



## New Approaches of Biotechnology in Textile Coloration

Heba Mohammed El-Sayed El-Hennawi

Dyeing, Printing and Textile Auxiliaries Department, Textile Industries Research Division,  
National Research Centre



### Abstract

The application of biotechnology in textile processing is gaining worldwide detection as one of the hopeful approaches to pollution issues and cost reduction. Hydrolase enzymes as amylases, pectinase, and cellulase have been successfully used in the wet process over the past 40 years. The application of these enzymes in one bath is a new challenge. Laccases enzymes are used in biosynthesis and decolorization of textile effluents. Application of enzyme in hydrolysis of starch thickeners which is the main component of the printing paste enzyme is one of the promising areas. Green synthesis of dyes, *in-situ* coloration, and polymer grafting are the new trends of enzymes application as it produces smart colored textile. The application of biotechnology in textiles is the solution to almost all the problems of this industry. The enzyme's researches illustrate the improvement in product quality at shorter times and lower energy and water consumption. This state of art highlights the novel approaches of application of enzymes in the textile coloration and industry covering both current types of research and pilot application.

**Keywords:** New approaches, Biotwcknology, Textile coloration

### 1. Introduction

Environmental considerations are fundamental factors during the choice of consumer goods including textiles. On the other hand, social

**Table 1.** The ecofriendly aspect in textiles is one of the considerable issues because textiles present another skin layer. Application of biotechnology (enzyme and yeast) attract the

pressures are increasing on textile processing units due to polluting effluents. [1, 2] The effect of chemicals used in wet processing on human health is listed in researches and textile factories to overcome the textile pollution problems that fulfilled the requirement of the customer and environment consideration.

**Table 1:** Health hazards associated with chemicals used in textile processing textile industries

Process	Chemicals used	Health hazards
<b>Singeing</b>	A small amount of exhaust gases, neglectable impact	-----
<b>Desizing</b>	Enzymes or H <sub>2</sub> SO <sub>4</sub> for starch, detergents, and alkali for PVA and CMC	Bloating and diarrhea Irritant to eyes and skin
<b>Scouring</b>	NaOH, Na <sub>2</sub> CO <sub>3</sub> , surfactants, chlorinated solvents	The nonionic detergent may cause bloating and diarrhea Irritant to eyes and skin
<b>Bleaching</b>	Hypochlorite, hydrogen peroxide, acetic acid	Chlorine gases released cause severe irritation of respiratory tract and eyes tract and eyes toxic gases
<b>Mercerization</b>	NaOH, surfactants, acid, liquid ammonium	-----

\*Corresponding author e-mail: [skybird740@yahoo.com](mailto:skybird740@yahoo.com).; (Heba El-hennawi).

Receive Date: 29 June 2020, Revise Date: 28 August 2020, Accept Date: 11 November 2020

DOI: 10.21608/EJCHEM.2020.34098.2714

©2021 National Information and Documentation Center (NIDOC)

## 2. Enzyme classification

The enzyme's role in fermentation was known in Ancient Greece. By the time more knowledge of enzyme theory, extraction and the appearance of thermostable enzymes added up the new potential of industrial processes. [3, 4] Classification of enzyme according to its application are listed in **Error! Reference source not found.**

### The added-value of using enzymes in the textile industry

- An enzyme acts as a catalyst, meaning it helps in proceeding the reaction, and at the same time faster by decreasing the activation energy of the substrate and in the same time it remains at the end of the reaction
- Each reaction has its condition of temperature and pH depending on the enzyme used but overall, all enzymes act at the same condition of its natural source (bacteria, fungi,...etc) which are modest conditions.
- According to the hypothesis describe the reaction of enzyme-substrate. Enzymes bind with the substrate and under the optimum condition, ES compound dissociates to the

**Table 2:** Classification of enzyme according to its application

Group of enzymes	Reaction catalyzed	Example
<b>Oxidoreductases</b>	Transfer of hydrogen and oxygen atoms or electron from one substrate to another	Dehydrogenases Oxidases
<b>Transferases</b>	Transfer of a specific group (a phosphate or methyl, etc) from one substrate to another	Transaminase Kinases
<b>Hydrolases</b>	Hydrolysis of a substrate	Esterases Digestive enzymes
<b>Isomerases</b>	Change of the molecular form of the substrate	Phosphor hexo isomerase Fumarase
<b>Lyases</b>	Nonhydrolytic removal or addition of a group to the substrate	Decarboxylases Aldolases
<b>Ligases (synthetases)</b>	Joining of two molecules by the formation of new bonds	Citric acid synthetase

## 4. Enzyme applications in textile wet process

The application of enzymes in textile manufacturing were shown in

**Figure 1**, and will be discuss in details.

### 4.1. Cotton desizing

Amylases enzymes are proficient in digesting glycosidic linkages found in starch. Truncated enzymes are higher stability and activity comparing to other amylase enzymes used due to its altering structural conformations, and increased affinity with the substrate. [4, 13]

The ultrasonic energy assists enzymatic desizing of cotton by reducing processing time to half with higher efficiency. [14]

required product only and the enzyme. That reduces pollution as no byproduct and the enzyme itself is biodegradable. [5, 6]

## 3. Industrial Enzymes: Production and Application

Novozyme Company was one of the leader's companies which have produced commercial enzyme formula ready for use in textile factories. Then different companies around the world have entered this field which increases the competition that affects the quality and the price of the commercial enzymes. Enzyme products are available at high concentrations suitable for the production of high-quality textiles and the products are compatible with most chemical auxiliaries.

Genetic science help in the production of the new enzyme as it can identify the responsible gene on enzyme secretion. [5-7]

Textile enzymes are classified under the category of technical enzymes based on application classification (see **Error! Reference source not found.**). [8, 9] Commercial enzymes used in textiles are mainly hydrolysis enzymes. [10-12]

### 4.2. Enzymatic scouring (bio-scouring)

The enzyme-catalyzed the removal of impurities from the surface of cellulosic fibers, in bleaching and dyeing improvement. Enzymatic scouring performed at neutral pH, low water consumption, and does not affect fiber strength and low percent of weight loss compared with processing in traditional ways. [15]

Combined desizing and scouring process for cotton fabric using each of the two commercial amylase enzymes namely BEISOL T2090 and BEISOL PRO were investigated. The optimum conditions were enzyme concentration at 2% of the weight of the fabric, time 30 minutes, temperature 90°C, and pH 7.5. [16]

Treatment with one of those couples enzymes (pectinase + cellulase), (pectinase +

protease), (pectinase + cutinase) are capable of removing cotton noncellulosic impurities completely when used on an industrial scale. [17, 18]

#### 4.3. Enzymatic bleaching (Biobleaching)

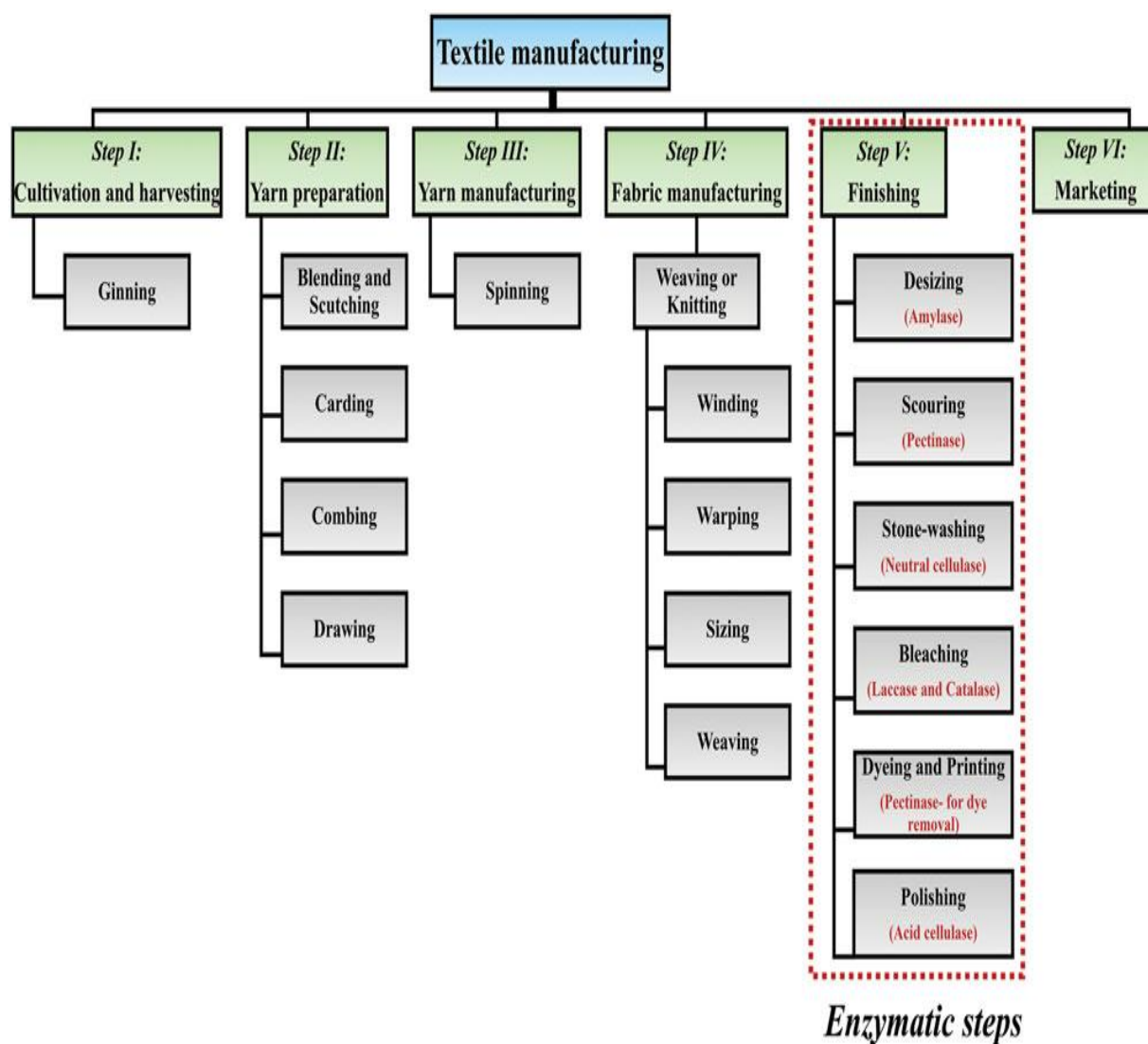
In the bleaching process, the natural coloring matter is removed which is directly related to the success of subsequent coloration operations such as dyeing and printing. Glucose is oxidized by glucose oxidase enzyme to hydrogen peroxide.

