Online ISSN: 2537-0979

### **ORIGINAL ARTICLE**

# **Evaluation of CD68 and MCP1 in Patients with Bacterial Prostatitis**

## <sup>1</sup>Zahraa D. Gheni, <sup>2</sup>Israa S. Abbas

<sup>1</sup>Department of Pathological Analysis, College of Applied Medical Sciences/University of Kerbala, Karbala/Iraq <sup>2</sup>College of Al-Amal Specialized Medical Sciences, Karbala/Iraq

#### **ABSTRACT**

Key words: Prostatitis, CD68, MCP-1, PSA, CRP

\*Corresponding Author:
Zahraa Dheyaa Gheni
Department of Pathological
Analysis, College of Applied
Medical Sciences/University of
Kerbala, Karbala/Iraq
Tel.: 07823201736
zahraa.dh@s.uokerbala.edu.iq
esraa.sultany@uokerbala.edu.iq

Background: Bacterial prostatitis is a medical condition that affects the prostate gland. It is considered one of the most common urinary tract infections in males. There are two types of this condition: acute and chronic. Both types present with urinary symptoms and pelvic pain, but they differ in severity and duration.CD68 is a marker on the surface of macrophages that significantly contributes to immune response recognition, while MCP-1 chemokine attracts monocytes to the site of inflammation. The elevated levels of both immunological biomarkers in the blood, along with other inflammatory markers that are widely linked to the immune response. Aim of Study: This study investigates the evaluation of immune and inflammatory markers in patients with bacterial prostatitis. Methodology: A case-control study was conducted at a private clinic in Karbala and at Al-Nassiriya Teaching Hospital in Al-Nassiriya city from October 2024 to May 2025. The study involved 120 male participants, divided into three groups: Eighty patients diagnosed with acute and chronic bacterial prostatitis, and forty healthy men with no prior history of prostate infections. The participants' ages ranged from 34 to 74 years; all men were diagnosed by a urologist based on patient history and NIH criteria.MCP-1 and CD68 have been measured in men by the ELISA technique. Results: Patients with ABP and CBP exhibited significantly higher levels of WBC, CRP, and PSA compared to the control group (p < 0.01). Immune markers CD68 and MCP-1 were effective in distinguishing between ABP and CBP infections (p < 0.01). ROC analysis showed that, in ABP, MCP-1 had the highest diagnostic accuracy with an AUC: 99.375%), followed by CD68 (AUC: 99.063%) and PSA (AUC: 96.906%). In CBP, MCP-1 also demonstrated superior performance (AUC: 99.219%), outperforming CD68 and PSA (AUCs: 92.906% and 95.094%, respectively). Conclusion: In both acute and chronic prostatitis, significantly elevated levels of MCP-1 and CD68 were observed, clearly reflecting the presence of inflammatory activity in prostate tissue. Furthermore, both MCP-1 and CD68 demonstrated high sensitivity and specificity compared to PSA, thereby enhancing their diagnostic accuracy.

## INTRODUCTION

Prostatitis is one of the most prevalent clinical conditions, characterized by a range of urinary symptoms such as urgency, frequency, pelvic pain, and sexual dysfunction <sup>1</sup>. Prostatitis includes acute bacterial prostatitis (ABP), chronic bacterial prostatitis (CBP), chronic prostatitis/chronic pelvic pain syndrome (CP/CPPS), and asymptomatic inflammatory prostatitis <sup>2</sup>. ABP often progresses to a chronic form, characterized by recurrent episodes interspersed with asymptomatic periods of varying duration <sup>3</sup>. The primary cause of prostate inflammation is the presence of pathogenic microorganisms. Bacterial prostatitis is frequently associated with infections caused by Enterobacteriaceae species, particularly *Escherichia coli*, as well as other well-known uropathogens<sup>4</sup>.

