

Zagazig J. Agric. Res., Vol. 43 No. (3) 2016

http://www.journals.zu.edu.eg/journalDisplay.aspx?Journalld=1&queryType=Master



IMPACT OF SOME PLANT ESSENTIAL OIL TREATMENTS ON CONTROLLING CHERRY TOMATOES SPOILAGE, IMPROVEMENT SHELF LIFE AND QUALITY ATTRIBUTES DURING STORAGE

Shaymaa M. Raafat^{1*}, M.I. Abou-Zaid², M.R. Tohamy² and H.E. Arisha³

1. Egyptian Plant Quarantine, Ismailia, Egypt

2. Plant Pathol. Dept., Fac. Agric. Zagazig Univ., Egypt

3. Hort. Dept., Fac. Agric. Zagazig Univ., Egypt

ABSTRACT

Food safety is one of the major issues to fresh fruits and vegetables. The preservation of freshness quality of cherry tomato (*Lycopersicon esculentum* Mill.) is relevant due to their economical impact. Under modified atmosphere (MA) (12% CO₂ + 6% O₂) cool storage conditions (7° C ± 1 and RH 90 ± 3%), some essential oils (black seed, fenugreek, cinnamon, spearmint, basil, and thyme) were used to treat, healthy and inoculated fruits with spoilage pathogens *Alternaria alternata* and *Botrytis cinerea*. The results indicated that fruits decay decreased in coated fruits treated with essential oils. However, as the storage period progressed, a general declining trend in all fruits quality parameters were observed for all treatments except for tomatoes treated with the essential oils, which, showing a lowest weight loss, firmness, Total Soluble Solids (TSS), pH, sugar fractions and antioxidant components (vitamin C., lycopene and phenolic fractions). The highest color changes were found in control group, inoculated fruits and untreated group. The obtained results suggested that essential plant oils may improve control decay of fruits, enhancement fruit shelf life and quality-related attributes on top of the well- documented antimicrobial protection during cold storage.

Key words: Essential oils, cherry tomato, quality parameters, spoilage, *Alternaria alternata, Botrytis cinerea*.

INTRODUCTION

Postharvest losses of fruits and vegetables is a serious problem caused by fungi, bacteria and unfavourable condition during marketing and storage. Fungal pathogens are mainly responsible for postharvest losses of fruits and vegetables (Korsten, 2006). Cherry tomato (Lycopersicon esculentum Mill. var. Catalena) is perishable and susceptible to postharvest diseases owing to the warm and humid climate. Alternaria alternata (Fr.) and Botrytis cinerea (Fr.Pers)., are a major saprophytic pathogens causing postharvest black rot and gray mold, respectively in cherry tomato (Feng and Zheng, 2007; Wang *et al.*, 2010; Tian *et al.*, 2011; Chen *et al.*, 2014 ; Yan *et al.*, 2014).

The use of synthetic preservatives as antimicrobial agents to control fungal spoilage of food has been practiced for many years. However, this had led a number of environmental and health problems because of the carcinogenicity, teratogenicity, high acute toxicity and long degradation periods of the synthetic preservatives (Lingk, 1991).

The antimicrobial properties of plant products such as essential oils, have been recognized and used for food preservation and in medicine since ancient times in the world (Tian *et at.*, 2011). Essential oils as antimicrobial agents have two

^{*}Corresponding author: Tel. : +201006276432 E-mail address: dr sh.m@hotmail.com

main advantages the first one, their natural origin means the majority of them are safer for consumers, and the second they have low risk of developing pathogen resistan strains (Cardile *et al.*, 2009).

A new worldwide trend to explore alternatives that control postharvest diseases, giving priority to decay- preventing methods with minimal impact on human health and environment. In addition, consumer have become more health- conscious regarding safety aspects concerning the handling of fruit and vegetables (Dulce *et al.*, 2010).

In this regard, treating fresh fruit and vegetables with edible coating and essential oils is an important approach that holds promise for the extension of storage life of fresh horticultural crops (Tabaestani *et al.*, 2013). These natural compounds are generally recognized as safe for environment and human health, so interest in their use in the goal for sustainable agriculture has increased and a lot of research has been done, proving, in many cases, that plant essential oils and extracts many have a role as pharmaceuticals and food preservations (Hammer *et al.*, 2001).

In recent years, numerous studies have documented the antifungal effect of plant essential oils to control food spoilage fungi (Tzortzakis, 2009; Dulce *et al.*, 2010; Shirzad *et al.*, 2011; Tian *et al.*, 2011; Xing *et al.*, 2011; Tabaestani *et al.*, 2013 ; Chen *et al.*, 2014; Yan *et al.*, 2014).

The essential oils of thyme, cinnamon, anise and spearmint have more effect on fungal development and subsequent mycotoxin production in wheat grains (Soliman and Badeaa, 2002).

Tian *et al.* (2011) reported that results of both *in vitro* and *in vivo* studies on fungal spoilage of cherry tomatoes indicate that employing dill (*Anethun graveolens* L.) oil as a significant potential fumigant against *A. alternata* and *Aspergillus* spp extended usual storage and transport period is very promising. Xing *et al.* (2011) suggested that microencapsulated cinnamon oil could reduce cherry tomato fruits decay and keep their quality.

In addition, cumin essential oil possess strong antifungal activity against *A. alternata*

and could be a promising natural products for use as anti - *A. alternata* agent to control black rot in cherry tomato (Tabaestani *et al.*, 2013).

In vivo A. alternata black rot disease incidence of cherry tomato treated with citronella essential oil was significantly reduced compared with the control treatment 5 days after storage at 25°C and 95% relative humidity (Chen *et al.*, 2014).

Evaluation of antifungal activity of thyme, ajowan, fennel and summer savory essential oils against postharvest gray mold (*Botrytis cinerea* PERS; FR) of kiwi fruits showed that increase in their tested concentrations and antifungal activity (Shirzad *et al.*, 2011).

The aim of this study was to improve cherry tomatoes storability by testing the effect of some essential oils on postharvest cherry tomatoes spoilage caused by *Alternaria alternata* and *Botrytis cinerea* and postharvest quality parameters.

MATERIALS AND METHODS

Plant Materials

Cherry tomatoes were harvested at red stage of maturity (red color covering between 60-90% of fruit surfaces) as described by (Helyes *et al.*, 2006). Healthy selected fruits were washed with distilled water then, disinfected by immersing them in ethanol 70% for 2 min. Sterilized fruits washed twice with sterile double – distilled water (5min each), and allowed to dry for 1 hr., in laminar flow.

Fungal Isolates and Detection the Tested Fungi

The tested fungi, *Alteraria alternata* and *Botrytis cinerea* were isolated from infected cherry tomato fruits showing typical symptoms of black rot and gray mold, respectively detected in Pl. Path. Lab. Fac. of Agriculture, Zagazig University according to key of Brown (1924) and Barnett (1962) and identified in Mycological Center of Assiout Univ. (MCAU). Identified stock cultures were maintained on Potato Dextrose Agar (PDA) slant medium at 4°C for sporulation.

Both fungi (*A. alternata* and *B. cinerea*) were grown on PDA medium and incubated at 25° C in the dark for 7 days. Resulted conidia were scrapped out from agar surface and suspended in sterile distilled water and fungal concentrations were adjusted to be 10^{6} conidia /ml using a haemacytometer.

Essential Oils

Pure essential oils of black seed (*Nigella sativa* L.), fenugreek (*Trigonella foenum-graecum* L.), cinnamon (*Cinnamomum zeylanicum* L.), spearmint (*Mentha peperita* L.) basil (*Ocimum basilicum* L.) and thyme (*Thymus vulgaris* L.), were purchased from International Flavor and Plant Oils Res Inst., Giza, Egypt. All plant oils were stored in dark bottles at 4°C for further studies.

In vivo Assessment of Antifungal Activity of some Essential Oils on Cherry Tomato Fruits Spoilage

The antifungal assay of the essential oils against spoilage cherry tomato fruits caused by *Alternaria alternata* and *Botrytis cinerea in vivo* was conducted as disease incidence and severity according to Tian *et al.* (2011) and Tabaestani *et al.* (2013).

The previously sterilized fruits were superficially wounded four times in the equator by the aid of sterilized stainless steel rod with a probe tip measured 1mm wide and 2 mm in length. Sterilized wounded fruits were dipped in spore suspension containing 10^6 spores / ml of *A. alternat*a and / or *B. cinerea* and incubated at 20°C for 24 hr., to resemble common fungal infections. Nevertheless wounded un inoculated, fruits were treated and preserved as a control treatments.

The plant oils treatments consisted of five minutes immersion of aforementioned fruits (inoculated wounded fruits) in plant oils individually (10% concentration) and / or in sterilized distilled water in case of control treatment. Fruits were then forced-air cooled to 5°C and placed in white plastic containers ($14 \times 11 \times 5$ cm) each contains 10 fruits.

The experiments conducted 36 treatments each, which were the combination of two fungi (*A. alternata* and *B. cinerea*), three times (15, 30

and 45 days) and 6 oil treatments was used in this experiment. In all treatments three replicates were used and experimental testes evaluation were contained out every 15 days for up to 45 days. All storage experiments were conducted in a cold room of Horticulture Dept., Fac. Agric., Zagazig Univ., which maintained at $7 \pm 1^{\circ}$ C and $90 \pm 3\%$ relative humidity (RH) under modified atmosphere conditions (MA) (12% CO₂+6% O₂).

Ouality parameters of oil – treated / untreated fruits were evaluated after 15, 30 and 45 days storage for fruit firmness (Firm) Kader (1991), Total Soluble Solids (TSS) Kader (1991), Ascorbic acid (Vitamin C.) Loucoss (1994), pH of fruits juice was measured using a pH meter (HI 198/30 Hanna instrument), sugars fractions by photometrical methods (Bernfeld, 1955; Miller, 1959) phenolic fractions was determined using the colorimetric method (Snell and Snell, 1953) and Folin-Denis reagent preparation (Guttinger, 1981), lycopene fruit content and color measurement (were, L : luminance, a: redness and b: yellowness.) with the help of (Tomato Meter) Cloroflex Ez, according to Tabaestani et al. (2013). The weight loss of tomato fruits was calculated by differences between initial weight and final weight divided by initial weight (Moneruzzaman et al., 2009; Gharezi et al., 2012). Shelf life was calculated by counting the days required to attain the last stage of ripening, but up to the stage when fruits remained still acceptable for marketing.

Statistical Analysis

Collected data were subjected to proper statistical analysis of variance according to Snedecor and Cochran (1980) and the differences among treatments were compared using LSD at 0.05 level.

