aureus found to be slightly greater samples (90%)compared minced meat (80%), while it was found to

be lower in luncheon samples (60%). All tested meat product samples were positive to molds (100%).

The results demonstrated in Table revealed that the minimum and maximum APC were ranged from 3.79 to 5.71, 4.46, to 6.18, and 3.28 to 4.79 Log<sub>10</sub> CFU/g in and minced meat, sausage minimum respectively. The and maximum psychrotrophic were fluctuated from 3.45 to 5.38, 3.99 to 5.85 and 3.04 to 3.96 CFU/g in minced  $Log_{10}$ sausage and luncheon, respectively. The minimum and maximum coliforms were ranged from 2.04 to 3.79, 2.18 to 4.18 and 2.66 to 3.18 Log<sub>10</sub> CFU/g in the three examined meat products, respectively. Meanwhile, for S. aureus such values ranged from 2.30 to 3.30, 2.60 to 3.78 and 2.00 to 2.85 Log<sub>10</sub> CFU/g for minced meat, sausage and luncheon, respectively. The minimum and maximum total mold count was varied from 1.85 to 2.70, 2.00 to 3.30 and 2.00 to 3.60 Log<sub>10</sub> CFU/g in minced meat, sausage and luncheon, respectively.

The results of mean counts (Table 2) illustrated that the highest APC, PBC, coliforms, S. aureus, and mold counts were recorded in sausage samples  $(5.25\pm0.21,$  $4.90\pm0.21$ ,  $3.35\pm0.22$ ,  $3.01\pm0.13$ ,  $2.73\pm0.20$  $log_{10}$ CFU/g, respectively) and the lowest counts were obtained in minced meat luncheon for  $(4.73\pm0.21)$  $\log_{10}$  CFU/g), PBC  $(3.57\pm0.10)$ , coliforms  $(2.92\pm0.09)$ and S. aureus  $(2.42\pm0.14 \log_{10} \text{ CFU/g})$ , respectively, and minced meat for mold  $(2.22\pm0.18\log_{10} CFU/g)$ .

Table 1: Prevalence of aerobic plate count, psychrotrophic bacteria count, coliform, S. aureus and mold counts in different types of meat product samples (no=60).

Types of meat	Microbial agents	<b>Positive Samples</b>	
products		No.	%
Minced meat	APC	20	100
	PBC	20	100
	Coliforms	20	100
	S. aureus	16	80
	Mold	20	100
Sausage	APC	20	100
	PBC	20	100
	Coliforms	20	100
	S. aureus	18	90
	Mold	20	100
Luncheon	APC	20	100
	PBC	20	100
	Coliforms	10	50
	S. aureus	12	60
	Mold	20	100

No, Number of samples; APC, Aerobic plate count; PBC, Psychrotrophic bacteria count; S. aureus, Staphylococcus aureus.

Table 2: Statistical analytical results (log<sub>10</sub> CFU/g) of aerobic plate count, psychrotrophic bacteria count, coliform, *S. aureus* and mold counts in examined meat products samples (n=20 for each meat product).

		Microbial count		
Types of meat products	Microbial agents	Minimum (Log <sub>10</sub> CFU/g)	Maximum (Log <sub>10</sub> CFU/g)	Mean ±SE
Minced meat	APC	3.79	5.71	4.73±0.21 b
	PBC	3.45	5.38	$4.35\pm0.20^{b}$
	Coliforms	2.04	3.79	$3.04\pm0.18^{a}$
	S. aureus	2.30	3.30	$2.74\pm0.11^{ab}$
	Mold	1.85	2.70	$2.22 \pm 0.18^{b}$
Sausage	APC	4.46	6.18	5.25±0.21 a
	PBC	3.99	5.85	4.90±0.21 a
	Coliforms	2.18	4.18	$3.35\pm0.22^{a}$
	S. aureus	2.60	3.78	3.01±0.13 a
	Mold	2.00	3.30	$2.73\pm0.20^{a}$
Luncheon	APC	3.28	4.79	4.06±0.17 °
	PBC	3.04	3.96	3.57±0.10 °
	Coliforms	2.66	3.18	$2.92\pm0.09^{a}$
	S. aureus	2.00	2.85	$2.42\pm0.14^{\ b}$
	Mold	2.00	3.60	$2.83\pm0.19^{a}$

