

Sensitivity of some *Fusarium* species to fungicides and *Trichoderma* isolates

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ABSTRACT

Three laboratory experiments were done in order to assess the effectiveness of both most commonly used fungicides and *Trichoderma* isolates against pathogenic *Fusarium spp.*, as well as the interaction between fungicides and *Trichoderma* isolates. In the first one: Eighteen of commonly used fungicides as chemical control for *Fusarium* diseases (thiabendazole, thiophanate-methyl, imazalil, monceren, carboxin, vitavax-200, vitavax-300, captan, tolclofos-methyl, triadimefon, penconazole, propiconazole, hexaconazole, mancozeb, ridomil MZ, ridomil plus, iprodione and homai) were evaluated against eight species of *Fusarium* which cause *Fusarium* diseases of different legumes (*Fusarium oxysporum f.sp. phaseoli*, *F. solani f.sp. phaseoli* from bean seeds "F1, F2"; *F. oxysporum f.sp. lupini*, *F. solani f.sp. lupini* from lupine seeds "F3, F4"; *F. oxysporum f.sp. ciceris*, *F. solani f.sp. ciceris* from chickpea seeds "F5, F6" and *F. moniliforme*, *F. solani* from cowpea seeds "F7, F8"). The results in this part showed that Imazalil was extremely highly effective fungicide since the EC_{50} values for all tested *Fusarium* species were ranged from less than $0.1 \mu\text{g ml}^{-1}$ to $0.22 \mu\text{g ml}^{-1}$. Thiabendazole, homai, and thiophanate-methyl were highly effective fungicides with EC_{50} values less than $10 \mu\text{g ml}^{-1}$ ($0.03 \mu\text{g ml}^{-1}$ for thiabendazole against F2 and $6.05 \mu\text{g ml}^{-1}$ for homai against f7), and generally we can arrange the eight species of *Fusarium* according to their sensitivity to the tested fungicides in ascending order as follows: F1, F8, F4, F2, F6, F5, F3, and F7. In the second part of work: efficacy of biological

antagonists was studied by using twenty four isolates of *Trichoderma* against the eight species of *Fusarium*, and we found that most of the *Trichoderma* isolates had antagonistic effect against most of the tested *Fusarium* species. Finally we examined the most effective tested fungicides against the most antagonists of *Trichoderma* against *Fusarium* species. The data revealed that most of *Trichoderma* isolates are insensitive (tolerant) than fungi of *Fusarium spp.* to all the tested fungicides. Imazalil was the most suitable fungicide because it was highly effective fungicide against *Fusarium* species (pathogenic fungi), while the *Trichoderma* (non pathogenic fungi) are insensitive to it without any exceptions, followed by thiabendazole with only one exception.

Keywords: *Fusarium* species, Fungicides, *Trichoderma* isolates, Antagonistic activity.

INTRODUCTION

Seeds of bean (*Phaseolus vulgaris* L.), cowpea (*Vigna sinensis* Erdl), chickpea (*Cicer arietinum* L) and lupine (*Lupinus termis* L) remain as important part of diet for both humans and animals in Egypt.

Different fungal species were reported as seed borne pathogens on legumes which causes different types of diseases and the major type of these pathogens is *Fusarium* i.e. *Fusarium oxysporum f.sp. lupini* (Morkunas *et al.*, 2004 & Morkunas *et al.*, 2005), *Fusarium acutatum* (Gopalakrishnan *et al.*, 2005), *Fusarium oxysporum f.sp ciceris* (Kaur and Mukhopadhyay, 1992; Hervas *et al.*, 1998 and Cho and Muehlbauer, 2004) and *Fusarium solani f.sp. phaseoli* (Estevez *et al.*, 2001).

Trichoderma spp. as *T. harzianum* and *T. Viride* are filamentous soil fungi and they are present in substantial numbers in nearly all agricultural

soil. Their mycoparasitism involves a complementary action of antibiosis, nutrient competition and cell wall degradation of the target fungi (Viterbo *et al.*, 2001). *Trichoderma* have been investigated extensively as bio-control agent of several plant pathogenic fungi especially *Fusarium spp.* *in vitro* and *in vivo* in legumes as chickpea (Kaur and Mukhopadhyay, 1992); cowpea (Ushamalini *et al.*, 1997) and bean (Estevez *et al.*, 2001).