Cluster of Differentiation 68 (CD68) is a transmembrane glycoprotein that belongs to the lysosome-associated membrane glycoprotein (LAMP) family, with a molecular weight of approximately 110 kDa. It is highly glycosylated and predominantly expressed on human monocytes, macrophages, and microglia. Initially identified as a macrophage surface marker, CD68 plays a vital role in the immune response, especially in macrophage interactions with low-density lipoproteins (LDL), and is involved in immune regulation <sup>5</sup>. Macrophages serve both immunological and physiological functions. In response to tissue injury or infection, monocyte-derived macrophages are recruited from the bloodstream. These macrophages exhibit plasticity, adapting to environmental signals, as described in the conventional paradigm <sup>6,7</sup>.

Monocyte Chemoattractant Protein-1 (MCP-1) is a key cytokine that regulates inflammatory and immune responses. It is secreted by monocytes, endothelial cells, smooth muscle cells, and fibroblasts in response to inflammatory mediators such as IL-1 and TNF-α. Moreover, bacterial infections may trigger circulating Toll-like receptor ligands, leading to increased MCP-1 production by mesenchymal stem cells in the bone marrow, thereby enhancing the innate immune response. MCP-1 binds to CCR2, which is essential for maintaining monocyte homeostasis in the bloodstream even in the absence of inflammation. It also facilitates the migration of inflammatory monocytes from the bone marrow into the bloodstream and subsequently to sites of infection <sup>8,9,10</sup>.

## **METHODOLOGY**

A case-control study was carried out in a private clinic in Karbala and at Al-Nassiriya Teaching Hospital in Al-Nassiriya city from October 2024 to May 2025. There were 120 subjects in the study. Male participants were divided into three groups Acute Bacterial Prostatitis (ABP), Chronic Bacterial Prostatitis (CBP), and controls, with forty participants in each group. The diagnosis of ABP and CBP was made according to the National Institutes of Health.(NIH). The ages ranged between (34 - 74 years), Mean  $\pm$  SD(53.64  $\pm$  10.60 years ). Gentile urine examination 2 (G.U.E.2) was collected after a prostatic massage done by a physician. The urine samples were processed immediately after collection and examined under 10x and 40 magnification of the field microscope to detect pus cells. A sample of blood was centrifuged to obtain serum for the measurement of PSA and CRP levels. The

sandwich *ELISA* was used to evaluate the biomarker MCP-1 and CD68 in blood.

#### Statistical analysis

Statistical analysis was carried out by IBM SPSS Statistics software version 23. Additionally, the mean and standard deviation were calculated, ANOVA and Kruskal-Wallis tests have been utilized to discern statistically significant differences across multiple groups. Receiver operating characteristic (ROC) was plotted, and cut-off, area under the curve(AUC), sensitivity (%), and specificity(%). Accuracy is indicated determined with a P value < 0.01.

#### RESULTS

The results of the current study showed Mean  $\pm$  SD of WBCS in blood in the control, ABP, and CBP groups were  $\pm$  7.87 $\pm$ 0.79, 10.04 $\pm$ 1.32, and 9.55 $\pm$ 1.72, respectively. The Mean  $\pm$  SD of pus cells in the control group was 0.00 $\pm$ 0.00, while in the ABP and CBP groups, it was 37.00 $\pm$ 8.53 and 30.00  $\pm$  9.60, respectively. The CRP Mean  $\pm$  SD in the control group was 0.72  $\pm$  2.82, whereas in the ABP and CBP groups, it was 23.12  $\pm$  3.62 and 17.62  $\pm$  3.08, respectively. with a highly statistically significant P < 0.01.

MCP-1 level in serum showed a significant increase in the ABP group, with a mean  $\pm$  sd of 338.38  $\pm$  38.88, compared to the control group68.18  $\pm$  9.82, and the CBP group,  $302.56 \pm 50.28$ , p-value <0.01. CD68 level also showed higher in ABP group  $78.55 \pm 8.10$  compared to the CBP  $66.29 \pm 12.06$ , and the control group (30.49  $\pm$  4.86) with p-value <0.01. PSA levels were lowest in the control group,  $2.82 \pm 0.72$ , and highest in the ABP,  $23.12 \pm 3.62$ , compared to the CBP17.62  $\pm$  3.08, with a p-value of 0.00003.

