

## ALGAL FLORA OF AIN HELWAN I. ALGAE OF THE WORM SPRING

Sanaa M. M. Shanab

*Botany department, Faculty of science, Cairo University*

### **Abstract**

Seasonal water samples were collected during the period of April 2004 to January 2005 and physico-chemical analyses were performed for each sample. Isolation and identification of the different algal species were carried out. Concerning the physical water characteristics, temperature of the water samples, during the period of study, ranged 26 - 30 °C, pH values ranged 6.9 – 7.78 and light intensities, received by the surface of shallow water of the spring, ranged 26.4-115 Klux. On the other hand the values of chemical water characteristics (anions, cations, micronutrients) were seasonally fluctuated and no detection of cobalt and cadmium were recorded. The total identified algal flora in the water samples of the four seasons recorded 209 species and 116 genera belonging to 8 algal divisions; Cyanobacteria, Chlorophyta, Bacillariophyta, Chrysophyta, Xanthophyta, Euglenophyta, Dinophyta and Charophyta. Cyanobacteria were the maximally represented division (91 species and 29 genera) during the period of study, followed by Chlorophyta (59 species and 45 genera), Bacillariophyta (52 species and 35 genera), Chrysophyta and Euglenophyta were represented each by (2 species and 2 genera). While, the least representative algal divisions were Charophyta, Dinophyta and Xanthophyta (1 species and 1 genera). The largest algal biomass was recorded in winter sample (138 species and 92 genera) followed in descending order by autumn (87 species and 56 genera), summer (71 species and 45 genera) and spring samples (48 species and 25 genera). It seemed that the diversity of algal flora in water samples was mainly controlled by and significantly correlated with some physico-chemical characteristics of water samples.

**Key Words:** Algal flora, Ain Helwan, Physico-chemical analyses, Worm Spring.

### **Introduction**

Many researches have been reported in different parts of the world concerning thermal springs as in Czechoslovakia, Greece, Italy, India, Japan, Pakistan, Spain, Saudia Arabia, South Africa, USA and USSR (Kullberg, 1971; Ali *et al*, 1983; Nikitina, 1983; Foged, 1984; Schoeman and Archibald, 1988; Arif, 1989; Seoane, 1990; Jha and Kumar, 1990). Egypt has a fair number of thermal springs distributed in Cairo, Suez, Sinai and Western desert. A lot of geochemical, hydrogeological studies (El Ramly, 1969; Issar *et al*, 1971; Swanberg *et al.*, 1976 and 1984; El Kiki *et al.*, 1978; Abdou, 1994) and phycological researches (Shaaban and El Habibi, 1978; Shaaban, 1985; Abou-El kheir and Ismail, 1986; Abou-El kheir and Mekky, 1986; Hamed 1995) were done dealing with separate and or combind comparative studies on the different springs and recording their algal communities.

The algal flora of Ain Helwan was partly studied by Abou-El Kheir and Mekky, 1986 (identified 34 taxa belonging to Bacillariophyta (25 species), Cyanobacteria (5 species) and Chlorophyta (4 species), Hamed, 1995 (compared the algal flora of 10 Egyptian thermal springs), and El-Gamal and Salah El-Din,

1999 identified a new species from Ain Helwan spring (*Compsopogon helwanii*). These studies concluded that each spring in Egypt has its own chemical characteristics and algal flora.

From the historical point of view, Ain Helwan spring is a permanent spring which started flowing in 1939 during the construction of the Cairo-Helwan railway and still flowing till now. This spring and many old and new sulphur springs ( $\approx 8$ ) lie in an area east of the Nile River and 25 km South of Cairo between El-Maasara in the North and El-Tabbin in the south.

Water temperature range (26-30 °C) did not show large changes with time which indicates that these springs receives most of their water from great depth (originate from the Nubian sandstone aquifer and water moves upward through a thick fractured limestone strata along fault planes affecting the area, feeding these springs with their water). Chemical analysis of spring's water, from previous studies, (Abdou, 1994, Hamed 1995), showed that it contained large amounts of dissolved salts (considered as brackish water as it contained 0.4 % NaCl), small amounts of metals (Cu, Zn and Al), and radioactivity within the permissible dose. Based on the physico-chemical properties, Helwan Springs have long been used for natural mud therapy and its water is useful in treatment of chronic articular disease and certain skin disorders (Abdou, 1994).

The studies conducted on the water quality and floral composition of Ain Helwan hot spring over many years have resulted in questioning whether there are changes in algal species composition due to change in water quality of the spring. Accordingly this work was designed to investigate the seasonal changes in population of different thermophilic algal species inhabiting the water of the spring.

### ***Material and Methods***

The worm spring of Ain Helwan (Figure 1), can be divided into three sites, the original site (1) surrounded by hexagonal cement building of 5 meters height and 3.5 meters in diameter, (its temp. was 30 °C) Figure (2a), then the water of the spring pass through an underground passage ( $\approx 100$  meters long) then it pass upward on the ground surface through an exposed granite part (temp. 28 °C, it termed site 2), which is about 1 meter large and high and of more than 100 meters long. The first 15 meters of the passage was more covered with fixed slimy colored algal growths (grey filaments, dark blue-green and brown patches and spots), as seen in Figure (2b and c). The granite part, leads to the second exposed shallow, narrow and muddy passage of the spring (called site 3). It is of 0.5 meter large and about 0.3-0.5 meter deep in some parts of its passage and more than 100 meters long. Its temperature was 26 °C and it was inhabited by dense algal growths (tufts and aggregation of long green filaments were obviously seen on water surface together with epipellic blue green and/or brown growths) Figure (2d). The worm spring is terminated by a shallow lake-like part (160 m<sup>2</sup>) where many children were swimming since four years ago. But nowadays, this lake was inhabited by dense hydrophytic plants, (Figure 2e and f) such as

*Phragmites* sp. (+++), *Juncus subulatus* Forssk (+), *Typha domingensis* Pers. (+++), *Cyperus laevigatus* L. (++) and *Cynodon* sp. (++)

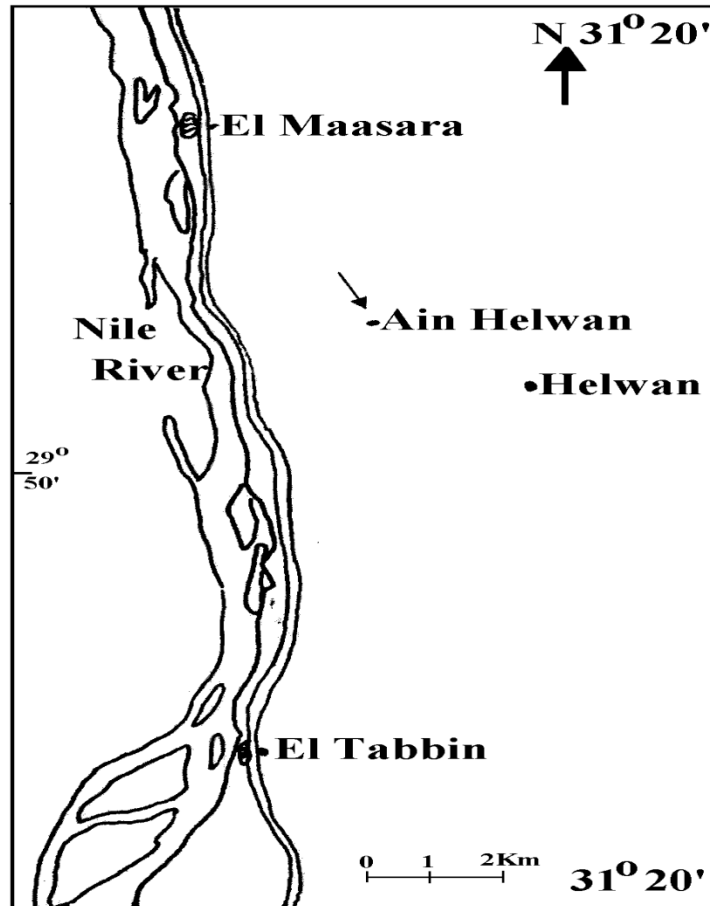
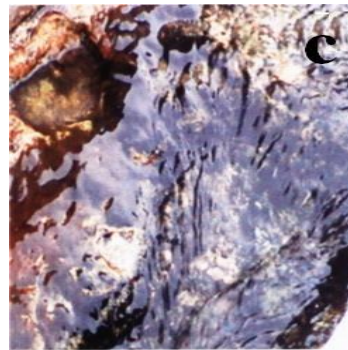


Figure (1): Location of Ain Helwan Spring From (Abdou, 1994)

To study the different characteristics of the warm spring of Ain Helwan and its algal flora, samples were collected from the three sites of the spring and physico-chemical analysis were performed as well as microscopic examination of algal taxa and their identification. No algal presence in water sample from site (1), while the exposed site (2) have always fixed slimy unchangeable algal flora on the walls and bottom of the passage. Samples from these algae were collected by scratching with a knife and Table (2) showed the identified algal flora of site (2) of the spring. The exposed muddy site (3) have free living and epipellic algal flora, which was found by continuous sample examination, to undergo different seasonal changes. So, seasonal water samples were collected from site (3) of the spring to study the seasonal variations in the algal flora as affected by both physical and chemical water characteristics.



a- The original site with cement building surrounding the spring



b and c- The 1<sup>st</sup> exposed granit site



d- The 2<sup>nd</sup> exposed muddy site

e and f- The terminal lake-like inhabited with different hydrophytes

Figure (2): The Warm Spring of Ain Helwan

### 1. Collection of water samples from the three sites of the spring

Water samples were collected from the three sites of Ain Helwan according to the method of Bourrelly (1970); its physico-chemical characteristics and algal identification were performed.