We realize that *in vitro* screening with our arbitrary rating system for integrated control including chemical control with fungicides and biological effective antagonists against soil borne plant pathogens especially *Fusarium spp.* is a simplistic approach to understanding a small sector for integrated control systems in disease control. So this research aimed to find out one or more fungicide possess high potential fungicidal property against the pathogenic fungi (*Fusarium spp.*) meanwhile the non-pathogenic fungi (*Trichoderma spp.*) is insensitive or tolerant toward them.

MATERIALS AND METHODS

1- Source of isolates

a- Fungi

The tested eight species of fungi were isolated, purified and then identified in the Department of Plant Pathology, Faculty of Agriculture, Alexandria University, Egypt and in Agriculture Research Center, Ministry of Agriculture, Cairo, Egypt as follow: F1: *Fusarium oxysporum f.sp.phaseoli* and F2: *Fusarium solani f.sp. phaseoli* from bean seeds, F3: *Fusarium oxysporum f.sp. lupini* and F4: *Fusarium solani f.sp. lupini* from lupine seeds, F5: *Fusarium oxysporum f.sp ciceris* and F6: *Fusarium solani f.sp ciceris* from chickpea seeds, F7: *Fusarium moniliforme* and F8: *Fusarium solani* from cowpea seeds.

b- Trichoderma

Trichoderma isolates were isolated from diseased plants as follow: isolates No. 1 to 6 from bean, 7 to 13 from lupine, 14 to 21 from chickpea and isolates No. 22 to 24 from cowpea.

2- Fungicides used

Eighteen different commonly used fungicides are: thiabendazole, thiophanate-methyl, imazalil, monceren, carboxin, vitavax-200, vitavax-300, captan, tolclofos-methyl, triadimefon, penconazole, propiconazole, hexaconazole, mancozeb, ridomil MZ, ridomil plus, iprodione and homai.

3- Antifungal assay

a- against *Fusarium* spp

The antifungal activity of eighteen common fungicides against eight species of *Fusarium* was tested using the radial growth technique method (Zambonelli *et al.*, 1996). Appropriate volumes of the stock solutions of the tested fungicides were added to molten potato dextrose agar (PDA) media to obtain a range of concentrations (0.1, 1, 5, 10, 100, 1000 µg ml⁻¹) immediately before pouring into the Petri dishes (9.0 cm diameter) at 40-45 °C. Each concentration was tested in triplicate and all the experiment was repeated twice. The discs of mycelial felt (0.5 cm diameter) of the tested pathogenic fungi, (eight species of *Fusarium*) or non-pathogenic fungi (four isolates of *Trichoderma*) taken from 8-day-old cultures on PDA plates, were transferred aseptically to the centre of Petri dishes. The treatments were incubated at 25 °C in the dark. Colony growth diameter was measured after the fungal growth in the control treatments (without any fungicides) had completely covered the Petri dishes. Percentage of mycelial growth inhibition was calculated from the formula: Mycelial growth inhibition = [(DC-DT) /DC] × 100 (Pandey *et al.*, 1982), where DC and DT are average diameters of fungal colony of control and treatment, respectively. The

concentration of the fungicide that inhibiting the mycelial growth by 50 % (EC_{50}), was determined by a linear regression method, (Finney, 1971).

b- against *Trichoderma* isolates

The most effective fungicides (thiabendazole, thiophanate-methyl, imazalil, vitavax-200 and homai) were evaluated against four *Trichoderma* isolates (the most antagonistic isolates "one for each plant": T5, T12, T18 and T22) using the previously mentioned method. EC_{50} values were calculated and the relative effectiveness was expressed as tolerance or insensitivity. This parameter (tolerance or sensitivity) was estimated by dividing the EC_{50} of fungicide against *Trichoderma* over EC_{50} of fungicide against fungi.