 $<sup>^{</sup>a,b,c}$  Means carrying different superscript letters within each microbial category are significantly different at P < 0.05 among the examined minced meat, sausage, and luncheon. No, Number of samples; APC, Aerobic plate count; PBC, Psychrotrophic bacteria count; S. aureus, Staphylococcus aureus.

### **Discussion**

such as Meat products minced meat, luncheon, and sausage may be contaminated at any stage of the processing, packing, and shipping processes with various pathogens. The meat products become hazardous customers and inappropriate for human consumption due to these pathogens. The hygienic status of these products can be assessed using a variety of indicators; Coliforms are frequently used to assess the safety and cleanliness of meat products [9, 28].

The microbiological quality and safety of meat in any country requires continual and regular monitoring and updating. It is also necessary for the industry to ensure that consumers are provided with meat and meat products that are of high quality and safe for consumption [29].

PBC, Aerobic plate count (APC), coliforms. Staphylococci, and mold counts were conducted in this study in evaluate microbiological the quality of meat product samples. These samples included minced meat, sausage, and luncheon. All of the samples demonstrated high levels of APC, with values being  $4.73\pm0.21$ , mean  $5.25\pm0.21$ , and  $4.06\pm0.17 \log_{10}$  CFU/g for the minced beef, sausage, and luncheon samples, respectively. It is of the utmost importance to evaluate the hygienic conditions under which a food item has been created, handled, and stored [30]. Although the APC of any food item is not a definitive indicator of whether or not it is safe for ingestion, APC is frequently utilized evaluate to the overall microbiological quality of meat as well as its shelf life [31, 32]. It additionally provides an indication of the hygienic quality of a product throughout the entire process of processing and distribution

The findings that were presented in Table 2 demonstrated that the APC results in minced meat agreed with the findings that were presented by Erdem-Ayten *et al.* 

[34]. On the other hand, researchers found lower levels in minced meat [35], as well luncheon as in sausage and [36]. minced However. meats had higher microbial previous research counts in [37]. Furthermore. aforementioned study indicated that the mean values  $(log_{10}$ CFU/g) of the APC/g were  $6.1\pm0.1$  in minced meat, 4.8±0.1 in sausage, and 4.2±.1 in luncheon [38]. A high APC could be the result of the product being contaminated from a variety of sources, or it could be the result of processing that was not adequate, or it could be the result conditions that were storage suitable [39].

As a result of their capacity to thrive at low temperatures, psychotrophic bacteria are the primary culprits responsible for the spoilage of meat products that are stored at refrigeration temperatures. It is possible to obtain valuable information preservation regarding the quality certain meat products by utilizing the total According **PBC** [5]. to the current investigation, the presence of PBC was detected in all samples, with significant variations observed (P < 0.05). The mean value of their counts in the samples of sausage was  $4.90\pm0.21$   $\log_{10}$  CFU/g as well as minced meat  $(4.35\pm0.20 \log_{10}$ CFU/g) and luncheon samples (3.57±0.10 log10 CFU/g). In this study, the obtained minimum and maximum values of the total psychotrophic count (log<sub>10</sub> CFU/g) meat samples (minced the meat, luncheon) and were sausage, nearly similar to other investigations that were conducted previously [40-44]. The latter reports showed these results were more favorable. The presence of high PBCs in study may be an indication during inappropriate hygienic practices slaughtering, processing, retail or handling [11]. Psychrotrophic bacteria are cold able thrive in environments because they have developed adaptations that allow them to do that. These adaptations include proteins that are able to function at low temperatures increased quantities of unsaturated fatty

acids in their cell membranes. Despite the fact that their presence in food might cause it to go bad or put people at danger getting sick, they also play important the process part in biodegradation natural ecosystems, in during particularly the winter months [45]. On the other hand, our findings regarding PBCs seemed to be in direct opposition to those obtained by previous researchers [5, 40].