### 2. Collection of Seasonal water samples (from site 3).

Water samples from site (3) of the spring were seasonally collected, in 1L glass-bottles at approximately the same morning time of each season [spring sample was collected at 14 April 2004, summer sample at 14 July 2004, autumn sample at 14 October 2004, winter sample at 14 January 2005 and were given the signs L1, L2, L3 and L4, respectively]. Each water sample was divided into three parts:

The 1<sup>st</sup> part was preserved in 4% formalin and used for the identification of the algal species.

The 2<sup>nd</sup> part was kept in the incubator at  $20 \pm 1^\circ \text{C}$  under continuous illumination of  $30 \mu\text{E} / \text{m}^2 / \text{s}$  (using cool white fluorescent lamps).

The 3<sup>rd</sup> part was used for the determination of the physico-chemical analyses.

### 3. Physical parameters

The following physical analyses were carried out:

- a- pH; *in situ* by pH meter (ORION) model 230 A portable pH / ISE Meter Instruction Manual.
- b- Light intensities were measured *in situ* using LT Lutron Lx-101 digital Lux Meter Operation Manual Digital Lux Meter.
- c- Temperatures were measured *in situ* using thermometer.

### 4. Chemical analyses

Different methods of chemical analyses were performed for water samples. Soluble anions, cations, chemical oxygen demand (COD), biological oxygen demand (BOD), macro- and micro-nutrients and heavy metals (Cu, Ni, Zn, Co, Cd and Pb) were also determined according to Doeck and Tracy (1956), Boyed (1984), and APHA (1985).

### 5. Algal identification and Photography

Identification of the algal species was carried out according to Desickachary (1959), Bourrelly (1968, 1970 and 1972), Prescott (1978 and 1982), Foged (1980) and Cox (1996). The algal species were sketched and photographically recorded using Nikon Labo-Photo (AFX-II) microscope combined with camera (Nikon Fx-35, Japan). Most photos were magnified by (x 400) except otherwise cited in the plates.

## Results and Discussion

The Physico-chemical characteristics of water samples of the worm spring (Table 1) revealed that the covered (control) original site (1) and the two exposed sites (2, 3) have almost similar values of total soluble salts (TSS), pH, soluble anions, BOD and cations (except a slight increase in  $\text{Mg}^{2+}$  and decrease in  $\text{Na}^+$  of water sample in site 3) while variations in temperatures and COD values were recorded.

COD increased markedly in sites (2 and 3) and this increase was much more pronounced in site (2). The temperature of the three sites varied and fluctuated between 30, 28 and 26 °C in sites (1, 2 and 3), respectively. Phosphorus decreased in site (3) and increased in site (2) as compared with its value in site (1). Iron value was slightly decreased in site (3) while it was completely absent in site (2). It is worthy to mention that the data of copper content revealed its absence in sites 1 and 2, while it is unexpectedly present in site (3).

**Table (1): Physico-chemical analyses of water samples from sites (1, 2 and 3) of Ain Helwan worm spring.**

Chemical Analysis	Water Samples		
	Site(1)	Site (2)	Site (3)
Total soluble salts ppm	4.90	4.85	4.90
EC as mM / mL	3136	3104	3136
pH	6.84	7.00	7.42
Water temperature (°C)	30	28	26
<b>Soluble Anions (mg/L)</b>			
Carbonate CO <sub>3</sub> <sup>2-</sup>	—	—	—
Bicarbonate HCO <sub>3</sub> <sup>-</sup>	4.50	4.40	4.50
Chloride Cl <sup>-</sup>	43.26	44.03	43.45
Sulfates SO <sub>4</sub> <sup>2-</sup>	3.36	3.65	2.94
<b>Soluble Cations (mg/L)</b>			
Ca <sup>++</sup>	10.50	10.39	9.95
Mg <sup>++</sup>	9.40	9.44	10.77
Na <sup>+</sup>	30.87	31.92	29.82
K <sup>+</sup>	13.65	12.87	13.65
COD (mg / L)	29.00	49.00	38.00
BOD (mg / L)	9.00	9.00	9.00
<b>Macro- and Microelements (mg/L)</b>			
N <sup>+++</sup> (%)	0.06	0.05	0.05
P <sup>+++</sup>	36.50	41.40	31.30
Fe <sup>++</sup>	52.30	—	41.85
Mn <sup>++</sup>	0.95	0.25	0.75
Zn <sup>++</sup>	—	—	—
Cu <sup>++</sup>	—	—	0.102

(Samples were collected in winter where the air temperature was 15 °C).

Microscopic examination of the water sample from the three sites of the spring of Ain Helwan showed no algal presence in the covered original site (1), which might be mainly due to the unavailability of natural light penetration, a cause of the hexagonal cement building which surround the origin of the spring (inspite of the suitability of other physical and chemical characteristics of this water for algal growth). A scratched sample from the exposed granite site 2 showed an aggregation of gray filaments fixed on the walls and identified as

*Lyngbya birgie* together with other Cyanobacteria, Chlorophyta and Bacillariophyta species, which were mainly present on the bottom of the passage and recorded in Table (2). These algal species might be thermophilic and pollution tolerant species as site 2 receive always pollutants (as detergents and others) from the visitors who use the water of this place and the polluted water pass to site (3) altering some of its chemical characteristics (phosphorus, iron, copper and COD contents) and consequently its algal flora.

**Table (2): Algal Flora of the exposed granite place (2) of the warm Spring of Ain Helwan (all over the year).**

N <sup>o</sup>	Cyanobacteria	Bacillariophyta	Chlorophyta
1	<i>Chroococcus minor</i> (Kütz.) Naegell	<i>Amphora clevei</i> Grun.	<i>Cerasterias staurastroides</i> West & West
2	<i>Isocystis messanensis</i> Borzi	<i>Cymbella cistula</i> (Hempr.) Kirvh.	<i>Enteromorpha compressa</i> (Linnaeus) Nees
3	<i>Lyngbya birgie</i> G.M. Smith	<i>Denticula tenuis</i> Kuetz.	<i>Mougeotia scalaris</i> Hassal
4	<i>L. erugineo-coerulea</i> (Kütz.) Gommont	<i>Fragilaria capucina</i> Desmazieres	<i>Pseudopleurococcus printzii</i> Vischer
5	<i>L. tenue</i> Gommont	<i>Gyrosigma acuminatum</i> (Kuetz.) Rabenhorst	-
6	<i>L. perelegans</i> Lemmermann	<i>Navicula angusta</i> Grunow	-
7	<i>Microcoleus Lacustris</i> (Rab.) Forlow	<i>Nitzschia linearis</i> (C.A. Agardh) W. Smith	-
8	<i>Myxosarcina spectabilis</i> Geitler	<i>N. frustulum</i> (Kütz.) Grunow	-
9	<i>O. acuminata</i> Gommont	<i>N. obtuse</i> W. Smith	-
10	<i>O. limnetica</i> Lemmermann	<i>Stauroneis parvula</i> var. proninula Grunow	-
11	<i>O. tenuis</i> var. tergestina Rabenhorst	<i>Surirella oblonga</i> Her.	-
12	<i>O. rubescens</i> De Cand	-	-
13	<i>Synechocystis aquatilis</i> Sauvageau	-	-
Total species	13	11	4
Total genera	7	9	4

Cyanobacteria was represented by (7 genera and 13 species), Chlorophyta by (4 genera and 4 species) and Bacillariophyta by (9 genera and 11 species). Algal flora of this exposed granite site (2) of the spring was found to be extremely stable all over the year and could be controlled mainly by the worm temperature of the water. These were in conformity with the findings of Lund (1965) and Sompong *et al.* (2005).

The second exposed muddy passage (site 3) of the spring have different algal flora which are affected by seasonal physico-chemical changes of the water. On the contrary to sites (1 and 2), site (3) showed seasonally different algal flora. So this study was directed and concentrated on the seasonal algal growths in

water of passage (3) as affected by the seasonal physico-chemical characteristics of water.

The limited variations in seasonal temperatures (26-30 °C) recorded in this study (Table 3), and confirmed by the previous investigations of Abdou (1994) and Hamed (1995), indicated that, this spring (considered worm according to Meinzer, 1923) receives its water from great depths. Any increase or decrease in algal populations seemed to be correlated with fluctuations in water temperatures. This was in accordance with the results obtained by Lund (1965), Thompson and Guo (1992) and Kobbia *et al.* (1995).

**Table (3): Seasonal physico-chemical analyses of water samples (site 3) of Ain Helwan during 2004 –2005.**

Parameters		Water Samples			
		L1 (Spring)	L2 (Summer)	L3 (Autumn)	L4 (Winter)
Physical Analyses	Temperature (°C)	28.5	30	27.5	26
	pH	7.28	7.68	7.78	6.92
	Light intensity (klux)	60	115	46.3	26.4
Chemical Analyses	<b>Anions (mg / mL)</b>				
	PO <sub>4</sub> -P	0.00003	0.093	0.145	0.072
	NH <sub>4</sub> -N	0.047	0.009	0.0005	0.045
	NO <sub>3</sub> -N	0.023	0.0005	0.002	0.006
	SiO <sub>3</sub> <sup>-2</sup>	0.098	0.111	0.082	0.048
	Cl	0.035	0.060	0.050	0.040
	CO <sub>3</sub> <sup>-</sup>	2.4	1.8	5.1	6.0
	HCO <sub>3</sub> <sup>-</sup>	9.0	11.4	8.4	6.0
	<b>Heavy metals (µg / mL)</b>				
	Zn	0.02	0.02	0.04	0.04
	Pb	0.02	0.09	0.132	0.010
	Cu	0.03	0.120	0.101	0.16
	Ni	0.146	0.00	0.00	0.392
	Co	0.00	0.00	0.00	0.00
Cd	0.00	0.00	0.00	0.00	

The recorded pH values of the water in this study were mostly shifted to the alkaline side (6.92 – 7.78). This tendency could be mainly due to the higher activity of photosynthetic processes of the dense algal populations which were controlled by the seasonal water characteristics. Such assumption seemed to be in conformity with the findings of Kobbia *et al.* (1993).