Table 1: Comparison between Inflammatory and hematology parameters in patients, stratified by their control

Tuble II comparison between innammator.		y and nematorogy parameters in patients, strucined by their control					
Parameters	Control		ABP		СВР		
	Mean	Std± Deviation	Mean	Std±Deviation	Mean	Std± Deviation	P. value
Blood WBCs (cells/µL)	7.87	0.79	10.04	1.32	9.55	1.72	0.00006**
Pus in urine (cells/HPF)	0.00	0.00	37.00	8.53	30.00	9.60	0.00002**
CRP ( mg/L)	2.82	0.72	23.12	3.62	17.62	3.08	0.00003**

Blood WBCs (white blood cell), CRP (C-reactive protein), ABP (Acute bacterial prostatitis), CBP (Chronic bacterial prostatitis).

Table 2: Comparison between Immunological parameters in patients, stratified by their controls

Parameters	Control		ABP		CBP		D volue
	Mean	Std ±Deviation	Mean	Std± Deviation	Mean	Std± Deviation	P. value
MCP1(pg/dL)	68.18	9.82	338.83	38.88	302.56	50.28	0.00003**
CD68(pg/dL)	30.49	4.86	78.55	8.10	66.29	12.06	0.00004**
PSA(ng/mL)	2.82	0.72	23.12	3.62	17.62	3.08	0.00007**

MCP-1 (Monocyte chemotactic protein -1), CD68 (Cluster of Differentiation 68), PSA(Prostate Specific Antigen)

<sup>\*\*</sup> The mean difference is significant at the 0.01 level

<sup>\*\*</sup> The mean difference is significant at the 0.01 level

In both acute and chronic bacterial prostatitis, significant positive correlations were observed between markers, blood WBCs, MCP-1, and CD68 (R=0.649, 0.717), respectively, with a p-value < 0.01. Also, PSA showed a strong correlation with MCP-1 and CD68 (R=0.811, 0.804), respectively, with p-value <0.01. Furthermore, pus in urine with MCP-1 and CD68 (R=0.839, 0.779, respectively, p-value <0.01. Additionally, significant correlation between MCP-1 and CD68 (R=0.846 with p-value 0.00).

ROC curve in ABP visualizes the trade-off between true positive rate (sensitivity) and false positive rate.

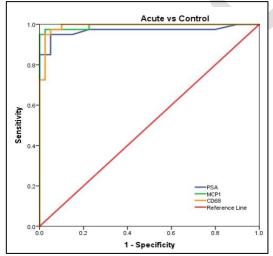
The overall AUC, sensitivity, and specificity for PSA, MCP-1, and CD68 were as follows: (92.000%, 95.000%, 95.094%), (99.000%,100.000%,99.219%), (92.000%, 95.000%, 92.906%), respectively, indicating excellent diagnostic ability. As in Figure 1.

The ROC curve in CBP visualizes the trade-off between true positive rate (sensitivity) and false positive rate. The overall AUC, sensitivity, and specificity for PSA, MCP-1, and CD68 were as follows: (92.000%, 95.000%, 95.094%), (99.000%,100.000%,99.219%), (92.000%, 95.000%, 92.906%), respectively, indicating excellent diagnostic ability.

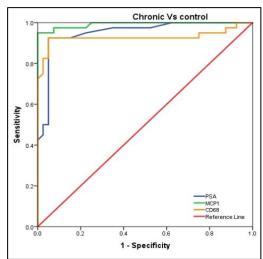
**Table 3: Patients Correlation Coefficient Among Research Parameters According to the patients (Acute and Chronic bacterial prostatitis)** 

Parameters	Value	CRP	PSA	Pus in Urine	MCP1	CD68	Age
Blood WBCs	R. value	.709**	.799**	.737**	.649**	.717**	.083
	P. value	.000	.000	.000	.000	.000	.466
CRP	R. value	1.000	.866**	.855**	.811**	.804**	419 <sup>**</sup>
	P. value		.000	.000	.000	.000	.000
PSA	R. value		1.000	.849**	.767**	.788**	185
ISA	P. value			.000	.000	.000	.101
Pus in Urine	R. value			1.000	.830**	.779**	132
r us in Office	P. value				.000	.000	.242
MCP1	R. value				1.000	.846**	330**
MCFI	P. value					.000	.003
CD68	R. value					1.000	272*
CD06	P. value						.014

<sup>\*\*</sup> Significant correlation at the 0.01 level.



