

Spatial and Temporal Distribution of Plankton in a Tropical Reservoir, southwestern Nigeria

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ABSTRACT

Spatial and temporal variations in the distribution of plankton of a tropical reservoir were investigated for six months. Chlorophyceae was richest in the phytoplankton community in term of the number of species and individuals. They were more abundant in the late rainy /early dry season with *Pediastrum simplex* and *Coelastrum chodati* being the dominant species. Constant species occurred only in Chlorophyceae, which included *Spirogyra*, *Closterium* sp., *Coelastrum chodati*, *Pediastrum simplex*, *Ulothrix zonata* and *Scenedesmus quadricauda*. Cyanophyceae developed mainly in the late dry season months (reached peak in March) with *Microcystis flos-aqua* being dominant. In the zooplankton community, Crustacea had the highest number of individuals and were more abundant in the late dry season. *Camptocercus* sp., *Bosmina* sp. and *Daphnia magna* were the main species. Rotifers had the highest species number with peaks observed in the late rainy/early dry season. The common species encountered included *Chromagaster* sp., *Epiphanes* sp., *Gastropus* sp., *Nolthoca* sp., *Trichocerca* sp., and *Brachionus* sp. The common protozoan species were *Ichthyophthrius*, *Chilodonella*, *Prorodon* and *Colpoda*. The diversity indices calculated varied spatially; phytoplanktonic organisms were more diverse in Station 1 with the least evenness, highest evenness occurred in Station 2. The increase in total number of taxa encountered in this study compared to previous studies and implication of *Microcystis flos-aqua* being the dominant blue- green algae were discussed.

INTRODUCTION

The quality and quantity of water in reservoirs are very important especially in reservoirs with multifunctional roles. The health of the aquatic ecosystem of lakes are very sensitive issues and lakes in different regions of the world particularly in developing countries are facing a variety of problems associated with anthropogenic activities and unsustainable use of their resources as demonstrated by **El-Serehy et al. (2018)**. Deterioration of water quality in reservoirs results from excessive nutrients inputs, eutrophication, acidification, heavy metal contaminants, organic pollutants and obnoxious fishing practices as asserted

by **Gupta (2014)**. The monitoring of water quality of reservoirs is important as it helps with the management of the eutrophication and productivity of the water body. Eutrophication has a considerable impact on the two main components of the plankton communities (phytoplankton and zooplankton) causing many changes in their abundance and species composition and affecting the relationships between them. Changes in the plankton community structure in relation to physicochemical parameters may be a first sign of deterioration in the water quality (**Ochocka and Pasztaleniec, 2016**). They also suggested that both plankton elements (phytoplankton and zooplankton) should be taken into account in the ecological status assessment of lakes. Studies on the structure and functioning of planktonic communities in reservoir ecosystems provide opportunities to investigate patterns of responses to cyclical variations and episodic disturbances as suggested by **Nogueira (2001)**. Biomass, taxonomical composition and diversity of phytoplankton are part of the criteria on which the trophy of a water body might be assessed as suggested by **OECD (1982)**. The habitat-template approach proposes that algal attributes are matched to opportunities provided by the environment and that there is an encouraging fit of phytoplankton species to the range of habitats described by the trophic spectrum as written by **Akin-Oriola (2003)**. The changes in the phytoplankton community can be particularly useful as an assessment tool, due to their rapid response to environmental stress. The qualitative and quantitative studies of phytoplankton may provide good indices of water quality and capacity of water to sustain heterotrophic communities as suggested by **Agarwal et al. (2018)**

Zooplankton is one of the most important biotic elements that impact all functional aspects of aqueous ecosystems including food chains and trophic networks, energy flow, and the circulation of matter as asserted by **Paturej et al. (2017)**. Species composition and abundance of zooplankton communities can be influenced by a number of physical, chemical and biological factors as demonstrated by **Sampaio et al. (2002)**. The composition and abundance of zooplankton have been reported to be significantly impacted by the trophic state of reservoirs (**Pinto-coelho et al., 2005; Paturej, 2006; Kudari and Kanadami, 2008 and Gonzalez et al., 2011**)

Awba Reservoir, a man-made lake on the Awba Stream, University of Ibadan serves as a source of water supply to the water treatment plant for domestic uses in the University. The Awba stream however receives untreated effluents from the staff and student residences, Zoological Garden, Faculty of Science laboratories and its environment. Recently, the lake has been earmarked for ecotourism alongside water supply; water-based tourism activities include boating, sailing, motor boating, swimming, skiing, and fishing activities, these are often less detrimental to water quantity and quality compared to many other human uses of water like agriculture, industry etc. as suggested by **Long (2012)**. Previous studies on the ecology of the reservoir were recorded, including **Mombeshora et al. (1981)** on the trace metal level; **Ugwumba and Adebisi (1992)**, on food and feeding relationship in fish; **Akin-Oriola (2003)**, **Chukwuka and Uka (2007)**; **Anago et al. (2013)** on plankton composition and **Aderogba & Ayoade (in press)** on trophic status. The aim of this paper was to update knowledge on the current status of limnology of the reservoir through assessment of the plankton composition and abundance of the Awba reservoir, Ibadan.

MATERIALS AND METHODS

The Awba reservoir on latitude 7° 26' - 7° 27' N and longitude 3° 53' - 3° 54' E (Figure 1) is located in the University of Ibadan, south western region of Nigeria about 160 km from the Atlantic Ocean coast at an altitude of 185 m above sea level as written in **Akin-Oriola (2003)**. The reservoir was constructed in 1964 with a maximum depth of 5.5 m and surface area of about 6 ha. The annual rain occurs from April to October with a characteristic August break during which the rain abates. The water of the lake is still with occasional multi-directional water movements due to wind effects. Wind action in the reservoir is minimal in the dry season and the high temperatures at this period result in thermal stratification of the water. It is surrounded by a modified tropical rain forest vegetation with aquatic macrophytes such as *Pistia stratiotes*, *Canna indica*, *Nymphae lotus*, *Banahima* sp. on its edges as written in **Akin-Oriola (2003)**.