RESULTS AND DISCUSSION

Isolation and Identification

The fungi isolated from cherry tomato fruits exhibiting typical black rot and gray mold were detected as on *Alternaria alternata* (FR.) Keissl *Botrytis cinerea* (Fr.Res)., respectively according to Mycological Center of Assiout Univ. (MCAU).

In vivo Assessment of Antifungal Activity of some Essential Oils on Cherry Tomato Fruits Spoilage

The antifungal activity of some essential oils *in vivo* on cherry tomato is shown in Tables 1- a, b, and c.

From Table 1-a Alternaria alternata and Botrytis cinerea, isolated from cherry tomato fruits, showed complete growth inhibition when exposed to black seed (Nigella satival L.) and fenugreek (Trigonella foenum – graecum L.) 15 days after storage. In addition, the same result was observed after 30 days of storage against A. alternata and B. cinerea when treated with fenugreek oil.

Treating fruits with black seed oil reduced deeply fruits decay incidence and percentage severity of *A. alternata* which recorded 1.66, 8.33% for disease incidence and percentage severity, compared with the control (0.66% and 1.66%). Also recorded (0.33, 8.33%, respectively for disease incidence and percentage severity of *B. cinerea*, respectively compared with the control (1 and 25%) after 45 days of storage.

The black seed oil treatment caused evident reduction of *A. alternaria* and *B. cinerea* spoilage (disease incidence and percentage severity) of cherry tomatoes fruits than the others of all untreated fruits after 30 and 45 days of storage.

Data also indicated that, treating cherry tomato fruits with fenugreek oil was the most effective treatment which significantly decreased the disease incidence and percentage severity of *A. alternata* and *B. cinerea* spoilage than the control treatments after 30 days of storage.

Table (1- b) shows the effect of cinnamon and spearmint oils treatment on inoculated and uninoculated cherry tomato fruits with spoilage pathogens (*A. alternata* and *B. cinerea*). After 15 days of storage, the essential oil treatments caused complete prevention of fruit spoilage diseases incidence and percentage severity at all treatments of the experiment which evenness the healthy and inoculated control treatment. The same result was observed for both cinnamon and spearmint oils treatment against *A. alternaria* spoilage and the same pattern was recorded for spearmint against *B. cinerea* cherry tomato spoilage after 30 days of storage, when compared with control treatment and also the inoculated fruits.

In general, after 45 days of storage spearmint and cinnamon essential oils treatment caused allowanced spoilage incidence and severity against *A. alternaria* and *B. cinerea*. In addition, the essential oils tested had (cinnamon and spearmint) highest effect against *A. alternaria* being 1% and 0.33% of disease incidence in inoculated fruits than *B. cinerea* being 1 and 0.66% for cinnamon and spearmint oil, respectively at 45 days after storage. On the other hand, spearmint oil caused the highest decrease in disease incidence and percentage severity.

The beneficial effect of cinnamon oil to control postharvest diseases has been recorded by Cruz (2003) who reported that more disease control were achieved by immersing banana fruits into the essential oil of (*Cinnamomum zeylaniam*) against *Colletotrichum musae*.

In addition, Sajad *et al.* (2011) evaluated the effect of natural components (oils of *Thyme vulgaris* and *Mentha piperita*) on citrus green mould control and they found that such treatment improved postharvest quality and decreased decay (27%) compared to the control.

Result obtained from Table 1-c reveal that using essential oils treatments of basil and thyme completely inhibited spoilage of cherry tomato fruits caused by *A. alternata* and *B. cinerea*, after 15 days of storage.

Spraying or dipping fruits in lemongrass and thyme essential oils for controlling postharvest diseases of several fruits were reported by several investigators (Somda et al., 2007; Tzortzakis and Economakis, 2007). Thyme oils vapors was the most effective treatment which reduced strawberry gray mold and inhibited including various fungi food spoilage, mycotoxin producing fungi and postharvest pathogenic fungi as mentioned by Nguefak et al. (2004), Sun Og et al. (2007) and Abd - Alla et al. (2011).

	Alternaria alternata								Botrytis cinerea							
Da	iys	15		3	30		45	1	5	3	0	4	15			
Treatment	D)	S	D	S	D	S	D	S	D	S	D	S			
Control	0		0	0.66	1.66	1	16.66	0	0	1	25	1.33	33.33			
Inoculated (Inc.) 0		0	1	16.66	2.66	33.33	0	0	1	8.33	3.33	33.33			
Black seed	0		0	0.66	1.66	1.66	8.33	0	0	0.33	8.33	1.66	33.33			
Black seed +Inc	. 0		0	0.66	8.33	1.66	33.33	0	0	0.66	8.33	2.33	33.33			
Fenugreek	0		0	0	0	0.66	8.33	0	0	0	0	0.33	16.66			
Fenugreek +Inc	. 0		0	0	0	1.33	16.66	0	0	0	0	0.33	16.66			

Table 1-a. Effect of black seed and fenugreek essential oil treatments on spoilage disease incidence and severity percentage caused by *Alternaria alternata* and *Botrytis cinerea* under modified atmosphere storage conditions

LSD at 5% for Disease [D (%)]: NS for Severity [S (%)]: NS

a- Effect of *A. alternata* and *B. cinerea* on spoilage disease incidence and severity (%)

b- Effect of storage time	(days) on	spoilage
disease incidence and sev	erity (%)	

Fungi	D	S	Days	D	S
A. alternata	0.666	8.055	15	0	0
B. cinerea	0.759	12.5	30	0.5	7.222
F. test	NS	NS	45	1.638	23.611
LSD 5%			F. test	NS	NS
			LSD 5%		

c- Effect of oil treatments on spoilage disease incidence and severity (%)

Treatment	D	S
Control	0.666	12.777
Inoculated (Inc.)	4	15.277
Black seed	2.166	8.611
Black seed +Inc.	2.666	15.277
Fenugreek	0.833	4.166
Fenugreek +Inc.	1.166	5.555
F. test	**	*
LSD 5%	0.577	8.581

Thyme oil exhibited a high degree of inhibition of *A. alternaria* and *B. cinerea* in tomato (Bouchra *et al.*, 2003 ; Feng and Zheng, 2007).

It has been observed that thyme oil treatments inhibited completely the percentage of spoilage cherry tomato fruits incidence and severity, which recorded 00.00% for *A. alternata* and *B. cinerea* after 30 days of storage. The same result was recorded when treated cherry tomato fruits with basil oil against *B. cinerea* gray mold, while thyme oil treatment caused the highest effect against *B. cinerea* compared to basil oil treatment Table (1-c).

Results also show that highest reduction of cherry tomato spoilage incidence caused by *A. alternata* and *B. cinerea* when fruits treated with both essential oils (thyme and basil) after 15 and 30 days of storage.

Similar observations were also recorded by Bouchra *et al.* (2003) who demonstrated that *Thymus glandulosus* Req. was the most efficient in controlling *Botrytis cinerea*, a most critical pathogen in postharvest. Also, the essential oil and crude extract of wild basil were tested on the mycelial growth of *Alternaria alternata* which lead to total inhibition of the fungal mycelial growth (Benini *et al.*, 2010). In addition, thyme and oregano oil reduced rot development in tomatoes caused by *B. cinerea* and *A. alternate* (Plotto *et al.*, 2003).

In conclusion, mechanism underlying the action of essential oil enrichment on the switch between vegetative and reproductive phases of fungal development remains to be fully understood. The negative impact of essential oils on fungi sporulation may reflect the effect of the volatiles emitted by oils on surface mycelial development and / or the precipitation / transduction of signals involved in the switch from vegetative to reproductive development.

Nevertheless, suppression of spore production by essential oils could play a major role in limiting pathogen spread by lowering spore load under storage atmosphere and on surfaces of plant (Tzortzakis and Economakis, 2007).

Fruits and vegetables are highly perishable products particularly once they have been harvest. Being metabolically active, they tend to

lose energy reserves through respiration and water through transpiration. In addition to the biochemical changes, there may be losses in quality through mechanical damage, pests and diseases, as well as physiological disorders induced by high or low temperature or incorrect storage atmosphere. Improving the existing practices of handling and storage as well as developing new techniques in order to maintain post-harvest quality has always been a challenge to the researchers (Debeaufort et al., 1998). In this regard, treating fresh fruits and vegetables (cherry tomatoes in our study) with edible coating is an important approach that holds promise for the extension of storage life of fresh horticultural crops similarly to the results obtained by Tabaestani et al. (2013).

Total Soluble Solids (TSS)

Total soluble solids (TSS) is an indication of soluble sugars level in fruits and therefore sweetness (Saltveit, 2005).

It is evident from results in Table 2-a that total soluble solids of cherry tomatoes was found to be affected by coating treatment with fenugreek and black seed essential oils, especially fenugreek oil treatment, while inoculated fruits with A. alternata and / or B. cinerea lead to decline of TSS value especially after 30 and 45 days from storage. However, as the storage progressed a general declining trend in the TSS was observed for all treatment (Table 2-a). The changes in TSS were significantly influenced by the duration of storage. In yet other studies revealed that gradual decrease in TSS of tomatoes associated with increasing storage time has been reported (Kumarrai et al., 2012; Kagan-Zur and Mizrahim, 1993).

The same trend was observed when treated cherry tomatoes with cinnamon and spearmint essential oils Table (2-b) and also in case of basil and thyme oils treatment Table (2-c).

Firmness

The texture of tomato is a very important major quality feature for consumers. Texture in tomato is a function of skin toughness. Also, sufficient softening is necessary to lend the fruit's edibility and to release the cell content during chewing for the olfactory perception of the fruit aroma eating and flavor. It is commonly accepted that consumer preference is for firm fruit which do not lose juice during eating or slicing (Frankel and Jen, 1989).

