# **High-Throughout Techniques Assessing the Molecular Diagnosis**

# Eman E.A. Mohammed\*, Ghada M. M. Al-Ettribi, Tamer H.A. Ammar

for Neurometabolic Disorders: A Comprehensive Review

Medical Molecular Genetics Department, Human Genetics and Genome Research Institute, National Research Centre, Cairo, Egypt

## **ABSTRACT**

Key words: Neurometabolic disorders, High-Throughput techniques, Next-Generation sequencing, NGS, WES, WGS

\*Corresponding Author:
Eman E.A. Mohammed, PhD,
Medical Molecular Genetics
Department,
Human Genetics and Genome
Research Institute,
National Research Centre,
Cairo, Egypt.
Tel: 002-01012802875
em.mohammed99@gmail.com
ORCID: 0000-0001-8860-0980

Neurometabolic diseases associated with neurological manifestations are a group of heterogeneous genetic disorders that share the alteration in specific aspects of cellular metabolism, ultimately leading to the disease. Often first described by pediatricians, the more recently reported adult-onset forms have phenotypes considerably different, sometimes, from the pediatric ones and might mimic other more common adults neurological disorders. Adult-onset neurometabolic diseases are individually rare, heterogeneous and frequently show complex clinical presentations. These reasons, in addition to the myriad of specialized biochemical diagnostic tools available, account for a significant diagnostic delay and under-diagnosis. However, unlike many other neurogenetic diseases, a substantial part of neurometabolic diseases can be successfully treated, with both conservative and more recently approved innovative therapeutics. Early recognition and diagnosis of a treatable neurometabolic disease should have major impacts, supports the stabilization of the disease or even the regression of some signs and symptoms, halting unnecessary diagnostic investigations, and allowing for family screening and treatment of pre-symptomatic carriers. An overview of adult-onset neurometabolic diseases will be outlined, starting from important general considerations to phenotypical descriptions focusing on treatable diseases. A diagnostic approach for the adult neurologist will be developed to help decision making when suspecting a neurometabolic disease. Neurometabolic Syndromes (NMS) are rare, heterogeneous genetic disorders that primarily affect the central nervous system (CNS) due to defects in specific metabolic pathways. Accurate, early diagnosis is critically important for initiating disease-modifying therapies, such as enzyme replacement or gene therapy, before irreversible neurological damage occurs.

# **INTRODUCTION**

Inborn errors of metabolism (IEMs) are also referred to as congenital metabolic diseases (CMDs) or inherited metabolic disorders (IMDs). They are a group of metabolic disorders caused by a genetic mutation that leads to deficiency of an enzyme required for the formation of a protein or for the catalysis of a biochemical reaction in the body. The problem arises usually from accumulation of substrate (toxic) or absence of an important product of the reaction. IEM can appear at birth or later in life such as phenylketonuria, albinism, lactose intolerance, Mucopolysaccharidosis (MPS), Gaucher disease, fabry disease ....etc <sup>1</sup>.

Genetic mutations that affect the activity of the acid hydrolases causes what are known as lysosomal storage diseases (LSD). These mutations block the normal metabolism causing macromolecules accumulation inside the lysosomes that eventually lead to severe physiological damage. Lysosomal storage diseases have a rare incidence of 1 in 7000 live birth <sup>2</sup>.

LSDs symptoms differ according to the affected enzyme, the type of accumulated substance and the type of cells that substance accumulates in. Also clinical picture can vary markedly within the same disorder <sup>3, 4</sup>.

Online ISSN: 2537-0979

Lysosomal storage disorders (LSDs) constitute a group of >50 inherited disorders characterized by the accumulation of specific undegraded metabolites in the lysosomes. This over storage is commonly caused by a deficient or absent activity of one of the many lysosomal hydrolases or, in a few cases, by the deficiency of other non-enzymatic lysosomal proteins. Although singularly considered rare, the combined birth prevalence of LSD is estimated from 7.5 to 23.5 per 100,000 live births. Clinical signs and symptoms may occur from the prenatal period to adulthood, and may develop at different progression rate, according to the pathology, leading to a wide spectrum of disease forms, from mild to extremely severe, that in most cases affect the neurologic compartment, hence the neurometabolic disorders 4,5

Generally, the diagnostic approach includes an accurate clinical evaluation that leads to the formulation

of a suspicion for one or more of the neurometabolic disorders. This is followed by biochemical tests, aimed to detect the storage products in the body fluids, the biochemical results might be oriented by enzymatic analyses' aspects. Finally, if an enzyme deficiency is detected, genetic analysis is performed for the suspected gene <sup>6</sup>. However, this diagnostic route presents several limitations.