4- Antagonistic activity

Agar discs, 7mm diam., taken from the growing margin of the pathogen and *Trichoderma* isolates were transferred on malt extract agar (MEA) medium (17 %) in 9 cm diameter Petri dishes, containing 10 ml. medium. Inocula of antagonists and tested fungi were plated on the opposite sides on the surface of MEA medium and incubated at 25 - 27 °C. Five replicates per each treatment were prepared. Degree of antagonism was scored according to Bell *et al.*, (1982) using a scale from 1 – 5 classes as the following: class1: *Trichoderma* completely overgrew the pathogen and covered the entire medium surface, class 2: *Trichoderma* overgrew at least two – thirds of medium surface, class 3: *Trichoderma* and the pathogen each colonized approximately one – half of the medium surface (more than one – third and less than two – thirds) and neither organism appeared to dominate the other, class4: The pathogen colonized at least two – thirds of the medium surface and appeared to withstand encroachment by *Trichoderma*, and class 5: the pathogen overgrew the *Trichoderma* and occupied the entire medium surface.

RESULTS AND DISCUSSIONS

Fungicidal activity against *Fusarium spp.*

The activities of the tested fungicides in terms of radial growth inhibition are summarized in Table (1). Imazalil is extremely highly effective fungicide since the EC_{50} values of it against all tested species of *Fusarium* was less than $1\mu\text{g ml}^{-1}$ and these values ranged from less than $0.1\mu\text{g ml}^{-1}$ to $0.22\mu\text{g ml}^{-1}$. Thiabendazole, homai, and thiophanate methyl were highly effective fungicides and arranged after imazalil in their effectiveness with EC_{50} less than $10\mu\text{g ml}^{-1}$ ($0.03\mu\text{g ml}^{-1}$ for thiabendazole against F2 and $6.05\mu\text{g ml}^{-1}$ for homai against F7). This result is in agreement with the finding of Ilyas *et al.*, (1992) who found that Thiophanate-methyl was effective against *Fusarium oxysporum f.sp ciceris*.

Triazole fungicides include propiconazole, hexaconazole and penconazole were ranged from extremely highly effective to effective fungicides with EC_{50} value equal to $0.06\mu\text{g ml}^{-1}$ for penconazole against F4 and $33.21\mu\text{g ml}^{-1}$ for propiconazole against F7, concerning to the activity of propiconazole against *Fusarium spp.* of chickpea. The obtained data are coincide with Ilyas *et al.*, (1992). Triadimefon as triazole fungicide arrange in the order thirteen in the effectiveness since the EC_{50} value reached to more than $1000\mu\text{g ml}^{-1}$ in most cases except against *Fusarium solani* of cowpea (F8) it was $0.91\mu\text{g ml}^{-1}$. The formula of carboxin with captan (vitavax-300) and thiram (vitavax-200) were moderately effective fungicides (EC_{50} values $\leq 100\mu\text{g ml}^{-1}$ in most cases) while carboxin alone was non effective fungicide with EC_{50} value reached to $\geq 1000\mu\text{g ml}^{-1}$ in many cases, and this was the same which found by Estevez *et al.*, (2001) using captan and vitavax-200 against *Fusarium solani f.sp. phaseoli*. We can arrange the remaining fungicides according to their effectiveness in descending order as follow: Iprodione, ridomil plus, mancozeb, monceren, tolclofos-methyl, ridomil MZ and captan. Our data for Tolclofos-methyl is

Table (1): Fungicidal Activity of tested fungicides against different Fusarium species.