**Fig. 1:** A Receiver Operating Characteristic (ROC) curve was applied as a graphical diagram to show the accuracy of a binary classification model across all possible classification thresholds.



**Fig. 2:** A Receiver Operating Characteristic (ROC) curve was used to illustrate the performance of a binary classification model across all possible classification thresholds.

## **DISCUSSION**

The occurrence of acute and chronic bacterial prostatitis leads to an increase in the number of white blood cells in both acute and chronic infections. Also, the number of white blood cells (WBC) increases during prostatitis as a result of the immune response triggered by bacteria in the prostate. <sup>11,12</sup>

An increase in the number of pus cells in the urine above the normal limit is a sign of an immune response, where innate immune factors play a role in the appearance of symptoms. <sup>13</sup> However, it can be observed that a large number of these cells may not appear in some cases, such as in cases of acute and chronic prostatitis infections, possibly because the infection does not occur in the deep tissues<sup>14</sup>

Acute prostatitis and associated with a significant increase in CRP levels, indicating an acute inflammatory response, compared to control and chronic, indicating a rapid inflammatory response. Reported that both acute and chronic bacterial prostatitis lead to elevated prostate-specific antigen (PSA) levels. This increase results from inflammation and tissue damage within the prostate, which facilitates PSA leakage into the bloodstream. 17,18 The immune response during inflammation, including increased CD68 expression, contributes to tissue remodeling and may result in prostate enlargement, accompanied by symptoms such as frequent and painful urination.<sup>19</sup> During acute bacterial infections, macrophages detect pathogen-associated molecular patterns (PAMPs) and endogenous danger signals through pattern recognition receptors (PRRs). This triggers the activation of M1-like macrophages and the release of proinflammatory mediators <sup>20</sup>. In this study, bacterial infections significantly increased blood CD68 levels (p < 0.01), supporting the finding that both acute and chronic prostatitis are associated with higher CD68 expression compared to healthy controls 21 Monocyte Chemoattractant Protein-1 (MCP-1) levels were significantly elevated in patients with acute bacterial prostatitis compared to both chronic cases and healthy controls <sup>22</sup>. MCP-1 levels rise during bacterial infections and remain elevated in chronic conditions, indicating ongoing immune activity <sup>23</sup>. Inflammation of the prostate stimulates MCP-1 expression, and its release from stromal cells increases in chronic inflammation, contributing to the development of benign prostatic hyperplasia (BPH)<sup>24</sup> · The present study demonstrated that bacterial infection-induced elevations in MCP-1 and WBC levels signify a robust immune response. MCP-1 plays a central role in recruiting monocytes to the site of infection, and its elevated levels in blood reflect systemic inflammation. <sup>25,26</sup>

A strong and statistically significant correlation was observed between C-reactive protein (CRP) and MCP-1 (p < 0.01), aligning with previous studies that reported a

similar association during inflammatory responses <sup>27,28</sup>. The correlation between CRP and CD68 was also highly significant (p < 0.05), in agreement with prior research suggesting that elevated CRP levels enhance macrophage polarization (CD68), promoting an M1 proinflammatory phenotype. <sup>29,30</sup>

There was a strong correlation between PSA and MCP-1 (R=0.767), indicating that increased MCP-1 levels are associated with elevated PSA, likely due to inflammation-induced disruption of prostatic tissue barriers <sup>31</sup>. Furthermore, PSA levels also correlated significantly with CD68 expression (p<0.05), suggesting that increased PSA may contribute to heightened macrophage activity in the bloodstream <sup>32</sup>.