				Alterna	ria alter	nata				Botryt	is ciner	ea	
]	Days		15	3	60	2	15	1	5	3	60	2	15
Treatment]	D	S	D	S	D	S	D	S	D	S	D	S
Control		0	0	0.66	33.33	1	8.33	0	0	1	25	1.66	33.33
Inoculated (In	c.)	0	0	1.33	16.66	2	100	0	0	1.66	25	3.33	33.33
Cinnamon	(0	0	0	0	1	8.33	0	0	0.33	1.66	1	16.66
Cinnamon +In	ic.	0	0	0.33	8.33	1.66	25	0	0	0.66	8.33	2.33	25
Spearmint	(0	0	0	0	0.33	1.66	0	0	0	0	0.66	16.66
Spearmint +In	ic.	0	0	0.66	6.66	1	16.66	0	0	0.33	16.66	1.66	33.33

Table 1-b. Effect of cinnamon and spearmint essential oil treatments on spoilage disease incidence and percentage severity caused by *Alternaria alternata* and *Botrytis cinerea* under (MA) conditions

LSD at 5% for Disease [D (%)]: NS for Severity [S (%)]: NS

a- Effect of *A. alternata* and *B. cinerea* on spoilage disease incidence and severity (%)

b- Effect of storage time (days) on spoilage disease incidence and severity (%)

Fungi	D	S	Days	D	S
A .alternata	0.555	13.333	15	0	0
B. cinerea	0.814	9.018	30	0.583	5.75
F. test	*	NS	45	1.472	27.777
LSD 5%	0.367		F. test	**	NS
			LSD 5%	0.475	

c- Effect of oil treatments on spoilage disease incidence and severity (%)

Treatment	D	S
Control	0.722	12.666
Inoculated (Inc.)	1.388	25.277
Cinnamon	0.388	4.222
Cinnamon +Inc.	0.833	9.833
Spearmint	0.166	5.555
Spearmint +Inc.	0.611	9.5
F. test	**	*
LSD 5%	0.295	7.706

Table 1-c. Effect of basil and thyme essential oil treatments on spoilage disease incidence and percentage severity caused by *Alternaria alternata* and *Botrytis cinerea* under (MA) conditions

		Alte	ernaria	altern	ata				Botryt	is ciner	ea	
Days	15	5	3	30	2	45	1	5		30		45
Treatment	D	S	D	S	D	S	D	S	D	S	D	S
Control	0	0	0.66	16.66	1	16.66	0	0	1	8.33	1	16.66
Inoculated (Inc.)	0	0	1	16.66	2	33.33	0	0	1	33.33	3.33	33.33
Basil	0	0	0	0	1	25	0	0	0.33	8.33	1	33.33
Basil +Inc.	0	0	0.66	8.33	1.66	16.66	0	0	0.66	16.66	2.66	33.33
Thyme	0	0	0	0	0.66	16.66	0	0	0	0	1	16.66
Thyme +Inc.	0	0	0	0	1	33.33	0	0	0.33	1.66	1.66	8.33

LSD at 5% for Disease [D (%)]: NS

for Severity [S (%)]:18.87

a- Effect of *A. alternata* and *B. cinerea* on spoilage disease incidence and severity (%)

b- Effect of storage time (days) on spoilage disease incidence and severity (%)

Fungi	D	S	Days	D	S
A. alternata	0.537	13.333	15	0	0
B. cinerea	0.777	9.018	30	0.472	9.166
F. test	NS	NS	45	1.5	23.611
LSD 5%			F. test	NS	NS
			LSD 5%	_	

C- Effect of oil treatments on spoilage disease incidence and severity (%)

Treatment	D	S
Control	0.611	9.722
Inoculated (Inc.)	1.222	19.444
Basil	0.388	11.111
Basil +Inc.	0.944	12.5
Thyme	0.277	5.555
Thyme +Inc.	0.5	7.222
F. test	*	N.S
LSD 5%	0.589	—

In this study, as revealed by the puncture test values of different essential oils coating treatments, it has made statistically great impact to maintain firmness for long time during experiment.

According to Tables 2-a, b and c, all tested essential oil treatments enhanced the firmness of healthy and treated infected fruits compared to untreated infected ones. In addition inoculated fruits treated with essential oils tented to be firmer than healthy ones.

Also, it was found that oil untreated fruits were softer than the other oil treatments at the end of storage period (45 days).

Generally, after 45 days of storage (the end of storage period), firmness values observed for healthy fruits and treated with black seed, fenugreek, cinnamon, spearmint, basil and thyme recorded 300, 370, 417, 466.66, 403 and 330, respectively. However, inoculated control fruits with *A. alternata* being 276.66, 300 and 200, respectively. While the inoculated control fruits with *B. cinerea* bing 267.66, 200 and 300, respectively. However, firmness values of healthy fruits and treated with aforementioned oils recorded 333.33, 403.33, 393.33, 440, 416.66 and 466.66, respectively.

According to the result obtained in Tables 2a, b and c, all oil treatments enhanced the firmness of healthy and inoculated fruits compared to untreated fruits in control treatment. In addition, inoculated fruits tended to be firmer than healthy ones. Also, it was found that oil untreated fruits were softer than all treated ones. In this respect some reports revealed that essential oils coating fruits significantly delayed the softening of whole tomato during storage (Tabaestani *et al.*, 2013). Also, Serrano *et al.* (2005) reported that treated sweet cherry fruits with essential oils such as eugenol, thymol and menthol showed benefit in maintenance fruit firmness comparing with control treatments.

pH values

From data in Tables 2-a, b and c, significant decrease was observed in pH values of all treatments at the end of storage time compared with the initial storage time.

The fenugreek essential oil treatment caused clear increasing in pH values than the all other

treatments being 3.16, 3.63 and 2.26 for control treatment, coated with fenugreek oil and coated and inoculated with *A. alternate*, respectively after 15 days of storage, while at the end of storage time (45 days) being 1.2, 2.2 and 1.1, respectively.

The same trend was observed in fruits inoculated with *B. cinerea* and treated with fenugreek oil which recorded 2.2, 2.96 and 1.8 after 15 days of storage time and 0.93, 0.7 and 0.5 after 45 days of storage, respectively.

The results presented in Table (2 - c) indicate that all basil oil treatments at all storage time recorded lowest values than the control treatment and / or inoculated cherry tomato fruits with *A. alternata* while, only pH value was increased at all storage time than those of other treatments when cherry tomatoes were inoculated with *B. cinerea*.

The present results suggest that using cinnamon essential oil as an edible coating, the pH values increased at all storage time in inoculated cherry tomato fruits with *A. alternaria* and / or *B. cinerea* than those of other treatments. Our results confirmed other studies carried by Deirdre *et al.* (1999) Tzortzakis (2007) and Marjanlo *et al.* (2009).

Sugar Fractions

The sugar fractions values of cherry tomatoes inoculated with *A. alternate* and / or *B. cinerea* and/or non inoculated and treated with different essential oils, then stored under modified atmosphere (MA) conditions were presented in Table 3.

According to data in Table 3, inoculated fruits with spoilage pathogens caused significant reduction over all total sugar percentage which had 4.37 and 4.84 compared with healthy fruits (control) as a result of inoculated with *A. alternata* and *B. cinerea* respectively.

In addition healthy fruits treated with black seed, fenugreek and thyme oils lead to significant increase in all sugar fraction amounts in the case of *A. alternata*. Fruits inoculated with *A. alternata* and coated with thyme oil caused increase of all sugar fraction concentrations (total, reducing and non reducing). The same trend was also observed in case of fruits inoculated with *B. cinerea* and coated with cinnamon and fenugreek oil, respectively.

 Table 2-a. Effect of black seed and fenugreek essential oils treatments on total soluble solids (TSS), firmness and pH of storage cherry tomatoes under (MA) conditions

			Ŀ	llterna	ria ali	terna	ta						Botryt	is cir	ierea			
Day	s	15			30			45			15			30			45	
Treatment	Firm	TSS	pН	Firm	TSS	pН	Firm	TSS	pН	Firm	TSS	pН	Firm	TSS	pН	Firm	TSS	pН
Control	523.33	4.66	3.16	426.66	3.00	1.96	320	2.25	1.2	583.33	5.50	2.2	390	3.41	1.53	296.66	2.16	0.93
Inoculated (Inc.)	473.33	4.83	2.03	360	2.83	1.46	276.66	1.66	1.13	493.33	4.75	1.24	306.66	2.50	1.06	276.66	2.00	0.33
Black seed	523.33	5.00	2.43	493.33	3.91	1.7	300	2.25	1.1	496.66	5.33	2.5	446.66	3.58	2.00	333.33	3.00	0.8
Black seed +Inc.	453.33	5.16	1.28	346.66	2.41	1.23	250	1.00	0.93	436.66	5.33	1.93	400	1.1	1.1	300	1.9	0.73
Fenugreek	600	5.25	3.63	586.66	4.58	2.73	370	3.50	2.2	660	5.66	2.96	510	2.33	2.33	403.33	3.00	0.7
Fenugreek +Inc.	570	5.33	2.26	523.33	3.91	1.73	346.66	2.66	1.1	386.66	4.16	1.8	436.66	1.00	1.00	306.66	2.00	0.5
LSD at 5% for Fi	rm= 6	1.97]	rss=	= NS						pH=	NS				

a- Effect of *A. alternata* and *B. cinerea* on total soluble solids (TSS), firmness and pH

b-	Effect	of	storage	time	(days)	on	total
	soluble	soli	ds (TSS),	firm	less and	pН	

Fungi	Firm	TSS	pН
A. alternata	430.185	3.563	1.874
B. cinerea	413.148	3.735	1.453
F. test	N.S	N.S	**
LSD 5%			3.364

			_
Days	Firm	TSS	pН
15	516.666	5.073	2.358
30	433.333	3.59	1.627
45	315	2.283	1.005
F. test	**	**	**
LSD 5%	47.262	0.411	0.185

c- Effect of oil treatments on total soluble solids (TSS), firmness and pH

Treatment	Firm	TSS	рН
Control	423.333	3.481	1.833
Inoculated (Inc.)	364.444	3.097	1.283
Black seed	427.777	3.847	1.755
Black seed +Inc.	364.444	3.191	1.277
Fenugreek	521.666	4.569	2.438
Fenugreek +Inc.	428.333	3.708	1.394
F. test	**	**	**
LSD 5%	25.301	0.377	0.159

Table 2-b.Effect of cinnamon and spearmint essential oils treatments on total soluble solids (TSS), firmness and pH of storage cherry tomatoes under (MA) conditions

				Alterna	uria ali	ternat	a						Botryt	is cine	erea			
Day	/\$	15			30			45			15			30			45	
Treatment	Firm	TSS	pН	Firm	TSS	pН	Firm	TSS	pН	Firm	TSS	pН	Firm	TSS	pН	Firm	TSS	pН
Control	480	5.00	3.1	440	3.08	2.16	376.66	2.58	1.7	610	4.58	2.93	406.66	3.00	2.00	296.66	1.75	0.9
Inoculated (Inc	.) 470	4.75	2.63	376.66	3.08	2.06	300	2.08	0.63	526.66	3.08	2.06	386.66	3.00	1.76	200	1.5	1.16
Cinnamon	606.66	4.83	3.26	570	4.66	2.56	417	3.00	1.9	636.66	5.08	2.73	583.33	4.58	2.20	393.33	2.83	1.3
Cinnamon +Inc	. 503.33	5.5	2.26	443.33	3.66	1.56	360	2.66	1.1	500	4.41	1.83	400	3.5	1.26	260	2.00	0.56
Spearmint	543.33	5.25	3.03	493.33	4.75	1.86	466.66	3.16	0.9	583.33	4.41	2.63	543.33	4.00	1.83	440	3.08	0.93
Spearmint +Inc	. 483.33	5.33	2.33	423.33	3.25	1.23	333.33	2.00	0.9	506.66	4.41	2.03	400	3.08	1.50	330	2.75	0.63
LSD at 5% fo	r Firm	n = NS	S				TSS	= NS						pH=	NS			

a- Effect of *A. alternata* and *B. cinerea* on total soluble solids (TSS), firmness and pH