Integrated desizing–bleaching–process for cotton towel then dyed with reactive dye without discharging each spent bath has revealed better eco-sustainability and economy. The desizing effluent discharge treated with Amyloglucosidase–pullulanase enzyme given glucose that will be oxidized by glucose oxidase enzyme generating hydrogen

peroxide. In the next step, the catalase enzyme was added before reactive dyes to remove residual hydrogen peroxide. This method saves 400% and 50% of water and energy respectively. [10, 19]

Brewer's yeast enzymes (lipase, amylase, and cellulase) were applied on bamboo and bamboo/cotton knitted fabrics. The natural dye uptake increased for the two treated fabrics by improving the wettability. [20]

In another study combination of Brewer's yeast enzymes and microwave irradiation treatment of semi-finished rami fabric was investigated. The colorimetric data revealed that the treatment of rami fabrics in the microwave imparted bright color with excellent colorfastness properties at optimum temperature 60°C, pH 4 for 20 min. [21]



**Figure 1:** Application of enzymes in textile manufacturing

## 5. Denim fashion look

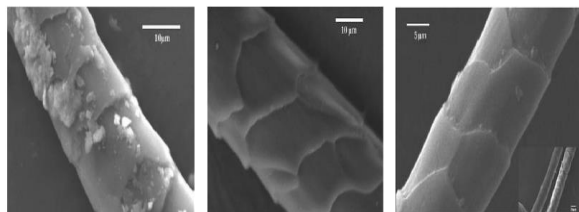
Denim washing technology is mainly used to make it soft, supple, and smooth which improves wearer's comfort besides modifying the appearance outlook. [22, 23] Sandblasting was the first finish method applied on denim fabric in which sand is sprayed at high pressure on the selected area of the denim fabric. The sprayed sand removes color from the required area of the denim fabric. [24]

In 2017 aloe gel was used for desizing cotton fabric as it contains enzymes like amylase, lipase, and cellulase besides different minerals like manganese. The Enzymatic cotton desizing conditions were 50-60 g/l aloe gel concentration with 5 g/l sodium chlorite at temperature 70-80°C for 60 minutes. The results of aloe gel desizing are compared with that of enzyme and there was a high amount of weight loss in aloe gel desizing which proves that this process is more efficient than synthetic enzyme desizing and also provides antibacterial finish. [25]

## 6. Enzymatic treatment of wool

Raw wool can't be colored either by dyeing or printing owing to the hydrophobic impurities of the epicuticular surface membrane. Convention chemical treatment used sodium carbonate for alkaline scouring or pretreatment using potassium permanganate, sodium sulfite or hydrogen peroxide. [26]

Treatment of wool fiber with different concentrations of protease enzyme improves its physical and chemical properties compared to untreated fabric with a slight increase in weight loss and tensile strength (see **Figure 2**). [27]



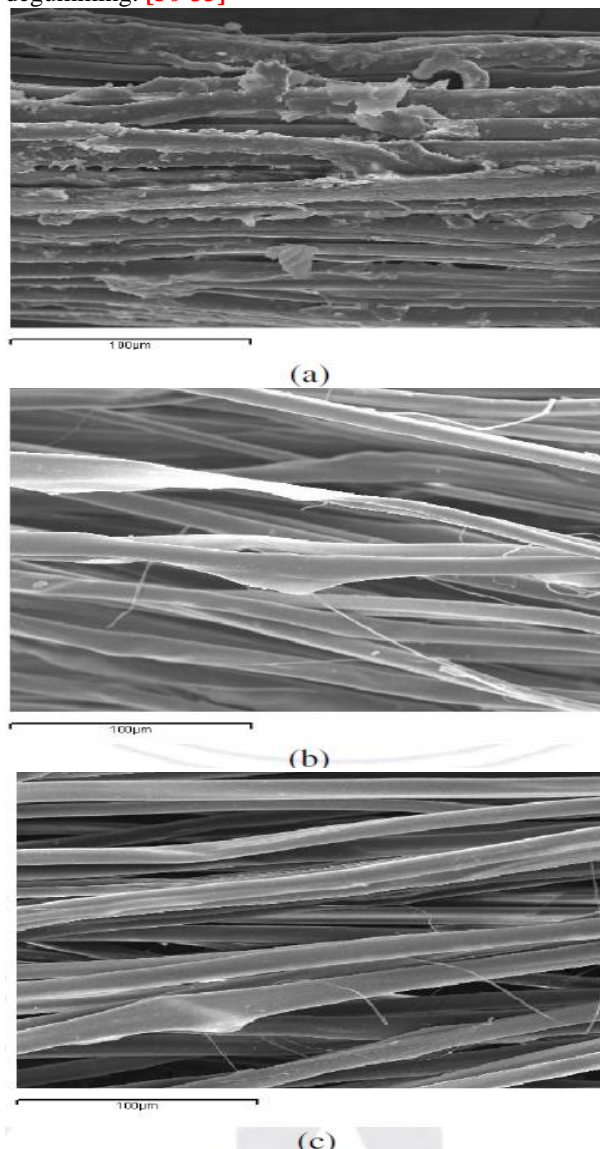
**Figure 2:** SEM of raw wool, chemically treated and protease treated wool [27]

Wool fabric has the affinity to shrink by wet processing. This is approved by the chemical commercial process using chlorine-Hercosett but its main drawback contamination of wastewater effluent with organic halogens (AOX). [28]

Glycerol diglycidyl ether was used to form cross-linking bridges in the wool at high temperature curing after protease treatment. This method produces complete shrink-resistance wool fabric dyed with reactive dyes. [29]

## 7. Enzymatic treatment of silk

Raw silk structure contains the outer sericin layer which consists of amino acids that cover the fiber and decrease its wettability. The degumming process is typically performed in detergent at high temperature and alkaline pH to emulsify sericin. This method affects the silk's physical properties along with the release of detergent effluent in the environment (see **Figure 3**). A variety of commercially available proteolytic enzymes as lipases and bacterial proteases are being explored for degumming. [30-33]



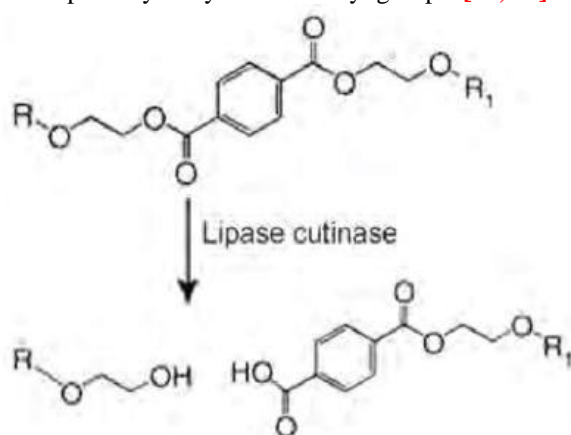
**Figure 3:** SEM of (a) raw silk (b) after degumming soap/alkali (c) degumming with protease enzyme

## 8. Surface modification of synthetic fibers

The hydrophobicity nature of synthetic polymers as polyamide (PA) and polyester (PET) make them uncomfortable to wear. [34]

Sodium hydroxide was used in the traditional chemical process to improve the hydrophilicity and flexibility of the synthetic textile but due to the increase in weight loss and yellowing of fibers, this process was unacceptable. Many proteases enzymes with a different commercial name such as Protex Gentle L, were used to induce changes in the nylon 6,6 polymer. Protease treatment of Nylon 6,6 showed significant high reactive and acid dye bath exhaustion. [35-37]

The role of enzymatic treatment of polyester is to improve the hydrophilicity by hydrolysis of ester bond using lipase and polyesterases enzyme (see **Figure 4**). [38-40] Hydrolysis of ester bond by cutinases, lipases, and esterases enzymes formed active polar hydroxyl and carboxyl groups. [41, 42]



**Figure 4:** Enzymatic hydrolysis of polyester [38-40]

## 9. Textile coloration

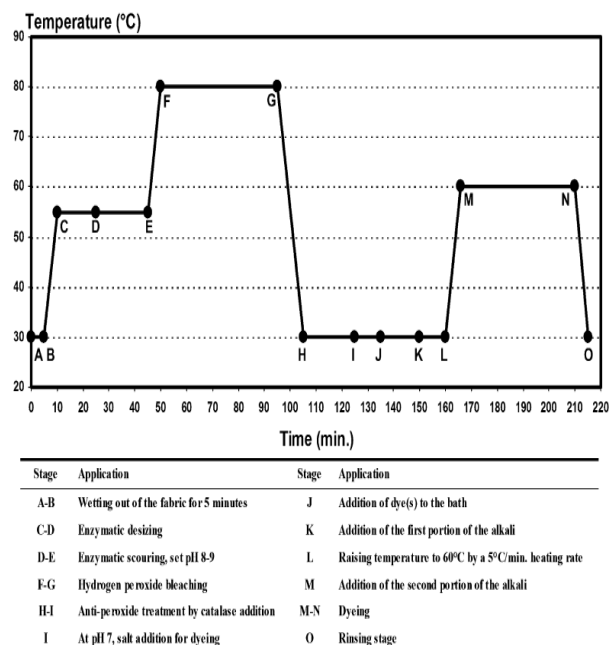
### 9.1. Dyeing

Dyeing is one of the coloration methods of biotreated textile samples. The quality of dyed samples is measured by color evenness. Investigations were carried out to optimize one bath pretreatment and dyeing as it reduces the consumption of water, shorter process time, and economy. [43-47] Rapid Enzymatic Single-bath Treatment (REST) is a developed a new method to dye untreated woven cotton with reactive dye using a single bath combined process as shown in **Figure 5**. All tasks were completed in half the time of the conventional dyeing and without water replacement. The fastness properties of dyed cotton samples are very good and no effect on the tensile strength. [48]