Field sampling was conducted from October 2017 to March 2018 (October- January, late rainy/early dry season; February – March, late dry season) at monthly interval from three selected sampling stations on the reservoir. Plankton were collected with a 55 µm bolting silk plankton net and preserved in 4% formalin. One mL of a sub-sample was randomly taken from the bottle by using pipette and placed into a Sedgewick Rafter counting cell and slowly covered with a cover slip. Sorting, identification, and counting were carried out under a compound microscope. Plankton identification was analysed at the lowest possible taxonomic level according to the standard taxonomic references (**Whitford and Schumacher 1973; Needham and Needham 1974; Jeje and Fernando 1987 and Nwankwo 2004**).

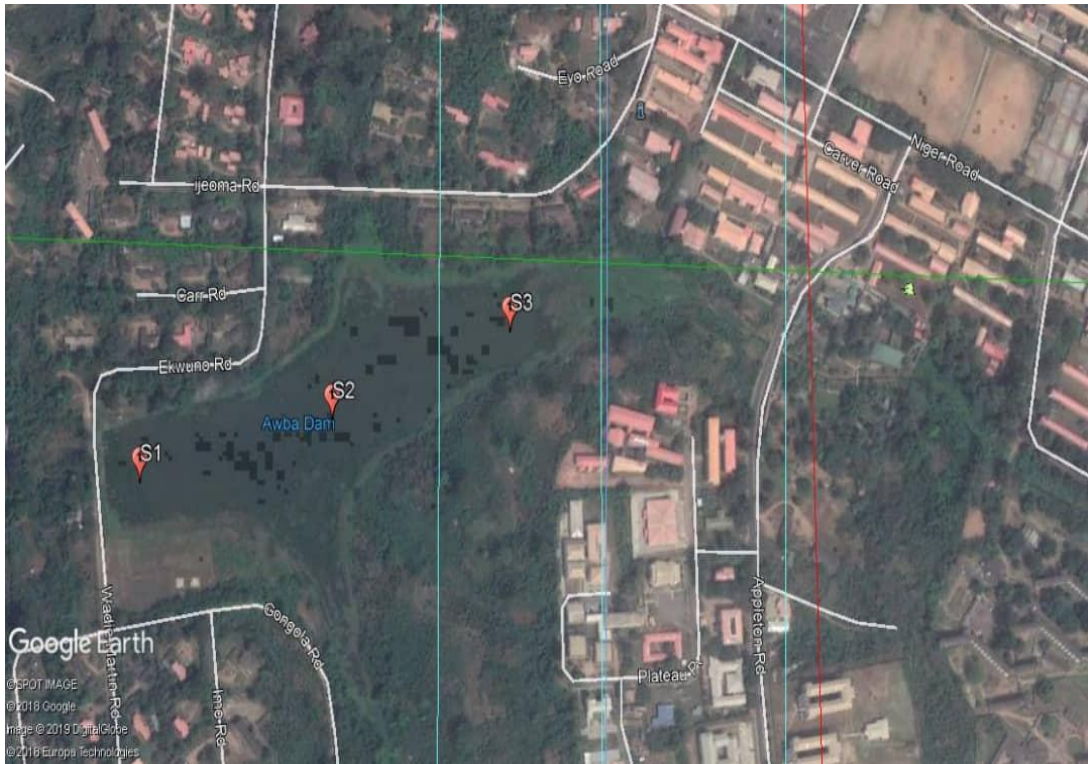
Diversity was evaluated using the Shannon Wiener index (**Pielou 1975**). Margalef's Index (d) known as species richness index was calculated as given by **Margalef (1958)**. Evenness (E) expresses the degree of uniformity in the distribution of individual among the taxa was also calculated as given by **Magurran (1988)**. The Sørensen index by **Sørensen (1948)** was used to compare the plankton species compositions of the different stations. Species were classified according to their frequency of occurrence as: constant (50% or more), common (between 10% and 50%) or rare (below 10%), as described in **Gomes (1989)**.

RESULTS

Phytoplankton Composition and Abundance

The phytoplankton composition of Awba reservoir during the study period was represented by 7 families, 83 genera and 126 species. Chlorophyceae was the dominant form in the community in term of number of species (70) and abundance (9.63×10^5 organisms/mL). This was followed by Cyanophyceae with 28 species and 4.79×10^5 organisms/mL. Most abundant species were *Pediastrum simplex* (Meyen) Lemmermann (green algae, 1.78×10^5 organisms/mL), *Coelastrum chodatii* Duceillier (green algae, 1.71×10^5 organisms/mL), *Closterium gracile* Breb. (desmid, 1.49×10^5 organisms/mL) and *Microcystis flos-aqua* (Wittr.) Kirchner (blue green algae, 1.04×10^5 organisms /mL) Table 1. The total number of species varied from 76 in Station 1 to 89 in Station 2. However, the highest phytoplankton abundance was recorded in Station 1 (7.24×10^5

organisms /mL) and the least in Station 3 (4.24×10^5 organisms /mL). Generally there was a gradual decrease in abundance of major phytoplankton group from Station 1 to Station 3 except Chrysophyceae and Euglenophyceae (Figure 2).