The fluctuated seasonal light intensities (26.4- 115 Klux), received by the water surface of the shallow Spring during different light/dark cycles and the recorded algal flora indicated that light and temperature of the Spring water could be the most controlling physical factors affecting algal productivity. These results



were in harmony with the results obtained by Carvalho and Malcata (2003); Sompong *et al.* (2005) and Abrontes *et al.* (2006) who emphasized the predominant role of water temperature and light in phytoplankton succession.

According to the chemical characteristics, water of Ain Helwan Spring is classified as brackish water habitat contained 0.4 % NaCl as reported by Hamed (1995) and this was confirmed by the higher abundance of mesohalobien algal species [*Enteromorpha compressa* (Green), *Oscillatoria tenuis* (Cyanobacteria) and *Nitzschia palea* (Diatom)]. The filamentous green alga, *E. compressa* was found persisted in all seasonal water samples, and it was normally found in marine habitats, confirmed the brackish nature of the spring water which stimulated the growth of certain  $\alpha$ -mesohalobien species (Hamed, 1995).

The total identified algae/year were 209 species and 116 genera belonging to 8 algal divisions; Cyanobacteria, Chlorophyta, Bacillariophyta, Chrysophyta, Euglenophyta, Xanthophyta, Charophyta and Dinophyta.

Winter samples recorded the maximum algal flora followed in descending order by autumn, summer and spring ones as recorded in Table 4 and 5 ( $L4 > L3 > L2 > L1$ ). These results were in confirmity with the findings of Reederers *et al.* (1998); Ali (2000); Gaballah *et al.* (2000) and Domingues *et al.* (2005) who reported the importance of light and micronutrient factors influencing phytoplankton distribution.

Cyanobacteria dominated by 91 species and 29 genera, which represented 43.54% and 25 % of the total recorded algae, respectively and as illustrated in Plates (I and II) and Tables (4 and 5). The prokaryotic Cyanobacterial cells were characterized by its tolerance and adaptability to grow under wide range of seasonal physico-chemical water variabilities. This was obviously noticed in Cyanobacterial dominance all over the year (Table 5). These results were in accordance with the results obtained by Kobbia *et al.* (1993).

Chlorophyta Plate (III and IV) with 59 species and 45 genera represented 28.23 % and 38.79 % subdominated Cyanobacteria not only in total algal mass production/year but also in seasonal samples (except for winter sample where diatoms subdominated the Cyanobacteria). Higher temperature and light intensities in summer and spring seasons were inhibitory to members of Chlorophyta, Bacillariophyta and those of other divisions.

Bacillariophyta followed Chlorophyta was represented by 52 species and 35 genera of 24.88 % and 30.17 % as shown in Plate (V) and Tables (4 and 5). Diatoms flourished in winter and subdominated Cyanobacteria in this season.

This was confirmed by the marked decrease in silicate content in winter. Pennate forms always predominated the centric forms; this oligotrophic tendency of pennate forms was also obtained by Kobbia *et al.* (1990 and 1995). Abundance of mesohalobien diatoms (*Synedra* and *Nitzschia*) were recorded together with some oligohalobien ones (*Amphora* and *Navicula*).

Division Euglenophyta, Chrysophyta, Dinophyta, Charophyta and Xanthophyta were very poorly presented in the spring seasonal water samples

during the period of study. They were found during autumn and winter samples which were considered the most favourable seasons for algal growths not only for the three main algal divisions (Cyanobacteria, Chlorophyta and Bacillariophyta) constituting the water algal community but also to members of other divisions.

**Table (4): Seasonal algal composition of the water samples of Ain Helwan Spring during 2004-2005.**

Algal Species	Seasonal Water Samples			
	L1 (Spring)	L2 (Summer)	L3 (Autumn)	L4 (Winter)
<b>Cyanobacteria</b>				
<i>Albrightia tortusa</i> Copeland	—	—	—	+
<i>Anabaena viguieri</i> Denis and Fremey	—	—	—	+
<i>Anabaena spiroides</i> Klebahn	+	—	—	+
<i>Anabaena spiroides</i> var. <i>crassa</i> Lemmermann	+	—	—	+
<i>Anabaena helicoidea</i> Bernard	+	—	—	+
<i>Anabaena iyengari</i> Bharad	+	—	—	—
<i>Anabaena affinis</i> Lemmermann	+	—	—	—
<i>Anabaena wisconsinense</i> Prescott	+	—	—	—
<i>Aphanothece castagnei</i> (Bréb.) Rabenhorst #	+	+	+	+
<i>Aphanothece elabens</i> (Bréb.) Elenkin	—	—	+	+
<i>Aphanothece caldariorum</i> Richter #	+	+	+	+
<i>Aphanothece elachista</i> (West and West) Starmach	—	+	+	+
<i>Aphanothece nidulans</i> Richter	—	—	—	+
<i>Aphanocapsa delicatissima</i> West and West	—	+	—	—
<i>Borzia trilocularis</i> Cohn	—	+	—	—
<i>Chroococcus minor</i> (Kütz.) Naegeli	—	+	—	+
<i>Chroococcus turgidus</i> (Kütz.) Naegeli	+	—	—	—
<i>Cyanareus hamiformis</i> Pascher	—	—	+	—
<i>Cyanoptyche gloeocystis</i> Pascher fo. <i>minor</i> Starmach	—	+	—	+
<i>Dactylococcopsis acicularis</i> Lemmermann #	+	+	+	+
<i>Dactylococcopsis fascicularis</i> Lemmermann #	+	+	+	+
<i>Gomphosphaeria aponina</i> Kuetzing	—	—	—	+
<i>Lithococcus schizodicwarmorum</i> Copeland	—	+	+	+
<i>Lyngbya valderianum</i> Gomont	+	+	—	+
<i>Lyngbya tenue</i> Gomont	+	+	—	+
<i>Lyngbya turidum</i> Gomont	—	+	—	+
<i>Lyngbya lucidum</i> Kütz.	—	+	+	+
<i>Lyngbya contorta</i> Lemmermann	—	—	—	+
<i>Lyngbya lagerheimii</i> (Moebius) Gomont	—	—	+	+
<i>Lyngbya perelegans</i> Lemmermann #	+	+	+	+
<i>Lyngbya aerugineo-caerulea</i> (Kuetzing) Gomont	—	—	+	+
<i>Lyngbya bipunctata</i> Lemmermann #	+	+	+	+
<i>Lyngbya spirulinoides</i> Gomont	+	+	—	—
<i>Lyngbya tylorii</i> Drouet and Strickland	—	—	+	+
<i>Lyngbya birgei</i> G.M. Smith	+	+	+	—
<i>Lyngbya latissima</i> Prescott	—	—	+	—
<i>Lyngbya limnetica</i> Lemmermann	—	—	+	—
<i>Lyngbya putealis</i> Montagne	+	+	—	—
<i>Lyngbya autumnale</i> (Agardh) Gomont	+	—	—	—
<i>Mastigocoleopsis obtuse</i> (Carter) Geitler	—	—	—	+
<i>Microcystis aeruginosa</i> Kütz.	+	+	—	—
<i>Microcystis elachista</i> (West and West) Starmach #	+	+	+	+

Algal Flora of Ain Helwan I. Algae of The Worm Spring

<i>Microcystis viridis</i> (A. Braun) Lemmermann	—	—	+	—
<i>Microcoleus vaginatus</i> (Vaucher) Gomont	—	+	+	+
<i>Microcoleus acutissimus</i> Gardner	—	+	—	—
<i>Myxosarcina dubia</i> Ercegovic	—	—	+	+
<i>Myxosarcina spectabilis</i> Geitler and Ruttner	—	—	—	+
<i>Nodularia spumigena</i> Mertens	—	—	—	+
<i>Nodularia implexa</i> (Bornet and Flah) Nordstedt and Wittrock	+	—	—	—
<i>Nodularia harveyana</i> Thuret	+	—	—	—
<i>Nostoc piscinale</i> Kütz.	—	+	—	—
<i>Nostoc paludosum</i> Kütz.	+	—	—	+
<i>Nostoc linckia</i> (Roth) Bornet and Flah	+	—	—	+
<i>Nostoc carneum</i> C.A. Agardh	—	—	—	+
<i>Nostoc parmelioides</i> Kütz.	+	—	—	—
<i>Oscillatoria platensis</i> (Nordstedt) Geitler	—	—	—	+
<i>Oscillatoria tenuis</i> C.A. Agardh	—	—	+	+
<i>Oscillatoria limnetica</i> Lemmerman	+	—	—	—
<i>Oscillatoria rubescens</i> De Cand.	—	—	+	—
<i>Oscillatoria limosa</i> (Roth) C.A. Agardh.	—	—	+	—
<i>Oscillatoria sancta</i> (Kütz.) Gomont	—	—	+	—
<i>Oscillatoria chalybea</i> Mertens	—	—	+	—
<i>Oscillatoria tenuis</i> var. <i>tergestina</i> (Kütz.) Rabenhorst	—	—	+	—
<i>Oscillatoria hamelii</i> Frémeu	—	—	+	—
<i>Oscillatoria okeni</i> Agardh	—	+	—	—
<i>Oscillatoria acuminata</i> Gomont	—	+	—	+
<i>Oscillatoria margaritifera</i> Kütz.	—	+	—	—
<i>Oscillatoria lacustris</i> (Klebahn) Geitler	—	+	+	—
<i>Phormidium retzii</i> (C.A. Agardh) Gomont	—	—	+	—
<i>Phormidium ambiguum</i> Gomont	—	+	+	+
<i>Phormidium mucicola</i> Naumann and Huber Pestalozzi	—	—	+	—
<i>Plectonema nostocorum</i> Bornet #	+	+	+	+
<i>Plectonema purpureum</i> Gomont	—	+	—	+
<i>Plectonema malayense</i> Biswas	+	—	+	—
<i>Plectonema notatum</i> Schmidle #	+	+	+	+
<i>Plectonema wollei</i> Farlow	—	+	—	—
<i>Pseudanabaena catenata</i> Lauterb.	+	—	—	—
<i>Rhodostichus expansus</i> Geitler and Pascher	+	+	—	—
<i>Schizothrix lacustris</i> A. Braun	—	—	—	+
<i>Schizothrix muelleri</i> Naegeli	—	—	—	+
<i>Spirulina laxa</i> G.M.Smith	—	+	—	—
<i>Spirulina major</i> Kütz.	—	—	—	+
<i>Spirulina subsalsa</i> Oersted	—	+	+	+
<i>Synechococcus vantioghiemi</i> Pringsheim	—	+	+	+
<i>Synechococcus nidulans</i> (Pringsheim) Komarek	—	—	—	+
<i>Synechococcus cedrorum</i> Sauvageau	—	+	—	—
<i>Synechocystis aquatilis</i> Sauvageau #	+	+	+	+
<i>Stigonema turfaceum</i> (Berkeley) Cooke	—	—	—	+
<i>Stigonema mamillosum</i> (Lyngb) C.A. Agardh	—	—	—	+
<i>Stigonema minutum</i> (Agardh) Hass.	+	—	—	+
<i>Stigonema mesentericum</i> Geitler	+	—	—	—
<i>Tubiella elenkini</i> Hollerbach.	+	—	—	+
<b>Chlorophyta</b>				
<i>Actinastrum hantzschii</i> Lagerheim	—	+	—	—
<i>Ankistrodesmus fractus</i> (West and West) Brunn	+	—	—	—
<i>Ankistrodesmus braunii</i> (Naegeli) Brunn	—	—	+	—
<i>Apatococcus lobatus</i> (Chodat) Boye Petersen	—	+	+	—