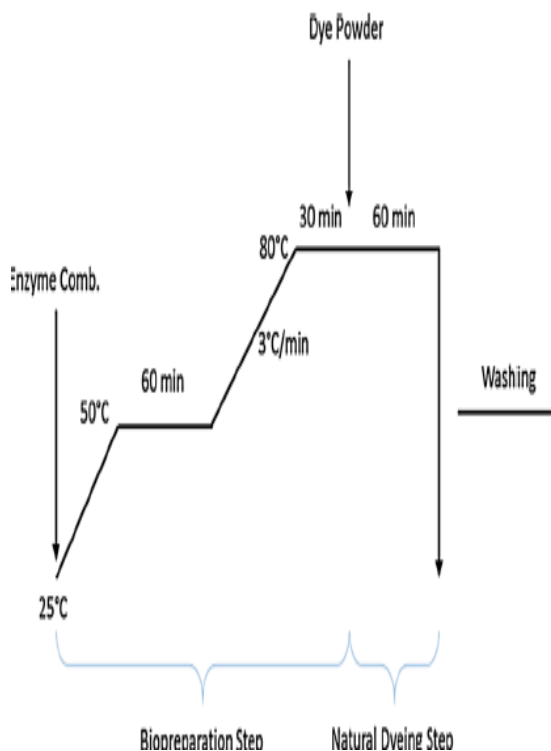
Further experimental was carried out using an ultrasonic bath was used in biotreatment and dyeing of cotton fabric with natural dyeing (see **Figure 6**). The ultrasonic energy provided gave high color efficiency at lower water consumption and energy. [49]

Another experiment was carried out on cotton /polyester blend, at any ratio, shows that it could be dyed with no cleaning process. The

colorfastness properties results are very good and hand feel was also found acceptable. This experiment could be exploitable on other blends of cellulosic/polyester fibers. [50]



**Figure 5:** Temperature and time condition of REST [48]

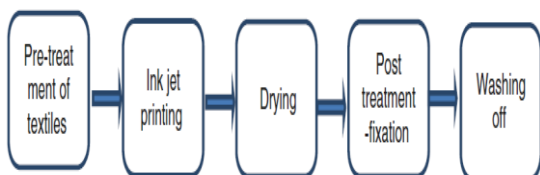


**Figure 6:** Experimental condition of biotreatment and dyeing cotton with natural dyes [49]



## 9.2. Digital printing

Digital printing has more advantages than screen printing as it is flexible, creative since there are no limits of colors or designs, and eco-friendly. The main drawback is the high price of used inks (see **Figure 7**).



**Table 3.** The optimum condition used was pH adjusted at 4 and 10 at optimum 70°C and 50°C

**Table 3:** Effect of enzymes concentration on color strength of biotreated digital printing fabrics

Enzyme used	Concentration	Fabric					
		Linen/polyester		Linen		Polyester	
		K/S	%K/S	K/S	%K/S	K/S	%K/S
Untreated	-	0.76	100	1.76	100	12.02	100
Brewer's yeast	50 ml/kg	0.88	115.79	0.55	31.25	0.28	2.33
	100 ml/kg	0.93	122.37	0.73	41.48	0.49	4.08
	300 ml/kg	1.01	132.90	0.92	52.28	0.72	5.99
	600 ml/kg	2.40	315.79	2.17	123.30	3.62	30.11
	900 ml/kg	4.01	527.63	3.11	176.71	7.60	63.22
Valumax A356	50 g/kg	1.56	205.26	3.20	181.82	9.51	79.12
	100 g/kg	1.70	223.68	3.25	184.66	9.72	80.86
	200 g/kg	2.78	356.79	4.10	232.95	11.23	93.43
	220 g/kg	2.60	342.11	3.48	197.73	10.75	89.43
	240 g/kg	2.53	332.89	2.84	161.36	10.47	87.10
	260 g/kg	2.21	290.79	2.78	157.95	10.28	85.52
Valumax A828	50 g/kg	1.19	156.58	3.05	173.30	7.11	59.15
	100 g/kg	1.45	190.79	3.16	179.55	8.28	68.88
	200 g/kg	3	394.74	4.16	236.36	12.69	105.57
	220 g/kg	2.62	344.74	3.62	205.68	10.75	89.43
	240 g/kg	2.23	293.42	2.83	160.80	10.55	87.77
	260 g/kg	1.69	222.37	2.10	119.31	9.93	82.61

## 10. Decolorization of dyes

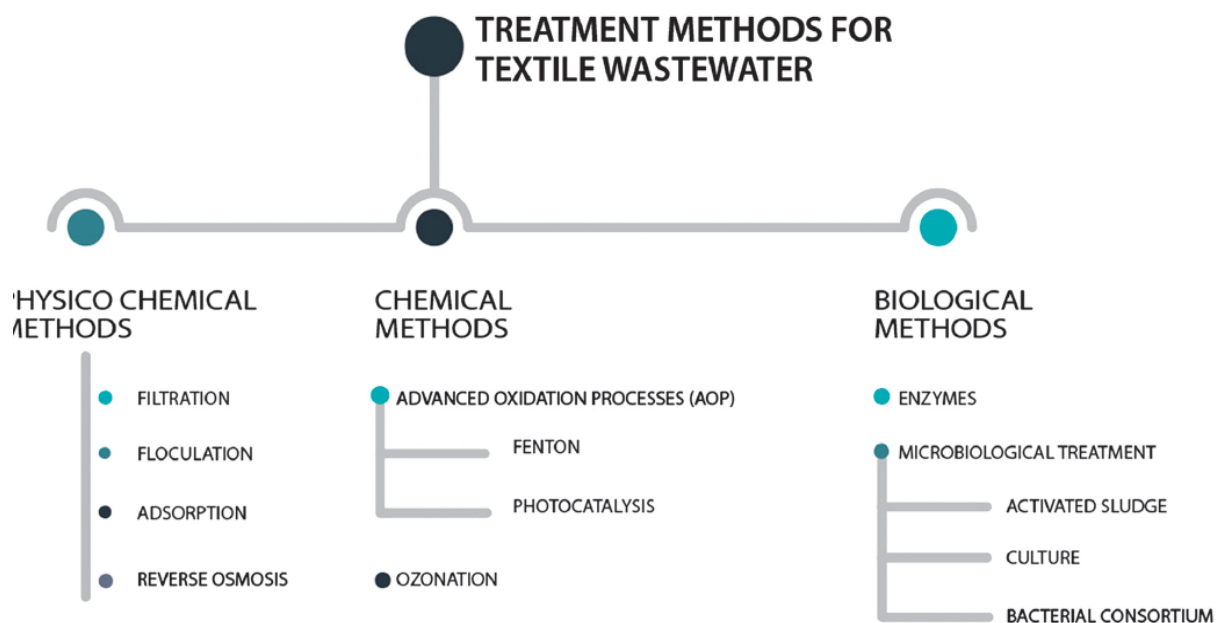
The textile industry consumes about 60 % of the dyestuff market. [54] The reactive azo dyes come at the top of the dyes used that is related to the high demand for bright colored cotton fabrics. The washing effluent of the colored garment after dyeing or printing contains an different metal ions and unfixed hydrolyzed dye that released into the

**Figure 7:** Digital printing steps

Polyester fabric is widely used in inkjet printing due to its characters as high strength and flexibility but on the other hand, it had low wettability that gives dull colors and even bleeding. Enzymatic pretreatment of polyester meant for inkjet printing was carry out using different enzymes like lipase, cellulase, and brewer's yeast. The results show a significant increase in the color strength and its percentage over 200% of biotreated inkjet printed polyester, linen, and linen polyester blend fabrics as shown in

for 60 min for Brewer's yeast suspension upon using the two Valumax enzymes relatively. [51-53]