There is no evidence that coliform bacteria are the major cause of spoilage; rather, their presence is indicative of insufficient hygiene or the possibility of cross-contamination during preparation or storage of the food [11]. Regarding the evaluation of coliforms in this study, the findings indicated that the presence of coliforms was more abundant the samples of minced meat sausage (100%) compared to the samples of luncheon (50%) with mean values of  $3.35\pm0.22$ ,  $3.04\pm0.18$ , and  $2.92\pm0.09$ respectively. Similar  $\log_{10}$ CFU/g, findings were obtained by previous researchers [38, 46-50]. Coliform counts` results this investigation in were  $3.04\pm0.18$ ,  $3.35\pm0.22$ , and  $2.92\pm0.09$ log<sub>10</sub> CFU/g in minced meat, sausage, and luncheon, respectively. When compared to the results found in minced meat  $(2.80\pm0.10 \log_{10} \text{ CFU/g})$  [35], luncheon and  $3 \log_{10} \text{ CFU/g}$ and sausage (3.1 respectively) [33], these findings almost identical to those seen in minced meat. On the other hand, higher results were obtained in minced meat (4.5 x 107 [34]. Furthermore, log10 CFU/g) the mean values of coliforms expressed as  $log_{10}$  CFU/g was found to be 2.6±0.1 in luncheon, 3.1±0.1 in minced meat, and 2.9±.01 in sausage, as reported elsewhere [9]. Furthermore, the statistical findings demonstrated that there is no significant difference (P >0.05) in the coliform between the samples of (minced meat, sausage, products luncheon). It is possible that the nature of coliforms, which may not flourish efficiently under cold storage conditions

as psychrotrophic bacteria, is the reason for lower coliform counts as compared to APC and PBC. Coliforms are a specialized subset that comprise a lesser fraction of the entire microbial community in meat [11].

The data that were obtained and presented in Table 2 showed that the minimum and maximum counts ( $log_{10}$  CFU/g) of S. aureus in the investigated meat products varied from 2.30 to 3.30 in minced meat, 2.60 to 3.78 in sausage, and 2.0 to 2.85 in luncheon, which are consistent with the findings of previous studies [48, 51, 52]. Likely, Shaltout et al. [9] observed that the mean values  $(\log_{10} \text{ CFU/g})$  of S. aureus counts/g were 2.2 ±.1 in minced meat, 2.6  $\pm$ .1 in sausage, and 2.5  $\pm$ .1 in luncheon. Furthermore, Shaltout et al. [5] and Younis et al. [53] obtained data that comparable to the luncheon samples. Both of these studies conducted on minced meat. aureus count was found to be the highest samples sausage  $(3.01\pm0.13),$ of followed by mined meat  $(2.74\pm0.11),$ while the lowest level was found in luncheon  $(2.42\pm0.14)$ . It is possible that the reason for the high S. aureus counts is the fact that the majority of markets are typical open-air marketplaces that do not have air conditioning; these factors likelihood of enhance the bacterial contamination and subsequent growth [54]. In the meantime, lower results were recorded by several writers [43, 55]. the other hand, S. aureus had not been detected in minced meat as reported by other studies [56, 57].

Mold contaminations of meat and meat products can occur during the slaughtering of animals, transportation, or processing of meat products through the contaminated equipment use of contaminated additives and spices, which are considered to be the most significant source of contamination in meat products [58]. Mold was found in every single sample of product that meat examined, without exception. As shown in Table 2, the total mold count (log10 CFU/g) ranged from 1.85 to 2.70 for minced meat, 2.00 to 3.30 for sausage, and 2.00 to 3.60 for luncheon. A slightly elevated count of mold was seen in the luncheon samples  $(2.83\pm0.19)$  $log_{10}$ CFU/g) and sausage  $(2.73\pm0.20$  $log_{10}$ CFU/g), in contrast to the samples of minced meat, which had the lowest count  $\log_{10}$  $(2.22\pm0.18)$ CFU/g). Mold count samples results from luncheon were essentially identical to those obtained by other reporters [4, 59-61]. On the other hand, a higher prevalence of mold was recorded in meat samples [62, 63]. results of sausage samples were nearly similar to those obtained previously [60, and were significantly lower 641 those obtained bv Brr et Furthermore, the obtained results in this study are nearly similar to those obtained by Saad et al. [66] and were lower than those reported previously [63, 67].