Legend: S1 – Station 1, S2- Station 2, S3- Station 3

Figure 1: A map of University of Ibadan Showing Awba Reservoir and Sampling Stations

Table 1: Relative abundance of phytoplankton organisms of Awba Reservoir, Ibadan, Nigeria

CHLOROPHYCEAE	Total Abundance (Organism/ mL)	%
<i>Actinastrum hantschii</i>	4000	0.23
<i>Ankistrodesmus</i> sp.	5000	0.29
<i>Asterococcus limneticus</i>	8000	0.46
<i>Binuclearia tatrana</i>	1000	0.06
<i>Botryococcus</i> sp.	9000	0.52
<i>Chlamydomonas feneserata</i>	5000	0.29
<i>Chlorella vulgaris</i>	32000	1.83
<i>Cladophora</i> sp.	1000	0.06
<i>Clostridium retigerum</i>	30000	1.72
<i>Coelastrum angustae</i>	39000	2.23
<i>Coelastrum chodatii</i>	171000	9.78
<i>Coelastrum morus</i>	28000	1.60
<i>Coelastrum probescideum</i>	1000	0.06
<i>Coleochaete orbicularis</i>	2000	0.114

<i>Coleoclaete scutata</i>	1000	0.06
<i>Crucigenia crucifera</i>	1000	0.06
<i>Crucigenia quadrata</i>	1000	0.06
<i>Dispora crucgenibides</i>	1000	0.06
<i>Dysmorphococcus variabilis</i>	1000	0.06
<i>Eudorina elegans</i>	2000	0.114
<i>Geminella minor</i>	2000	0.114
<i>Gloeocystis botryoides</i>	10000	0.57
<i>Gonphosphaeria wichurae</i>	18000	1.03
<i>Groenbladia neglecta</i>	1000	0.06
<i>Hematococcus lacustris</i>	5000	0.29
<i>Hormidium rivulare</i>	4000	0.23
<i>Hormidium subtle</i>	2000	0.114
<i>Hydrodictyon</i> sp.	1000	0.06
<i>Menoidium gracile</i>	1000	0.06
<i>Microspora amoena</i>	22000	1.26
<i>Mougeotia</i> sp.	3000	0.17
<i>Myrecia globosa</i>	1000	0.06
<i>Oedogonium</i> sp.	7000	0.40
<i>Ophiocytium</i> sp.	1000	0.06
<i>Oocystis vulgaris</i>	1000	0.06
<i>Palmellococcus minutus</i>	1000	0.06
<i>Palmellococcus protothecoides</i>	4000	0.23
<i>Palmodictyon varium</i>	6000	0.34
<i>Pandorina morum</i>	2000	0.114
<i>Pediastrum boryanum</i>	4000	0.23
<i>Pediastrum duplex</i>	47000	2.69
<i>Pediastrum simplex</i>	178000	10.18
<i>Protococcus</i> sp.	1000	0.06
<i>Pseudendodonium</i> sp.	1000	0.06
<i>Quadrigula closteriodes</i>	1000	0.06
<i>Scenedesmus abundans</i>	7000	0.40
<i>Scenedesmus acuminatus</i>	2000	0.114
<i>Scenedesmus armatus</i>	27000	1.55
<i>Scenedesmus bijuga</i>	1000	0.06
<i>Scenedesmus oahuensis</i>	5000	0.29
<i>Scenedesmus oblige</i>	2000	0.114
<i>Scenedesmus opoiensis</i>	3000	0.17
<i>Scenedesmus polifica</i>	7000	0.40
<i>Scenedesmus quardricauda</i>	30000	1.72
<i>Schroederia setigera</i>	1000	0.06
<i>Schitochlamys gelatinosa</i>	1000	0.06
<i>Sorastrum</i> sp.	1000	0.06

<i>Spirogyra</i> sp.	34000	1.95
<i>Spirotaenia</i> sp.	3000	0.17
<i>Stigmeoclonium carolinianum</i>	1000	0.06
<i>Tetraspora</i> sp.	11000	0.63
<i>Trebouria cladoniae</i>	8000	0.46
<i>Ulothrix amphigranulata</i>	13000	0.47
<i>Ulothrix terierrima</i>	1000	0.06
<i>Ulothrix variabilis</i>	29000	1.66
<i>Ulothrix zonata</i>	58000	3.32
<i>Volvox dissipathrix</i>	3000	0.17
<i>Volvox perglobator</i>	6000	0.34
<i>Volvox proiliticus</i>	16000	0.92
<i>Zygnema</i> sp.	26000	1.49
Subtotal	963000	55.09
DESMIDACEAE		
<i>Cosmarium cucumis</i>	32000	1.83
<i>Closterium gracile</i>	149000	8.52
<i>Gonatozygon aculeatum</i>	8000	0.46
<i>Micraterias thomasi</i>	2000	0.114
SUBTOTAL	191000	10.93
CYANOPHYCEAE		
<i>Anabaena azolla</i>	13000	0.74
<i>Aphanocapsa delicatissima</i>	36000	2.06
<i>Aphanocapsa elachista</i>	3000	0.17
<i>Aphanocapsa pulchra</i>	59000	3.38
<i>Capsosira</i> sp.	1000	0.06
<i>Chroococcus pactidus</i>	1000	0.06
<i>Chrysococcus turgidus</i>	1000	0.06
<i>Coelospharium kuetzingianum</i>	19000	1.09
<i>Coelospharium naegeliarium</i>	8000	0.46
<i>Cyanoptycha gloccystis</i>	9000	0.52
<i>Glosocapsa alpicola</i>	13000	0.74
<i>Glosocapsa granosa</i>	4000	0.23
<i>Glosocapsa lacustris</i>	1000	0.06
<i>Glosocapsa magna</i>	1000	0.06
<i>Lyngbya aerugineocarulea</i>	2000	0.114
<i>Merismopedia convolute</i>	6000	0.34
<i>Microcystis aeruginosa</i>	90000	5.15
<i>Microcystis flos-aqua</i>	104000	5.95
<i>Microcystis pulverea</i>	17000	0.97
<i>Nostoc parmelioides</i>	4000	0.23
<i>Nostoc piscinade</i>	9000	0.52
<i>Oscillatoria prolifica</i>	11000	0.63