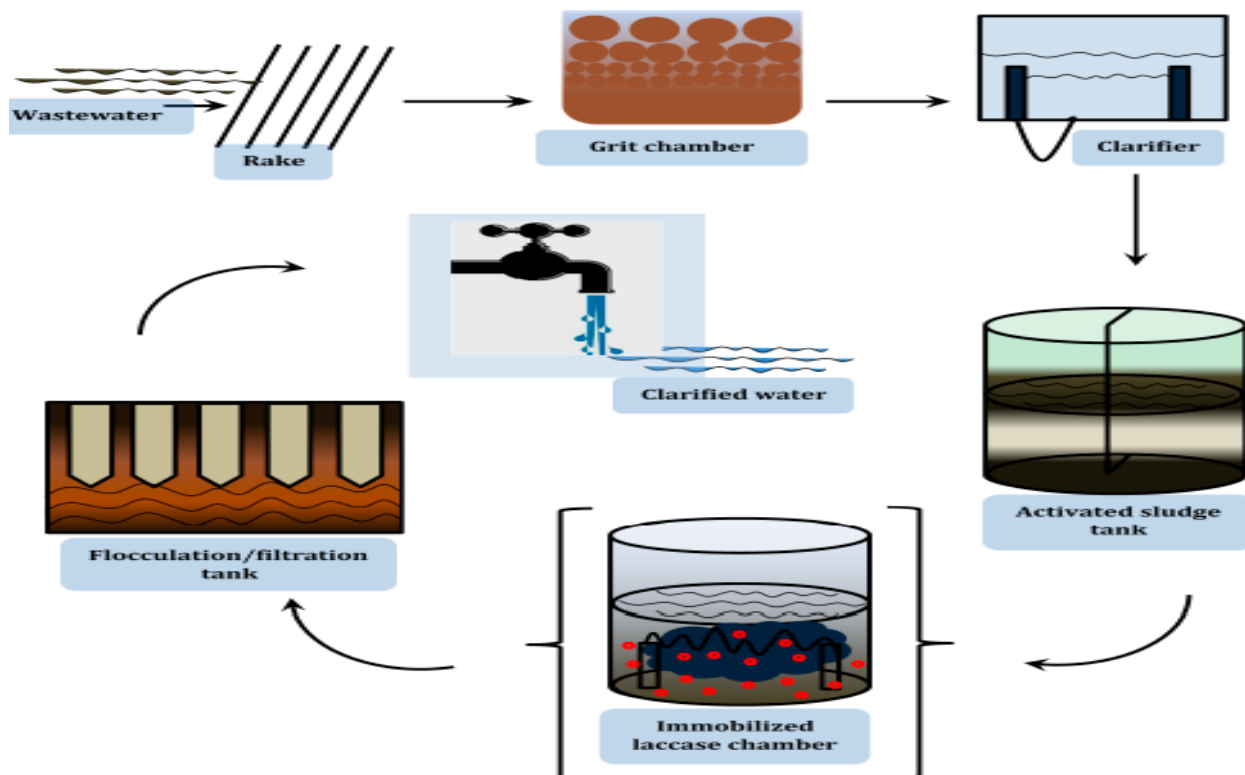
environment. [55, 56] Sometimes the color of the effluent can be noticed, due to the high percentage of dye, given colored streams or lakes. [57] These effluents are toxic for aqua life beside it prevents the sunlight leading to reduce photosynthesis and percentage of dissolved oxygen. Several physical and chemical treatments are used in the wastewater treatment but they have their limitations which render them unattractive (see **Figure 8**). [58, 59]



**Figure 8:** Textile wastewater treatment methods [58, 59]

Enzymatic degradation is viewed as a promising process for waste treatment if designed to be cost-effective. Oxidative enzyme as Laccase can convert the mean pollutants into less toxic or insoluble compounds that can be removed from

effluents. [60-63] Studies state the capability of laccase to degrade textile effluent dyes but its application on industrial processes is not always successful due to the loss of the enzyme of stability and activity. [64] The use of redox mediators and emulsions can improve catalytic reactions. Immobilized enzyme provides several economic advantages included the improvement of thermal stability and possible reuse (Figure 9). [65]



**Figure 9:** Suggested scheme of wastewater treatment using immobilized Laccase enzyme [65]

Commercial laccase formula produces from Novozyme Company for denim washing was used for decolorization of simulated textile effluent consist of a mixture of three reactive dyes and auxiliaries. The results were promising as Laccase catalyzed degradation of around 55% of dyes. Effluent treated analysis as COD, BOD, and toxicity were found to be in the permitted limits of wastewaters. [66]

### 11. Enzymes used in the detergent industry

Detergents which are a prime application area for enzymes represent 25–30 % of the total enzyme sales. The enzymatic containing detergent entered the market in 1960. [67] Since then the detergent companies started to introduce more enzymes in its detergent to reduce the phosphate percentage to an eco-friendly limit along with the bleaching agent. The new trend in the detergent using a mix of enzymes (lipase, protease, and amylase) and softener. Application of detergent is not limited to laundry only, it was extended to others as dishwashing and industrial cleaning (see

**Table 4).** [10, 68-70]

**Table 4:** The role of different enzymes in the detergent formula

Name of enzyme	Effective as stain remover for
Proteases	Grass, blood, egg, sweat stains
Lipases	Lipstick, butter, salad oil, sauces
Amylases	Spaghetti, custard, chocolate
Cellulases	Color brightening, softening, soil removal

## 12. New trends in the application of enzyme in textile

### 12.1. Immobilization

Fabrics (cotton, polyester, and polyamide) are attractive support for enzyme immobilization as they are flexible, cheap, and lightweight. Immobilized enzymes are thermostable, easy to recover, and reused several time. The using methods for immobilization are illustrated in **Table 5**. Immobilization enzymes are used in several industries manly food but still need to be induced in textile especially wet operations. [71-75].

**Table 5:** Methods for immobilization of different enzymes on textile

Enzyme	Fabric type	Activation/binding material
Peroxidase (HRP)	Polyester, polyethylene	Glutaraldehyde, plasma
Catalase	Polyester, polyamide 6,6	Photochemical
Catalase	Cotton	Oxidation by sodium periodate
Tyrosinase	Silk fibroin, polyamide 6,6	Glutaraldehyde
Laccase	Polyamide 6,6	Enzymatic hydrolysis, Glutaraldehyde, and spacer
Glucose oxidase	Silk fibroin, polyamide 6, viscose, polyester, polypropylene	Various activation strategies
Alkaline phosphatase	Silk fibroin	Low-temperature plasma
Lysozyme	Cotton	Esterification with glycine/ glutaraldehyde
Organophosphate hydrolase	Wool	Glutaraldehyde
	cotton	Esterification with glycine/ glutaraldehyde
Thrombin	Polyester	Ethylenediamine

### 12.2. Biodischarge printing

Discharge printing based on the destruction of the ground dye in the printed areas to obtain the required pattern. The commonly used discharging agents are based on formaldehyde sulphonylates and thiourea dioxide. Formaldehyde sulphonylate ( $\text{NaHSO}_2\text{CH}_2\text{O}\cdot 2\text{H}_2\text{O}$ ). Those agents are carcinogenic as it dissociates producing formaldehyde (see **Figure 10**).

The optimum condition of applying the Peroxidase enzyme in textile discharge printing was at pH 8.5 at 70°C for 60 min. [76] In another research, the commercial enzyme formula of Laccase and cellulase beside brewer's yeast suspension has been applied successfully in discharge printing. The higher percent removal of ground dye follows the order laccase enzyme, Brewer's yeast suspension, and cellulase enzyme commercial product at the end. [77] Laboratory experiments reached the optimum conditions (temperature and treatment time, pH) of all used enzymes and brewer's yeast filtrate for their utilization as discharging agents. The next step was to apply the results on an industrial scale at Egyptian Knitted Service Company in comparison with rongalite C. Laccase enzyme removed 94.8, 95.0, and 96.8% of the ground color on using sunzol yellow and sunzol blue dischargeable reactive dyes. All the aforementioned biomaterials were used either alone or mixed with rongalite C. Samples with different halftones have been obtained by changing the nature of the biomaterial and/or its ratio with rongalite C in the mixture. [78]

In another lab study, the laccase enzyme was used in the discharge printing of cotton, silk, and wool dyed with natural dyes. The laccase was added to the printing paste then printed on the naturally dyed samples. Halftone samples were obtained at the optimum conditions of enzymes (pH 4.5, treatment temperature 70°C for 1 hour). [79]





**Figure 10:** The action of cellulases in discharge printing (a), White discharge printing (b) and Halftone (c)

### 12.3. Biodegradation of starch thickener

Different kinds of starches were subjected to gelatinization under the action of sodium **Table 6**):

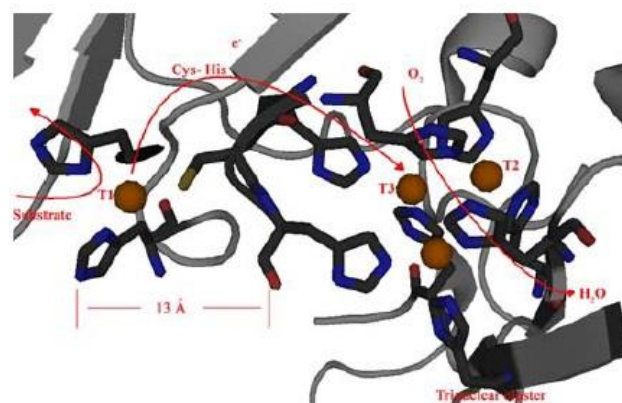
- *Group 1:* direct oxidation of phenolic derivatives
- *Group 2:* Using mediator (compound of low molecular weight and low redox potential) to assist the oxidation of phenolic and non-phenolic substrates
- *Group 3:* homomolecular reaction by coupling the produced reactive radicals.

hydroxide solution then treated with an alpha-amylase enzyme in the gelatinized form. The modified starches were evaluated as thickeners in printing wool fabrics with acid dye and polyester using disperse dye. The results demonstrated that increasing the apparent viscosity by increasing starch concentration.