A particularly strong correlation (R = 0.846, p < 0.01) was observed between the immunological markers MCP-1 and CD68. Both markers increase in response to inflammation, and MCP-1 may enhance the activity of CD68-positive macrophages. MCP-1 levels are further influenced by macrophage polarization during inflammatory processes, reinforcing the close relationship between these two markers <sup>33,34,35</sup>

Receiver Operating Characteristic (ROC) curve analysis confirmed the diagnostic accuracy and specificity of the immunological markers. While PSA was useful for identifying prostatic infection in both ABP and CBP, its diagnostic accuracy was limited due to overlapping elevations across various prostatic disorders. In contrast, MCP-1 showed high sensitivity and specificity in diagnosing bacterial prostatitis <sup>35</sup>. This finding supports the conclusion that MCP-1 is a reliable biomarker for identifying bacterial infections <sup>36,37</sup>. Similarly, CD68 demonstrated notable diagnostic accuracy and sensitivity in both acute and chronic conditions.

#### **CONCLUSION**

In both acute and chronic prostatitis, significantly elevated levels of MCP-1 and CD68 were observed, clearly reflecting the presence of inflammatory activity in prostate tissue. Furthermore, both MCP-1 and CD68 demonstrated high sensitivity and specificity compared to PSA, thereby enhancing their diagnostic accuracy.

Ethical approval and consent of patients: This study was approved by the ethics committee of the Department of Clinical Laboratories/College of Applied Medical Sciences/University of Kerbala.Reference No: CLAMSKU/13. After the Ministry of Health and Environment granted permission to conduct the study, samples were collected, and the study started .The study participants gave their permission to collect sociodemographic data as well as undertake experiments on the selected samples with respect for patient confidentiality.

## **Funding Statement: Nil funding**

**Conflict of Interest declaration:** We declare that we have no affiliations with or involvement in any organization or entity with any financial interest in the subject matter or materials discussed in this manuscript.

**Data Access Statement:** Research data supporting this publication are available at the location

## **REFERENCES**

- 1. Lam JC, Lang R, Stokes W. How I manage bacterial prostatitis. Clin Microbiol Infect. 2023;29(1):32-37. doi:10.1016/j.cmi.2022.05.035.
- 2. Hua L, Gao X, Zhan J, Wu X, Liu H. Prostatitis and male infertility. Aging Male. 2025;28(1):2494550. doi:10.1080/13685538.2025.2494550.
- 3. Lupo F, Lang R, Stokes W, et al. The immune system fails to mount a protective response to Gram-positive or Gram-negative bacterial prostatitis. J Immunol. 2020;205(10):2763-2777. doi:10.4049/jimmunol.2000587.
- 4. Oseni SO, Naar C, Pavlović M, Asghar W, Hartmann JX, Fields GB, Kumi-Diaka J. The molecular basis and clinical consequences of chronic inflammation in prostatic diseases: prostatitis, benign prostatic hyperplasia, and prostate cancer. Cancers. 2023;15(12):3110. doi:10.3390/cancers15123110.
- 5. Korzhevskii DE, Kirik OV. Brain microglia and microglial markers. Neurosci Behav Physiol. 2016;46:284-290. doi:10.1007/s11055-016-0231-z.
- Vasquez JDS, Nkongolo S, Traum D, Sotov V, Kim SC, Mahamed D, Mehrotra A, Patel A, Chen D, Fung S, Gaggar A, Feld JJ, Chang KM, Wallin JJ, Jansen HLA, Gehring AJ. Virus-associated inflammation imprints an inflammatory profile on monocyte-derived macrophages in the human liver. J Clin Invest. 2025;135(8). doi:10.1172/JCI17524.
- 7. Shukla S, Telraja J, Yadav M, Prakash H. Modulation of macrophage signaling pathways during bacterial infections. Front Cell Infect Microbiol. 2021;11:689759. doi:10.3389/fcimb.2021.689759.
- 8. Serbina NV, Pamer EG. Monocyte emigration from bone marrow during bacterial infection requires signals mediated by chemokine receptor CCR2. Nat Immunol. 2006;7(3):311-317. doi:10.1038/ni1309.
- 9. Balamayooran G. Essential role of monocyte chemoattractant protein-1 in gram-negative bacterial pneumonia. [Dissertation]. Louisiana State University; 2012. Available from: <a href="https://repository.lsu.edu/gradschool/dissertations/3/407">https://repository.lsu.edu/gradschool/dissertations/3/407</a>.