<i>Bracteacoccus minor</i> (Chodat) Petrova	—	—	—	+
<i>Cerasterias irregularis</i> G.M. Smith	—	—	+	+
<i>Chaetophora elegans</i> (Roth.) C.A. Agardh	—	—	+	+
<i>Chlorococcum humicola</i> (Naegeli) Rabenhorst	—	—	+	+
<i>Chlorococcum wimmeri</i> Rabenhorst emend Starr	—	—	+	+
<i>Chlorella vulgaris</i> Beyerinck	—	+	+	+
<i>Chloronomala palmelloides</i> Mitra	—	—	—	+
<i>Chlorosarcina consociata</i> (Klebs) G. M. Smith	—	—	—	+
<i>Closterium lumuta</i> (Müller) Nitzsch	—	+	—	—
<i>Coelastrum cambricum</i> Archer	—	+	—	+
<i>Coelastrum microsporum</i> Naegeli	—	—	—	+
<i>Crucigenia tetrapedia</i> (Kirchner) West and West	+	+	+	—
<i>Cyanoptycha gloeocystis</i> Pascher fo. minor Starmach	—	+	+	—
<i>Dactylococcus infusionum</i> Naegeli	—	—	+	+
<i>Desmococcus vulgaris</i> Brand	—	—	+	—
<i>Enteromorpha compressa</i> (Linnaeus) Nees #	+	+	+	+
<i>Eremosphaera viridis</i> De Bary	—	—	—	+
<i>Genticularia spirotaenia</i> De Bary	—	—	—	+
<i>Hormidiella parvula</i> Iyengar and Kanthamma	—	—	—	+
<i>Hyalidella polytomoides</i> Pascher	—	—	—	+
<i>Kentrosphaera gloeophila</i> (Bohlin) Brunthaler	—	—	—	+
<i>Kirchneriella elongata</i> G.M. Smith #	+	+	+	+
<i>Kirchneriella contorta</i> (Schmidle) Bohlin	—	—	+	+
<i>Kirchneriella lunaris</i> (Kirchner) Moebius	+	+	+	—
<i>Microthamnion strictissimum</i> Rabenhorst	—	—	—	+
<i>Mougeotia floridana</i> Trans	—	—	+	+
<i>Mougeotia genuflexa</i> (Dillw.) C.A. Agardh	—	—	+	—
<i>Mougeotia scalaris</i> Hassall	—	+	—	—
<i>Muriella terrestris</i> Petersen fo. major	—	—	+	+
<i>Murelliopsis pyrenigera</i> Reisingl	—	—	+	—
<i>Myremecia aquatica</i> G.M. Smith	—	—	—	+
<i>Neochloris aquatica</i> Starr	—	—	+	+
<i>Palmodictyon varium</i> (Naegeli) Lemmermann	—	+	—	—
<i>Pandorina morum</i> (Müller) Bory	—	—	—	+
<i>Planktosphaerella terrestris</i> Reisingl	—	—	—	+
<i>Polytoma obtusum</i> Pascher	—	—	+	—
<i>Pseudopleurococcus printzii</i> Vischer	—	—	+	+
<i>Rhizoclonium hookeri</i> Kütz.	—	—	—	+
<i>Rhopalocystis oleifera</i> Schuss	—	—	—	+
<i>Schizomeris leibleinii</i> Kütz.	—	—	+	+
<i>Schroederiella papillata</i> Korch	—	—	—	+
<i>Spirotaenia condensate</i> Breb.	—	—	—	+
<i>Spirogyra porticalis</i> (Muell.) Cleve	+	+	+	—
<i>Spirogyra rhizobrachialis</i> Jao.	—	—	+	—
<i>Spirogyra majuscula</i> Kütz.	—	+	—	—
<i>Spirogyra varians</i> (Hassall) Kütz.	—	+	—	—
<i>Tetraedron muticum</i> (A. Braun) Hansgirg	—	—	—	+
<i>Tetraedron minimum</i> (A. Braun) Hansgirg	—	—	—	+
<i>Tetraedron obesum</i> (West and West) Wille	—	—	—	+
<i>Trochiscia obtusa</i> (Reinsch) Hansgirg	—	—	+	+
<i>Tussetia elaiogenetes</i> Moewus	—	—	+	—
<i>Ulothrix subtilissima</i> Rabenhorst	—	—	+	+
<i>Ulothrix tenuissima</i> Kütz.	—	+	—	—
<i>Ulothrix aequalis</i> Kütz.	—	+	—	—
<i>Uronema elongatum</i> Hodgetts	—	—	—	+
<b>Bacillariophyta</b>				
<i>Actinocyclus</i> sp. Ehrenberg #	+	+	+	+

Algal Flora of Ain Helwan I. Algae of The Worm Spring

<i>Achnanthes lanceolata</i> (Bréb.) Grunow	—	—	—	+
<i>Amphora ovalis</i> Kütz.	+	+	—	+
<i>Amphora clevei</i> Grunow #	+	+	+	+
<i>Amphora libyca</i> Ehrenberg	—	—	—	+
<i>Asterionella Formosa</i> Hassall	—	—	—	+
<i>Bacillaria paradoxa</i> Gmelin	—	—	—	+
<i>Biddulphia laevis</i> Ehrenberg	—	—	+	+
<i>Brebissonia boeckii</i> (Ehrenberg) Grunow	+	+	—	+
<i>Campylodiscus noricus</i> var. <i>hibernicus</i> (Ehrberg) Grunow	—	—	+	—
<i>Cocconeis placentula</i> (Her.) var. <i>euglypta</i> (Her.) Cleve	—	—	—	+
<i>Coscinodiscus lacustris</i> Grunow #	+	+	+	+
<i>Cymbella cistula</i> (Ehrenberg) Kirchner	—	+	+	—
<i>Denticula elegans</i> Kütz.	—	+	+	+
<i>Denticula tenuis</i> Kütz.	—	—	+	—
<i>Denticula Kuetzingii</i> Grunow	—	—	—	+
<i>Diatoma mesodon</i> (Ehrenberg) Kütz.	—	—	—	+
<i>Diatomella</i> sp. Greville	—	—	+	—
<i>Fallacia hudsonis</i> (Grunow and Cleve) A.J. Strickle and D.G. Mann	—	—	—	+
<i>Fragillaria capucina</i> Desmazières	—	—	+	+
<i>Fragillariaforma virescens</i> (Ralfs) Williams and Round	—	—	—	+
<i>Gomphoneis herculeana</i> (Ehrenberg) Cleve	—	—	—	+
<i>Gomphonema lanceolatum</i> var. <i>insignis</i> (Greg.) Cleve	—	—	—	+
<i>Gyrosigma acuminatum</i> (Kuetzing) Rabenhorst	—	+	+	+
<i>Hantzchia amphioxys</i> (Ehrenberg) Grunow	—	—	—	+
<i>Mastogloia dansei</i> Thwaites and W. Smith	—	—	+	+
<i>Melosira italica</i> (Ehrenberg) Kuetzing	—	—	+	—
<i>Melosira granulata</i> (Ehrenberg) Ralfs	—	+	—	+
<i>Meridion circulare</i> (Greville) Agardh	—	—	—	+
<i>Navicula certa</i> Hust.	—	+	—	—
<i>Navicula digitoradiata</i> fo. <i>minor</i> Foged.	+	+	—	+
<i>Navicula halophila</i> (Grun.) Cleve	—	—	—	+
<i>Navicula viridula</i> (Kuetzing) var. <i>rostellata</i> (Kuetzing) Cleve	—	—	—	+
<i>Navicula tuscula</i> (Ehrenberg) Grunow fo. <i>rostrata</i> Hustedt	—	—	—	+
<i>Navicula slesvicensis</i> Grunow	—	—	—	+
<i>Neidium magellanicum</i> Cleve	—	—	—	+
<i>Nitzschia interrupta</i> (Reichelt) Hust.	—	+	—	—
<i>Nitzschia bilobata</i> W. Smith	—	—	+	—
<i>Nitzschia recta</i> Hantzsch and Rabenhorst	—	—	—	+
<i>Nitzschia ovalis</i> Arnott	—	—	—	+
<i>Nitzschia valdestriata</i> Aleem and Hustedt	—	—	—	+
<i>Nitzschia amphibia</i> Grunow	—	—	—	+
<i>Nitzschia palea</i> (Kütz.) W. Smith	—	—	—	+
<i>Opephora martyr</i> Herib.	—	—	+	—
<i>Rhoicosphenia curvata</i> (Kütz.) Grunow	—	—	+	—
<i>Rhopalodia gibba</i> var. <i>ventricosa</i> (Ehrenberg) O. Mütter	—	+	—	+
<i>Stauroneis parvula</i> var. <i>prominula</i> Grunow	—	+	—	—
<i>Surirella angusta</i> Kütz.	—	—	—	+
<i>Surirella oblonga</i> Ehrenberg	—	—	+	+
<i>Synedra rumpens</i> Kütz.	—	—	—	+