It is possible that the fluctuations in the overall number of molds found in samples are the result of varying degrees cleanliness during the manufacturing and storage processes. The highest total mold count in luncheon may be related to poor handling storage circumstances, or whereas the mold count in sausage may composition attributed the to sausage, which is minced meat packed in the intestines of animals that may not have been sufficiently cleansed [4].

#### Conclusion

The findings of this research showed that product samples that analyzed (minced meat. sausage, luncheon) substantially were contaminated with a variety of different microorganisms. species of This contamination was a result of faulty and inadequate sanitary practices an environmental that occurred state throughout the handling, transportation, processing, and storage of the materials. APC and psychrotrophic species are the most prevalent types of bacteria that were found in the investigated samples. As a imperative that stringent result, it is

hygiene procedures be adhered to throughout the processing and storage stages. In addition, stringent laws ought to be enacted in order to encourage the production of goods that have excellent preservation qualities.

### Conflict of interest: None

### References

- [1] Owusu, M; Basnet, A. and Kilonzo-Nthenge, A. (2024): Antibiotic resistant bacteria in goat meat and hygienic practices among retail stores in Nashville, Tennessee. Front. Sustain Food Syst, 8:1460350.
- [2] Shawish, R. and Tarabees, R. (2017): Prevalence and antimicrobial resistance of Bacillus cereus isolated from beef products in Egypt. Open Vet J, 7(4): 337-341.
- [3] Sofos, J.N. (2014): Meat and Meat Products. In: Food Safety Management. Elsevier Inc., Oxford, 119-162.
- [4] Habashy, A.H.A; Darwish, W.S; Hussein, M.A. and El-Dien W.M.S. (2019): Prevalence of different mold genera in meat and meat products with some reduction trials using essential oils. Adv Anim Vet Sci, 7(s2): 79-85.
- [5] Shaltout, F.A; Bar,r A.A.H. and Abdelaziz, M.E. (2022): Pathogenic Microorganisms in Meat Products. Biomed J Sci Tech Res, 41(4): 32836-32843.
- [6] Elgaffry, O.; Darwish, W.S.; El Bayoumi, R.M.; Hussein, MA.; Megahed, A.; Reda, L.M.; Basiony A. and Dawod, Y.F. (2024): Jun Biogenic amines' residues in meat products with a reference to their microbial status. Open Vet J, 14(6):1394-1402.
- [7] Serhan, M; Hourieh, H; El Deghel, M.and Serhan, C. (2022): Hygienic sanitary risk and microbiological quality of meat and meat-contact surfaces in traditional butcher shops and retail establishments—Lessons from a developing country. Int J Environ Health Res, 34, 600–610.
- [8] Kim, H.J.; Kim, D.; Kim, H.J.; Song, S.O.; Song, Y.H. and Jang, A. (2018):