Neurometabolic disorders are largely overlapping in their phenotypic manifestations, hence their identification requires deep clinical phenotyping. Moreover, the biochemical methods are laborious, and they are often subject to high variability, taking into consideration that multiple enzymatic assays should be expensive. Also, not all of these disorders present with

elevated levels of storage products. This may delay the diagnosis that could be even difficult to reach <sup>7</sup>.

The diagnosis of neurometabolic disorders is a demanding task for clinicians due to the phenotype and penetrance variability, the shared signs and symptoms, and the uncertainties related to biochemical enzymatic assay results <sup>6</sup>.

Advanced gene sequencing and analysis techniques constitute the most reliable current approach for diagnosing, preventing, and treating this complex group of disorders. High-Throughput Technologies, such as Next-generation sequencing (NGS)<sup>8,9</sup>; including whole-exome sequencing (WES) and targeted gene panels have revolutionized this field.

Table 1: Examples of Genes that will be Included in the Panel and Their Associated Neurometabolic disorders<sup>7</sup>

Gene	Syndrome	Enzyme/substrate
SGSH	MPS IIIA, Sanfilippo	N-Sulfoglucosamine
	syndrome A	Sulfohydrolase (Heparan sulfate)
NAGLU	MPS IIIB, Sanfilippo	e N-Acetyl-α-glucosaminidase (Heparan sulfate)
	syndrome B	
HGSNAT	MPS IIIC, Sanfilippo	Heparan-α-glucosaminideN-acetyltransferase (Heparan
	syndrome C	sulfate)
GNS	MPS IIID, Sanfilippo	N-acetylglucosamine-6-
	syndrome D	Sulfatase (Heparan sulfate)
GLA	Fabry disease	α-Galactosidase A (Globotriaosylceramide)
GLB1	GM1 gangliosidosis:	β-Galactosidase
	type I, II, III	(GM1 ganglioside, keratan sulfate & oligosaccharides)
HEXA	GM2 gangliosidosis,	β-Hexosaminidase
	Tay–Sachs disease	(GM2 ganglioside, Glycosphingolipids& oligosaccharides)
HEXB	GM2 gangliosidosis,	β-Hexosaminidase
	Sandhoff disease	(GM2 ganglioside, GA2 glycolipid & oligosaccharides)
GALC	Krabbe disease (KD)	galactocerebrosidase

Neurometabolic disordersis clinically heterogeneous group in which the neurological manifestations are prominent clinical Lysosome facilitates the degradation of various products of the cellular turn-over that are mainly derived from lysosomes through endocytosis. Alternative pathways for substrate entry into lysosomes are phagocytosis. Mucopolysaccharidoses comes from abnormalities in the turnover of keratin sulfate, heparin, and dermatan sulfate and Fabry disease results from alpha-galactosyl sphingolipids oligosaccharides. GM1 gangliosidosis is the results of a beta-galactosidase deficiency and the accumulation of ceramides caused by an acid ceramidase deficiency that causes Farber disease. GM2 gangliosidosis as Tay-Sachs disease (an alpha subunit of beta hexosaminidase deficiency); Sandhoff disease (a beta subunit of beta-hexosaminidase deficiency); and the GM2 AB variant are other types of lysosomal storage diseases. The complex lipid of gangliosides is

found predominantly in gray matter in the brain. Classic forms of gangliosidoses present in early and late infancy and are usually fatal during these periods. In the early infantile form of GM1, gangliosidoses dysmorphic features may present at birth and often hepatosplenomegaly is noted ``.