Fungi	Firm	TSS	РН
A.alternata	450.555	3.824	1.946
B.cinerea	444.629	3.361	1.666
F. test	NS	*	NS
LSD 5%		0.202	

b- Effect of storage time (days) on total soluble solids (TSS), firmness and pH

Days	Firm	TSS	pН
15	537.5	4.673	2.58
30	457.5	3.652	1.811
45	347.777	2.451	1.027
F. test	**	**	**
LSD 5%	38.933	0.244	0.234

c- Effect of oil treatments on total soluble solids (TSS), firmness and pH

Treatment	Firm	TSS	рН
Control	435	3.333	2.133
Inoculated (Inc.)	380.555	2.916	1.722
Cinnamon	534.444	4.055	2.227
Cinnamon Inc.	411.111	3.652	1.45
Spearmint	511.666	4.111	1.866
Spearmint +Inc.	412.777	3.486	1.438
F. test	**	**	**
LSD 5%	46.206	0.382	0.445

 Table 2-c. Effect of basil and thyme essential oils treatments on total soluble solids (TSS), firmness and pH of storage cherry tomatoes under (MA) conditions

	Altern					ernat	a						Botryt	is cine	erea			
Day	s	15			30			45			15			30			45	
Treatment	Firm	TSS	pН	Firm	TSS	pН	Firm	TSS	pН	Firm	TSS	pН	Firm	TSS	pН	Firm	TSS	pН
Control	413.33	4.58	2.83	390	3.41	1.9	316.66	1.66	1.66	1.00	2.25	3.23	413.33	3.58	2.93	376.66	2.58	2.33
Inoculated (Inc.)	393.32	4.58	1.83	306.66	2.5	1.16	200	1.03	1.30	0.9	5.66	2.63	373.33	3.00	2.3	300	2.08	2.00
Basil	533.33	5.33	3.13	523.33	3.58	2.16	403	3.08	3.08	0.93	5.08	2.53	473.33	4.58	2.36	416.66	3.00	1.56
Basil +Inc.	446.66	4.25	2.03	430	3.33	1.36	250	1.75	1.75	0.33	4.83	2.33	420	2.75	1.63	360	2.66	1.2
Thyme	660	4.91	2.9	470	5.41	1.86	330	2.00	2.00	0.76	6.00	3.23	466.66	4.58	2.66	466.66	3.16	2.06
Thyme+ Inc.	456.66	3.41	1.7	510	4.16	1.26	320	2.16	2.16	0.66	5.75	2.56	443.33	4.00	2.13	333.33	2.00	1.46
LSD at 5% for	Firm=	Firm= 75.24 TSS=1.06						06	pH= NS									

a-Effect of *A. alternata* and *B. cinerea* on total soluble solids (TSS), firmness and pH

Fungi	Firm	TSS	pН
A. alternata	408.518	3.418	1.611
B. cinerea	442.407	3.916	2.294
F. test	*	*	*
LSD 5%	32.456	0.49	0.442

b- Effect of storage time (days) on total soluble solids (TSS), firmness and pH

Days	Firm	TSS	pН
15	503.055	4.972	2.58
30	433.888	3.763	1.98
45	339.444	2.266	1.297
F. test	**	**	**
LSD 5%	25.987	0.408	0.065

c- Effect of oil treatments on total soluble solids (TSS), firmness and pH

Treatment	Firm	TSS	рН
Control	400.555	3.513	2.372
Inoculated (Inc.)	339.444	3.2	1.805
Basil	495	4.111	2.155
Basil +Inc.	400.555	3.263	1.483
Thyme	493.888	4.333	2.25
Thyme +Inc.	423.333	3.583	1.65
F. test	**	**	99
LSD 5%	30.716	0.435	0.178

Treatment		Alternaria	alternata		Botrytis ci	inerea
	Total	Reducing	Non-reducing	Total	Reducing	Non-reducing
Control initial time	3.5	1.24	2.26	3.5	1.24	2.26
Control end time	5.30	2.06	3.24	6.53	2.36	4.17
Inoculated (Inc.)	4.37	1.61	2.76	4.84	1.96	2.88
Black seed	5.68	2.27	3.41	3.85	1.36	2.49
Black seed+ Inc.	4.76	2.16	2.60	4.19	1.45	2.74
Fenugreek	6.07	2.43	3.64	6.10	2.24	3.86
Fenugreek +Inc.	4.34	1.59	2.75	5.92	2.15	3.77
Cinnamon	4.62	1.91	2.71	5.95	2.13	3.82
Cinnamon + Inc.	3.94	1.36	2.58	6.11	2.20	3.91
Spearmint	3.76	1.36	2.40	3.96	1.44	2.52
Spearmint + Inc.	3.86	1.44	2.42	4.13	1.58	2.55
Basil	4.24	1.83	2.41	3.90	1.38	2.52
Basil + Inc.	3.83	1.54	2.29	3.30	1.39	1.91
Thyme	5.30	2.05	3.25	4.17	1.55	2.62
Thyme + Inc.	6.10	2.75	3.35	4.80	1.63	3.17

 Table 3. Sugar fractions of cherry tomatoes stored under (MA) cold conditions affected by the interaction of spoilage pathogens inoculation (Alternaria alternata and Botrytis cinerea) and essential oil treatments

LSD at 5% for: Total sugars = 0.037 Reducing sugars = 0.024

Non-reducing sugars = 0.023

These results were in agreement with those reported by Ozkaya *et al.* (2009) who reported that strawberries had higher glucose amount after 10 days of (MA) storage compared with control.

It was observed that the increase in reducing, non-reducing and total sugar with the storage long time (From 3.5% at initial time to 5.3% and 6.53% at the end time of storage). The obtained results of sugar content in cherry tomato are in agreement with the findings of (Moneruzzaman et al., 2009; Gharezi et al., 2012). Increased total sugar content might be due to conversion of starch into sugars (Tsuda et al., 1999). Also, Vesaltalab and Gholami (2012)and Nabifarkhani et al. (2015), they suggested that total sugar content increased during storage period that may be due to the dehydration and decomposition of organic acids (used as an energy source) in the fruits.

It is evident from Table 3 that control treatment recorded significantly higher mean reducing, non-reducing and total sugar contents over all other treatments. This result was agree with the previous report carried out by Gharezi *et al.* (2012).

Vitamin C. amount

Data in Tables 4-a, b and c show, general significant decrease in vitamin c. content of inoculated cherry tomato fruits with both *A. alternata* and / or *B. cinerea*. Storage cherry tomatoes (healthy and inoculated) and/or coated with tested essential oil treatments caused a greatest relative decrease in vitamin c. content.

Also, gradual decrease in content of vitamin c. of all treatments influenced by the duration of the storage time. The fenugreek oil treatment

Table 4-a. Changes in some antioxidant contents (Vitamin C. and Lycopene) of cherry tomatoestreated by black seedand fenugreek essential oil and inoculated with spoilagepathogens (Alternaria alternata and Botrytis cinerea) during (MA) storage

		Alternaria alternata						Botrytis cinerea					
Day	s 1	5	3	80	4	45	1	5	3	60	4	5	
Treatment	Vit. C.	Lycop.	Vit. C.	Lycop.	Vit. C.	Lycop.	Vit. C.	Lycop.	Vit.C.	Lycop.	Vit. C.	Lycop.	
Control	13.03	90	10.93	85	8.5	76	10.13	83	8.53	77	5.66	60	
Inoculated (Inc.)	10.63	78	98.23	67	5.3	56	8.83	75	6.56	64	4.9	56	
Black seed	11.33	80	8.66	78	6.1	60	12.63	87	9.76	72	7.06	63	
Black seed + Inc.	9.23	71	6.83	69	3.9	52	9.53	69	7.06	50	5.36	46	
Fenugreek	13.6	64	9.06	55	7.36	50	12.66	54	9.6	42	8.9	33	
Fenugreek + Inc.	10.86	62	7.96	47	6.53	39	9.86	64	7.76	59	7.6	53	

LSD at 5% for Vit. C = 0.597

a- Effect of *A. alternata* and *B. cinerea* on Vitamin C. and Lycopene

Fungi	Vit. C. mg/ 100 g juice	Lycopene mg/ 100 g fresh weight
A. alternata	8.785	65.5
B. cinerea	8.433	61.5
F. test	NS	-
LSD 5%		-

b- Effect of storage time (days) on Vitamin C. and Lycopene

Days	Vit. C. mg/ 100g juice	Lycopene mg/ 100g fresh weight
15	10.975	73.083
30	8.402	63.75
45	6.441	53.666
F. test	**	-
LSD 5%	0.664	-

c- Effect of black seed and fenugreek oil treatments on Vitamin C. and Lycopene

Treatment	Vit. C. mg/100g juice	Lycopene mg/100g fresh weight
Control	9.483	78.5
Inoculated (Inc.)	7.3	66
Black seed	9.261	73.333
Black seed+ Inc.	6.961	59.5
Fenugreek	10.2	49.666
Fenugreek +Inc.	8.433	54
F. test	**	-
LSD 5%	0.597	-

caused slight increase in vitamin c value compared with the other treatments especially at 15 days after storage as a result of inoculated fruits with *A. alternata* and/or *B. cinerea* (Table 4-a). The same trend was observed in case of cherry tomato fruits inoculated by *B. cinerea* and treated with thyme oil. while, thyme oil treatment caused increase in vitamin c content at all days of storage being 11.36, 9.06 and 7.2 after 15, 30 and 45 days of storage, respectively (Table 4-c).

According to the results in Table 4-b cinnamon oil treatment caused insignificant increase of vitamin c content among the postharvest oil fruit treatment at all stages of storage (15, 30 and 45 days of storage being 12, 10.36 and 7.73 mg/100g, respectively.

An increase in ascorbic acid (vitamin c) content in fruits is thought to be an indication that the fruits are still in the ripening stage, while a decrease indication senescent fruit (Erip-Roberts *et al.*, 2002). In addition, Miller and Evans (1997) reported that phenolic substances have been found to play a protective effect on tissue ascorbic acid content. The presence of phenolic in the fruit cells may help to maintain the ascorbic acid content value.