<i>Tabellaria fenestrata</i> var. <i>asterionelloides</i> Grunow	—	—	—	+
<i>Tabellaria flocculosa</i> (Roth) Kütz.	—	—	—	+
<b>Charophyta</b>				
<i>Tolypella intricata</i> (trentep.) N. leonhardi	—	—	—	+
<b>Chrysophyta</b>				
<i>Chrysamoeba radians</i> Klebs	—	+	+	+
<i>Chrysidiastrum catenatum</i> Lanterborn.	—	—	+	+
<b>Xanthophyta</b>				
<i>Tribonema bombycinum</i> var. <i>tenue</i> Hazen	—	—	+	—
<b>Dinophyta</b>				
<i>Amphidinium klebsii</i> Kof.	—	—	+	+
<b>Euglenophyta</b>				
<i>Euglena polymorpha</i> Dangeard	—	—	—	+
<i>Phacus orbicularis</i> var. <i>caudatus</i> Skvortzow	—	—	—	+

+ = presence of algal species; — = absence of algal species. # = persisting species in the four seasonal samples.

**Table (5): Seasonally isolated algal genera and species belonging to different divisions from water samples (site 3) during 2004-2005.**

Algal Taxa	Water Samples					
	L1 Spring	L2 Summer	L3 Autumn	L4 Winter	Total algae per year	% per total algae
<b>Cyanobacteria</b>						
Species	36	39	38	53	91	<b>43.54</b>
Genera	15	18	14	24	29	<b>25</b>
<b>Chlorophyta</b>						
Species	6	17	28	38	59	<b>28.23</b>
Genera	5	13	23	33	45	<b>38.79</b>
<b>Bacillariophyta</b>						
Species	6	14	17	41	52	<b>24.88</b>
Genera	5	12	16	29	35	<b>30.17</b>
<b>Chrysophyta</b>						
Species	—	1	2	2	2	<b>0.96</b>
Genera	—	1	2	2	2	<b>1.72</b>
<b>Euglenophyta</b>						
Species	—	—	—	2	2	<b>0.96</b>
Genera	—	—	—	2	2	<b>1.72</b>
<b>Xanthophyta</b>						
Species	—	—	1	—	1	<b>0.86</b>
Genera	—	—	1	—	1	<b>0.48</b>
<b>Charophyta</b>						
Species	—	—	—	1	1	<b>0.86</b>
Genera	—	—	—	1	1	<b>0.48</b>
<b>Dinophyta</b>						
Species	—	—	1	1	1	<b>0.96</b>
Genera	—	—	1	1	1	<b>1.72</b>
<b>Seasonal Isolated Algal Species</b>	<b>48</b>	<b>71</b>	<b>87</b>	<b>138</b>	<b>209</b>	<b>—</b>
<b>Seasonal Isolated Algal Genera</b>	<b>25</b>	<b>45</b>	<b>56</b>	<b>92</b>	<b>116</b>	<b>—</b>

Chrysophyta was the only division of the previously mentioned five ones recorded in summer sample (1 genus and 1 species); this means that certain

members of golden-brown algae could tolerate adverse physico-chemical characteristics of spring water in this season (*Chrysamaeba radians*).

From Table (4) certain species were found to be persisted in all seasonal samples and marked (#). Ten Cyanobacteria, two Chlorophyta and three Bacillariophyta species were recorded allover the year, which generally confirmed the high tolerance of Cyanobacteria species as well as certain species belonging to the other two divisions. However, it was not possible to correlate community structure with any specific environmental factor (Davey and Rothery, 1993) since each taxon varied independently to the others.

### **References**

- Abdou, I. I. M.** (1994). Geology and hydrogeology of the Helwan area, Egypt M.Sc. thesis, Geology department, Faculty of science, Menoufia University.
- Abou-El Kheir, W. S. and Mekky, L.** (1986). Studies on the algal floras inhabiting different water sources in Egypt. 2-Lakes and Springs. *Phytologia*, **61(5):285-296**.
- Abou-El kheir, W. S and Ismail, G. H.** (1986). Notes on the aquatic habitats of macrophytes and associated algae in various regions in Egypt. 1- El-Fayum region. *Phytologia*, **60:469-482**.
- Abrontes, N.; Antunes, S. C.; Pereira, M. J. and Goncalves, F.** (2006). Seasonal succession of cladocerans and phytoplankton and their interactions in a shallow eutrophic lake (Lake Vela, Portugal. *Acta Oecologica*, **29(1):54-64**.
- Ali, S. S.; Jafri, S.I.H.; Leghari, S.M. and Thebow, S.** (1983). Studies on the flora and fauna of a hot sulphur spring at Lakki (district Dadu), Sind, Pakistan. *Kar. Univ.J. Sci.*, **11(2):185-197**.
- Aly, M.S.** (2000). Ecological studies on phytoplankton in closed lakes of Wadi-El-Natron. *Egypt. J. Phycol.*, **1:203-209**.
- APHA** (1985). Standard methods for the examination of water and wastewater, 16<sup>th</sup>ed. American public health association, Washington, DC, USA, **1268 pp**.
- Arif, I. A.** (1989). Algal distributions in a warm spring of Saudia Arabia. *Arab Gulf J. Scient. Res.*, **7(2):145-154**.
- Bourrelly, P.** (1968). Les algues d'eau douce, initiation à la systématique. Tome II. Les Algues Jaunes et brunes, Chrysophycees, Xanthophycees et Diatomees. Ed. N. Boubée and Cie, Paris, France, **pp.438**.
- Bourrelly, P.** (1970). Les algues d'eau douce, initiation à la systématique. Tome III. Les algues bleues et rouges. Ed. N. Boubée and Cie, Paris, France, **512 pp**.
- Bourrelly, P.** (1972). Les algues d'eau douce, initiation à la systématique. Tome I. Les algues vertes. Ed. N. Boubée and Cie, Paris, France, **511 pp**.

- Boyed ,C.E. (1984)** Water quality in warm water fish ponds .Auburn University Agri .Exp. Sta .,Auburn, Alabama .
- Carvalho, A. P. and Malcata, F. X.** (2003). Kinetic modeling of the autotrophic growth of *Pavlova lutheri*, study of the combined influence of light and temperature. *Biotechnol. Prog.*, **19:1128–1135**.
- Cox, E. J.** (1996). Identification of fresh water diatoms from live material- Chapman and Hall, 2-6-Boundary Row, London SE 1 8 HN, U.K., **156 p**.
- Davey, M.C. and Rothery, P.** (1993). Primary colonization by microalgae in relation to spatial variation in edaphic factors on Antarctic fellfield soils. *J. Ecol.*, **81:335-343**.
- Desikachary, T.V.** (1959). Cyanobacteria, Monographs on Algae Academic press, New York and London.
- Doeck, K. and Tracey, M.V.** (1956). Modern methods of plant analysis-Springer Verlag, Berlin.
- Domingues, R.B.; Barbosa, A. and Galvao, H.** (2005). Nutrients, light and phytoplankton succession in a temperate estuary (the Guadiana, southwestern Iberia). *Estuarine, Coastal and Shelf Science*, **64(2-3):249-260**.
- El-Gamal, A. D. and Salah El-Din, R. A.** (1999). New species of the genus *Compsopogon montagne* (*C. helwanii*) from Egypt. *Phykos*, **38(1and2):37-42**.
- El-Kiki, J. E.; Mabrook, B. and Swailem, F.M.** (1978). Evaluation of trace elements and tritium content in some mineral springs in Egypt. *Isotope and Rad.Res.*, **10(1):55-82** .
- El- Ramly, I. M.** (1969). Recent review of investigations on the thermal and mineral springs in the UAR. *XXIII Int .Geol. Cong.*, **16: 201-213**.
- Foged, N.** (1980). Diatoms in Egypt. In: Nova Hedwigia Band XXXIII-Braunschweig 1980. *J. Cramer*, **629-707**.
- Foged, N.** (1984). The diatom flora in springs in Jutland, Denmark. *Bibliotheca Diatomologica*, **4: 1-118**.
- Gaballah, M. M.; Touliabah, H. E. and Sorage, M. S.** (2000). Diatom communities associated with some aquatic plants in polluted water courses, Nile Delta. *Egypt. J. Phycol.*, **1:211-224**.
- Hamed, A. F.** (1995). Studies on the algal flora of some thermal springs in Egypt. Ph.D.Thesis Ain Shams University, Faculty of Science, Botany Department, **161 P**.
- Issar, A.; Rossenthal, E.; Eckstein, I. and Bogoch, R.** (1971). Formation waters, warm springs and mineralization phenomena along the eastern shore of the Gulf of Suez. *Bull. Int. Assoc. Sci. Hydrology*, **16(3):25-44**.
- Jaha, M. and Kumar, H. D.** (1990). Cyanobacterial flora and physico-chemical properties of Saptadhara and Brahma Kund warm springs of Rajgir, Bihar, India. *Nova hedwigia*, **50(3-4 ):529-534**.
- Kobbia, I. A.; Shabana, E. F.; Dowidar, A. E.; and El-Attar, S. A.** (1990). Changes in physico-chemical characters and phytoplankton structure of