Fabry disease is transmitted as X-linked traits however most neurometabolic disorders are transmitted in autosomal recessive inheritance. Fortunately, enzyme replacement therapy (ERT) for Fabry disease is available. The early timing of therapeutic interventions is of utmost importance because of the limited regenerative capacity of the brain <sup>11</sup>.

Targeted sequencing is an appealing approach to implement in routine diagnostic strategy, given its low sequencing costs and short sequencing time <sup>9,12-14</sup>. However, a good coverage must be ensured and, when this is not reached, validation by Direct sequencing needs to be performed on the proband and on the

parents as final step 15. Moreover, the possibility to fill the gaps in the panel design must be guaranteed, especially in case of strong suspicion for a specific disease <sup>16</sup>. Sanger sequencing still remains a reliable sequencing technique, and it should be considered an important support to NGS approaches, especially for confirmation of variants with a coverage below the good coverage threshold<sup>14</sup>. Therefore, each laboratory should have a diagnostic flow chart, providing appropriate molecular genetic tools to address the clinical suspicion <sup>17</sup>. We believe that the application of the panel or, in a near future, of a WES or WGS analysis will be the first or one of the first steps in diagnostic route, supported by a thorough phenotyping of the patients <sup>16</sup>. A tight collaboration between clinics and laboratory could increase the yield of the diagnostic process of neurometabolic disorders.

Genomics takes a comprehensive view that implicates all the genes within an organism, including protein-coding genes, RNA genes, cis- and transelements, etc. It is a data-driven science involving the high-throughput technological development of next-generation sequencing (NGS) that generates the entire DNA data of an organism. These techniques include whole genome sequencing (WGS), whole exome sequencing (WES), and transcriptomic and proteomic profiling <sup>18–23</sup>. With the recent rapid accumulation of these omics data, increased attention has been paid to bioinformatics and machine learning (ML) tools with established superior performance in several genomics implementations <sup>24</sup>.

## 1. The Power of High-Throughput Omics

High-throughput technologies, collectively known as 'omics', allow for the simultaneous measurement of thousands of molecules in a single biological sample (blood, urine, CSF, or tissue). This shifts diagnosis from searching for a single missing enzyme to identifying an entire molecular signature of the disease.

#### 1.1 Genomics and Transcriptomics

Whole Exome Sequencing (WES) & Whole Genome Sequencing (WGS) rapidly identify mutations in all known disease-causing genes and can uncover novel genetic variants. WES/WGS drastically reduces the diagnostic odyssey for NMS patients, whose clinical symptoms often overlap with common neurodevelopmental disorders.

**Transcriptomics (RNA-Seq) me**easures the expression levels of all RNA molecules. This helps in understanding the functional consequence of a mutation—for example, if a variant causes abnormal splicing or reduced expression of a critical enzyme.

# 1.2 Metabolomics and Proteomics

**Metabolomics technique** <sup>25</sup> measures small molecule metabolites (e.g., amino acids, organic acids, lipids, GAGs). This is arguably the most critical high-throughput technology for NMS, as the accumulation of specific, undegraded metabolites (like heparan sulfate in

Sanfilippo syndrome) is the direct cause of the pathology. High-resolution mass spectrometry (HRMS) enables the simultaneous detection and quantification of hundreds of these molecules.

**Proteomics approach** <sup>26</sup> measures the quantity and modification of all proteins. For NMS, this can reveal downstream effects, such as elevated inflammatory proteins (cytokines) or changes in lysosomal proteins due to storage stress.

# 2. ML/AI in the Diagnostic Pipeline

The immense volume and complexity of high-throughput data generated by 'omics (terabytes of genomic data, hundreds of metabolites per sample) require sophisticated computational tools. AI and ML algorithms are essential for extracting meaningful, actionable insights from this noise <sup>27,28</sup>.

#### 2.1 Pattern Recognition and Classification

ML models, such as **Support Vector Machines** (SVM) and **Random Forests**, excel at binary classification (Disease vs. Healthy) or multi-class classification (Subtype A vs. Subtype B). AI automatically identifies the most important 'features' (e.g., a specific ratio of three metabolites plus two gene expression values) that distinguish a disease state, effectively simplifying complex biological reality into a diagnostic signature <sup>29</sup>.