<i>Oscillatoria sancta</i>	51000	2.92
<i>Phormidium angustisi</i>	2000	0.114
<i>Phormidium corium</i>	7000	0.40
<i>Phormidium teune</i>	3000	0.17
<i>Rivularia</i> sp.	2000	0.114
<i>Tolypothrix fragalis</i>	2000	0.114
SUBTOTAL	479000	27.4
EUGLENOPHYCEAE		
<i>Euglena caudate</i>	26000	1.49
<i>Euglena acus</i>	8000	0.46
<i>Euglena deses</i>	12000	0.69
<i>Euglena fusa</i>	6000	0.34
<i>Euglena pronaina</i>	1000	0.06
<i>Euglena spiroides</i>	3000	0.17
<i>Euglena varibilis</i>	1000	0.06
<i>Trachelomonas ensifera</i>	3000	0.17
<i>Trachelomonas horrida</i>	1000	0.06
<i>Trachelomonas volgensis</i>	1000	0.06
SUBTOTAL	62000	3.55
BACILLARIOPHYCEAE		
<i>Amphipleura pellucida</i>	2000	0.114
<i>Cymbella</i> sp.	2000	0.114
<i>Cyclotella</i> sp.	5000	0.29
<i>Gyrosigma</i> sp.	1000	0.06
<i>Mastigloia</i> sp.	6000	0.34
<i>Navicula</i> sp.	7000	0.40
<i>Nitzschia</i> sp.	7000	0.40
<i>Rhizosolenia longiseta</i>	1000	0.06
<i>Synedra</i> sp.	1000	0.06
SUBTOTAL	32000	1.83
CHRYSOPHYCEAE		
<i>Amphichrysis compressa</i>	3000	0.17
<i>Chrysamoeba radian</i>	3000	0.17
<i>Chrysosphaera pacudosa</i>	1000	0.06
<i>Uroglenopsis</i> sp.	1000	0.06
SUBTOTAL	8000	0.42
XANTHOPHYCEAE		
<i>Tribonema</i> sp.	13000	0.74
SUBTOTAL	13000	0.74

Chlorophyceae dominated the phytoplankton community in late rainy /early dry season. Cyanophyceae were encountered mainly in the late dry season months and reached peak in March (3.59×10^5 organisms/mL). Euglenophyceae also attained peak in abundance (7 species and 4.6×10^4 organisms/ mL) in late dry season months Figure 3. Constant

species according to criteria used occurred only in Chlorophyceae which included *Spirogyra*, *Closterium* sp, *Coelastrum chodatii* Duceillier, *Pediastrum simplex*, *Ulothrix zonata* (Weber & Mohr) Kutz. and *Scenedesmus quadricauda* (Turp.) Brebisson. The common blue-green algae included *Coelosphaerium*, *Polycystis*, *Nostoc*, *Anabaena*, *Aphanocapsa*, *Microcystis flos-aqua*, *Phormidium corium* (Ag.) Gom., *Oscillatoria prolifica* (Grev.) Gom., and *Glosocapsa granosa* (Berk.) Kutz. A large number of species were considered rare and represented by 35 green algae, 10 blue-green algae, 7 diatoms, 5 euglenoids, 3 golden algae and 1 yellow green algae.

Zooplankton Composition and Abundance

Three zooplankton taxa were identified and Crustacea made up the largest percentage (41.22% of total zooplankton number) with Protozoa being the least (19.85% of total zooplankton number), as in Table 2. However, rotifers (12) had the highest species number followed by protozoan (10). The species that were relatively more abundant were *Camptocercus* sp. (Cladocera, 12.98%), *Bosmina* sp. (Cladocera, 12.21%), *Daphnia magna* Straus, 1820 (Cladocera, 12.21%), *Chromagaster* sp. (Rotifer, 11.45%), *Notholca* sp. (Rotifer, 5.34%), *Ichthyophthrius* sp. and *Chilodonella* sp. (Protozoa, 4.58% each) and *Epiphanes* sp. (Rotifer, 4.58%). The highest zooplankton abundance occurred in Station 2(40%) and least in Station 1(29.6%) as in Figure 4. Rotifers dominated the zooplankton community in the late rainy/early dry season, while crustacean were more abundant in late dry season (Figure 5). The constant zooplankton according to criteria used was only microcrustaceans, *Bosmina* sp., and *Camptocercus* sp., while *Daphnia*, *Coleps* and *Cypridopsis* were common. Among the rotifers, the common species included *Chromagaster*, *Epiphanes*, *Gastropus*, *Nolthoca*, *Trichocerca*, and *Brachionus*. The common protozoan species were *Ichthyophthrius*, *Chilodonella*, *Prorodon* and *Colpoda*. The species considered to be rare were two crustaceans, three rotifers and six protozoans.

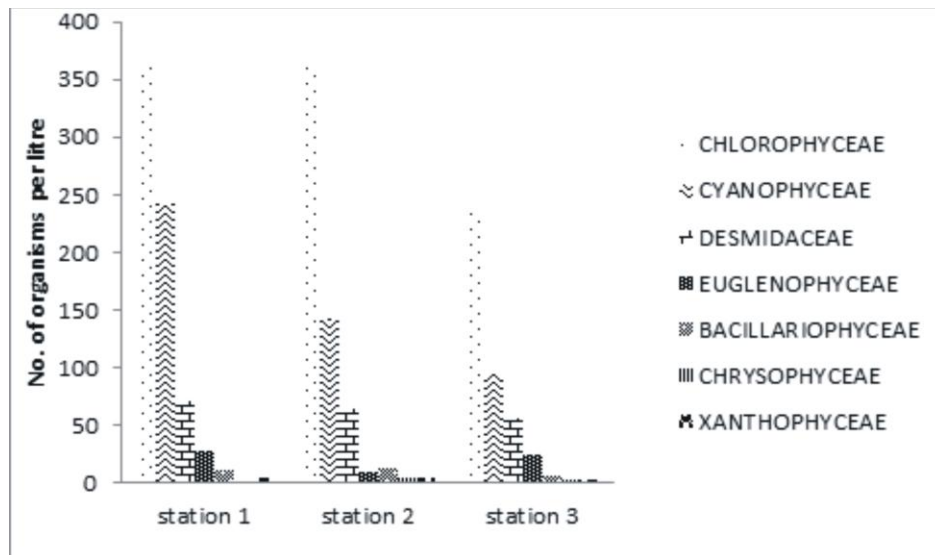


Figure 2: Spatial variation in abundance of major phytoplankton groups of Awba Reservoir