- Evaluation of the Microbiological Status of Raw Beef in Korea: Considering the Suitability of Aerobic Plate Count Guidelines. Korean J Food Sci Anim Resour, 38(1):43-51.
- [9] Shaltout, F.A; Salem, A.M; Khater, D.F. and Lela, R.A. (2016): Studies on bacteriological Profile of some meat products. Benha Vet Med J, 31(1): 43-49.
- [10] Mousa, M.M; Ahmed, A.A. and El-Shamy, S.Y. (2014): Microbiological Criteria of Some Meat Products. Alex J Vet Sci, 42: 83-89.
- [11] Al-Mazrouei, M.A; Al-Kharousi, Z.S; Al-Kharousi, J.M and Al-Barashdi, H.M. (2024): Microbiological Evaluation of Local and Imported Raw Beef Meat at Retail Sites in Oman with Emphasis on Spoilage and Pathogenic Psychrotrophic Bacteria. Microorganisms, 12(12):2545.
- [12] Wakabayashi, R; Aoyanagi, A. and Tominaga, T. (2024): Rapid counting of coliforms and Escherichia coli by deep learning-based classifier. J Food Saf, 44: e13158.
- [13] Shaltout, F.A.; Nasief, M. Z.; Lotfy, L.M. and Gamil, B. T. (2019): Microbiological status of chicken cuts and its products. Benha Vet Med J, 37(1): 57-63.
- [14] Santos, E.C.C.D.; Castro, V.S.; Cunha-Neto, A.; Santos, L.F.D.; Vallim, D.C.; Lisbôa, R.C.; Carvalho, R.C.T.; Junior, C.A.C. and Figueiredo, E.E.S. (2018): Escherichia coli O26 and O113:H21 on Carcasses and Beef from a Slaughterhouse Located in Mato Grosso, Brazil. Foodborne Pathog Dis,15(10):653-659.
- [15] Salem, A.M.; Nassif, M. and Mohammed, B. (2020): Safety of meat served at a university hostel. Benha Vet Med J, 38(2): 80-83.
- [16] Rodríguez-Lázaro, D; Oniciuc, E.A; García, P.G; Gallego, D; Fernández-Natal, I; Dominguez-Gil, M; Eiros-Bouza, J.M;Wagner, M; Nicolau, A.I. and Hernández, M. (2017): Detection and characterization of Staphylococcus aureus and methicillin-resistant S. aureus

- in foods confiscated in EU borders. Front Microbiol, 8: 288041.
- [17] Ahmadi, S.A; Aziz, A; Afshar, M.F. and Abi, A.J. (2025): Exploring Staphylococcus aureus Prevalence and Antimicrobial Resistance in Ready-to-Eat Meat Products in Kabul City. Egypt J Vet Sci, 15: 1-6.
- [18] CFS: Centre for Food Safety. Food and Environmental Hygiene Department. Vol. 43. Hong Kong, China: F. Queensway Government Offices; 2014. Microbiological Guidelines for Food (for ready-to-eat food in general and specific food items) pp. 12.
- [19] Pitt, J.I. and Hocking, A.D. (2009): Fungi and food spoilage. 3rd Ed. Springer Science - Business Media, LLC, 233 Spring Street, New York, NY 10013, USA, 2009.
- [20] Dagnas, S. and Membré, J.M. (2013): Predicting and preventing mold spoilage of food products. J Food Prot, 76(3):538-51.
- "International [21] **ISO** Standards Organization" 6887-2 / (2003): Microbiology of food and animal feeding stuffs, preparation of test samples, initial suspension and decimal dilutions for microbiological examination. Part 2: Specific rules for the preparation of meat and meat products.
- "International [22] **ISO** Standards Organization" 4833-1 (2013): Microbiology of food chain- Horizontal for method the enumeration microorganisms. Part I; Colony count at 30C by the pour plate technique. International Standards Organization, Geneva, Switzerland.
- [23] FAO "Food and Agriculture Organization"(1992): Manual of Food Quality Control. 4Rev.1.Microbiological analysis (Andrews, W.Ed) FAO food and nutrition paper No.14/4 Rev
- [24] ICMSF "International Commission on Microbiological Specifications for Foods" (1986): In: Microorganisms in Foods. 1. Sampling for Microbiological Analysis. 2. Principles and Scientific