Fungicides	F1	F2	F3	F4	F5	F6	F7	F8
				EC ₅₀ (µg ml ⁻¹)				
Thiabendazole	1.58(B)	0.03(A)	1.43(B)	0.44(A)	3.81(B)	0.47(A)	2.03(B)	0.22(A)
Thiophanate-methyl	2.43(B)	3.48(B)	1.12(B)	2.56(B)	4.67(B)	0.52(A)	3.34(B)	4.48(B)
Imazalil	≤0.1(A)	≤0.1(A)	0.14(A)	≤0.1(A)	0.17(A)	0.22(A)	≤0.1(A)	≤0.1(A)
Monceren	43.25(C)	12.74(C)	≥1000(E)	8.57(B)	≥1000(E)	84.45(C)	≥1000(E)	87.94(C)
Carboxin	14.05(C)	100.00(C)	556.31(D)	101.97(D)	153.31(D)	≥1000(E)	≥1000(E)	≥1000(E)
Vitavax 200	3.07(B)	0.92(A)	8.46(B)	1.48(B)	12.74(C)	2.11(B)	12.93(C)	1.32(B)
Vitavax 300	6.70(B)	5.39(B)	17.56(C)	7.24(B)	25.00(C)	31.77(C)	36.12(C)	6.21(B)
Captan	33.15(B)	≥1000(E)	≥1000(E)	648.81(D)	≥1000(E)	188.25(D)	≥1000(E)	≥1000(E)
Tolclofos-methyl	≥1000(E)	3.10(B)	≥1000(E)	2.32(B)	≥1000(E)	7.33(B)	≥1000(E)	75.47(C)
Triadimefon	17.40(C)	15.26(C)	422.89(D)	13.70(C)	9.91(B)	226.56(D)	≥1000(E)	0.91(A)
Penconazole	0.26(A)	0.13(A)	31.13(C)	0.06(A)	23.96(C)	12.66(C)	31.46(C)	0.09(A)
Propiconazole	0.08(A)	0.23(A)	3.73(B)	0.13(A)	5.33(B)	18.53(C)	33.21(C)	0.21(A)
Hexaconazole	0.50(A)	3.70(B)	14.02(C)	1.80(B)	2.77(B)	6.12(B)	20.65(C)	0.11(A)
Mancozeb	0.08(A)	279.76(D)	167.45(D)	2.74(B)	43.67(C)	0.59(A)	119.05(D)	4.27(B)
Ridomil MZ	7.73(B)	486.76(D)	152.07(D)	17.84(C)	87.17(C)	899.34(D)	353.96(D)	17.98(C)
Ridomil plus	0.42(A)	75.14(C)	47.11(C)	16.65(C)	140.07(D)	1.22(B)	703.48(D)	11.54(C)
Iprodione	31.72(C)	66.04(C)	16.41(C)	36.27(C)	52.86(C)	5.50(B)	0.68(A)	4.66(B)
Homai	2.45(B)	0.77(A)	2.10(B)	1.27(B)	3.20(B)	0.48(A)	6.05(B)	3.50(B)

F1: *Fusarium oxysporum f.sp. phaseoli* (Bean)

F2: *Fusarium solani f.sp. phaseoli* (Bean)

F3: *Fusarium oxysporum f.sp. lupini* (Lupine)

F4: *Fusarium solani f.sp. lupini* (Lupine)

F5: *Fusarium oxysporum f.sp. ciceris* (Chickpea)

F6: *Fusarium solani f.sp. ciceris* (Chickpea)

F7: *Fusarium moniliforme* (Cowpea)

F8: *Fusarium solani* (Cowpea)

A: Extremely highly effective fungicide (EC₅₀ ≤ 1 µg ml⁻¹)

B: Highly effective fungicide (EC₅₀ ≤ 10 µg ml⁻¹)

C: Effective fungicide (EC₅₀ ≤ 100 µg ml⁻¹)

D: Less effective fungicide (EC₅₀ ≤ 1000 µg ml⁻¹)

E: Non effective fungicide (EC₅₀ ≥ 1000 µg ml⁻¹)

in-contrast with the finding of Ilyas *et al.*, (1992). Also, from Table (1), we can arrange the eight species of *Fusarium* according to their sensitivity to the tested fungicides in ascending order as follow: F1, F8, F4, F2, F6, F5, F3, and F7. The obtained results are supported by many researches of many authors who used our fungicides against another *Fusarium spp.* (Buchvarova *et al.*, 1989; Champawat and Pathak, 1991; Gopinath and Shetty, 1992; Vuksa *et al.*, 1994; Allen, *et al.*, 2004 and Raja *et al.*, 2004).