- Nile water in the vicinity of iron and steel factory at Helwan (Egypt). *Egypt. J. Bot.*, **33(3):215-233**.
- Kobbia, I. A.; Dowidar, A. E.; Shabana, E. F. and El-Attar, S. A.** (1993). Succession Biomass level of phytoplankton in the Nile water near the starch and glucose factory at Giza (Egypt). *Egypt. J. Microbiol.*, **28(1):131-143**.
- Kobbia, I. A.; Metwali, R. M. and El-Adel, H. M.** (1995). Studies on freshwater at Qalubia province (Egypt) in relation to some physico- chemical factors. *Egypt. J. Bot.*, **35(1):25-43**.
- Kullberg, R. G.** (1971). Algal distribution in six thermal spring effluents. *Trans. Amer. Micros. Soc.*, **90(4):412-434**.
- Lund, J. W. G.** (1965). The ecology of the freshwater phytoplanktons. *Biol. Rev.*, **40:231**.
- Meinzer, O. F.** (1923). Outline of ground water hydrology with definitions. U.S. Geol. Survey Water Supply Paper 494, Washington, D.C., USA, **71 pp**.
- Nikitina, V. N.** (1983). Blue green algae of mineral and warm springs of the Kronotski reservation Russian ,USSR. *Vestnik Leningradskogo Universiteta Biologiya*, **3: 47-53** .
- Prescott, G.W.** (1978). How to know the fresh water algae. Brown Company publishers Dubuque, Iowa, USA, **pp. 12-267**.
- Prescott, G.W.** (1982). Algae of the western great lakes area W.M.C. Brown Publ. Dubuque, Iowa, **pp.1-977**.
- Reeders, H. H.; Boers, P. C. M.; Van der Malen, D. J. and Helmerhorst, T. H.** (1998). Cyanobacteria dominance in the lakes veluwemeer and wolderwijd, the Netherlands. *Water Science and Technology*, **37(3):85-92**.
- Schoeman, F. R. and Archibald, R. E. M.** (1988). Taxonomic notes on the diatoms (Bacillariophyceae) of the Gross Barmen thermal springs in South West Africa/Namibia. *S. Afr. J. Bot.*, **54(3):221-256**.
- Seoane, A. N.** (1990). Phycological study of the thermal spring of Torneiros Levios , Spain. *Anales del Jardin Botanico de Madrid*, **47(2):295-300**.
- Shaaban, A. S.** (1985). The algal flora of Egyptian Oases. 11- On the algae of Siwa Oasis. *Proc. Egypt. Bot. Soc.*, **4:1-10**.
- Shaaban, A. S. and El Habibi, A.** (1978). The algal flora of Egyptian Oases .1- The algal flora of Kharga Oasis. *Bull. Inst. Dest.*, **28(1):227-232** .
- Sompong, U.; Hawkins, P. R.; Besley, C. and Peerapornpisal, Y.** (2005). The distribution of cyanobacteria across physical and chemical gradients in hot springs in northern Thailand. *FEMS Microbiology Ecology*, **52(3):365-376**.
- Swanberg, C. A.; Morgan, P.; Hennin, S. F.; Daggett, P.H.; Melic, Y.S. and El-Sherif, A. A.** (1976). Preliminary report on the thermal springs of Egypt, *Proc. Mediterranean area. Natl. Univ., Athens ,Greece*, **2: 540-554**.
- Swanberg, C. A.; Morgan, P. and Boulous, F.K.** (1984). Geochemistry of the ground waters of Egypt. *Annals of the Geological Survey of Egypt*. **14:127-150**.

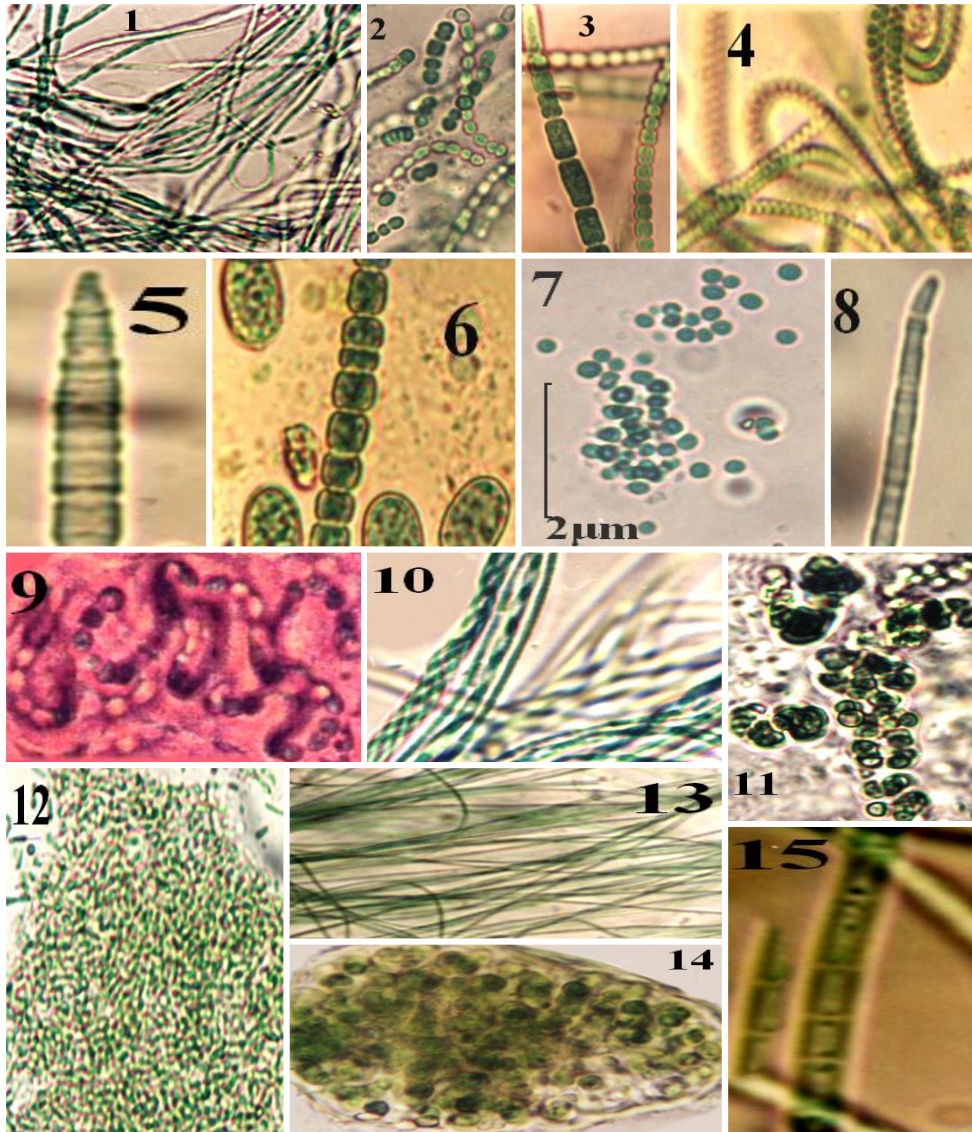
**Thompson, P.A. and Guo, M. (1992).** Effects of variation in temperature I- On the biochemical composition of eight species of marine phytoplankton. *J. Phycol.*, **28:481-488.**