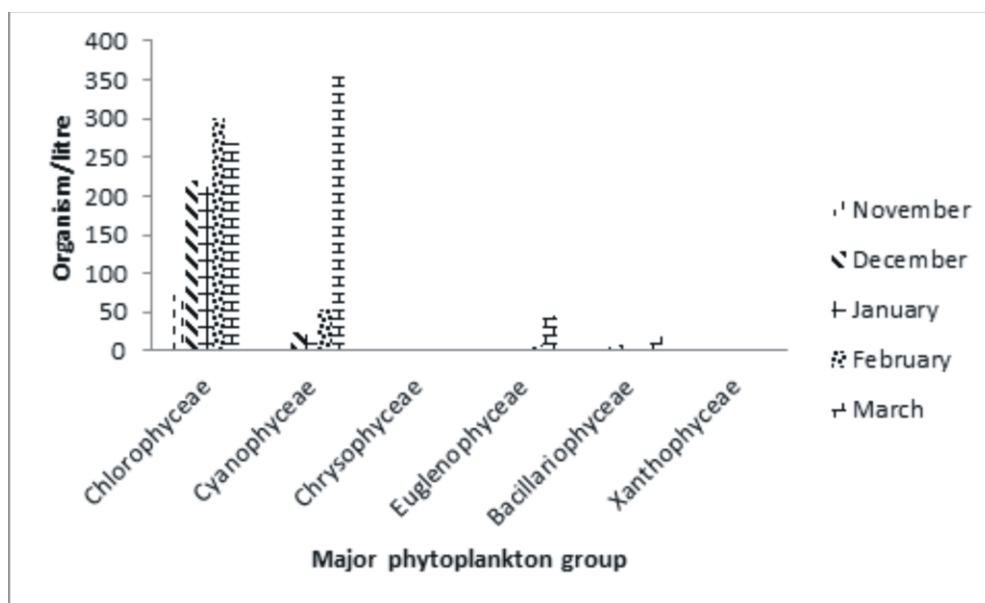


Figure 3: Temporal variation in abundance of major phytoplankton groups of Awba Reservoir

Table 2: Relative abundance of zooplankton organisms of Awba Reservoir, Ibadan, Nigeria

CRUSTACEA	Total Abundance	
	Organism/ mL	%
<i>Bosmina</i> sp.	16000	12.21
<i>Camptocercus</i> sp.	17000	12.98
<i>Daphnia magna</i>	16000	12.21
<i>Simocephalus</i> sp.	1000	0.76
<i>Cyclops</i> sp.	1000	0.76
<i>Cypridopsis</i> sp.	3000	2.29
SUB TOTAL	54000	41.22
PROTOZOA		
<i>Blepharisma</i> sp.	1000	0.76
<i>Chilodonella</i> sp.	6000	4.58
<i>Coleps</i> sp.	3000	2.29
<i>Colpoda</i> sp.	3000	2.29
<i>Frontonia</i> sp.	1000	0.76

<i>Ichthyophthrius</i> sp.	6000	4.58
<i>Lacrymaria</i> sp.	1000	0.76
<i>Prorodon</i> sp.	4000	3.05
<i>Spirostomum</i> sp.	1000	0.76
<i>Urostyla</i> sp.	1000	0.76
SUB TOTAL	26000	19.85
ROTIFERA		
<i>Brachionus urceolaris</i>	2000	1.53
<i>Chromagaster</i> sp.	15000	11.45
<i>Epiphanes</i> sp.	6000	4.58
<i>Euchlanis</i> sp.	3000	2.29
<i>Gastropus</i> sp.	5000	3.82
<i>Keratella</i> sp.	3000	2.29
<i>Notholca</i> sp.	7000	5.34
<i>Ploesoma</i> sp.	1000	0.76
<i>Rotaria</i> sp.	3000	2.29
<i>Syndiaeta</i> sp.	1000	0.76
<i>Testudinella</i> sp.	2000	1.53
<i>Trichocerca</i> sp.	3000	2.29
SUB TOTAL	51000	38.93

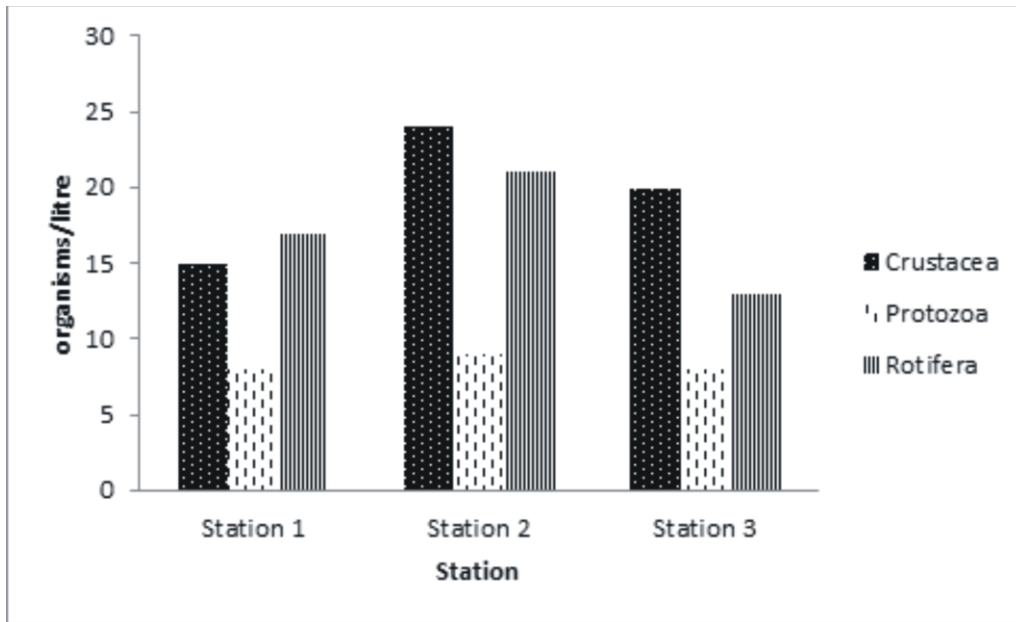


Figure 4: Spatial variation in abundance of major zooplankton groups of Awba Reservoir