### 12.4. Green catalysts for Synthesis of dyes

A wide range of dyes is produced by the oxidation reaction of colorless compounds as phenols derivatives using laccase enzymes. **[80, 81]**

The oxidation mechanism of laccase depends on the presence of copper atom (II) at its core (see **Figure 11**). The scientist named them according to their spectroscopic adsorption band to T1, T2, and T3. The T1 copper is responsible for the blue color of laccase as it has an absorption band at 610 nm. **[82-86]** The oxidation-reduction reaction catalyzed by laccase depends on two factors the pH of the medium and the redox potential of the substrate. Phenols consider the best substrate of laccase because of its low redox potentials. The role of the laccase enzyme copper's atoms in proceeding the oxidation-reduction mechanism of phenol derivatives start its attach to T1 copper permit the electron transfer from the substrate to copper T1 given phenoxyl radical then the second Step is the electron transfer, T2/T3 coppers, given the reduced enzyme form. **[80, 87, 88]** The produced phenoxyl radical is the starter of further reaction. The chemical reactions catalyzed by laccase can be divided into three groups (



**Figure 11:** Oxidation–reduction mechanism of laccase

#### 1. Phenolic and non-phenolic dyes

Dyes of wide color range from yellow, brown to red, and blue are the result of the phenoxyl radicals produced from laccase oxidation reaction with the phenolic or non-

phenolic compound as derivative of benzoic acid. [89-94]

## 2. Phenoxazinone

Phenoxazinone compounds are colorful, non-toxic and some if they have fluorescence properties used as probes in the field of molecular biology and can be used in textiles for dyeing of natural fabrics. [95, 96]

## 3. Azo dyes

Synthesis of azo dye without using the forbidden amino acid could be achieved by using the laccase enzyme. it has been reported that red azo dye is the end product oxidation reaction, catalysis with methoxy phenols with benzothiazoles. These results lead to the eco-friendly synthesis of azo dyes. [89, 97, 98]

**Table 6:** laccase assist the synthesis of dyes using different chemicals

Substrate of laccase + reaction partner	Type of reaction product
<b>Phenolic and non-phenolic dyes</b> Phenoxy- and aminoderivatives of benzene- and naphthalene-sulfonic acid; dihydroxynaphthalene; phenylphenol; pyridine	Yellow, orange, red, purple, brown and green/grey products phenoxazinone dye
Phenoxy-, amino-, methoxy-, methyl-, sulfo- and carboxy-derivatives of benzoic acid	Yellow, orange, red, purple, brown, green/grey, violet and blue products
Indole + TEMPO + dioxygen over pressure	Trimer 2,2-bis(3'-indolyl)-indoxyl yellow compound
Catechol + polyoxometalate	Dark coloured heterogenic polymers
Ferulic acid + hydro-organic biphasic system	Yellow product (possible food colourants)
3-Methyl-2-benzothiazolinone hydrazone + 3-dimethylaminobenzoic acid	Indamine dye (blue) (for enzymatic activity measuring)
Alkylated pyrogallol derivatives	Benzotropolone structures (orange solids)
3-Methylcatechol + primary linear amines with alkyl chain lengths (C4—C9)	Secondary amines as red solid (pharmaceutically valuable)
Natural phenols (gallic acid + syringic acid; catechin + catechol; ferulic acid + syringic acid)	Yellow, orange, red, purple, brown and black products
Aromatic amine	Binuclear and trinuclear aromatic compounds
ABTS	Cation radical ABTS <sup>•+</sup> dication ABTS <sup>2+</sup> purple and green colouration of fabrics
<b>Phenoxazinone dyes</b> 3-Hydroxyanthranilic acid	Cinnabarinic acid
4-Methyl-3-hydroxyanthranilic acid	2-Amino-4,6-dimethyl-3-phenoxazinone-1,9-carboxylic acid (actinomycin-like cytotoxic derivative)
3-Hydroxyorthanilic acid	2-Amino-3-oxo-3H-phenoxazine-1,9-disulfonic acid water soluble red dye (LAO)
3-Amino-4-hydroxybenzenesulfonic acid	2-Amino-3-oxo-3H-phenoxazine-7-sulfonic acid from yellow to red water soluble phenoxazinone dye
3-Amino-4-hydroxybenzenesulfonic acid	2-Amino-3-oxo-3H-phenoxazine-7-sulfonic acid phenoxazinone dye
2-Amino-3-hydroxy-benzenephosphoric acid	2-Amino-3-oxo-3H-phenoxazine-1,9-diphosphonic acid (aminophenoxazinone water soluble dye)
<b>Azo dyes</b> 3-Methyl-2-benzothiazoline hydrazone + methoxyphenols	Azo dye (for enzymatic activity measuring)
Anthraquinonic acid dye (Acid Blue 62)	Azo dye (Acid Red 1)
Anthraquinonic acid dye (Acid Blue 62)	Azoanthraquinone dye (LAR 1)
4-Methylaminobenzoic acid	Coloured azo intermediates

## 12.5. Biosynthesis of Indigo dyes

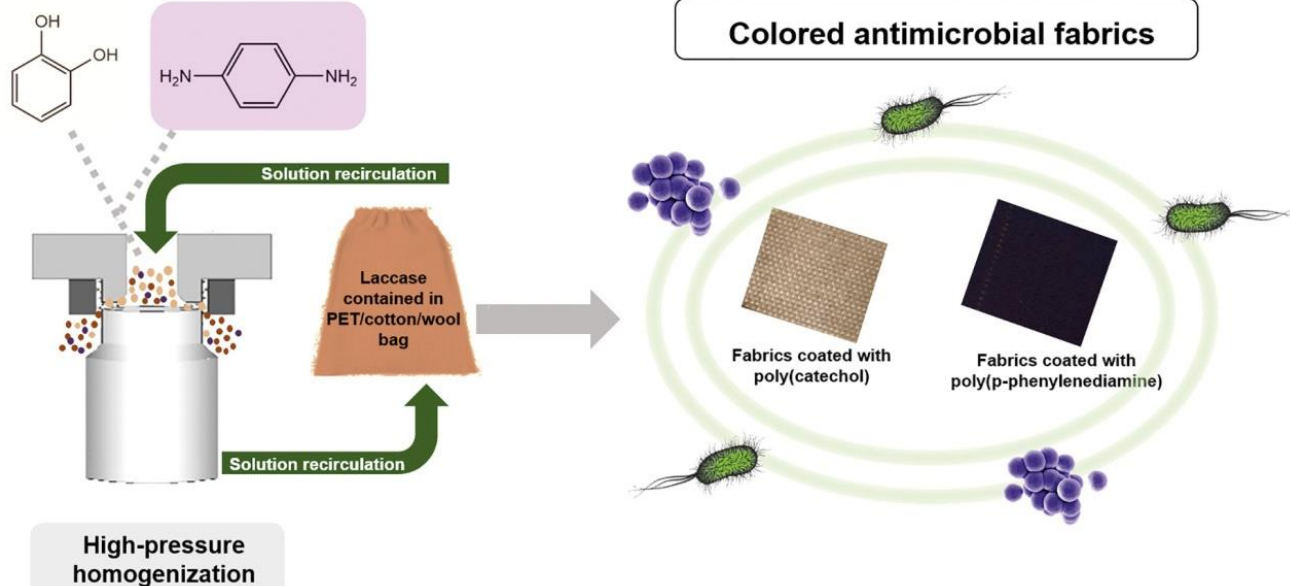
Indigo dyes were first extracted from the plants but this method isn't economic as it gives a limited amount of dye after long extraction time. Synthesis of indigo dye in the lab was the best solution by increasing demand on vat dyes. Indigo

and indirubin are classified carcinogenic compounds besides the last medical reports state that it causes Alzheimer's disease. [99] Successful attempts of producing indigo from the microbial process have been proceeding but they give low dye yield. [100, 101] A promising method was produced using

*Pseudomonas* sp. HOB1 and anionic surfactant and the indigo produced dye used in dyeing cotton. [102]

### 12.6. In situ coloration

Flavonoids such as quercetin, rutin, and morin are colorless compounds that undergo oxidation and polymerization on the surface of natural fibers given colored yellow to brown colored samples. [103] this was applied in textile printing by adding the laccase enzyme, flavanoid as rutin, buffer to the gelatinous thickener. the cotton samples were printed using a screen-printed technique then put in



**Figure 12:** Methodology of preparation of colored antimicrobial fabrics

### 12.7 Application of Enzymes in grafting functional polymer

Producing fabrics with UV protection, antimicrobial, and/or antioxidant activities are an added value to the fabrics. [105]

The phenomena of yellowing of wool by exposure to sunlight have attracted the researchers. [106] Wool fabric was treated in aqueous-ethanol medium 80/20 (v/v%) with aquanordihydroguaiaretic acid (NDGA) in the presence laccase to enhance the formation of inters/intramolecular bonds and cross-links in wool were efficient to protect wool against the undesirable photo yellowing. [107]

laccase-catalyzed grafting of functional molecules on cellulose confers new properties to fiber in an eco-friendly manner. [108]

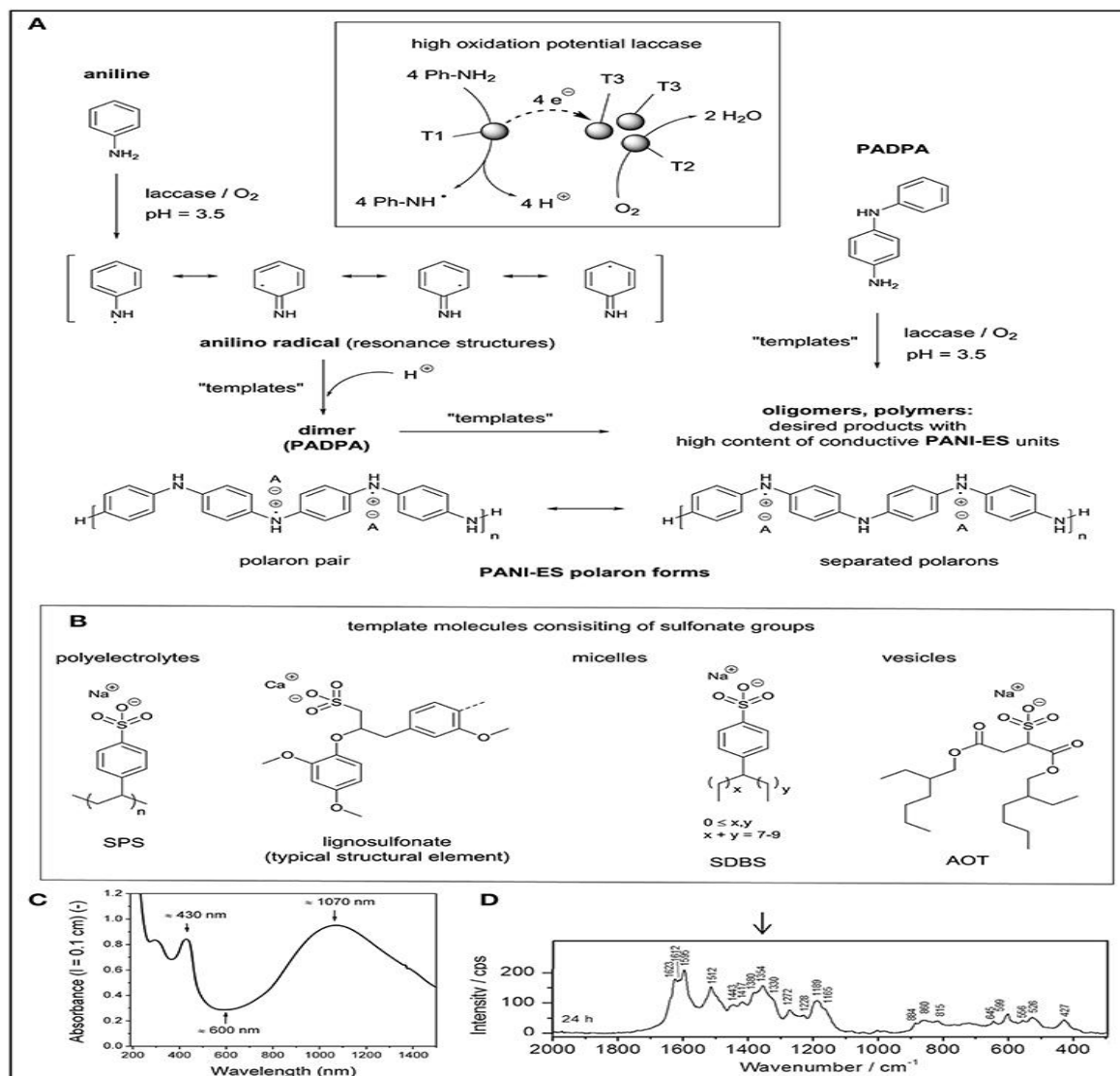
the oven at the optimum temperature and time to have colored samples.