Antagonistic activity

Interactions between isolates of *Trichoderma* and *Fusarium* species were recorded (Table 2). Antagonism was not apparent against F2 and F1 (rating ≥ 3), only 50 % of *Trichoderma* isolates had a high level of antagonism (rating ≤ 2). Hundred percentages of isolates of *Trichoderma* were antagonistic against F4 and 57 % against F3 and the remaining isolates rated ≥ 3 . One hundred percent and 37.5 % of *Trichoderma* isolates appeared to be antagonistic against F6 and F5, respectively. Only one isolate of *Trichoderma* was non antagonistic against F8, but the remaining isolates were antagonistic to F8 and F7. Concerning to the antagonistic activity of *Trichoderma* isolates, our data are supported by Thangavelu *et al.*, (2004) who found that *Trichoderma harzianum* was effective in inhibiting the growth of *Fusarium in vitro*; Estevez *et al.*, (2001) who found that *Trichoderma harzianum* was effective against *Fusarium solani f.sp. phaseoli*; Hervas *et al.*, (1998) showed that *Trichoderma harzianum* effectively suppress wilt disease of chickpea caused by *Fusarium oxysporum f.sp. ciceris* and Kaur and Mukhopadyay (1992) who demonstrated that the chickpea wilt complex caused by several soil borne fungi including *Fusarium oxysporum f.sp. ciceris* was effectively controlled by *Trichoderma harzianum*.

Table (2): Effectiveness of Trichoderma isolates against Fusarium species.

Trichoderma isolates	F1	F2	F3	F4	F5	F6	F7	F8
T1	Class 3	Class 3	-	-	-	-	-	-
T2	Class 3	Class 3	-	-	-	-	-	-
T3	Class 3	Class 3	-	-	-	-	-	-
T4	Class 2	Class 3	-	-	-	-	-	-
T5	Class 1	Class 3	-	-	-	-	-	-
T6	Class 1	Class 3	-	-	-	-	-	-
T7	-	-	Class 2	Class 2	-	-	-	-
T8	-	-	Class 3	Class 2	-	-	-	-
T9	-	-	Class 2	Class 2	-	-	-	-
T10	-	-	Class 2	Class 2	-	-	-	-
T11	-	-	Class 3	Class 2	-	-	-	-
T12	-	-	Class 1	Class 2	-	-	-	-
T13	-	-	Class 3	Class 2	-	-	-	-
T14	-	-	-	-	Class 3	Class 1	-	-
T15	-	-	-	-	Class 2	Class 1	-	-
T16	-	-	-	-	Class 3	Class 1	-	-
T17	-	-	-	-	Class 2	Class 1	-	-
T18	-	-	-	-	Class 1	Class 1	-	-
T19	-	-	-	-	Class 3	Class 1	-	-
T20	-	-	-	-	Class 3	Class 1	-	-
T21	-	-	-	-	Class 3	Class 2	-	-
T22	-	-	-	-	-	-	Class 2	Class 1
T23	-	-	-	-	-	-	Class 2	Class 3
T24	-	-	-	-	-	-	Class 2	Class 2

F1: *Fusarium oxysporum f.sp. phaseoli* (Bean)
 F2: *Fusarium solani f.sp. phaseoli* (Bean)
 F3: *Fusarium oxysporum f.sp. lupini* (Lupine)
 F4: *Fusarium solani f.sp. lupini* (Lupine)
 F5: *Fusarium oxysporum f.sp. ciceris* (Chickpea)
 F6: *Fusarium solani f.sp. ciceris* (Chickpea)
 F7: *Fusarium moniliforme* (Cowpea)
 F8: *Fusarium solani* (Cowpea)

T1 – T6 : Trichoderma isolated from diseased bean
 T7 – T13: Trichoderma isolated from diseased lupine
 T14 – T21: Trichoderma isolated from diseased chickpea
 T22 – T24: Trichoderma isolated from diseased cowpea

Class 1: Trichoderma completely overgrew the pathogen and covered the entire medium surface
 Class 2: Trichoderma overgrew at least two – thirds of medium surface
 Class 3: Trichoderma and the pathogen each colonized approximately one – half of the medium surface (more than one – third and less than two – thirds) and neither organism appeared to dominate the other.