ML models can be trained on improving newborn screening data (dried blood spots) to differentiate between true positive cases and common false positives, thus reducing unnecessary follow-up tests and anxiety for families.

### 2.2 Biomarker Discovery and Validation

Many NMS lack effective, non-invasive biomarkers, especially those reflecting CNS involvement. ML is transforming this search, **multi-Omics integration by using the** AI models use techniques like **Factor Analysis** or **Deep Neural Networks** to integrate different 'omics layers (e.g., genomic variants, plasma metabolites, and clinical symptoms) to discover novel **panels of biomarkers** that are more sensitive and specific than single-molecule markers. For example; identifying a panel of neuro-inflammatory cytokines (from proteomics) combined with a specific urine GAG profile (from metabolomics) that together indicate advanced stages of neurodegeneration in MPS III <sup>30</sup>.

ML analyzes sensor data (wearable devices, sleep monitors) to quantify clinical features like gait, hyperactivity, or sleep disruption, generating **digital biomarkers** that correlate with disease progression and therapeutic response.

# Analysis of Neurometabolic Disorder Gene Interactions Using GeneMANIA

The GeneMANIA database <sup>31</sup> was utilized as a key bioinformatic tool to systematically explore and visualize the functional associations and interaction networks among the genes and proteins implicated in

the ten Neurometabolic Disorders (NMDs) detailed in Table 1.

This approach provides a contextual framework for understanding the molecular complexity underlying these heterogeneous syndromes. The resulting

comprehensive network, presented in Figure 1, illustrates various modes of interaction, including; physical interactions, co-expression, co-localization, genetic interactions, and pathway membership.

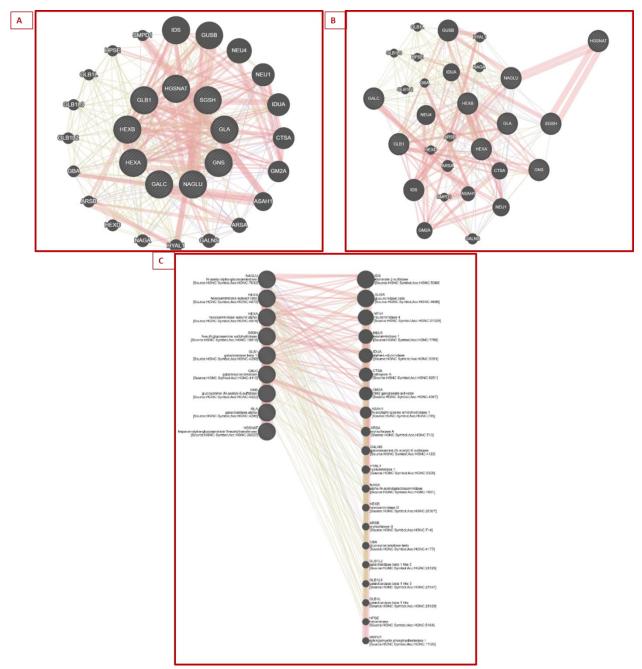


Fig. 1: GeneMANIA analysis showing the functional associations among the genes and proteins related to the ten Neurometabolic disorders (NMDs) in Table 1. Represent integrated network evidence derived from physical interactions, co-expression, co-localization, genetic interactions, and pathway membership.

Online ISSN: 2537-0979

The construction of this network serves to identify key hub genes that share interaction partners across multiple NMDs. This insight is crucial for revealing shared molecular mechanisms and identifying potential convergence points in the disease pathology, which may suggest common diagnostic markers or therapeutic targets across seemingly distinct disorders.

The data derived from the GeneMANIA analysis, visually represented in Figure 1, provides a powerful, evidence-based foundation for interpreting the functional relationships of the proteins associated with the NMDs listed in Table 1, thereby enriching our understanding of their underlying etiology.