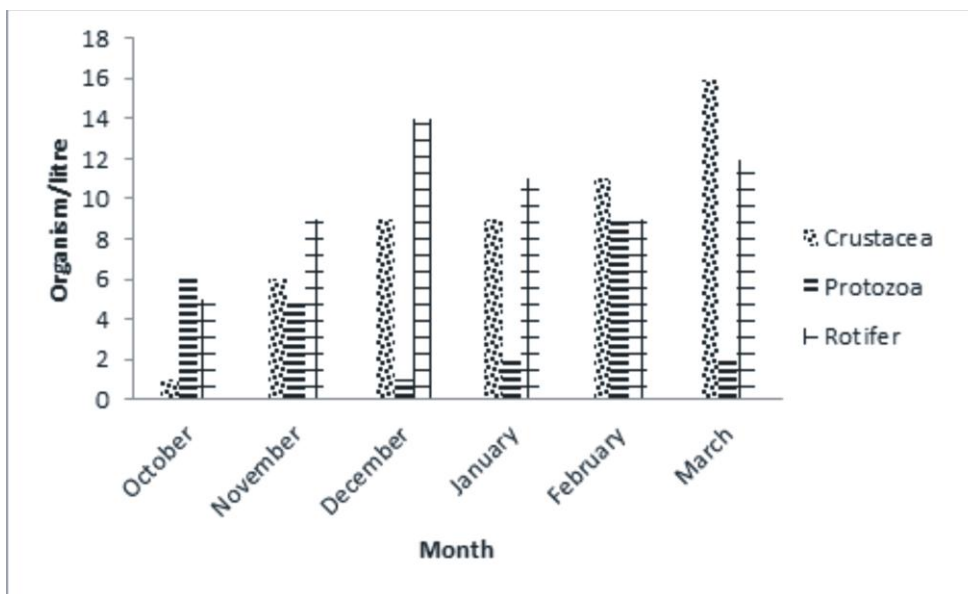


Figure 5: Monthly variation in abundance of major zooplankton groups of Awba Reservoir

Diversity Indices

Monthly variation occurred in diversity indices of phytoplankton of Awba reservoir; Shannon- Wiener index H' ranged from 2.25 to 3.47 being lowest in the early dry season months. Lower values of Margalef's index D and evenness E were also obtained during early dry season months (Figure 6). The diversity indices calculated varied spatially; phytoplanktonic organisms were more diverse in Station 1 with the least evenness, highest evenness occurred in Station 2 (Figure 7). Zooplankton diversity indices obtained were lowest in Station 3; highest D occurred in Station 2 (Figure 8). Sorenson's coefficient of community similarity showed that phytoplankton (0.64 – 0.68) and zooplankton (0.61-0.70) in the stations were alike/identical.