Effectiveness of the fungicides against trichoderma isolates

The effect of the most effective fungicides against the eight *Fusarium* species, (thiabendazole, thiophanate-methyl, imazalil, vitavax-200 and homai) are examined against four isolates of *Trichoderma* (the most antagonistic isolates of *Trichoderma* "one for each plant ": T5, T12, T18 and T22) using radial growth technique. The obtained data expressed as EC_{50} are shown in Table 3. From the obtained data, it was found that most of *Trichoderma* isolates are insensitive (tolerant) than *Fusarium spp.* to all the tested fungicides. The values of tolerance or insensitivity are shown in Table (4). The insensitivity ranged from 1.41 to 81 for T1; 1.65 to 17.2 for T2; 2.24 to 18.15 for T3 and 1.15 to 7.36 for T4. From Table (4), we can conclude that imazalil is the most suitable fungicide because it is highly

Table (3): Effectiveness of certain fungicides against some *Trichoderma* isolates

Fungicides	Tb	Tl	Tch	Tc
	EC_{50} ($\mu\text{g ml}^{-1}$)			
Thiabendazole	2.43	2.37	8.53	1.62
Thiophanate-methyl	2.66	2.11	1.07	5.13
Imazalil	1.92	1.78	3.02	0.65
Vitavax 200	4.32	18.95	11.16	5.78
Homai	1.74	1.21	1.97	1.80

Tb (T5) : *Trichoderma* isolated from bean

Tl (T12) : *Trichoderma* isolated from lupine

Tch (T18): *Trichoderma* isolated from chickpea

Tc (T22) : *Trichoderma* isolated from cowpea

Table (4): Insensitivity (Tolerance) of *Trichoderma* isolates to fungicides (comparing with *Fusarium*)

Fungicides	Tb/F1	Tb/F2	Tl/F3	Tl/F4	Tch/F5	Tch/F6	Tc/F7	Tc/F8
Thiabendazole	1.54	81.00	1.65	5.39	2.24	18.15	0.80	7.36
Thiophanate-m	1.09	0.76	1.88	0.82	0.23	2.06	1.54	1.15
Imazalil	19.20	19.20	12.71	17.20	17.76	13.73	6.50	6.50
Vitavax 200	1.41	4.70	2.24	12.8	0.88	5.29	0.45	4.38
Homai	0.71	2.26	0.58	0.95	0.62	4.10	0.30	0.51

T(b,l,ch,c): EC₅₀ (µg ml⁻¹) of fungicides against *Trichoderma* isolates

F(1,2,3,4,5,6,7,8) EC₅₀ (µg ml⁻¹) of fungicides against *Fusarium* species

effective fungicide against *Fusarium* species (pathogenic fungi) and the four *Trichoderma* (non pathogenic fungi) are insensitive to it without any exceptions, followed by thiabendazole which had one exception. The data of this point are in agreement with Montealegre and Henriquez (1990); Kay and Stewart (1994) and Figueras *et al.*, (1996).

REFERENCES

- Allen, T. W.; S. A. Enebak and W. A. Carey (2004). Evaluation of fungicides for control of species of *Fusarium* on longleaf pine seed. *Crop Protection*, 23 (10): 979-982.
- Bell, D. K; H. D. Wells and C. R. Markham (1982). *In vitro* antagonism of *Trichoderma* species against six fungal plant pathogens. *Phytopathology*, 72: 379-382.