#### **Egyptian Experiences**

Our team's research has centered on leveraging advanced computational and molecular methodologies to enhance the diagnosis and therapeutic understanding of rare genetic disorders, with a specific focus on lysosomal storage diseases. Our core contribution to Sanfilippo syndrome research <sup>32–34</sup> lies in the development and validation of a novel gene-specific Bayesian Gaussian mixture achine learning (ML) model to predict the missense variant pathogenicity of Sanfilippo syndrome <sup>35</sup>. This hybrid computational tool is specifically designed to accurately predict the pathogenicity of novel missense variants across all known subtypes of the syndrome. This predictive capability is critical for accelerating molecular diagnosis and clarifying the clinical significance of variants of uncertain significance (VUSs) identified during highthroughput genetic screening.

We have conducted comprehensive studies providing genetic insights into Fabry disease <sup>36</sup>. Furthermore, our team has successfully secured approval and executed therapeutic experiments aimed at evaluating and validating potential treatment strategies for this X-linked lysosomal storage disorder.

Our work extends beyond these specific disorders, encompassing several other genetic conditions <sup>37–42</sup>. A primary strategic focus moving forward is the expanded application of Artificial Intelligence (AI) and Machine Learning (ML) techniques. This integration will be instrumental in Improving Diagnostic Accuracy by utilizing AI/ML for the analysis of complex genetic and clinical data to significantly increase the precision and speed of molecular diagnosis across a broader spectrum of rare diseases.

# **CONCLUSION**

The convergence of high-throughput 'omics and AI/ML is transforming the diagnosis of neurometabolic syndromes. By moving beyond single-gene or single-enzyme defects to interpret holistic, multi-layered molecular signatures, these technologies are paving the way for faster, more accurate diagnoses, better patient stratification, and the discovery of novel therapeutic targets. This ensures that life-saving interventions can

be delivered at the earliest possible stage, significantly improving outcomes for children affected by these rare disorders.

Applying computational models to screen, prioritize, and predict the efficacy of novel compounds, thereby streamlining the development and personalization of future therapeutic interventions. This ongoing research portfolio demonstrates our commitment to transforming the field of rare disease diagnostics and treatment through the strategic convergence of genomics, bioinformatics, and artificial intelligence.

#### REFERENCES

- Jeanmonod R, Asuka E, Jeanmonod D. Inborn Errors Of Metabolism. In: StatPearls [Internet]. Treasure Island (FL): StatPearls Publishing; 2022 [cited 2022 Dec 11]. Available from: http://www.ncbi.nlm.nih.gov/books/NBK459183/
- Sanderson S, Green A, Preece MA, Burton H. The incidence of inherited metabolic disorders in the West Midlands, UK. Arch Dis Child. 2006 Nov;91(11):896–9.
- 3. Suzuki Y. β-Galactosidase deficiency (-galactosidosis): GM1 gangliosidosis and Morquio B disease. Metab Mol Bases Inherit Dis. 1995;2785–823.
- 4. La Cognata V, Guarnaccia M, Polizzi A, Ruggieri M, Cavallaro S. Highlights on Genomics Applications for Lysosomal Storage Diseases. Cells. 2020 Aug 14;9(8):1902.
- 5. Ferreira CR, Gahl WA. Lysosomal storage diseases. Transl Sci Rare Dis. 2017;2(1–2):1–71.
- Fernández-Eulate G, Carreau C, Benoist JF, Lamari F, Rucheton B, Shor N, et al. Diagnostic approach in adult-onset neurometabolic diseases. J Neurol Neurosurg Psychiatry. 2022 Apr 1;93(4):413–21.
- Zanetti A, D'Avanzo F, Bertoldi L, Zampieri G, Feltrin E, De Pascale F, et al. Setup and Validation of a Targeted Next-Generation Sequencing Approach for the Diagnosis of Lysosomal Storage Disorders. J Mol Diagn JMD. 2020 Apr;22(4):488– 502.
- 8. Slatko BE, Gardner AF, Ausubel FM. Overview of Next-Generation Sequencing Technologies. Curr Protoc Mol Biol [Internet]. 2018 Apr [cited 2022 Dec 11];122(1). Available from: https://onlinelibrary.wiley.com/doi/10.1002/cpmb.5
- Gulilat M, Lamb T, Teft WA, Wang J, Dron JS, Robinson JF, et al. Targeted next generation sequencing as a tool for precision medicine. BMC Med Genomics. 2019 Jun 3;12(1):81.
- 10. Regier DS, Proia RL, D'Azzo A, Tifft CJ. The GM1 and GM2 Gangliosidoses: Natural History