Figure 6: Monthly variation in phytoplankton diversity indices of Awba reservoir

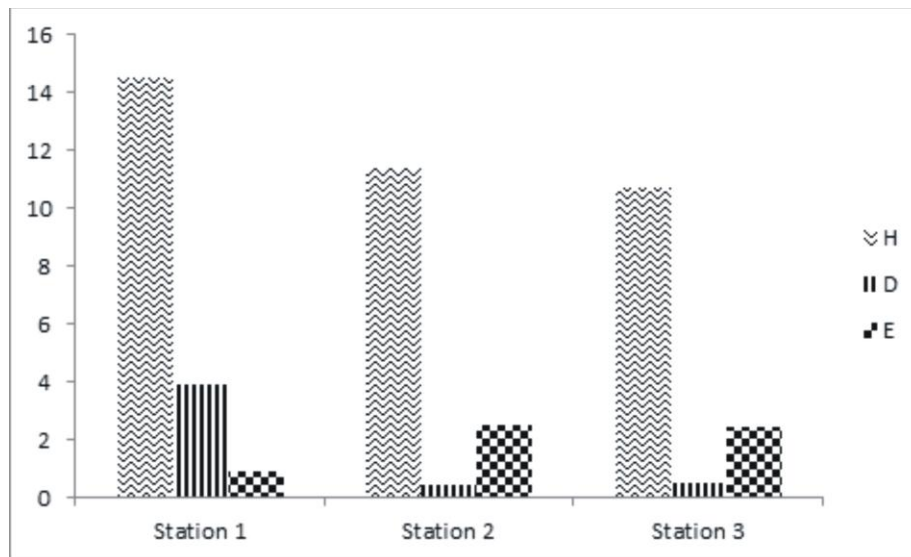


Figure 7: Spatial variation in phytoplankton diversity indices of Awba reservoir

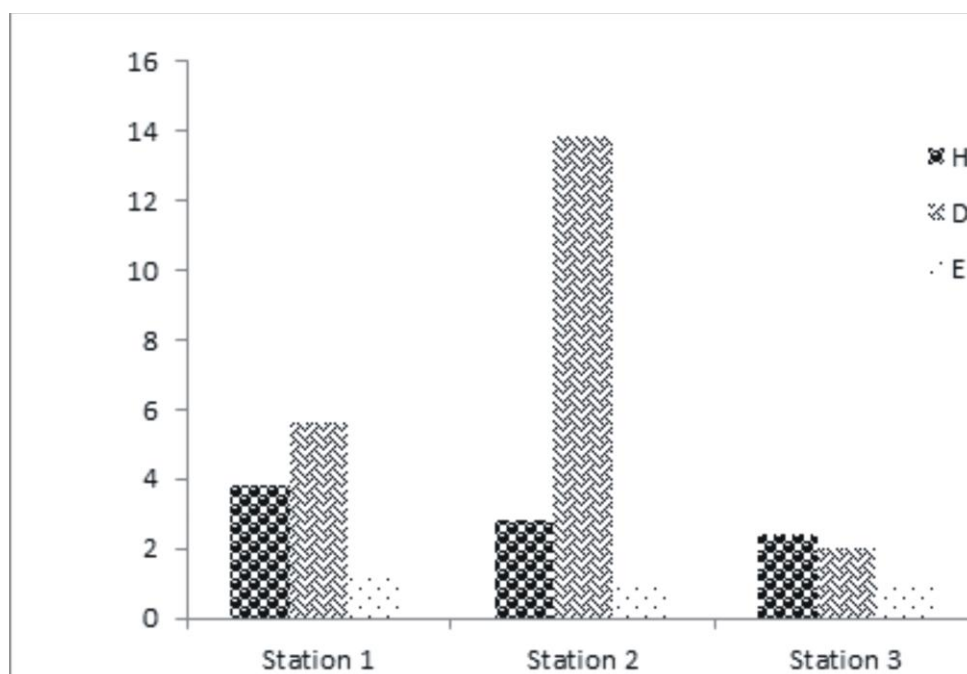


Figure 8: Spatial variation in zooplankton diversity indices of Awba reservoir

Table 3: WHO (1999) guidelines for algal bloom characterization

Acute Health Effects	Cyanobacteria (cells/mL)	Microcystin-LR* ($\mu\text{g/L}$)
Low	< 20,000	< 10
Moderate	20,000 to 100,000	10 to 20
High	100,000 to 10,000,000	20 to 2,000
Very High	> 10,000,000	> 2,000

*Microcystin-LR is commonly used to represent microcystin congeners due to widely available data and known toxicity

Source: Wyoming Department of Environmental Quality Water Quality Division 2018

DISCUSSION

Phytoplankton Composition and Abundance

Non-motile green algae formed a major component of the phytoplankton community of the Awba reservoir with the larger green algae *Pediastrum simplex* and *Coelastrum chodatii* being more abundant. According to **Shubert (2003)**, the non-motile greens are ubiquitous and widely distributed in aquatic habitats throughout the North American continent. Chlorophyceae were also best represented in Lake Awasa, Lake Skadar and Thomas dam (**Kebede and Belay 1994; Rakocevic-Nedovic and Hollert 2005; Ibrahim and Nafi'u 2017**). This is contrary to the report of **Akin-Oriola (2003)** and **Olagbemide (2011)** that blue-green algae dominated the phytoplankton community of

Awba reservoir, although the quantitative differences between the present study and the former may be due to expressing the abundance in organism/mL thus cells in the colonial organism were not counted in this study. Generally there is an increase in total number of taxa encountered in this study compared to previous studies: 14 taxa, **Akin-Oriola (2003)**; 43 taxa, **Olagbemide (2011)** and 49 taxa, **Anago et al. (2013)**. This could be due to difference in methods of collection and sampling stations and also this suggests gradual eutrophication of the Awba reservoir according to **Lepistö and Rosenström (1998)**, phytoplankton diversity in eutrophic waters is the highest compared to other types of trophic waters. Also several dominant species encountered in this study among the Chlorophyceae, Cyanophyceae (*Aphanocapsa*, *Anabaena*, *Microcystis*) and Euglenophyceae (*Euglena acus* Ehrenberg) are characteristic of eutrophic and mesotrophic lakes. The increase in the number of species according to the theory of intermediate disturbances **Reynolds et al. (1993)** was probably due to the slight changes in the ecosystem for example high nutrient concentration as reported by Aderogba and Ayoade (forthcoming).

Microcystis species, the predominant blue-green algae in the Awba reservoir are capable of rapid uptake of phosphate and nitrogen and hence producing large surface blooms and out competing other phytoplankton as asserted by **Xie et al. (2003)**. Under conditions of nutrient enrichment or eutrophication, the blue-greens are known to proliferate and form noxious blooms in freshwater environments (**Reynolds 1984, Stoyneva 2003**). The bloom proportion reached by the *Microcystis flos aqua* and *M. aeruginosa* (Kütz) in the reservoir during the late dry season is supported by the stratified nature of the reservoir (**Ganf 1974; Akin-Oriola 2003**). The formation of blooms also depends on retention time, type and age of water body as well as calm weather conditions with low turbulence of water (Bucka 1989). The development of phytoplankton blooms in eutrophic lakes is attributed to their ability to accommodate reduced nitrogen to phosphorus ratios, low edibility due to their large colony sizes coupled with large herbivore regulation of other taxa (**Barica 1994, Paerl and Tucker 1995**). **Aderogba and Ayoade (forthcoming)** reported low TN/TP ratio and advanced eutrophication of the reservoir.

M. aeruginosa blooms cause several environmental problems, including bad odor, bottom-layer hypoxia and the problem of greatest concern is the production of hepatotoxic cyanotoxins called microcystins by **Harke et al. (2016)**. Cases of human poisoning as demonstrated by **Jochimsen et al. (1998)**, livestock intoxication (**Beasley et al. 1989**), and mass mortality of wildlife (**Miller et al. 2010**) caused by microcystin contamination have been reported. Moreover, recent studies suggest increasing frequency of toxic (microcystin-producing) *M. aeruginosa* blooms in response to climate change as (**Paerl and Otten 2013**). The cyanobacteria densities (479,000 organism / mL) recorded in this study indicate high acute health effects/risk according to **WHO (1999)** guidelines for algal bloom characterization (Table 3). The Wyoming Department of Health (2018) identifies threshold values for microcystin ($\geq 10 \mu\text{g/L}$) and cyanobacteria density ($\geq 20,000$ cells/mL), above which a recreational use advisory will be issued. Subsequently, Wyoming Department of Health will inform the water management agency, and notify local health authorities of the situation and provide information on common cyanobacteria and cyanotoxin related symptoms. Also, Wyoming Department of Environmental Quality (WDEQ) will notify public water supplies with intakes located on

the surface water or downstream of surface water. The excessive abundance or blooming of eutrophic species has detrimental effects on the domestic, industrial and recreational uses of water and is in many cases a direct motivation for restorative measures (**Bryant 1994**). Thus, Awba reservoir may not be suitable for recreation/ecotourism in its present state.