- Applications. Vol.2, 2nd ed., Univ. of Toronto Press, Toronto. pp. 181-196.
- [25] FDA "Food and Drug Administration" (2001): Staphylococcus aureus. Bacteriological analytical manual. 8th Ed. Chapter12. Academic Press, Gaithersburg, UK.
- [26] ISO "International Standards Organization" 21527-1 (2008): Microbiology of food and animal feeding stuffs: Horizontal method for the enumeration of yeasts and molds (21527-1:2008).
- [27] Snedecor, G.W. and Cochran, W.G. (1982): Statistical Methods. 8th ed. Iowa, USA: Ames, Iowa State University Press.
- [28] Darwish, A.M; Niazi, Z.M. and Zaki, E.M. (1991): Escherichia Coli in meat products. Vet Med J, 39(3): 841-851
- [29] Al-Amri, I; Kadim, I.T; AlKindi, A; Hamaed, A; Al-Magbali, R. Khalaf, S; Al-Hosni, K. and Mabood, F. (2021): Determination of residues of pesticides, steroids, antibiotics, anabolic and antibacterial compounds in meat products Oman by liquid in chromatography/mass spectrometry and enzyme-linked immunosorbent assay. Vet World, 14(3):709-720.
- [30] Jay, J.M. 1997. Modern food microbiology, 4th Ed., Chapman and Hall, International Thomson Publishing, New York.
- [31] Allen, M.J; Edberg, S.C and Reasoner, D.J. (2004): Heterotrophic plate count bacteria--what is their significance in drinking water? Int J Food Microbiol, 92(3):265-74.
- [32] Ercolini, D; Russo, F; Nasi, A; Ferranti, P. and Villani, F. (2009): Mesophilic and psychrotrophic bacteria from meat and their spoilage potential in vitro and in beef. Appl Environ Microbiol, 75(7):1990-2001.
- [33] Yao, L; Champagne, C.P; Deschênes, L; Raymond, Y; Lemay, M.J. and Ismail, A. (2021): Effect of the homogenization technique on the enumeration of psychrotrophic bacteria in food absorbent pads. J Microbiol Methods, 187:106275.

- [34] Erdem-Ayten, k; Saglam, D; Ozer, D. and Ozcelik, E. (2014): Microbiological Quality of Minced Meat Samples Marketed in Istanbul. YYU Veteriner Fakultesi Dergisi, 25(3): 67 –70.
- [35] Salem-Amany, M; Amin-Reham, A. and Afifi-Gehan, S.A. (2010): Studies on Antimicrobial andAntioxidant Efficiency of Some Essential Oils in Minced Beef. J American Sci, 6:691-700.
- [36] El-Dosoky, H.F.A.; Shafik, S. and Baher, M. (2013): Detection of spoilage and food poisoning bacteria in some ready to eat meat products in Dakahlia governorate. Assiut Vet Med J, 59:55-59.
- [37] Gönülalan, Z. and Köse, A. (2003): Kayseri ilinde satışasunulan sığır kıymalarının mikrobiyolojikkalitesi. FÜ Sağlık Bil Derg, 17: 49–53.
- [38] Shaltout, F.A; Maarouf, A.A.A; El-Kewaiey, I.A. and Heweidy, A.Y.A. (2016): Prevalence of some food borne microorganisms in meat and meat products. Benha Vet Med J, 31(2): 213-219.
- [39] Zaharan-Dalia, A; Bassma, A.H. and El. Hifnawi, H.N. (2008): Incidence and radiation sensitivity of Bacillus cereus, Listeria monocytogenes and their toxins in some chicken products. World Appl Sci J, 5:182-188.
- [40] Karaboz, I. and Dincer, B. (2002): Microbiological investigation on someof the commercial frozen meat inIZMIR. Turk Elect J Biotechnol., 1(si): 18-23.
- [41] Badawi, M. (2008): Further studies on imported frozen meat Ph.D. Thesis (Meat Hygiene), Fac. Vet. Med., Alex. University, Egypt.
- [42] Ibrahim, M. A. R. (2011): Microbiological evaluation of retail frozen meat .M.V.SC., Thesis, Meat Hygiene, Fac. Vet. Med., Alex. University, Egypt.
- [43] Ali, M. (2016): Bacterial Contamination of Imported Frozen Meat M.V. Sc., Thesis (Meat Hygiene), Fac. Vet. Med., Alex. University, Egypt.
- [44] Khalafallah, B.M; Abd El-Tawab, A.A; Nada, S. and Elkhayat ,M.E. (2020): Phenotypic and genotypic