- Buchvarova, R.; I. Filipova and B. Atanasova (1989). Efficiency of fungicides against Fusarium wilt (*Fusarium oxysporum f.sp. dianthi*) in carnations. *Rasteniev'dni-Nauki*, 26 (9): 56-60.
- Champawat, R. S. and V. N. Pathak (1991). Effect of fungicidal seed treatment on wilt disease of cumin. *Journal of Turkish Phytopathology*, 20 (1): 23-26.
- Cho, S. and M. Muchlbauer (2004). Genetic effect of differentially regulated fungal response genes on resistance to necrotrophic fungal pathogens in chickpea (*Cicer arietinum L.*). *Plant Pathology*, 64 (2): 57-66.
- Estevez, J. C.; J. A. Percich and P. H. Graham (2001). Integrated management strategies of bean root rot with *Bacillus subtilis* and *Rhizobium* in Minnesota. *Field Crops Research*, 74: 107-115.
- Figueras, R. M.; C. Cristani and G. Vannacci (1996). Sensitivity of *Trichoderma* isolates and selected resistant mutants to DMI fungicides. *Crop Protection*, 15 (7): 615-620.
- Finney, D. J. (1971): *Probit analysis*, 3rd edn., Cambridge University Press, London, pp 318.
- Gopalakrishnan, S.; M. H. Beale, J. L. Ward and R. N. Strange (2005). Chickpea wilt identification and toxicity of 8-O-methyl fusarubin from *Fusarium acutatum*. *Phytochemistry*, 66 (13): 1536-1539.
- Gopinath, A. and H. S. Shetty (1992). Evaluation of fungicides for control of grain mould in sorghum. *Indian Phytopathology*, 45 (2): 232-23.

- J. Pest Cont. & Environ. Sci.* 13(1): 117-131 (2005).
- Hervas, A.; B. Landa, L. E. Datnoff and R. M. Jimenez-Diaz (1998). Effect of commercial and indigenous microorganisms on *Fusarium* wilt development in chickpea. *Biological Control*, 13 (3): 166-176.
- Kaur, N. P. and A. N. Mukhopadhyay (1992). Integrated control of chickpea wilt complex by trichoderma and chemical methods in India. *Tropical Pest Management*, 38 (4): 372-375.
- Kay, S. J. and A. Stewart (1994). The effect of fungicides on fungal antagonists of onion white rot and selection of dicarboximide-resistant biotypes. *Plant Pathology*, 43 (5): 863-871.
- Ilyas, M. B.; M. J. Iqbal and K. Iftikar (1992). Evaluation of some fungicides against *Fusarium oxysporum f.sp. ciceri* and chickpea wilt. *Pakistan Journal of Phytopathology*, 4: 1, 5-8.
- Montealegre, J. R. and J. L. Henriquez (1990). Possibilities of integrated control of *Sclerotium rolfsii* Sacc. with *Trichoderma* strains and fungicides. *Fitopatologia*. 25 (2): 68-74.
- Morkunas, I.; L. Marczak; J. Stachowiak and M. Stobiecki (2005). Sucrose induced lupine defence against *Fusarium oxysporum*, Sucrose stimulated accumulation of isoflavonoids as defense response of lupine to *Fusarium oxysporum*. *Plant Physiology and Biochemistry*, 43: 363-373.
- Morkunas, I.; W. Bednarski and M. Kozolowska (2004). Response embryo axes of germinating seeds of yellow lupine to *Fusarium oxysporum*. *Plant Physiology and Biochemistry*, 42: 493-499.

- Pandey, D. K.; N. N. Tripathi; R. D. Tripathi and S. N. Z. Dixit (1982). Fungitoxic and phytotoxic properties of essential oil of *Hyptis suaveolens*. PflKrankh PflSchutz, 89: 344-349.
- Raja, R. S.; D. D. Manuja and D. Mukherjee (2004). Evaluation of fortified fungicide solutions in managing corn rot disease of gladiolus caused by *Fusarium oxysporum*. Crop Protection, 23 (9): 783-788.
- Thangavelu, R.; A. Palaniswami and R. Velazhahan (2004). Mass production of *Trichoderma harzianum* for managing *Fusarium* wilt. Agriculture, Ecosystem & Environment, 103 (1): 259 – 263.
- Ushamalini, C.; K. Rajappan and K. Gangadharan (1997). Inhibition of *Macrophomina phaseolina* and *Fusarium oxysporum* by antagonists under *in vitro* conditions. Indian Phytopathology, 12 (2): 168-170.
- Viterbo, A.; S. Haran; D. Friesem; O. Ramot and I. Chet (2001). Antifungal activity of novel endochitinase gene (chit 36) from *Trichoderma harzianum* Rifai TM. FEMS Microbiology Letters 200: 169-174.
- Vuksa, V. P.; A. D. Bojovic and S. J. Adamovic (1994). Effect of fungicides on *Fusarium graminearum* Schwabe. Plant Protection today and tomorrow, Drustvo za zastitu bilja Srbje, Pp.461-470.
- Zambonelli, A.; A. Zechini D'Aulerio; A. Bianchi and A. Albasini (1996). Effects of essential oils on phytopathogenic fungi *in vitro*. J. Phytopathology, 144: 491-494.