In other research fabrics like cotton, wool, and (PET) undergo polymerization reaction catalysis by laccase under high pressure. The phenol derivatives used were catechol and p-phenylenediamine. **Figure 12** shows a promising methodology approach proposed for *in situ* coloration of fabrics assisted by laccase to produce colored antimicrobial fabrics with enormous potential applications in different fields including medical. [104]

Antibacterial and antioxidant linen fabric was obtained by its pretreatment by laccase enzyme to produce free radical on the fabric surface from the oxidation of nature phenols derivatives then chitosan was added followed by catechin and laccase. [109]

Lipase enzymes assist the synthesis of polyester onto the cotton surface to render it hydrophobic in a novel bio-approach to change the physical properties of cellulosic fabrics. [110]

**Figure 13** shows a suggested mechanism of laccase catalyze synthesis of conductive PANI-ES (A) aniline oxidation follow dimer (PADPA) and polymeric formation. (B) using template molecules containing sulfonate groups (C) IR spectra of the obtained PANI-ES-type polymers. All polymeric products consisting of conductive PANI-ES units that could use in producing conductive textiles. [111, 112]



**Figure 13:** Suggested reaction of synthesis of PANI-ES6 catalyze by laccase

### 13. Future Directions

Further research is needed for:

- Producing of the commercial enzyme such as cutinase, esterase, and polyesterase for the modification of synthetic fibers.
- Search for new enzyme-producing micro-organisms.
- Different immobilized enzymes that could be applicable in either wastewater treatment or fiber treatment.
- One bath treatment of cellulosic fabrics using enzymes will solve almost all the environmental problems associate with the wet process.
- Enzyme assist synthesis of dyes and in-situ coloration needs to get more attention to get it from the lab to the pilot scale.
- The future of textile industries is in the production of colored functional fabrics due to its high price and high demand.
- Producing of biological detergent formula work at low temperature.
- The biosynthesis of indigo dyes is very promising but needs more work on reducing production cost and time and increase the yield.
- The polymeric products obtained from aniline are difficult to isolate to be analyzed despite their promising electrochemical properties. Further researches on PADPA separation, the effect of templates, and its application on textiles are needed.

## 14. References

1. **Salama, M., Hassabo, A.G., El-Sayed, A.A., Salem, T. and Popescu, C.**, Reinforcement of Polypropylene Composites Based on Recycled Wool or Cotton Powders. *Journal of Natural Fibers*, 1-14 (2017)
2. **Elshemy, N.S., Nassar, S.H., El-Taieb, N.M., Shakour, A.A.A., Elmekawy, A.M. and Hassabo, A.G.**, Effect of Different Fabrics Types on the Adsorption of Air Pollution in Residential and Industrial Atmosphere in Cairo-Egypt. *Letters in Applied NanoBioScience*, **9**(4) 682 - 691 (2019)
3. **Haki, G.D. and Rakshit, S.K.**, Developments in Industrially Important Thermostable Enzymes: A Review. *Bioresour Technol*, **89**(1) 17-34 (2003)
4. **Cavaco-Paulo, A. and Gubitz, G.**, Textile Processing with Enzymes. 1<sup>st</sup> Edition ed. USA: Woodhead Publishing (2003)
5. **Ruttloff, H. and Behr's, B.**, Industrielle Enzyme. Hamburg: Verlag GmbH & Co., 97, 81-82 (1995)
6. **Behnke, U., H. Uhlig:** Enzyme Arbeiten Für Uns. Technische Enzyme Und Ihre Anwendung. München, Wien Carl Hanser Verlag, 36, 415-416 (1992)
7. **Boutros, S., Tulio, M., Anderson, M., Rodrigo, D. and Vasco, A.**, Up-to-Date Insight on Industrial Enzymes Applications and Global Market. *Journal of Bioprocessing & Biotechniques*, **4** 1-20 (2012)
8. **Chaplin, M.F. and Bucke, C.**, Enzyme Technology. 1st edn ed. England: Cambridge University Press (1990)
9. **Beilen, J.B.V. and Li, Z.**, Enzyme Technology: An Overview. *Curr Opin Bio-technol.*, **13** 338–344 (2002)
10. **Shaikh, M.A.**, Enzymes: A Revaluation in Textile Processing. *Pakistan Textile Journal*, **59** 48-51 (2010)
11. **Kirk, O., Borchert, T.V. and Fuglsang, C.C.**, Industrial Enzyme Applications. *Current Opinion in Biotechnology*, **13**(4) 345-351 (2002)
12. **Howard, R.L., Abotsi, E., Jansen van Rensburg, E.L. and Howard, S.**, Review: Lignocellulose Biotechnology: Issues of Bioconversion and Enzyme Production. *African Journal of Biotechnology*, **2**(12) 602-619 (2003)
13. **Eijsink, V.G., Vaaje-Kolstad, G., Vårum, K.M. and Horn, S.J.**, Towards New Enzymes for Biofuels: Lessons from Chitinase Research. *Trends Biotechnol*, **26**(5) 228-35 (2008)
14. **Wang, W.-m., Yu, B. and Zhong, C.-j.**, Use of Ultrasonic Energy in the Enzymatic Desizing of Cotton Fabric. *Journal of Cleaner Production*, **33** 179-182 (2012)
15. **Vigneswaran, C.**, Biovision in Textile Wet Processing Industry - Technological Challenges. *Journal of Textile and Apparel, Technology and Management*, **17**(1)(2011)
16. **Teli, M.D. and Adere, T.T.**, Short and Efficient Desizing and Scouring Process of Cotton Textile Materials. *International Journal of Engineering Trends and Technology*, **35**(6) 256 - 268 (2016)
17. **Battan, B., Dhiman, S., Sheoran, S., Mahajan, R. and Sharma, J.**, Application of Thermostable Xylanase of *Bacillus Pumilus* in Textile Processing. *Indian journal of microbiology*, **52** 222-9 (2012)
18. **Kalantzi, S., Mamma, D., Kalogeris, E. and Kekos, D.**, Improved Properties of Cotton Fabrics Treated with Lipase and Its Combination with Pectinase. *Fibres & Textile in Eastern Europe*, **18**(5) 82 - 92 (2010)
19. **Ali, S., Khatri, Z., Khatri, A. and Tanwari, A.**, Integrated Desizing–Bleaching–Reactive Dyeing Process for Cotton Towel Using Glucose Oxidase Enzyme. *Journal of Cleaner Production*, **66**(Complete) 562-567 (2014)
20. **El-Khatib, H.S., Badr, A.A., Diyab, W.A. and Atia, R.M.**, Part I: Enzymatic Treatment of Bamboo, Bamboo/Cotton Knitted Fabric Using Brewer's Yeast Suspension. *Alexandria Engineering Journal*, **58**(2) 819-825 (2019)
21. **El-Hennawi, H., Elshemy, N., Haggage, K., Zaher, A. and Shahin, A.**, Treatment and Optimization of Unconventional Heating to Enhance the Printability of Rami Fabric by Using Brewer's Yeast Enzyme. *Biointerface Research in Applied Chemistry*, **10**(2) 5174-5181 (2020)
22. **Pazarhoğlu, N.K., Sarişik, M. and Telefoncu, A.**, Laccase: Production by *Trametes Versicolor* and Application to Denim Washing. *Process Biochemistry*, **40**(5) 1673-1678 (2005)
23. **Khalil, E. and Sarkar, J.**, Effect of Industrial Bleach Wash and Softening on the Physical, Mechanical and Color Properties of Denim Garments. *IOSR Journal of Polymer and Textile Engineering (IOSR-JPTE)*, **1** 46-49 (2014)
24. **Mozumder, S.**, Effects of Sand Blasting with Industrial Enzyme Silicon Wash on Denim Apparel Characteristics. *Daffodil International University Journal of Science and Technology*, (2010)
25. **L, A. and X, T.**, Textile Bio Processing Using Aloe Gel. *Industrial Engineering & Management*, **06**(02)(2017)
26. **Rahman, M. and Nur, M.**, Feasible Application of Modern Eco-Friendly Treatment of Wool