- and Progress toward Therapy. Pediatr Endocrinol Rev PER. 2016 Jun;13 Suppl 1(Suppl 1):663–73.
- Bokhari SRA, Zulfiqar H, Hariz A. Fabry Disease.
   In: StatPearls [Internet]. Treasure Island (FL): StatPearls Publishing; 2022 [cited 2022 Dec 11]. Available from: http://www.ncbi.nlm.nih.gov/books/NBK435996/
- 12. Komlosi K, Sólyom A, Beck M. The Role of Next-Generation Sequencing in the Diagnosis of Lysosomal Storage Disorders. J Inborn Errors Metab Screen. 2016 Jan 1;4:2326409816669376.
- 13. Ying S, Zhihua Z, Yucan Z, Yu J, Qian L, Bixia Z, et al. Molecular Diagnosis of Panel-Based Next-Generation Sequencing Approach and Clinical Symptoms in Patients With Glycogen Storage Disease: A Single Center Retrospective Study. Front Pediatr. 2020;8:600446.
- Encarnação M, Coutinho MF, Silva L, Ribeiro D, Ouesleti S, Campos T, et al. Assessing Lysosomal Disorders in the NGS Era: Identification of Novel Rare Variants. Int J Mol Sci. 2020 Sep 1;21(17):6355.
- 15. Crossley BM, Bai J, Glaser A, Maes R, Porter E, Killian ML, et al. Guidelines for Sanger sequencing and molecular assay monitoring. J Vet Diagn Invest. 2020 Nov;32(6):767–75.
- 16. Tarailo-Graovac M, Wasserman WW, Van Karnebeek CDM. Impact of next-generation sequencing on diagnosis and management of neurometabolic disorders: current advances and future perspectives. Expert Rev Mol Diagn. 2017 Apr 3;17(4):307–9.
- 17. Fernández-Marmiesse A, Morey M, Pineda M, Eiris J, Couce ML, Castro-Gago M, et al. Assessment of a targeted resequencing assay as a support tool in the diagnosis of lysosomal storage disorders. Orphanet J Rare Dis. 2014 Apr 25;9:59.
- Auffray C, Imbeaud S, Roux-Rouquié M, Hood L. From functional genomics to systems biology: concepts and practices. C R Biol. 2003;326(10– 11):879–92.
- 19. Honoré B, Østergaard M, Vorum H. Functional genomics studied by proteomics. Bioessays. 2004;26(8):901–15.
- 20. Kulasingam V, Pavlou MP, Diamandis EP. Integrating high-throughput technologies in the quest for effective biomarkers for ovarian cancer. Nat Rev Cancer. 2010;10(5):371–8.
- 21. Goldfeder RL, Priest JR, Zook JM, Grove ME, Waggott D, Wheeler MT, et al. Medical implications of technical accuracy in genome sequencing. Genome Med. 2016;8(1):24.