In general, essential oils coating cherry tomato fruits could inhibit vitamin c loss by cutting as an abiotic elicitor generating reactive oxygen species, or due to the protection of antioxidant in essential oils (Kumarrai *et al.*, 2012; Xing *et al.*, 2011).

While, tomato fruits inoculated with *A. alternata* and treated with thyme oil being 84, 80 and 64 mg/100g fresh weight after 15, 30 and 45 days of storage, respectively. These results are in agreement with those obtained by Ali and Keith (1998) and Tabaestani *et al.* (2013).

Lycopene contents

The lycopene content and antioxidant activity of tomatoes varies between cultivars and is highest in cherry or small, cocktail fruits (Kaur *et al.*, 2004; Molyneux *et al.*, 2004). There were a correlation between fruit color and total antioxidant concentration observed with lycopene content which increase from the rose to red color stages (Brand *et al.*, 2006; Helyes *et*

al., 2006). This explained the fact that mature green tomatoes might having a significantly lower total antioxidant content than red tomatoes (Wold *et al.*, 2004).

Because of importance of antioxidant to human health (Madhavi and Salunkhe, 1998), antioxidants may be considered a valuable quality attribute of tomatoes and it is important to minimize losses of these compounds during the post – harvest period (Passam *et al.*, 2007).

It is clear from data presented in Tables 4-a, b and c that all lycopene amount were decreased during storage time of all experimental treatments. Also, inoculated fruits with *A. alternata* and *B. cinerea* and untreated fruits with the essential oils caused clear decreasing in lycopene amounts with progressing of storage time.

Phenolic Fractions

As shown in Table 5, cherry tomato fruits coated with basil and or cinnamon essential oils had the slight increase amounts of total phenols being 7mg/100g fruits fresh weight of both above mentioned oils compared with untreated one (6.75 mg/100g). On the other hand, treated fruits (inoculated by *A. alternata* and / or *B. cinerea*) with thyme, cinnamon, black seed and fenugreek oils lead to slightly increase of total phenolic compounds.

With the exception of cherry tomato fruits coated with spearmint, it was observed that significantly increase in free phenolic values for all inoculated fruits by *A. alternata*. The same trend was observed in case of fruits inoculated with *B. cinerea*, except of fruits coated with cinnamon oil. It may be concluded that infection with the spoilage pathogens enhanced the natural defense mechanism of plant tissues. Our result in term of phenolic fractions was in agreement with other studies (Capassoll and Torresll, 2013; Petriccione *et al.*, 2015).

Regarding phenolic content in tomato fruits of all treatments, it was noticed that significant decrease of phenolic compounds value occurred by prolonging the time of storage. These results are consistent with the findings of other researches (Tian *et al.*, 2011; Tabaestani *et al.*, 2013; Chen *et al.*, 2014; Xu *et al.*, 2014).

Table 4-b. Changes in Vitamin C. (mg/100g juice) and Lycopene (mg/100 g F.w.) of cherry tomatoes treated by cinnamon and spearmint essential oil and inoculated with spoilage pathogens (*Alternaria alternata* and *Botrytis cinerea*) during (MA) storage

		Alter	naria d	alterna	ta				Botrytis	s cinerea	ı	
Days	1:	5	3	60	4	45	1	.5	3	60	2	15
Treatment	Vit.c	Lyco.	Vit.c	Lyco.	Vit.c	Lyco.	Vit.c	Lyco.	Vit.c	Lyco.	Vit.c	Lyco.
Control	11.36	92	9.63	80	7.6	79	11.3	79	8.73	45	6.2	37
Inoculated (Inc.)	10.5	78	9.46	67	6.3	63	10.63	67	7.00	65	5.33	44
Cinnamon	12.00	59	10.36	54	7.73	45	10.3	84	8.23	78	6.46	56
Cinnamon +Inc.	10.03	84	8.6	80	6.4	64	9.3	70	7.43	67	6.00	54
Spearmint	9.16	78	7.9	71	6.53	68	11.2	64	9.26	50	7.93	47
Spearmint+ Inc.	8.73	75	7.63	59	5.16	47	9.53	68	7.43	54	6.03	44

LSD at 5% for Vit. C = 0.62

Fungi

A. alternata

B. cinerea

F. test LSD 5%

a- Effect of *A. alternata* and *B. cinerea* on Vitamin C. (mg/100g juice) and Lycopene (mg/100 g F.w.)

b- Effect of storage	time	(days) on	Vitamin	C.
(mg/100g juice)	and	Lycopene	(mg/100	g
F.w.)				

		,		
Vit. C. mg/ 100g Juice	Lycopene mg/ 100g fresh weight	Days	Vit. C. mg/100g Juice	Lycopene mg/ 100g fresh weight
8.696	69.055	15	10.458	75.583
8.261	60.111	30	8.475	64.166
NS	-	45	6.502	54
	-	F. test	**	-
		LSD 5%	0.725	-

c- Effect of cinnamon and spearmint oil treatments on Vitamin C. (mg/100g juice) and Lycopene (mg/100 g F.w.)

Treatment	Vit. C. mg/100g juice	Lycopene mg/100g fresh weight
Control	9.227	68.666
Inoculated (Inc.)	8.261	65.5
Cinnamon	9.183	62.666
Cinnamon +Inc.	7.961	69.833
Spearmint	8.816	63
Spearmint+ Inc.	7.422	57.833
F. test	**	-
LSD 5%	0.62	-

		Alt	ernari	a alteri	nata			1	Botrytis	cinere	a	
Days	s 1	15		30		45]	15	3	0	4	5
Treatment	Vit.c	Lyco.	Vit.c	Lyco.	Vit.c	Lyco.	Vit.c	Lyco.	Vit.c	Lyco.	Vit.c	Lyco.
Control	10.2	75	8.96	60	6.26	50	11.4	79	9.36	62	6.5	58
Inoculated (Inc.)	9.46	62	7.66	54	4.76	44	9.33	54	7.8	46	5.63	33
Basil	9.06	92	8.16	85	6.13	73	10.76	75	8.4	69	5.93	55
Basil + Inc.	8.76	67	6.9	51	5.2	43	8.63	67	5.76	58	3.86	47
Thyme	11.36	50	9.06	45	7.2	31	11.63	84	8.33	71	4.33	62
Thyme + Inc.	9.4	83	8.06	80	5.63	64	826	71	6.36	64	4.1	55

Table 4-c. Changes in Vitamin C. (mg/100g juice) and Lycopene (mg/100 g F.w.) of cherry tomatoes treated by basil and thyme essential oil and inoculated with spoilage pathogens (*Alternaria alternata* and *Botrytis cinerea*) during (MA) storage

LSD at 5% for Vit. C = 0.675

a- Effect of *A. alternata* and *B. cinerea* on Vitamin C. (mg/100g juice) and Lycopene (mg/100 g F.w.)

b- Effect of storage	time	(days) on	Vitamin	C.
(mg/100g juice)	and	Lycopene	(mg/100	g
F.w.)				

Fungi	Vit.C. mg/ 100g Juice	Lycopene mg/ 100g fresh weight	Days	Vit. C. mg/ 100g Juice	Lycopene mg/ 100g fresh weight
A. alternata	7.902	61.611	15	9.858	71.583
B. cinerea	7.568	61.666	30	7.885	62.083
F. test	NS	-	45	5.463	51.25
LSD 5%	_	-	F. test	**	-
			LSD 5%	0.697	-

c- Effect of basil and thyme oil treatments on Vitamin C. (mg/100g juice) and Lycopene (mg/100 g F.w.)

Treatment	Vit. C. mg/100g Juice	Lycopene mg/100g fresh weight
Control	8.775	64
Inoculated (Inc.)	7.427	48.833
Basil	8.077	74.833
Basil + Inc.	6.522	55.5
Thyme	8.655	57.166
Thyme + Inc.	6.955	69.5
F. test	**	-
LSD 5%	0.675	-

Treatment	Al	ternaria a	lternata	Botrytis cinerea			
	Total	Free	Compound	Total	Free	Compound	
Control Initial time	8.75	5.00	3.75	8.75	5.00	3.75	
Control End time	6.25	2.50	3.75	6.50	2.50	3.75	
Inoculated (Inc.)	6.25	4.75	1.50	6.25	4.75	1.50	
Black seed	6.50	2.75	3.75	6.50	2.75	3.75	
Black seed+ Inc.	7.00	3.25	3.75	7.00	2.50	4.50	
Fenugreek	6.25	2.25	4.00	6.25	2.25	4.00	
Fenugreek+ Inc.	6.50	3.25	3.25	6.75	4.25	2.50	
Cinnamon	7.00	4.75	2.25	7.00	4.75	2.25	
Cinnamon + Inc.	6.75	4.75	2.00	6.50	2.25	4.25	
Spearmint	6.75	3.00	3.75	6.75	3.00	3.75	
Spearmint + Inc.	6.25	2.50	3.75	7.00	3.75	2.25	
Basil	7.00	3.50	3.50	7.00	3.50	3.50	
Basil + Inc.	7.00	3.50	3.50	7.00	4.00	3.00	
Thyme	6.75	3.75	3.00	6.75	3.75	3.00	
Thyme+ Inc.	7.00	4.50	2.50	7.25	5.00	2.25	

 Table 5. Impact of phenolic fractions affected by some essential oil treatments and inoculated with spoilage pathogens (Alternaria alternata and Botrytis cinerea) of cherry tomatoes

LDS at 5% for Total phenol = NS Free = 0.94 Compound = NS

Lycopersicon esculentum is one of the most widely consumed fresh and processed vegetables in the world. It contains bioactive key components as phenolic compounds profile which differ among tomato varieties, which were related to genetic features, cultivation conditions, handlings methods and postharvest treatments (Barros *et al.*, 2012). In addition, cherry tomato had a significantly higher content of total phenolic compounds than the other's tomato varieties (Marsic *et al.*, 2011).

Color (L*, a*, b*)

Fruit color is virtually a major quality characteristic in all fruits and vegetables. Also uniformity of color within tomatoes is principal requirement of the European Union (EU) quality standards for this crop. Since the deposition of carotenoid pigments is responsible for the characteristic color of ripe tomatoes (Fraser *et al.*, 2000), an understanding of carotenoid

synthesis in tomato is of immediate relevance to quality (Passam *et al.*, 2007). During fruit ripening maximum concentrations of α and β -carotene occur at the turning to breaking stages (Meredith and Purcell, 1966), after which lycopene accumulates (Davies and Hobson, 1981).

Fruit color is probably the most important attribute that determines overall quality as it affects consumer perception (Tabaestani *et al.*, 2013).