- Fabric before Coloration. *International Journal of Scientific and Research Publications*, **4**(2014)
27. **Mojsov, K.**, Enzymatic Treatment of Wool Fabrics - Opportunity of the Improvement on Some Physical and Chemical Properties of the Fabrics. *The Journal of The Textile Institute*, **108**(7) 1136-1143 (2017)
  28. **Ammayappan, L. and Moses, J.J.**, Study on Physical and Handle Properties of Wool/Cotton Union Fabric Treated with Enzymes and Different Pre-Polymer Finishing Chemicals. *Current Chemical Research*, **1**(1) 19 - 25 (2011)
  29. **Smith, E. and Shen, J.**, Surface Modification of Wool with Protease Extracted Polypeptides. *Journal of biotechnology*, **156** 134-40 (2011)
  30. **LGulrajani, M., Agarwal, R. and Grover, A.**, Degumming of Silk with Lipase and Protease. *Indian Journal of Fibre & Textile Research*, **25**(3) 69-74 (2000)
  31. **More, S., Khandelwal, H.B., Joseph, M.A. and Laxman, S.**, Enzymatic Degumming of Silk with Microbial Proteases. *Journal of Natural Fibers*, **10**(2013)
  32. **More, S., Chavan, S. and Prabhune, A.**, Silk Degumming and Utilization of Silk Sericin by Hydrolysis Using Alkaline Protease from *Beauveria Sp.* (Mtcc 5184): A Green Approach. *Journal of Natural Fibers*, **15** 1-11 (2017)
  33. **Shetty, P., Madanthyar, B., Ramasubramanian, S., Malickal, S. and Ramachandran, L.**, Pineapple: Potential Source of Proteolytic Enzymes for Degumming of Raw Silk. *Modern Concepts & Developments in Agronomy*, **4**(2019)
  34. **Jordanov, I., Stevens, D.L., Tarbuk, A., Magovac, E., Bischof, S. and Grunlan, J.C.**, Enzymatic Modification of Polyamide for Improving the Conductivity of Water-Based Multilayer Nanocoatings. *ACS omega*, **4**(7) 12028-12035 (2019)
  35. **El-bendary, M.**, Enzymatic Surface Hydrolysis of Polyamide Fabric by Protease Enzyme and Its Production. **37** 273-279 (2012)
  36. **Kanelli, M., Vasilakos, S., Ladas, S., Symianakis, E., Christakopoulos, P. and Topakas, E.**, Surface Modification of Polyamide 6.6 Fibers by Enzymatic Hydrolysis. *Process Biochemistry*, **59** 97-103 (2017)
  37. **Abo El-Ola, S., Moharam, M., Eladwi, M. and El-bendary, M.**, Optimum Conditions for Polyamide Fabric Modification by Protease Enzyme Produced by *Bacillus Sp.* *Indian Journal of Fibre and Textile Research*, **39** 65-71 (2014)
  38. **Schimper, C.B., Ibanescu, C., Keckeis, R. and Bechtold, T.**, Advantages of a Two-Step Enzymatic Process for Cotton-Polyester Blends. *Biotechnol Lett*, **30**(3) 455-9 (2008)
  39. **Nimchua, T., Eveleigh, D.E., Sangwatanaroj, U. and Punnapayak, H.**, Screening of Tropical Fungi Producing Polyethylene Terephthalate-Hydrolyzing Enzyme for Fabric Modification. *J Ind Microbiol Biotechnol*, **35**(8) 843-50 (2008)
  40. **Haji, A., mohammad ali malek, R. and Mazaheri, F.**, Comparative Study of Exhaustion and Pad-Steam Methods for Improvement of Handle, Dye Uptake and Water Absorption of Polyester/Cotton Fabric. *Chemical Industry and Chemical Engineering Quarterly*, **17** 359-365 (2011)
  41. **El-Shemy, N.S. and El-Hawary, N.S.**, Basic and Reactive-Dyeable Polyester Fabrics Using Lipase Enzymes. *Journal of Chemical Engineering & Process Technology*, **07**(01)(2015)
  42. **Kumar, J. and Kumar, M.**, A Study on Improving Dyeability of Polyester Fabric Using Lipase Enzyme. *Autex Research Journal*, (2019)
  43. **Abo-Shosha, M.H., Nassar, F.A., Haggag, K., El-Sayed, Z. and Hassabo, A.G.**, Utilization of Some Fatty Acid/Peg Condensates as Emulsifiers in Kerosene Paste Pigment Printing. *Research Journal of Textile and Apparel*, **13**(1) 65-77 (2009)
  44. **Hassabo, A.G., Mendrek, A., Popescu, C., Keul, H. and Möller, M.**, Deposition of Functionalized Polyethylenimine-Dye onto Cotton and Wool Fibres. *Research Journal of Textile and Apparel*, **18**(1) 36-49 (2014)
  45. **Hassabo, A.G., Erberich, M., Popescu, C. and Keul, H.**, Functional Polyethers for Fixing Pigments on Cotton and Wool Fibres. *Research & Reviews in Polymer*, **6**(3) 118-131 (2015)
  46. **Hebeish, A., Shaarawy, S., Hassabo, A.G. and El-Shafei, A.**, Eco-Friendly Multifinishing of Cotton through Inclusion of Motmorillonite/Chitosan Hybrid Nanocomposite. *Der Pharma Chemica*, **8**(20) 259-271 (2016)
  47. **Aboelnaga, A., Shaarawy, S. and Hassabo, A.G.**, Polyacetic Acid/Functional Amine/Azo Dye Composite as a Novel Hyper-Branched Polymer for Cotton Fabric Functionalization. *Colloids and Surfaces B: Biointerfaces*, **172** 545-554 (2018)
  48. **Öner, E. and Sahinbaskan, B.Y.**, A New Process of Combined Pretreatment and Dyeing: Rest. *Journal of Cleaner Production*, **19**(14) 1668-1675 (2011)
  49. **Benli, H. and Bahtiyari, M.**, Use of Ultrasound in Biopreparation and Natural Dyeing of Cotton Fabric in a Single Bath. *Cellulose*, **22**(2015)

50. **Rasel, M., Ahmed, M. and Farjana, S.**, Modification of Reduction Clearing Process of Polyester Blend Cotton Knitted Fabric Dyeing. *volume 9* 101-106 (2018)
51. **Ahmed, M. and An, S.**, Efficient Dyeing Mechanism of Cotton/Polyester Blend Knitted Fabric. *Fibers and Polymers*, **19** 2541-2547 (2018)
52. **Javoršek, D. and Javoršek, A.**, Colour Management in Digital Textile Printing. **127**(4) 235-239 (2011)
53. **El-Hennawi, H.M., Shahin, A.A., Rekaby, M. and Ragheb, A.A.**, Ink Jet Printing of Bio-Treated Linen, Polyester Fabrics and Their Blend. *Carbohydrate Polymers*, **118** 235-241 (2015)
54. **Hassabo, A.G.**, Preparation, Characterisation and Utilization of Some Textile Auxiliaries, El-Azhar University: Cairo, Egypt (2005)
55. **El-Zawahry, M.M., Abdelghaffar, F., Abdelghaffar, R.A. and Hassabo, A.G.**, Equilibrium and Kinetic Models on the Adsorption of Reactive Black 5 from Aqueous Solution Using Eichhornia Crassipes/Chitosan Composite. *Carbohydrate Polymers*, **136** 507-515 (2016)
56. **Hassabo, A.G. and Mohamed, A.L.**, Multiamine Modified Chitosan for Removal Metal Ions from Their Aqueous Solution *BioTechnology: An Indian Journal*, **12**(2) 59-69 (2016)
57. **Jamee, R. and Siddique, R.**, Biodegradation of Synthetic Dyes of Textile Effluent by Microorganisms: An Environmentally and Economically Sustainable Approach. *European journal of microbiology & immunology*, **9**(4) 114-118 (2019)
58. **Ray, J., Jana, S., Mondal, B. and Tripathy, T.**, Enhanced and Rapid Adsorptive Removal of Toxic Organic Dyes from Aqueous Solution Using a Nanocomposite of Saponified Polymethyl Acrylate Grafted Dextrin with Embedded Nanosilica. *Journal of Molecular Liquids*, **275** 879-894 (2019)
59. **Li, H., Wu, S., Du, C., Zhong, Y. and Yang, C.**, Preparation, Performances, and Mechanisms of Microbial Flocculants for Wastewater Treatment. *International journal of environmental research and public health*, **17**(4) 1360 (2020)
60. **Bento, R.M.F., Almeida, M.R., Bharmoria, P., Freire, M.G. and Tavares, A.P.M.**, Improvements in the Enzymatic Degradation of Textile Dyes Using Ionic-Liquid-Based Surfactants. *Separation and Purification Technology*, **235**(2020)
61. **Forootanfar, H., Moezzi, A., Aghaie-Khozani, M., Mahmoudjanlou, Y., Ameri, A., Niknejad, F. and Faramarzi, M.A.**, Synthetic Dye Decolorization by Three Sources of Fungal Laccase. *Iranian J Environ Health Sci Eng*, **9**(1) 27 (2012)
62. **Pramanik, S. and Chaudhuri, S.**, Laccase Activity and Azo Dye Decolorization Potential of *Podoscypha Elegans*. *Mycobiology*, **46**(1) 79-83 (2018)
63. **Vidya, S., Chinchu, C. and Meera, B.S.**, Dye Decolourization Using Fungal Laccase: A Review. *International Journal of Innovations in Engineering and Technology*, **8**(1) 118 - 123 (2017)
64. **Chapman, J., Ismail, A. and Dinu, C.**, Industrial Applications of Enzymes: Recent Advances, Techniques, and Outlooks. *Catalysts*, **8**(6) 238-264 (2018)
65. **Wehaidy, H.R., Abdel-Naby, M.A., El-Hennawi, H.M. and Youssef, H.F.**, Nanoporous Zeolite-X as a New Carrier for Laccase Immobilization and Its Application in Dyes Decolorization. *Biocatalysis and Agricultural Biotechnology*, **19** 101135 (2019)
66. **Cristóvão, R.O., Tavares, A.P.M., Loureiro, J.M., Boaventura, R.A.R. and Macedo, E.A.**, Treatment and Kinetic Modelling of a Simulated Dye House Effluent by Enzymatic Catalysis. *Bioresource Technology*, **100**(24) 6236-6242 (2009)
67. **Maurer, K.H.**, Detergent Proteases. *Curr Opin Biotechnol*, **15**(4) 330-4 (2004)
68. **Gurkok, S.**, Microbial Enzymes in Detergents: A Review Sumeyra Gürkök. *International Journal of Scientific and Engineering Research*, **10** 75-81 (2019)
69. **Saeki, K., Ozaki, K., Kobayashi, T. and Ito, S.**, Detergent Alkaline Proteases: Enzymatic Properties, Genes, and Crystal Structures. *Journal of Bioscience and Bioengineering*, **103**(6) 501-508 (2007)
70. **Hasan, F., Shah, A.A., Javed, S. and Hameed, A.**, Enzymes Used in Detergents: Lipases. *African Journal of Biotechnology*, **9**(31) 4836-4844 (2010)
71. **Madhu, A. and Chakraborty, J.**, Developments in Application of Enzymes for Textile Processing. *Journal of Cleaner Production*, **145**(2017)
72. **M. M. Elnashar, M.**, Review Article: Immobilized Molecules Using Biomaterials and Nanobiotechnology. **01**(01) 61-77 (2010)
73. **Hemalatha, V., Kalyani, P., Chandana Vineela, K. and Hemalatha, K.P.J.**, Methods, Applications of Immobilized Enzymes-a Mini Review. *International Journal of Engineering*