- characterization of pseudomonas species isolated from frozen meat. Benha Vet Med J, 39: 47-51.
- [45] Gounot, A.M. (1986): Psychrophilic and psychrotrophic microorganisms. Experientia, 42: 1192–1197.
- [46] Paulsen, P; Schopf, E. and Smulders, F.J.M.( 2006): Enumeration of total aerobic bacteria and E.coli in minced meat and on carcass surface samples with an automated most-probable-number method compared with colony count protocols. J Food Prot, 69: 2500 2503.
- [47] Stagnitta, P.V; Micalizzi, B. and Guzmán, A.M.S. (2006): Prevalence of some bacteria yeasts and molds in meat foods in san Luis, Argentina. Cent Eur J Publ Health, 14:141–144.
- [48] Ahmed-Alyaa, S.O.S. (2015): Quality of Native and Imported Meat in The Egyptians Markets M.V.Sc., Cairo University.
- [49] Al-Mutairi, M.F. (2011): The incidence of Enterobacteriaceae causing food poisoning in some meat products. Adv J Food Sci Technol. 3: 116-121.
- [50] Hamed, E.A; Ahmed, A.S. and Abd El-Aaty, M.F. (2015): Bacteriological hazard associated with meat and meat products. Egypt J Agric Res, 93: 385-393.
- [51] Plaatjies, Z; Lues, J. and Buys, E. (2004): Staphylococcal growth in fresh vacuum-packed red meat at various storage conditions, In: 8th World Congress on Environmental Health, Durban, South Africa.
- [52] Phillips, D; Jordan, D.; Morris, S.; Jenson, I. and Sumner, J. (2006): A national survey of the microbiological quality of beef carcasses and frozen boneless beef in Australia. J Food Prot, 69: 1113-1117.
- [53] Younis, O.; Ibrahim, H. M.; Hassan M.; Amin, R. and Maarouf, A.A. (2019): Demonstration of some foodborne pathogens in different meat products: acomparison between conventional and innovative methods. Benha Vet Med J, 36(2): 219-228.

- [54] Zhang, H.N; Hou, P.B; Chen, Y.Z; Ma, Y; Li, X.P; Lv, H; Wang, M; Tan, H.L. and Bi, Z.W. (2016): Prevalence of Food borne Pathogens in Cooked Meat and Seafood from 2010 to 2013 in Shandong Province, China. Iran J Public Health, 45(12):1577-1585.
- [55] Gibriel, A. Y; Ebeid, H. M; Khalil, H. I. and Abdel-Fattah, A. A. (2007): Application of Monascus purpureus pigments produced using some food industry wastes in beef sausage manufacture. Egypt J Food Sci, 35:27-45.
- [56] Tolba, K. (1994): Microflora in locallyprocessed frozen meat. Vet Med J Giza, 42(2): 99-105.
- [57] Abdel-Aziz, W.M. (2015):
  Bacteriological hazards associated withconsumption of street vended meat products in Kalyobia governorate, In:
  AbdEl-aziz Wafaa M (Edt.),
  M.V.SC.Thesis(meat hygiene), Fac. Vet.
  Med., Benha University, Egypt.
- [58] Jay, J.M.; Leossner, M.J. and Golden, D.V. (2005). Modern food microbiology. 7th Ed. Chapter 2, taxonomy, role and significance of microorganisms in foods. Springer Sci. pp.13- 37.
- [59] Ouf, J.M; Nagwa, I.M.K. and Shabana, E.S.E. (2010): Incidence of proteolytic and lipolytic molds and yeasts in some ready to eat meat products. Assuit Vet Med J, 56(126): 132-143.
- [60] Elsayed, M.E; Abdelazeem, M.A; El-Diasty, E.M; Abouelmaatti, R.R. and Abbas, S.M. (2018): Prevalence of Aspergillus spp and Aflatoxins in luncheon, minced meat and sausage. Glob Anim Sci J, 6(2):17-23.
- [61] Ashraf, A.A; Eman, M.E; Gamal, R.H; Fatma, I.E. and Abd El-hady, M.F. (2019): Molecular characterization of lipolytic fungi isolated from meat products. J Amer Sci, 15(4): 8-16.
- [62] Ali, M.H; Farghaly, M.R. and Hammad, A.M. (2005): Mycological investigations in beef and chicken luncheon. Beni-Suef Vet Med J, 15(2): 98- 102.
- [63] El-Tabiy, A.A (2006): Mycological study on some processed meat products