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حساسية بعض أنواع الفيوزاريوم للمبيدات الفطرية وعزلات من فطر الترايكوديرما

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تم اجراء ثلاثة تجارب معملية وذلك لتقدير كفاءة كل من معظم المبيدات الفطرية الشائعة الاستخدام وكذلك عزلات من الترايكوديرما ضد أنواع من الفيوزاريوم الممرضة، وكذلك التداخل بين المبيدات الفطرية وعزلات الترايكوديرما. بالنسبة للتجربة الأولى اختصت بتقييم ثمانية عشر مبيد فطري ذات استخدام شهير في مكافحة امراض الفيوزاريوم (ثيابندازول، ثيوفينيت ميثيل، اماناليل، مونسرين، كربوكسين، فيتافكس ٢٠٠، فيتافكس ٣٠٠، كابتان، تولكلوفوس ميثيل، ترائي ادميفون، بنكونازول، بروبيكونازول، هكسكونازول، مانكوزيب، ريدوميل MZ، ريدوميل بلس، ايرودايون والهوماي) ضد ثمانية انواع من الفيوزاريوم والتي تسبب امراض الفيوزاريوم للعديد من البقوليات (فيوزاريوم اكسي سبورم فاصيولاي، فيوزاريوم سولاني، فاصيولاي (الفاصوليا) & فيوزاريوم اكسي سبورم ليوبنای، فيوزاريوم سولاني ليوبنای (القمح) & فيوزاريوم اكسي سبورم ساسيرس، فيوزاريوم سولاني ساسيرس (الحمص) & فيوزاريوم مونيليفورم وفيوزاريوم سولاني (اللوبياء) F1، F2، F3، F4، F5، F6، F7، F8 على الترتيب. ووضحت النتائج في هذا الجزء ان مبيد الامازاليل كان شديد الفعالية على معظم الفطريات المختبرة بقيمة EC_{50} اقل من ١ ميكروجرام/مل حيث تراوحت القيمة من ٠,٩ الى ٠,٢٢ ميكروجرام/مل تلاه في الفعالية مبيد الثيابندازول، الهوماي والثيوفينيت ميثيل بقيمة EC_{50} اقل من ١٠ ميكروجرام/مل (٠,٠٣ ميكروجرام/مل للثيابندازول ضد F2 و ٦,٠٥ ميكروجرام/مل للهوماي ضد F7). بصفة عامة يمكن ترتيب انواع الفيوزاريوم من حيث حساسيتها للمبيدات الفطرية المستخدمة كما يلي: $F4 \leq F2 \leq F6 \leq F5 \leq F3 \leq F7$ $F1 \leq F8 \leq$

بالنسبة للتجربة الثانية وهي عبارة عن دراسة للتأثير البيولوجي المضاد لـ ٢٤ عزلة من الترايكوديرما ضد الثمانية انواع من الفيوزاريوم وقد وجد ان معظم تلك العزلات من الترايكوديرما كان لها تأثير مضاد لمعظم انواع الفطريات المستخدمة. وفي النهاية تم اختبار اكثر المبيدات كفاءة ضد انواع الفيوزاريوم (من التجربة الأولى) ضد اكثر العزلات من الترايكوديرما تضادا للفطريات (من التجربة الثانية) وقد وجد ان معظم عزلات الترايكوديرما تحت التجربة كانت متحملة عن الفطريات لكل المبيدات الفطرية المختبرة. وان المبيد الفطري اماناليل هو اكثر المبيدات الفطرية ملائمة حيث انه شديد الفعالية ضد الفيوزاريوم (الفطريات الممرضة) بينما كانت الترايكوديرما (الفطريات الغير ممرضة) غير حساسة (متحملة) له، تلاه في ذلك المبيد الفطري الثيابندازول.