The green algae being less abundant in the late dry season during the period of study could be due to thermal stratification of the water of Awba Lake in dry season (**Akin-Oriola 2003**), and once stratification is stabilized, the non-motile greens begin to sink and decline in the water column, and sedimentation increases. Non-motile greens appear to be restricted to a relatively short growth period defined by a narrow range of environmental conditions within which to successfully compete with a mixed assemblage of phytoplankton (**Happey-Wood 1988**). Thus seasonal succession was observed in the Awba reservoir and cyanophytes dominated in the late dry season.

The variation in the phytoplankton species richness and diversity with sampling points agreed with the findings of **Sekandende et al. (2004)** in the satellite lakes of lake Victoria basin (Tanzania side) and **Eyo et al. (2013)** on the great Kwa River.

Zooplankton Composition and Abundance

The total number of species encountered in the zooplankton community of Awba reservoir in the present study differed in number and composition from the 13 species as given by **Chukwuka and Uka 2007** and five species by **Anago et al. (2013)**. Species composition and abundance of zooplankton communities can be influenced by a number of physical, chemical and biological factors as demonstrated by **Sampajo et al. ((2002)** especially temperature, quality and availability of food, competition and predation. The dominance of cladocerans in the zooplankton community of Awba reservoir during period of study could result from selective feeding by the invertebrate predators including fish on small-sized zooplankton like rotifers. Predation by invertebrates has a greater impact upon microzooplankton than on macrozooplankton, frequently reducing the abundance of the former as asserted **Zaret (1980)**. Predation by fish may affect zooplankton structure, in accordance with the fish feeding mode: selective feeders, by differential capture of organisms, tend to eliminate large species, which are replaced by less vulnerable small forms (**Brooks and Dodson 1965**); filter-feeding planktophage fishes do not actively select their preys and therefore more evasive species avoid predation whereas small forms are captured, thus diminishing zooplankton densities (**Drenner et al. 1982**). Cladocerans were also dominant in Nigeen lake and Keenjhar lake (**Jan et al. 2015; Rao and Azmi 2019**), and attributed to temperature enhancing rapid hatching of eggs, high nutrient conditions and food availability as suggested by **Pandit (1989)**. However, **Chukwuka and Uka (2007)** reported rotifers as the dominant group and **Anago et al. (2013)** encountered the copepod *Thermocyclops* as the most abundant. Cladocerans have been claimed to be good indicators of trophic state in lentic ecosystems. In Europe, the size range of species has been used as an indicator of water quality. According to **Gannon and Stemberger (1978)**, species of *Bosmina* are good indicators of lake trophic state. Species such as *Bosmina longirostris* (O. E. Müller), having a great ability to utilize colonial cyanophyceae as food, exhibit a greater tolerance to their blooms as suggested by **Fulton and Paerl (1987)**, so that they become abundant

in such conditions and may be considered bioindicators of eutrophication (**Matsumura-Tundisi 1999**). Thus, the predominance of *Bosmina* and *Camptocercus* sp. in the reservoir suggests they feed on the colonial cyanophyceae and chlorophyceae that were dominant in the phytoplankton community and this further confirmed the eutrophic state of the reservoir. *Bosmina longirostris* has been observed in eutrophic environments such as Barra Bonita Reservoir (**Matsumura-Tundisi 1999**) and Billings Reservoir complex (**Sendacz & Kubo 1999**) both in Brazil, however, **Güntzel (2000)** observed that among the six reservoirs on Tietê River, *Bosmina hagmanni* Stingelin, 1904 was most abundant in the less eutrophic ones.

Temporal variation and succession in zooplankton species was observed in the reservoir. Cladocerans reached peak in late dry season months coincided with bloom of *Microcystis aeruginosa* and *M. flos aqua* that could serve as food source. According to **Campbell and Haase (1981)**, quality and quantity of food can alter species composition as well as the abundance of the species, since particular organisms are highly selective about the size and the type of phytoplankton they eat.

The frequently encountered protozoan species (*Chilodonella* spp., *Ichthyophthirius*) in the Awba reservoir could cause fish diseases. Protozoans exhibit rapid and exponential reproductive strategies (e.g. *Chilodonella* spp. and versatile, resilient life stages e.g. *Ichthyophthirius multifiliis* Fouquet which have allowed parasitic protozoans to colonise aquatic environments globally). Among fish protozoans, *Ichthyophthirius* and *Trichodina* are two of the most predominant genera globally (**Lom and Dyková 1992**). *Ichthyophthirius multifiliis* is one of the most contagious ciliophoran parasites of fishes (**Matthews 2005; Dickerson 2006**). This parasite accounts for significant economic losses in aquaculture, the ornamental fish trade and epidemics in wild fish populations, resulting in mass mortalities (**Matthews 2005**). The ciliated protozoan *Ichthyophthirius multifiliis* infects several species of freshwater fish worldwide (**Dickerson and Findly 2014**). It causes high mortality associated with ichthyophthiriasis (“white spot disease”) in farmed fish, while low-level infections occur in wild fish.

The genus *Chilodonella* includes free-living ciliated protozoa as well as pathogenic species for freshwater species, with *Chilodonella hexasticha* (**Kiernik, 1909**) and *Chilodonella piscicola* (**Zacharias, 1894**) being the most important ones. These parasites cause outbreaks with high mortalities among farmed freshwater fishes with great economic losses as demonstrated by **Li et al. (2018)**. Heavy infection resulted in emaciation and mass mortalities. Infection in the gills caused severe degeneration, necrosis and consequent degradation of the branchial epithelium and occlusion of the capillaries. Infection also induced massive proliferation of chloride and mucus cells and also caused hyperplasia of the lining filamental epithelium. **Paperna and Van As (1983)** reported pathological and histopathological changes induced by infection of *Chilodonella hexasticha* (Kiernik) in wild and cultured cichlid fishes from Israel and South Africa.

In conclusion, the phytoplankton and zooplankton community of the Awba reservoir have increased species composition and abundance during the study period compared to previous studies. The abundance/bloom of *Microcystis flos aqua* further confirmed the advanced eutrophication of the reservoir.

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