- 10. Zhu T, Liao X, Feng T, Wu Q, Zhang J, Cao X, Li H. Plasma monocyte chemoattractant protein 1 as a predictive marker for sepsis prognosis: a prospective cohort study. Tohoku J Exp Med. 2017;241(2):139-147. doi:10.1620/tjem.241.139.
- 11. Ahn HK, Koo KC, Chung BH, Lee KS. Comparison of the delta neutrophil index with procalcitonin, erythrocyte sedimentation rate, and C-reactive protein as predictors of sepsis in patients with acute prostatitis. Prostate Int. 2018;6(4):157-161. doi:10.1016/j.prnil.2018.05.001.
- 12. Lavoignet CE, Le Borgne P, Chabrier S, et al. White blood cell count and eosinopenia as valuable tools for the diagnosis of bacterial infections in the ED. Eur J Clin Microbiol Infect Dis. 2019;38(8):1523-1532. doi:10.1007/s10096-019-03583-2.
- 13. Vermassen T, Van Praet C, Poelaert F, Lumen N, Decaesteker K, Hoebeke P, Van Belle S, Rottey S, Delanghe J. Diagnostic accuracy of urinary prostate protein glycosylation profiling in prostatitis diagnosis. Biochem Med (Zagreb). 2015;25(3):439-449. doi:10.11613/BM.2015.045.
- 14. Sunjaya J, Bororing SR, Iryaningrum MR. Association between urine leukocytes and the types of urinary tract infection: A systematic review. Indones J Urol. 2022;29(1):57-64. doi:10.32421/juri.v29i1.746.
- 15. Banza MI, Kasanga TK. Prostatites aiguës sur prostate non tumorale aux cliniques universitaires de Lubumbashi: aspects épidémio-cliniques et thérapeutiques. Pan Afr Med J. 2020;37(1):227. Available from: <a href="https://www.ajol.info/index.php/pamj/article/view/227235">https://www.ajol.info/index.php/pamj/article/view/227235</a>.
- 16. Al Hadrawi KK, Aldhalimi HK, Al-Hadrawi HA. Evaluation of some immune parameters in infertile men. Lat Am J Pharm. 2024;43:831-835. Available from:
  - https://www.researchgate.net/publication/38226435
- 17. He J, Han Z, Luo W, et al. Serum organic acid metabolites can be used as potential biomarkers to identify prostatitis, benign prostatic hyperplasia, and prostate cancer. Front Immunol. 2023;13:998447. doi:10.3389/fimmu.2022.998447.
- 18. Lee HY, Wang JW, Juan YS, et al. The impact of urine microbiota in patients with lower urinary tract symptoms. Ann Clin Microbiol Antimicrob. 2021;20:23. doi:10.1186/s12941-021-00428-9.
- Nagaratnam N, Nagaratnam K, Cheuk G. Prostate gland and related disorders. In: Geriatric Diseases. Springer; 2018:297-307. doi:10.1007/978-3-319-33434-9 34.