- 22. Nariai N, Kolaczyk ED, Kasif S. Probabilistic protein function prediction from heterogeneous genome-wide data. Plos One. 2007;2(3):e337.
- 23. Van Dijk EL, Auger H, Jaszczyszyn Y, Thermes C. Ten years of next-generation sequencing technology. Trends Genet. 2014;30(9):418–26.
- 24. Ritchie MD, Holzinger ER, Li R, Pendergrass SA, Kim D. Methods of integrating data to uncover genotype–phenotype interactions. Nat Rev Genet. 2015;16(2):85–97.
- 25. Goodacre R, Vaidyanathan S, Dunn WB, Harrigan GG, Kell DB. Metabolomics by numbers: acquiring and understanding global metabolite data. TRENDS Biotechnol. 2004;22(5):245–52.
- 26. Pandey A, Mann M. Proteomics to study genes and genomes. Nature. 2000;405(6788):837–46.
- 27. Miotto R, Wang F, Wang S, Jiang X, Dudley JT. Deep learning for healthcare: review, opportunities and challenges. Brief Bioinform. 2018;19(6):1236–46.
- 28. Topol EJ. High-performance medicine: the convergence of human and artificial intelligence. Nat Med. 2019;25(1):44–56.
- 29. Alsaid Alyousef A. Combined supervised and unsupervised learning to identify subclasses of disease for better prediction. 2022;
- 30. Zhang X, Rahnavard A, Crandall KA. Machine learning enhances biomarker discovery: From multi-omics to functional genomics. Med Res Arch. 2025;13(8).
- 31. Franz M, Rodriguez H, Lopes C, Zuberi K, Montojo J, Bader GD, et al. GeneMANIA update 2018. Nucleic Acids Res. 2018;46(W1):W60–4.
- 32. Fateen E, Nosier SS, Aziz NNA, Radwan AM, Mohammed EE. Clinical, biochemical, and molecular characteristics of Sanfilippo a syndrome (MPS IIIA) in a cohort of Egyptian patients. Orphanet J Rare Dis. 2025;20(1):454.
- 33. Fateen E, Nosier SS, Radwan AM, Mohammed EE. Clinical, biochemical, and molecular characterization of a cohort of Egyptian patients with Sanfilippo B syndrome (MPS IIIB): Bayesian Gaussian mixture model. Mol Biol Rep. 2025;52(1):1–12.
- 34. Mohammed EE, Fateen EM. Identification of three novel Homozygous NAGLU mutations in Egyptian patients with Sanfilippo Syndrome B. Meta Gene. 2019;21:100580.
- 35. Mohammed EE, Fayez AG, Abdelfattah NM, Fateen E. Novel gene-specific Bayesian Gaussian mixture model to predict the missense variants pathogenicity of Sanfilippo syndrome. Sci Rep. 2024;14(1):12148.

- Fateen E, Mohammed EE, Abdel-Aziz NN, Abdallah ZY, Elhossini RM, Aglan M. Genetic insights and therapeutic implications in Egyptian patients with Fabry Disease. Egypt Pharm J. 2025; 25(1):11-22.
- 37. Mohammed EE, Mostafa MI, AlEttribi GM, Ammar TH, Abd Elazeem AF, Ahmed NE moataz B, et al. Clinical and molecular characterization of patients with Gardner syndrome using whole exome sequencing. Egypt Pharm J. 2025; 24(4):97–103.
- 38. Ammar TH, Elshebawy H, Hamed K, Mohammed EE, El-Hariri HM, El-Bassyouni HT. Key genetic variants with multiple sclerosis risk in Egyptian patients. Egypt Pharm J. 2025; 25(1):1-10.
- 39. Youssef SS, Abbas EAER, Hassany M, Shaaban ASR, Mohammed EEA, Elbaz T. Impact of differential detection of TM6SF2 rs58542926 mutation in circulating tumor DNA versus

- peripheral blood cells on hepatocellular carcinoma patients. Discov Oncol. 2025;16(1):1071.
- Elshebawy H, T El-Bassyouni H, Mohammed EE, Hamed K, Ramzy GM, El-Hariri H, et al. The association of GSTT1 deletion, HindIII C> G PAI-1, and rs11808092 polymorphisms with Parkinson's Disease susceptibility: A genetic study in an Egyptian Cohort. Egypt J Chem. 2025;68(13):473–82.
- 41. Khedr MA, Behairy BE, Basiouny HEDM, Zeitoon NA, Elfert AY, Zakaria HM, et al. Clinical and molecular study of Egyptian pediatric patients with Crigler-Najjar syndrome. Egypt Liver J. 2025;15(1):26.
- 42. Ammar TH, Al-Ettribi GM, Abo Hashish MM, Farid TM, Abou-Elalla AA, Thomas MM. Screening of GHSR, GHRHR, GH1 genes in isolated growth hormone deficiency disease in Egyptian patients. Egypt J Med Hum Genet. 2024;25(1):16.