- exposed for sale in markets. Assiut Vet Med J, 52(110): 121-131.
- [64] Fatema, A.H.D. (2016): Toxigenic fungi and their metabolites insome meat products. M.V.Sc. thesis, (Meat Hygiene) Fac.Vet. Med., Benha Univ., Egypt.
- [65] Brr, A.H; Moustafa, N.Y. and Edris, A.M. (2004): Incidence ofmolds and aflatoxin in some meat products. Benha Vet Med J, 15(2): 65-75.
- [66] Saad, M.S; Ramadan, M.S; Reham, A.A.A. and Khalifa, E.A.A. (2015): The using of essential oils in improving mycological statusof some meat products. Benha Vet Med J, 29(1): 85-96.
- [67] Eleiwa, N.Z. and El-Diasty, E.M. (2014): Antifungal activity of dill essential oil (Anethum graveolens L.) in minced meat. VRI Phytomed, 2 (1): 6-12.

## الملخص العربى تقييم الوضع الصحي لبعض منتجات اللحوم المسوقة بمدينة الزقازيق — جمهورية مصر العربية

علاء الدين محمد علي مرشدي، عبدالسلام السيد حافظ , أمنية عقيل فودة ، وجيه صبحي درويش قسم صحة وسلامة وتكنولوجيا الغذاء- كلية الطب البيطري - جامعة الزقازيق - مصر

تكتسب منتجات اللحوم شعبية متزايدة لأنها تُمثل وجبات سريعة وسهلة التحضير وبأسعار منخفضة، وتُمكّن المُصنّعين من تحويل أنواع اللحوم المختلفة إلى منتجات موحدة. يُعدّ تحديد الجودة الميكروبية وسلامة اللحوم أمرًا بالغ الأهمية نظرًا لاحتمالية احتوائها على مسببات الأمراض. هدفت هذه الدراسة إلى تقييم الحالة الصحية غير المستكشفة لمنتجات اللحوم المُسوّقة (اللحم المفروم، والسجق، واللانشون) في مدينة الزقازيق، محافظة الشرقية، مصر.

أجريت هذة الدراسة على عدد 60 عينة عشوائية من منتجات اللحوم (اللحوم المفرومة- السجق-اللانشون) وأجرى عليها التحليل البكتريولوجي حيث تم عزل وتعريف بعض مجموعات البكتيريا الممرضة وذات الخطورة على الصحة العامة مثل عدد الصفائح الهوائية (APC)، وعدد البكتيريا الباردة (PBC)، وعدد البكتيريا القولونية، والمكورات العنقودية، والعفن. كان معدل انتشار APC و 400 PBC في جميع عينات منتجات اللحوم المفحوصة، و 100 الكوليفورم في اللحوم المفرومة والسجق و 50% في عينات المنتقودية الذهبية في عينات منتجات اللحوم، في حين كان معدل في عينات المنتقودية الذهبية في عينات المعوية ( (APC في عينات اللانشون، و 80% و 90% و 60% على التوالي للمكورات العنقودية الذهبية في عينات المعوية ( (APC للمكورات العنقودية الذهبية ( (APC ± 1.0.0) 4.05±2.0.2) (APC + 1.0.0±2.0.2) (APC + 1.0.0