Inoculated cherry tomato frutis with *A. alternata* and/or *B. cinerea* caused clear decrease in L, a and b values than control treatments (un inoculated healthy fruits), (Tables 6 - a, b and c). The color values (L, a and b) of all experimental treatments were gradually decreased with progressing time of storage (Tables 6 -a, b and c). Our result was in agreement with those obtained by Ali *et al.* (2010) and Tabaestani *et al.* (2013).

In conclusion, the essential oil treatments enhanced the color values of healthy fruits compared to untreated one (control treatment) being 22.82, 11.70, 9.88 for, L, a and b, respectively for (black seed oil treatment) and 21. 47, 9.24, 9.79 for L, a and b, respectively for (cinnamon oil treatment) and 21.40, 14.15, 9.06 for L, a and b, respectively for (spearmint oil treatment) compared to untreated and inoculated fruits with *A. alternaria* at the end of storage time.

In addition inoculated fruits with *A*. *alternaria* and treated with fenugreek and thyme essential oils true up to be colored than inoculated ones. Also, the same result was obtained by *B*. *cinerea* inoculated fruits and treated with fenugreek, thyme and basil oils than only inoculated fruits (Tables 6-a, b and c).

A delay in color change due to oil treatments has been reported in tomato (Ali *et al.*, 2010). Coating of tomatoes with basil gum delayed color change (Tabaestan *et al.*, 2013).

Weight loss percentage

It is evident from (Fig. 1- a and b) that weight loss percentage increased significantly with the prolongation of the storage period for all treatments. Generally, the weight loss occurs during storage due to its respiration process, the transference of humidity and some process of oxidation (Ayranic and Tunc, 2003). However, all treatments significantly reduced weight loss of tomatoes during storage compared to control treatment.

Significant difference in weight loss of fruits was observed due to the effects of oil treatments and time of storage and their interaction.

Generally it was found from data in Fig. 1 that treated fruits with fenugreek, cinnamon and thyme oils resulted in the lowest fruits weight loss being 10.22, 11.41 and 12.43%, respectively compared with control treatments (16.57%) at the end of storage time (45 days).

A similar percentages trend of oils coating treatment was detected when stored inoculated cherry tomato fruits with *A. alternata* and /or *B. cinerea* and treated with all the essential oils tested, specially fenugreek, cinnamon and thyme oils, respectively.

The reduction in weight loss was probably due to the effects of the coating oils used as a semi-permeable barrier against O_2 , CO_2 , moisture and solute movement, through reducing respiration, water loss and oxidation rates.

A similar effect was observed by Ben-Yehoshua (1969) for orange fruits coated with wax and those of Banks (1984), who reported that sucrose ester-based coating on banana fruits extended their storage life.

A similar result was reported by Tabaestani *et al.* (2013) which suggest that the cherry tomatoes weight loss can be delayed and storage period life can be extended when tomatoes stored at 20° C.

Shelf life

Extending the shelf life of tomatoes is very important for domestic and export marketing.

For results in Fig. 2 overall, the application of essential oils as cherry tomatoes coating could be a promising treatment to extend the shelf life of cherry tomatoes. Fruit under control treatment was recorded to give the longest life (12 days). The highest shelf life (15.5 days) was observed in thyme oil treatment, followed by oils treatment of cinnamon (15 days) of healthy fruits, while the lowest shelf life was recorded by spearmint and basil oils (10 and 9 days) respectively, similar to those obtained by Moneruzzaman *et al.* (2009).

From point of view the lowest days of shelf life (2 days) was recorded by fruits inoculated with both *A. alternata* and *B. cinerea* and oils untreated (Fig. 2 a and b).

In conclusion, from all aforementioned results, sensory evaluation proved the efficacy of coating fruits with essential oils and combined with modified atmosphere and cold storage conditions by maintaining the overall quality of tomato fruit during the storage period compared uncoated fruits.

Obtained results were in agreement with those reported sby Tabaestani *et al.* (2013), who concluded that coating cherry tomato fruits using basil mucilage edible coating and cumin essential oil and stored under 20°c improved tomato storability and extended the shelf life of cherry tomatoes.

Table 6-a.	Effect of black seed and fenugreek essential oil treatments on color changes of inoculated
	cherry tomato fruits with (Alternaria alternata and Botrytis cinerea) stored under (MA)
	conditions

	Color	Alte	ernaria alte	ernata	Botrytis cinerea			
	L*,a*,b*		Storage da	iys	Storage days			
Treatment		15	30	45	15	30	45	
	L*	24.33	22.61	20.22	24.64	22.70	21.29	
Control healthy	a*	13.80	11.66	10.64	15.22	13.29	12.30	
	b*	10.07	9.47	8.37	11.70	10.00	9.43	
	L*	23.47	21.84	19.65	22.21	21.00	20.32	
Inoculated (Inc.)	a*	10.75	9.28	8.61	14.40	11.81	12.71	
	b*	10.52	9.20	7.76	8.50	7.20	6.79	
Black seed	L*	25.13	23.71	22.82	19.70	18.54	17.55	
	a*	15.24	13.77	11.70	10.66	9.33	8.15	
	b*	13.82	11.62	9.88	10.35	9.25	7.21	
	L*	20.27	19.57	17.85	23.19	21.10	20.26	
Black seed + Inc.	a*	15.45	13.50	12.47	15.05	13.50	12.43	
	b*	10.33	8.68	6.74	13.20	10.50	8.61	
	L*	19.55	17.20	16.17	23.50	21.50	20.66	
Fenugreek	a*	12.07	10.40	8.41	14.20	13.25	12.61	
	b*	10.52	7.82	6.68	10.35	9.44	8.54	
	L*	24.55	23.60	2236	21.40	20.07	18.75	
Fenugreek + Inc.	a*	14.20	13.27	11.49	11.70	11.00	10.31	
	b*	13.51	11.44	10.56	9.29	8.13	6.97	

804

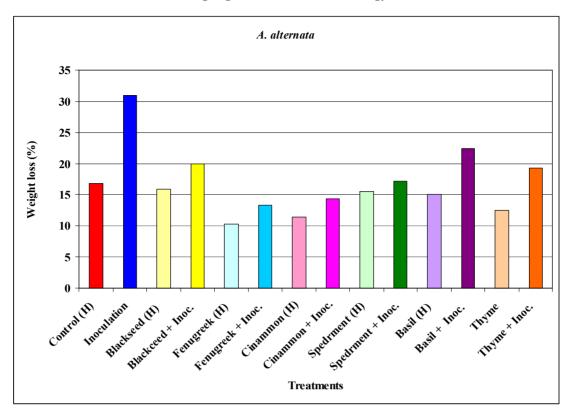
	Color	Alteri	naria alteri	nata	<i>Botrytis cinerea</i> Storage days			
	L*a*b*	St	orage day	8				
Treatment	-	15	30	45	15	30	45	
	L^*	24.80	21.95	20.29	20.80	19.95	18.78	
Control healthy	a*	10.65	9.32	8.48	17.40	10.34	9.70	
	b*	10.70	9.40	8.82	9.82	9.90	7.40	
	L*	20.40	18.21	17.36	23.50	20.40	18.70	
Inoculated (Inc.)	a*	10.33	9.50	9.23	12.23	11.04	10.43	
	b*	8.10	7.33	6.50	9.84	9.80	7.24	
	L^*	24.20	22.50	21.47	24.20	21.48	20.30	
Cinnamon	a*	11.40	11.01	9.24	12.15	10.19	12.04	
	b*	10.03	9.90	9.79	11.19	9.06	8.13	
	L*	20.22	20.00	18.80	22.32	20.00	19.64	
Cinnamon +Inc.	a*	13.74	12.11	11.84	14.34	13.24	9.65	
	b*	9.13	7.11	6.19	9.22	8.55	8.11	
	L*	24.44	20.50	21.40	20.82	18.21	17.36	
Spearmint	a*	15.33	14.20	14.15	11.24	9.45	8.76	
	b*	10.70	9.01	9.06	11.24	8.19	7.12	
	L*	21.50	20.18	19.81	17.80	17.00	16.22	
Spearmint + Inc.	a*	11.70	10.25	9.13	9.32	8.29	7.16	
	b*	9.18	8.02	7.31	9.30	7.55	6.31	

Table 6-b. Effect of cinnamon and spearmint essential oil treatments on color changes of inoculated cherry tomato fruits with (*Alternaria alternata* and *Botrytis cinerea*) stored under (MA) conditions

	Color L*a*b*	Alteri	aria alter	nata	<i>Botrytis cinerea</i> Storage days			
		St	orage day	ſS				
Treatment		15	30	45	15	30	45	
	L*	24.67	22.53	20.54	20.48	18.83	17.29	
Control healthy	a*	14.64	12.73	11.65	10.20	9.40	8.84	
	b*	9.53	8.59	8.45	10.03	9.35	8.24	
	L*	19.82	16.30	14.83	21.53	19.72	18.19	
Inoculated (Inc.)	a*	11.50	10.80	11.40	14.35	13.15	12.64	
	b*	9.11	9.40	7.50	8.73	7.40	6.47	
Basil	L*	22.43	20.48	21.53	25.83	22.83	21.97	
	a*	10.50	9.42	9.35	14.11	12.46	9.98	
	b*	9.25	9.00	8.72	10.50	9.98	9.83	
	L^*	21.97	20.73	19.84	21.55	19.73	19.18	
Basil + Inc.	a*	13.52	12.70	12.46	11.40	10.65	9.73	
	b*	10.42	9.55	8.54	10.35	8.54	7.52	
Thyme	L*	22.53	20.40	19.17	19.44	17.19	16.47	
	a*	14.25	13.00	12.75	9.24	9.01	7.98	
	b*	9.50	8.03	7.61	8.20	7.50	6.20	
	L*	24.83	22.40	20.98	22.71	20.77	19.76	
Thyme + Inc.	a*	15.20	13.19	12.99	17.42	15.44	14.68	
	b*	11.45	10.50	9.23	12.53	11.57	8.04	

 Table 6-c. Effect of basil and thyme essential oil treatments on color changes of inoculated cherry tomato fruits with (*Alternaria alternata* and *Botrytis cinerea*) stored under (MA) conditions

806



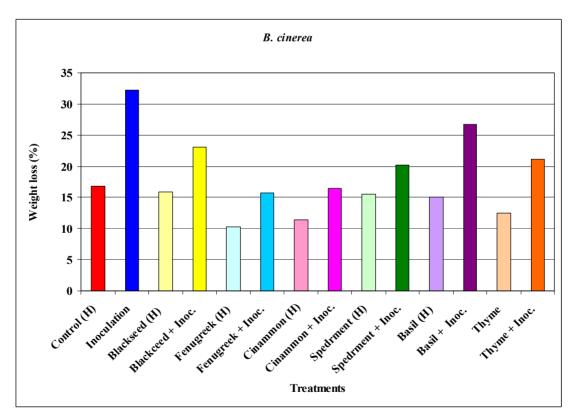
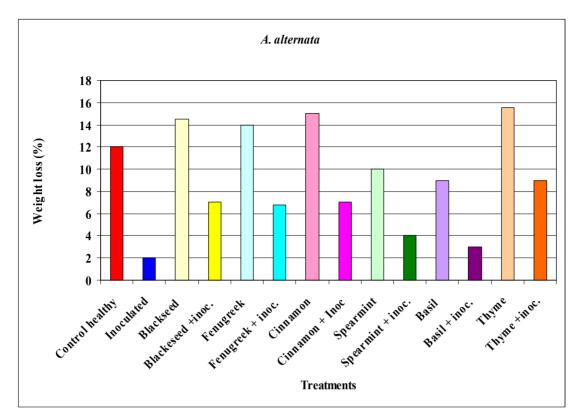


Fig. 1. Weight loss of cherry tomatoes uncoated (control) or coated with different essential oils and non or inoculated with fruit spoilage pathogens

H. = Healthy

Raafat, et al.