## الفلورا الطحلبية لعين حلوان 1- طحالب مياه العين الدافئه ثناء محمود متولى شنب

قسم النبات - كلية العلوم - جامعة القاهرة

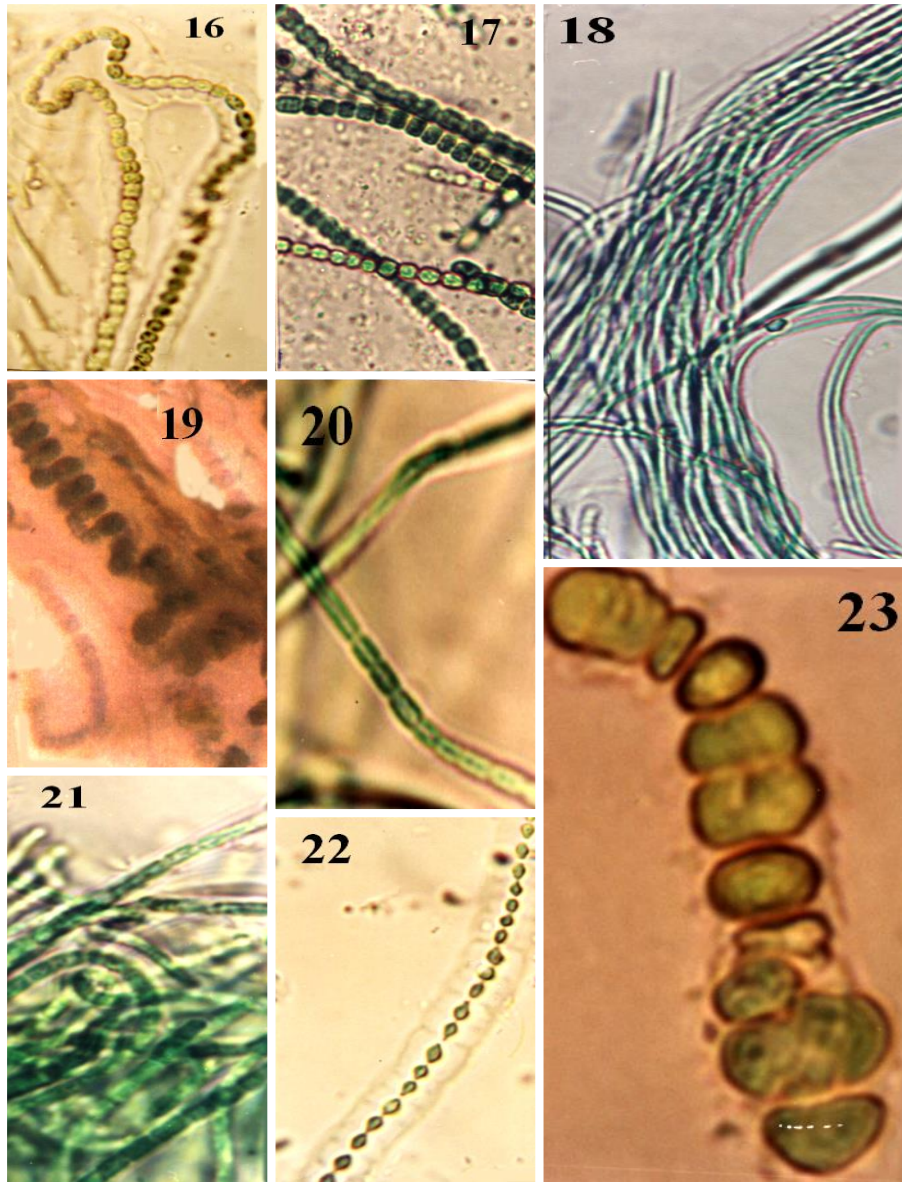
تم جمع العينات الموسمية لمياه عين حلوان خلال مدة دراسته من أبريل 2004 حتى يناير 2005 تم إجراء التحاليل الفيزيوكيميائية لكل عينه. كما تم عزل و التعرف على الانواع الطحلبيه المختلفه. بالنسبه للخواص الفيزيائيه لعينات الماء تراوحت درجات الحراره خلال مدة الدراسة بين 26-30 °م، الأس الهيدروجيني تراوح بين 6.9 - 7.78 و كانت شدة الاضاءة الساقطه على سطح الماء الضحل تتراوح بين 26.4 - 115 كيلولوكس. و من ناحيه أخرى فإن التحاليل الكيميائيه لعينات المياه (الإيونات و العناصر الغذائيه الصغرى) أظهرت تباينا و تذبذبا في الفصول المختلفه بالاضافه الى عدم وجود اى اثر للكوبلت و الكاديوم بها. العدد الكلى للفلورا الطحلبيه التى تم التعرف عليها فى عينات المياه كانت تحتوى على 209 نوع (116 جنس) منتميه الى 8 مجاميع طحلبيه هى السيانوبكتريا، الطحالب الخضراء، الدياتومات، الطحالب البنية-الذهبية، الطحالب الخضراء- المصفرة، اليوجلينيه، الدواره و الكاربه. تمثل العدد الاكبر من الفلورا الطحلبيه (خلال مده دراسته) فى السيانوبكتريا (91 نوع، 29 جنس) تليها الطحالب الخضراء (59 نوع، 45 جنس) ثم الدياتومات (52 نوع، 25 جنس) فالطحالب البنية-الذهبية و اليوجلينيه التى مثل كل منهما (2 نوع، 2 جنس). أما العدد الأدنى من الفلورا الطحلبيه فكان للطحالب الكاربه والخضراء-المصفرة والدواره التى تم التعرف على جنس واحد و نوعا واحدا من كل منهم فقط. أحتوت عينه المياه فى فصل الشتاء على أعلى نمو من الطحالب فى المجاميع المختلفه تليها تنازليا عينه الخريف فالصيف ثم الربيع. و لقد ظهر من نتائج هذه الدراسة وجود علاقة هامة بين التنوع فى الفلورا الطحلبية الفصلية وبعض الخواص الفيزيوكيميائية لعينات مياه العين.

**Plate I**  
**Certain Cyanobacteria species inhabiting water of Ain Helwan (Site 3) at different seasons during 2004-2005.**



- |  |   |
|--|---|
| 1. <i>Plectonema nostocorum</i> Bornet.        | 2. <i>Nostoc Linckia</i> (Roth) Bornet et Flah.       |
| 3. <i>Pseudanabaena catenata</i> Lauterb.      | 4. <i>Spirulina subsalsa</i> (Oersted) Agardh.        |
| 5. <i>Oscillatoria okeni</i> Agardh.           | 6. <i>Anabaena Iyengari</i> Bharad.                   |
| 7. <i>Microcystis aeruginosa</i> (Kütz.) Kütz. | 8. <i>Oscillatoria acuminata</i> Gomont.              |
| 9. <i>Nostoc paludosum</i> Kütz.               | 10. <i>Schizothrix lacustris</i> A.Braun.             |
| 11. <i>Stigonema minutum</i> (Ag.) Hass.       | 12. <i>Aphanothece caldarium</i> Richter.             |
| 13. <i>Lyngbya tenue</i> Gomont.               | 14. <i>Stigonema mamillosum</i> (Lyngb.) C.A. Agardh. |
| 15. <i>Lyngbya bipunctata</i> Lemm.            |   |

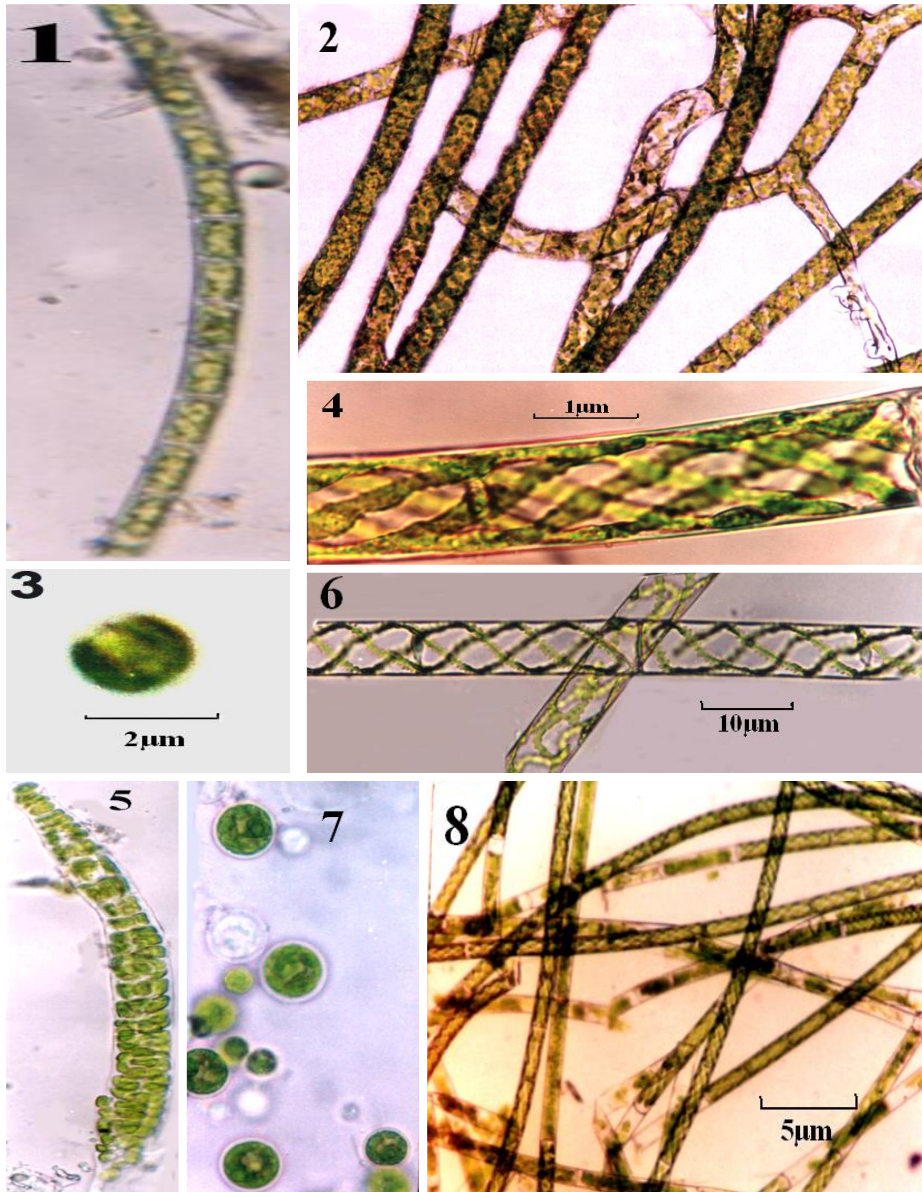
**Plate II**  
**Certain Cyanobacteria species inhabiting water of Ain Helwan (Site 3) at different seasons during 2004-2005.**



16. *Albrightia tortusa* Copeland (x 1000).      17. *Nodularia implexa* Bornet et Flah.  
18. *Microcoleus vaginatus* (Vaucher) Gomont.      19. *Anabaena helicoidea* Bernard.  
20. *Lyngbya turidum* Gomont. (x 1000).      21. *Lyngbya valderianum* Gomont. (x 1000).  
22. *Mastigocoleopsis obtusa* (Carter) Geitler.      23. *Nodularia harveyana* Thuret (x 1000).

**Plate III**

**Certain Chlorophyta species inhabiting water of Ain Helwan (Site 3) at different seasons during 2004-2005.**



1. *Mougeotia scalaris* Hassal.

3. *Chlorella vulgaris* Beijer.

5. *Schizomeris leibleinii* Kütz.

7. *Chlorococcum humicola* (Naeg.) Rabenhorst.