- 20. Song L, Lee C, Schindler C. Deletion of the murine scavenger receptor CD68. J Lipid Res. 2011;52(8):1542-1550. doi:10.1194/jlr.M015412.
- 21. Yong KK, Chang JH, Chien MH, et al. Plasma monocyte chemoattractant protein-1 level as a predictor of the severity of community-acquired pneumonia. Int J Mol Sci. 2016;17(2):179. doi:10.3390/ijms17020179.
- 22. Chamoun MN, Darville T, et al. Restriction of chronic Escherichia coli urinary tract infection depends upon T cell-derived interleukin-17, a deficiency of which predisposes to flagella-driven bacterial persistence. FASEB J. 2020;34(11):14572-14587. doi:10.1096/fj.202000760R.
- 23. Sherbet GV, editor. Growth factors and their receptors in cell differentiation, cancer, and cancer therapy. Elsevier; 2011. Available from: https://books.google.iq/books?id=9qlS8leBmJcC.
- 24. Balamayooran G, Batra S, Balamayooran T, et al. Monocyte chemoattractant protein 1 regulates pulmonary host defense via neutrophil recruitment during Escherichia coli infection. Infect Immun. 2011;79(7):2567-2577. doi:10.1128/IAI.00067-11.
- Kadim MM, AL-Dahmoshi HO, AL-Khikani FH. Sepsis biomarkers: current information and future visions. Microbes Infect Dis. 2024;5(1):201-210. doi:10.21608/mid.2023.205161.1508.
- 26. Mohamed AA, Salah W, Hassan MB, Eldeeb HH, Adaroas AS, Khattab RA, et al. MCP1, CRP and Procalcitonin as Novel Diagnostic Markers in Cirrhotic Patients with Spontaneous Bacterial Peritonitis. The Open Biomarkers Journal. 2022;12(1). doi:10.2174/18753183-v12-e2206270.
- 27. Pan SC, Wu YF, Lin YC, Cheng CM. Monocyte chemoattractant protein-1 detection in wound tissue fluids for the assisted diagnosis of wound infection. Surgery. 2024;176(1):154-161. doi:10.1016/j.surg.2024.03.003.
- 28. Devaraj S, Jialal I. C-reactive protein polarizes human macrophages to an M1 phenotype and inhibits transformation to the M2 phenotype. Arterioscler Thromb Vasc Biol. 2011;31(6):1397-1402. doi:10.1161/ATVBAHA.111.225508.
- Krizak J, Frimmel K, Bernatova I, Navarova J, Sotnikova R, Okruhlicova L. The effect of omega-3 polyunsaturated fatty acids on endothelial tight

- junction occludin expression in rat aorta during lipopolysaccharide-induced inflammation. Iran J Basic Med Sci. 2016;19(3):290-299.
- 30. Falagario U, Selvaggio O, Carrieri G, Barret E, Sanguedolce F, Cormio L. Prostatic inflammation is associated with benign prostatic hyperplasia rather than prostate cancer. J Gerontol Geriatr. 2018;66:178-182. Available from: <a href="https://www.jgerontology-geriatrics.com/article/view/67">https://www.jgerontology-geriatrics.com/article/view/67</a>.
- 31. Moser PL, Brunner A, Horninger W, Bartsch G, Mikuz G. Correlation between inflammatory cells (T and B lymphocytes, macrophages) in prostate biopsies and elevated PSA levels in a PSA screening population. Urology. 2002;59(1):68-72. doi:10.1016/S0090-4295(01)01493-5.
- 32. Atri C, Guerfali FZ, Laouini D. Role of human macrophage polarization in inflammation during infectious diseases. Int J Mol Sci. 2018;19(6):1801. doi:10.3390/ijms19061801.
- 33. Warford J, Lamport A-C, Kennedy B, Easton AS. Human brain chemokine and cytokine expression in sepsis: A report of three cases. Can J Neurol Sci. 2017;44(1):96-104. doi:10.1017/cjn.2016.310.
- 34. Fujita K, Ewing CM, Getzenberg RH, et al. Monocyte chemotactic protein-1 (MCP-1/CCL2) is associated with prostatic growth dysregulation and benign prostatic hyperplasia. Prostate. 2010;70(5):473-481. doi:10.1002/pros.21081.
- 35. Krisna DM, Hariatmoko H, Pinzon RT. Akurasi peningkatan prostate-specific antigen dalam darah terhadap keganasan prostat. JKKI: Jurnal Kedokteran dan Kesehatan Indonesia. 2018;9(1):24-29. doi:10.20885/JKKI.Vol9.Iss1.art5.
- 36. El-Toukhy N, Emam SM. Diagnostic and prognostic values of monocyte chemotactic protein1 in ascitic fluid of patients with spontaneous bacterial peritonitis. Egypt J Immunol. 2016;23(2):17-27. Available from: <a href="https://europepmc.org/article/med/28502130">https://europepmc.org/article/med/28502130</a>.
- 37. Chen Z, Li C, Yu J. Monocyte chemoattractant protein-1 as a potential marker for patients with sepsis: a systematic review and meta-analysis. Front Med (London). 2023;10:1217784. doi:10.3389/fmed.2023.1217784