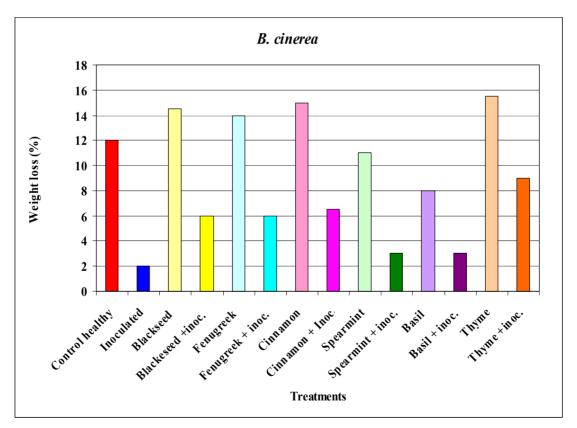


Fig. 2. Shelf life (days) of cherry tomatoes influenced by combined effect of essential oils coating, spoilage pathogens inoculation and storage conditions

REFERENCES

- Abd-Alla, M.A., M.M. Abd-El-Kader, F. Abd-El-Karem and R.S.R. El-Mohamedy (2011). Evaluation of lemongrass, thyme and picric acid against gray mold of strawberry fruit. J. Agric. Technol., 7 (6): 1775-1787.
- Ali, A., M. Maqbool, S. Ramachandran and P.G. Alderson (2010). Gum arabic as a novel coating for enhancing shelf-life and improving postharvest quality of tomato (*Solanum lycopersicum* L.) fruit. Postharvest Biol. and Technol., 58: 42- 47.
- Ali, B. and T. Keith (1998). Effect of modified atmosphere packing on postharvest qualities of Pink tomatoes. J. Agric. and Forestry, 22: 365-372.
- Ayranci, E. and S. Tunc (2003). A method for the measurement of the oxygen permeability and the development of edible films to reduce the rate of oxidative reactions in fresh foods. Food Chem., 80: 423- 431.
- Banks, N.H. (1984). Some effects of TAL-Prolong coating on ripening bananas. J. Exp. Bot., 35: 127 – 134.
- Barnett, H.L. (1962). Illustrated Genera of Imperfect Fungi, Second printing, Library of congress catalog, 60 (12998): 225.
- Barros, L., M. Duenas, A.M. Carvalho, C.S. Buelga and I.C. Ferreira (2012). Characterization and quantification of phenolic compounds in four tomato (*Lycopersicon esculentum* L.) farmers varieties in northeastern Portugal home gardens. Plant Food Hum Nut., 67 (3): 229-234.
- Ben-Yehoshua, S. (1969). Gas exchange, transportation and the commercial deterioration in storage of orange fruit, J. Am. Soc. for Hort. Sci., 94: 524 – 531.
- Benini, P.C., K.R.F. Schwam-Estrada, E.C. Klais, M.E.C. Crnz, A.t. Itako, R.M. Mesquini and J.B. Toliention Juniro (2010). *In vitro* effect on phytopathogens of essential oil and aqueous crude extract of *Ocimum gratissimum* harvest in the four seasons-Arquivos do Instituto Biologico., 77 : 677-683.

- Bernfeld, P. (1955). Mehtods in Enzymmologgy, Acad. Press, Inc.; New York, (1): 149-154.
- Bouchra, C., M. Achouri, L.M.I. Hassani and M. Humamouchi (2003). Chemical composition and antifungal activity of essential oils of seven *Moroccan labiatae* against *Botrytis cinerea* Pers: Fr., 89 (1): 165-169.
- Brand, S., Z. Pek, A. Lngasi and L. Helyes (2006). Lycopene content and color ripening tomatoes as affected by environmental conditions. J. Sci. Food and Agric., 86 : 568-572.
- Brown, W. (1924). A method of isolation single strains of fungi by cutting out a hyphal tip. Ann. Bot., 38: 402-404.
- Capassoll, F.S. and E.A.F. Torresll (2013). Antioxidant potential of tomatoes cultivated in organic and conventional systems. Brazilian of Biol. and Technol., 4.
- Cardile, V., A. Russo, C. Formisano, D. Rigano, F. Senatore and N.A. Arnold (2009). Essential oils of *Salvia bracteata* and *Salvia rubitolia* from Lebanon: chemical composition, antimicrobial activity and inhibitory effect on human melanoma cells. J. Ethnopharmacol., 126: 265- 272.
- Chen, Q., S. Xu, T. Wu, J. Guo and S. Sha (2014). Effect of citronella essential oil on the inhibition of postharvest *Alternaria alternata* in cherry tomato. J. Sci. Food Agric., 94 : 2441-2447.
- Cruz, E.M.S. (2003). Alternative products on the control of post harvest diseases in banana (*Musa paradisiaca* L.) apple (*Malus domestica* Barkh) and orange (*Citrus sinensis* L.). (Ph.D. Thesis). UEM: Universidada Estadual Maringa.
- Davies, J.N. and G.E. Hobson (1981). The constituents of tomato fruit: the influence of environment, nutrition and genotype. Critical Rev. in Food Sci. and Nut., 15: 205- 208.
- Debeaufort, F.J.A., Q. Gallo and A. Voilley (1998). Edible films and coating to packaging's: are view. Critical Rev. in Food Sci., 38: 299- 313.
- Deirdre, M., S. Holcroft and A.A. Kader (1999). Carbon dioxide-induced changes in color and

anthocyanin synthesis of stored strawberry fruit. Hort. Sci., 34 (7): 1244-1248.

- Dulce, M., C .Antunes and M.C. Ana (2010). The use of essential oils for postharvest decay control. A review: Flavor Fragr. J., 25 : 351-366.
- Erip-Roberts, B., A. Moraleja and B. Oleite (2002). Effect of storage temperature on ripening and postharvest quality of grape and mini-pear tomato. LWT. J. Food Sci. Technol., 43: 33 -38.
- Feng, W. and X.D. Zheng (2007). Essential oils to control *Alternaria alternata In vitro* and *In vivo*. Food Control, 18: 1126-1130.
- Frankel, C. and J.J. Jen (1989). Tomatoes. In Michael Eskin, N.A. (ed.), Quality and Preservation of Vegetables, CRC Press, Inc. Boca Raton, Florida, 58-68.
- Fraser, P.D., P. Bramleya and G.B. Seymsy (2000). Effect of the Cnr mutation on carotenoid formation during tomato fruit ripening. Phytoch., 58 : 75-79.
- Gharezi, M., N. Joshi and E. Sadeghin (2012). Effect of Post-harvest treatment on stored cherry tomatoes. Food Sci. and Nut., 2 (8): 150-157.
- Guttinger, T. (1981). Polyphenols in olive. J. Am. Oil Chem. Soc., 58: 966- 986.
- Hammer, K.A., C.F. Carson and T.V. Riley (2001). Antimicrobial activity of essential oils and other plant extracts. J. Appl. Microbiol., 86 (6): 985 – 990.
- Helyes, L., Z. Pek and A. Lugasi (2006). Tomato fruit quality and content depend on stage of maturity. Hort. Sci., 41: 1400-1401.
- Kader, A.A. (1991). Quality and its maintenance in relation to the postharvest physiology of strawberry. The strawberry into the 21st. Century: Proc. Third North Am. Strawberry Conf. Timber Press, Portland. OR, 145-152.
- Kagan-Zur, V. and Y. Mizrahim (1993). Long shelf-life small sized (cocktail) tomatoes may be picked in bunches. Scientia Hort., 56 : 31-41.

- Kaur, C., B. George, N. Deepa, B. Singh and H.C. Kapoor (2004). Antioxidant status of fresh and processed tomato. A Rev. J. Food Sci. and Technol., 41: 479-486.
- Korsten, L. (2006). Advances in control of postharvest diseases in tropical fresh produce. Int. J. Postharvest Technol. Innovat., 1 (1): 48-61.
- Kumarrai, G., R.A. Kumar, A.K. Singh, M.R. Rai, A.K. Chanturvedi and A.B. Rai (2012). Changes in antioxidant and phytochemical properties of tomato (*Lycopersicon esculentum* MILL.) under ambient condition. Pak. J. Bot., 44 (2): 667-670.
- Lingk, W. (1991). Health risk evaluation of pesticide contamination in drinking water-Gesunde. Pflanen, 43: 21-25.
- Loucoss, E.H. (1994). Determining ascorbic acid in large numbers of plant samples. Ind. Eng. Chem. Anal., 15: 649-652.
- Madhavi, D.L. and D.K. Salunkhe (1998). Tomato. In: Sakunkhe Dk, kadam SS (Eds.) Hand book of Vegetable Science and Technology: Production, Composition. Storage and Processing. Marcel Dekker Inc, New York, USA, 171- 202.
- Marjanlo, A., Y. Mostofi, Sh. Shoeibi and M. Fattahi (2009). Effect of cumin essential oil on postharvest decay and some quality factors of strawberry . J. Med. Plant, (8) 31: 25-43.
- Marsic, N.K., L. Gasperlin, V. Abram, M. Budic and R. Vidrih (2011). Quality parameters and total phenolic content in tomato fruits regarding cultivar and microclimatic conditions. Turk J. Agric., (35): 185-194.
- Meredith, F.I. and A.E. Purcell (1966). Changes in the concentration of carotenes of ripening Homestead tomatoes. Proceedings of the Am. Soc. for Hort. Sci., 89: 544- 552.
- Miller, G.L. (1959): Use of di nitro salicylic acid reagent for determination of reducing sugar. Am. Chem., 31: 426-428.
- Miller, N.J. and C.R. Evans (1997). The relative contributions of ascorbic acid and phenolic antioxidants to the total antioxidant activity

of orange and apple fruit juices and black currant drink. Food Chem., 60: 331- 337.