8. *Spirogyra acquinoctialis* G.S.West (x 100)

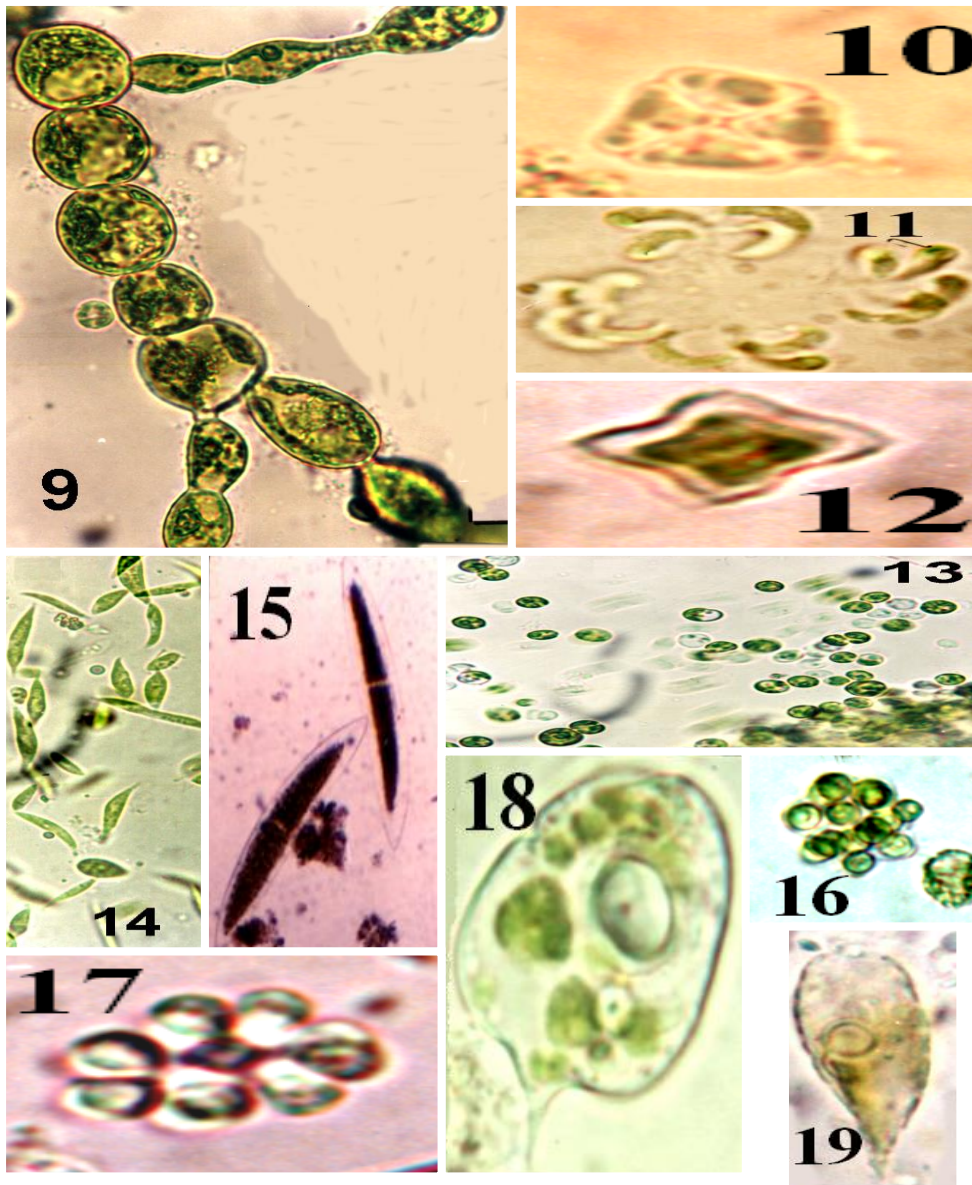
2. *Spirogyra rhizobrachialis* Jao. (x 200).

4. *Spirogyra porticalis* (Muell.) Cleve (x 1000).

6. *Spirogyra pratensis* Transeau (x 200).

**Plate IV**

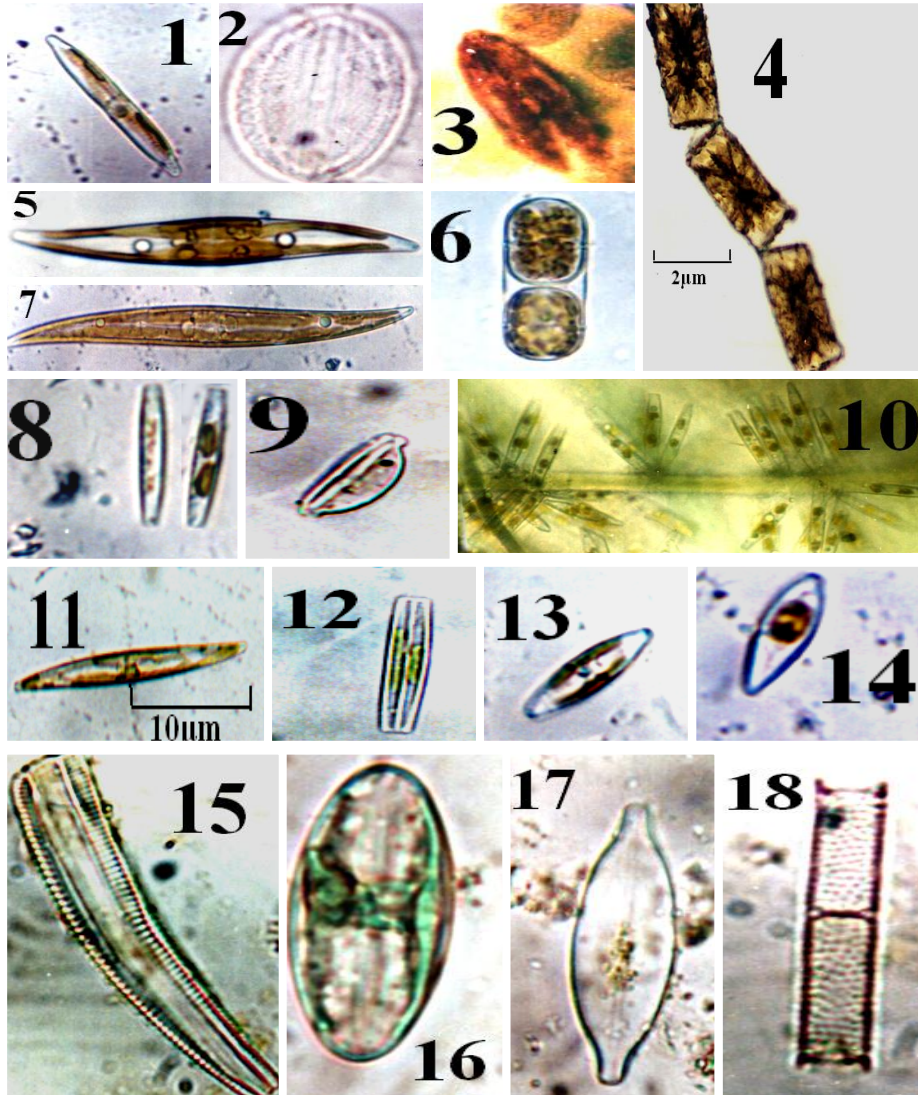
**Certain Chlorophyta and Euglenophyta species inhabiting water of Ain Helwan (Site 3) at different seasons during 2004-2005.**



9. *Pseudopleurococcus printzii* Vischer  
 10. *Crucigenia tetrapedia* (Kirch.) West and West  
 11. *Kirchneriella elongata* G. M. Smith  
 12. *Tetraedron minimum* (A. Braun) Hansgirg  
 13. *Muriella terrestris* Petersen fo. major  
 14. *Ankistrodesmus braunii* (Naeg.) Brunn. and *A. fractus* (West and West) Brunn.  
 15. *Closterium lunula* (Müller) Nitz.  
 16. *Coelastrum microsporum* Naegeli.  
 17. *Pandorina morum* (Müller) Bory.  
 18. *Phacus arbuticularis* var. *Caudatus* Skvortzow.  
 19. *Euglena polymorpha* Dangeard.

Plate V

Certain Bacillariophyta species inhabiting water of Ain Helwan (Site 3) at different seasons during 2004-2005.



- |  |   |
|--|---|
| 1. <i>Nitzschia recta</i> Hantzsch ex Rabenhorst.                            | 3. <i>Neidium magellanicum</i> Cleve.           |
| 2. <i>Cocconeis placentula</i> Ehr. var. <i>euglypta</i> (Ehr.) Cleve.       | 4. <i>Biddulphia Laevis</i> Ehrenberg.          |
| 5. <i>Gyrosigma acuminatum</i> (Kütz.) Rabenhorst.                           | 6. <i>Melosira dickiei</i> (Thwaites) Kuetzing. |
| 7. <i>Gyrosigma spencerii</i> (W.Smith) Cleve.                               | 8. <i>Nitzschia palea</i> (Kütz.) W.Smith.      |
| 9. <i>Cymbella cistula</i> (Hempr.) Grunow.                                  | 10. <i>Nitzschia ovalis</i> Arnott.             |
| 11. <i>Navicula digitoradiata</i> (Gregory) Ralfs.                           | 12. <i>Fragillaria capucina</i> Desmazières.    |
| 13. <i>Fallacia hudsonis</i> (Grunow and Cleve) A. J. Stickle and D.G. Mann. |   |
| 14. <i>Gomphonema olivaceum</i> (Hornemann) Brébisson (Photo8-14 are x 160). |   |
| 15. <i>Rhoicosphenia curvata</i> (Kütz.) Grun.                               | 16. <i>Amphora ovalis</i> Kütz.                 |
| 17. <i>Anomoeoneis spherophora</i> (Kütz.) Pfitzer.                          | 18. <i>Melosira granulate</i> (Ehr.) Ralfs.     |