- Molyneux, S.L., C.E. Lister and G.P. Savage (2004). An investigation of the antioxidant properties and color of glasshouse grown tomatoes. Int. J. Food Sci. and Nut., 55: 537-545.
- Moneruzzaman, K.M., A.B.M.S. Hossain, W. Sani, M. Saifuddin and M. Alenazi (2009).
 Effect of harvesting and storage condition on the postharvest quality of tomato (*Lycopersicon esculentum* Mill) CV. Roma VF. Australian J. Crop Sci., 3 (2): 113-121.
- Nabifarkhani, S.M., A.D. Garmakhany, E.G. Moghadam and M.A. Shakevi (2015). Effect of nano-composite and thyme oil (*Thymus vulgaris* L.) coating on fruit quality of sweet cherry (Takdaneh CV) during storage period. Food Sci. and Nut., 3 (4): 349-354.
- Nguefak, J., V. Leth, A. Zollo and S.B. Mathur (2004). Evaluation of five essential oil from aromatic plants of Cameroon for controlling food spoilage and mycotoxin producing fungi. Int. J. Food Microbiol., 94: 329-334.
- Ozkaya, O., D. Omur, C.S. Giulia and V. Giorio (2009). Evaluation of quality parameters of strawberry fruits in modified atmosphere packaging during storage. Afr. J. Biotech., 8 (5): 789-793.
- Passam, H.C., C.K. Joannis, J.B. Penelope and S. Dimitrios (2007). A review of recent research on tomato nutrition, breeding and postharvest technology with reference to fruit quality. The European. of Plant Sci. and Biotechnol., Global Sci. Books.
- Petriccione, M., F. Mastrobuoni, M.S. Pasquariello, M.S. Pasquariello, L. Zampella, E. Nobis, G. Capriolo and M. Scortichini (2015). Effect of chitosan coating on the postharvest quality and antioxidant enzyme system response of strawberry fruit during cold storage, Foods, 4: 501-523.
- Plotto, A., D.D. Roberts and R.G. Roberts (2003). The use of natural aromatic essential oils helps to maintain postharvest quality of crimson table grapes. Acta. Hort., 682 : 1723-1732.

- Sajad, F., J. Mehrdad, E. Shahin, R. Abbas and B. Hasan (2011). Effect of essential oils of *Thymuo vulgaris* and *Mentha piperita* on the control of green mould and postharvest quality of *Citrus sinensis* cv. Valencia. Afr. J. Biotech., 10 (66): 14932 – 14936.
- Saltveit, E.M. (2005). Fruit ripening fruit quality. In: Heuvelink, E (ed.), Tomatoes, CAB Int. Walling Ford, UK., 145-170.
- Serrano, M., D. Martinez- Romero, S. Castello, F. Guillen and D. Valero (2005). The use of natural antifungal compounds improves the beneficial effect of MAP in sweet cherry storage Jnnov. Food Sci. Emerg. Technol., 6 (1): 115- 123.
- Shirzad, H., H. Abbas, G. Youbert, F.A. Rasool and H.M. Mohammad (2011). Assessment of the antifungal activity of natural compounds to reduce postharvest gray mould (*Botrytis cinerea* Pers. Fr.) of kiwi fruits (*Actinidia deliciosa*) during storage. J. Pl. Prot. Res., 51: 1-6.
- Snedecor, G.W. and W.G. Cochran (1980). Statistical Method 7th Ed. Iowa State Univ. Press, Ames. Iowa, USA.
- Snell, R. and G. Snell (1953). Colorimetric Method of Analysis-Vol. III. 3rd Ed. New York, D. Van Nostrand Company Inc., P-225-233.
- Soliman, K.M. and R.I. Badeaa (2002). Effect of oil extracted from some medicinal plants on different mycotoxigenic fungi. Food and Chem. Toxicol., 40 (11): 1669-1675.
- Somda, I., V. Leth and P. Sereme (2007). Antifungal effect of *Cymbopogon citrates, Eucalyptus camldeulensis* and *Azadirachta indica* on sorgaum seed borne fungi. Asian J. Pl. Sci., 6 (8): 1182-1189.
- Sun Og, L., J.C. Gyung, S.J. Kyoung, K.L. He, Y.C. Kwang and K. Jin-Cheol (2007). Antifungal activity of five plant essential oils as fumigants against postharvest and soil borne plant pathogenic fungi. Pl. Pathol. J., 23 (2): 97-102.
- Tabaestani, H.S., N. Sedaghat, E.S. Pooya and A. Alipour (2013). Shelf life improvement and postharvest quality of cherry tomato

(*Solanum lycopersium* L.) fruit using basil mucilage edible coating and cumin essential oil. Int. J. Agron. and Pl. Prod., 4 (9): 2346-2353.

- Tian, J., X. Ban, H. Zeng, B. Huang, J. He and Y. Wang (2011). *In vitro* and *in vivo* activity of essential oil from dill (*Anethum* graveolens L.) against fungal spoilage of cherry tomatoes. Food Control, 22 : 1992-1999.
- Tsuda, T., K. Chachin and Y. Ueda (1999). Studies on keeping capacity of imported Carabo mango fruit from the Philippines. J. Jap. Soc. Hort. Sci., 69 (3): 669-674.
- Tzortzakis, N.G. (2007). Maintaining postharvest quality of fresh produce with volatile compounds. Innovative Food Sci. and Emerg. Technol., 8: 111- 116.
- Tzortzakis, N.G. and C.D. Economakis (2007). Antifungal activity of lemongrass (*Cymbopogon citrates* L.) essential oil against postharvest pathogen. Innovation Food Sci. and Emerg. Technol., 8 (2): 253-258.
- Tzortzakis, N.G. (2009). Impact of cinnamon oil - enrichment on microbial spoilage of fresh produce. Innovative Food Sci. and Emerg. Technol., 10: 97- 102.
- Vesaltalab, Z. and M. Gholami (2012). Effect of essence and extract of *Eugenia caryophyllata* on some qualitative characters of grape

during storage period. Irani. J. Hort., 43: 255-265.

- Wang, Y., X. Ren, X. Song, T. Yu, H. Lu, J. Wang and X.D. Zheng (2010). Control of postharvest decay on cherry tomatoes by marine yeast *Rhodosporidium paludigeneum* and Calcium Chloride. J. Appl. Microbiol. ISSN: 651-656.
- Wold, A-B., H.J. Rosenfeld, H. Badgered and R. Blomhoff (2004). The effect of fertilization on antioxidant activity and chemical composition of tomato cultivars (*Lycopersicon esculentum* Mill.). European J. Hort. Sci., 69: 167-174.
- Xing, Y., X. Li, Q. Xu, C. Shao and J. Yun (2011). Antimicrobial activity of microencapsulated cinnamon oil and its application of cherry tomato. Adv. Materials Res. Vols., 236 (238): 2307-2310.
- Xu, S., F. Yan, Z. Ni, Q. Chen and H. Zheng (2014). *In vitro* and *in vivo* control of *Alternaria alternata* in cherry tomato by essential oil from *Laurus nabilis* of Chinese origin. J. Sci. Food and Agric., 94 (7): 1403-1408.
- Yan, F., S. Xu, J. Guo, Q. Chen and Q. Meng (2014). Bio control postharvest *Alternaria alternata* decay of cherry tomatoes with rhamnolipids and possible mechanisms of action. J. Sci. Food Agric., 95:1469-1474.

مردود معاملة ثمار الطماطم الشيري ببعض الزيوت النباتية الطيارة علي مقاومة أعفان الثمار وإطالة. عمر الثمرة وتحسين صفات الجودة أثناء التخزين

تعد سلامة الغذاء أهم مشكلة تواجه الخضروات والفواكه الطازجة، والحفاظ على جودة ثمار الطماطم الشيرى تعد كاشفة للمردود الاقتصادى لها، عوملت ثمار الطماطم الشيرى (السليمة أو المحقونة بفطرى عفن ثمار الطماطم ألترناريا الترناتا وبوتريتس سينيريا) تحت ظروف التخزين فى الجو المعدل (١٢%ك أب + ٦% أب) والمبرد (٥٥م ± ١ ورطوبة نسبية ٩٠% ± ٣) ببعض الزيوت النباتية الطيارة لنباتات حبة البركة، الحلبة، القرفة، النعناع، الريحان والزعتر بتركيز ١٠%، حيث توصلت النتائج إلى خفض معدل حدوث عفن ثمار الطماطم المعاملة، انخفضت صفات جودة الثمار المصابة وغير المعاملة بالزيوت النباتية متأثرة بطول فترة التخزين ماعدا الثمار المعاملة بالزيوت حيث أبطأت من معدل وفيتامين جودة الثمار المعاملة بالزيوت النباتية متأثرة بطول فترة التخزين ماعدا الثمار المعاملة بالزيوت حيث أبطأت من معدل وفيتامين ج، الليكوبين، الفينولات) مقارنة بالثمار غير المعاملة بالزيوت حيث أبطأت من معدل وفيتامين ج، الليكوبين، الفينولات) مقارنة بالثمار غير المعاملة بالزيوت حيث أبطأت من معدل معاملة المقارنة وكذا معاملة الثمار المحقونة بالفطريات و غير المعاملة بالزيوت ديث أبطأت من معدل معاملة المقارة وكذا معاملة الثمار المعاد المائية الفقد فى الوزن، درجة الحموضة، السكريات، ومضادات الأكسدة وليتامين ج الليكوبين، الفينولات) مقارنة بالثمار غير المعاملة بالزيوت، و عليه أوضحت النتائج أن الزيوت النباتية معاملة المقارنة وكذا معاملة الثمار المحقونة بالفطريات و غير المعاملة بالزيوت، وعليه أوضحت النتائج أن الزيوت النباتية معاملة المقارة ويذا معاملة الثمار المحقونة بالفطريات و غير المعاملة بالزيوت، وعليه أوضحت النتائج أن الزيوت النباتية

المحكمون :

۱ ـ أ.د. محمود رزق الله عسران

۲ - أ.د. هاني محمد السعيد محمد

أستاذ ورئيس قسم أمراض النبات – كلية الزراعة – جامعة سوهاج

محمد الستاذ أمراض النبات المتفرغ – كلية الزراعة – جامعة الزقازيق