- Sciences & Research Technology*, **5**(11) 523 - 526 (2016)
74. **Kiehl, K., Straube, T., Opwis, K. and Gutmann, J.S.**, Strategies for Permanent Immobilization of Enzymes on Textile Carriers. *Engineering in Life Sciences*, **15**(6) 622-626 (2015)
  75. **Silva, C., Martins, M., Jing, S., Fu, J. and Cavaco-Paulo, A.**, Practical Insights on Enzyme Stabilization. *Critical Reviews in Biotechnology*, **38**(3) 335-350 (2018)
  76. **Karthikeyan, K. and Dhurai, B.**, New Method of Discharge Printing on Cotton Fabrics Using Horseradish Peroxidase. *Autex Research Journal*, **11** 61-65 (2011)
  77. **Ragheb, A.A., Haggag, K., Rekaby, M., Abd El-Thalouth, I., El-Hennawi, H.M. and Shahin, A.A.**, Bio-Discharge Printing on Cotton Knitted Fabrics Using Enzyme and Brewers Yeast. *Journal of Applied Sciences Research*, **9**(1) 205-225 (2013)
  78. **Rafat, B., Haggag, K., Abd El-Thalouth, J.I., Ragheb, A.A. and El-Shemi, N.**, Industrial Scale Production of Resist / Discharge Printed Cotton Knitted Garments Using Bio-Technique. *Journal of Applied Sciences Research*, **9**(1) 163 - 169 (2013)
  79. **Thalouth, J.I.A.E., Tawfik, S., Ragheb, A.A. and Mosaad, M.M.**, Technological Evaluation of Laccase Enzyme in Discharge Printing Using Natural Colours. *International Journal of Science and Research*, **4**(9) 501-509 (2015)
  80. **Polak, J. and Jarosz-Wilkolazka, A.**, Fungal Laccases as Green Catalysts for Dye Synthesis. *PROCESS BIOCHEMISTRY*, **47** 1295-1307 (2012)
  81. **Reiss, R., Ihssen, J., Richter, M., Eichhorn, E., Schilling, B. and Thöny-Meyer, L.**, Laccase Versus Laccase-Like Multi-Copper Oxidase: A Comparative Study of Similar Enzymes with Diverse Substrate Spectra. *PLoS ONE*, **8**(6) e65633 (2013)
  82. **Yin, Q., Zhou, G., Peng, C., Zhang, Y., Kües, U., Liu, J., Xiao, Y. and Fang, Z.**, The First Fungal Laccase with an Alkaline Ph Optimum Obtained by Directed Evolution and Its Application in Indigo Dye Decolorization. *AMB Express*, **9**(1)(2019)
  83. **Mandic, M., Djokic, L., Nikolaivits, E., Prodanovic, R., O'Connor, K., Jeremic, S., Topakas, E. and Nikodinovic-Runic, J.**, Identification and Characterization of New Laccase Biocatalysts from *Pseudomonas* Species Suitable for Degradation of Synthetic Textile Dyes. *Catalysts*, **9**(7) 629 (2019)
  84. **Prajapati, C.D., Smith, E., Kaneb, F. and Shen, J.**, Laccase-Catalysed Coloration of Wool and Nylon. *Coloration Technology*, **134** 423-439 (2018)
  85. **Zerva, A., Simić, S., Topakas, E. and Nikodinovic-Runic, J.**, Applications of Microbial Laccases: Patent Review of the Past Decade (2009–2019). *Catalysts*, **9**(12) 1023 (2019)
  86. **Patel, N., Shahane, S., Shivam, Majumdar, R. and Mishra, U.**, Mode of Action, Properties, Production, and Application of Laccase: A Review. *Recent Pat Biotechnol*, **13**(1) 19-32 (2019)
  87. **Polak, J. and Jarosz-Wilkolazka, A.**, Structure/Redox Potential Relationship of Simple Organic Compounds as Potential Precursors of Dyes for Laccase-Mediated Transformation. *Biotechnology Progress*, **28**(1) 93-102 (2012)
  88. **Strong, J. and Claus, H.**, Laccase: A Review of Its Past and Its Future in Bioremediation. *Critical Reviews in Environmental Science and Technology - CRIT REV ENVIRON SCI TECHNOL*, **41** 373-434 (2011)
  89. **Polak, J. and Jarosz-Wilkolazka, A.**, Whole-Cell Fungal Transformation of Precursors into Dyes. **9**(1) 51 (2010)
  90. **Kim, S., Silva, C., Evtuguin, D.V., Gamelas, J.A. and Cavaco-Paulo, A.**, Polyoxometalate/Laccase-Mediated Oxidative Polymerization of Catechol for Textile Dyeing. *Appl Microbiol Biotechnol*, **89**(4) 981-7 (2011)
  91. **Baisch, G., Wagner, B. and Oehrlein, R.**, An Efficient Chemo-Enzymatic Approach Towards Variably Functionalized Benzotropolones. *Tetrahedron*, **66** 3742-3748 (2010)
  92. **Herter, S., Mikolasch, A., Michalik, D., Hammer, E., Schauer, F., Bornscheuer, U. and Schmidt, M.**, C–N Coupling of 3-Methylcatechol with Primary Amines Using Native and Recombinant Laccases from *Trametes Versicolor* and *Pycnoporus Cinnabarinus*. *Tetrahedron*, **67**(48) 9311-9321 (2011)
  93. **Jeon, J.-R., Kim, E.-J., Murugesan, K., Park, H.-K., Kim, Y.-M., Kwon, J.-H., Kim, W.-G., Lee, J.-Y. and Chang, Y.-S.**, Laccase-Catalysed Polymeric Dye Synthesis from Plant-Derived Phenols for Potential Application in Hair Dyeing: Enzymatic Colourations Driven by Homo- or Hetero-Polymer Synthesis. **3**(3) 324-335 (2010)
  94. **Sousa, A.C., Martins, L.O. and Robalo, M.P.**, Laccase-Catalysed Homocoupling of Primary Aromatic Amines Towards the Biosynthesis of

- Dyes. *Advanced Synthesis & Catalysis*, **355**(14-15) 2908-2917 (2013)
95. **Bruyneel, F., D'Auria, L., Payen, O., Courtoy, P.J. and Marchand-Brynaert, J.**, Live-Cell Imaging with Water-Soluble Aminophenoxazinone Dyes Synthesised through Laccase Biocatalysis. *Chembiochem : a European journal of chemical biology*, **11**(10) 1451-1457 (2010)
96. **Forte, S., Polak, J., Valensin, D., Taddei, M., Basosi, R., Vanhulle, S., Jarosz-Wilkolazka, A. and Pogni, R.**, Synthesis and Structural Characterization of a Novel Phenoxazinone Dye by Use of a Fungal Laccase. *Journal of Molecular Catalysis B: Enzymatic*, **63**(3) 116-120 (2010)
97. **Enaud, E., Trovaslet, M., Bruyneel, F., Billottet, L., Karaaslan, R., Sener, M.E., Coppens, P., Casas, A.R., Jäger, I., Hafner, C., Onderwater, R.C.A., Corbisier, A.-M., Marchand-Brynaert, J., Vanhulle, S.J.D. and Pigments, A** Novel Azoanthraquinone Dye Made through Innovative Enzymatic Process. **85** 99-108 (2010)
98. **Martorana, A., Bernini, C., Valensin, D., Sinicropi, A., Pogni, R., Basosi, R. and Baratto, M.C.**, Insights into the Homocoupling Reaction of 4-Methylamino Benzoic Acid Mediated by *Trametes Versicolor* Laccase. **7**(11) 2967 (2011)
99. **Xiao, Z., Hao, Y., Liu, B. and Qian, L.**, Indirubin and Meisoindigo in the Treatment of Chronic Myelogenous Leukemia in China. *Leukemia & lymphoma*, **43** 1763-8 (2002)
100. **Wackett, L.**, Mechanism and Applications of Rieske Non-Heme Iron Dioxygenases. *Enzyme and Microbial Technology*, **31** 577-587 (2002)
101. **Parales, R.E., Lee, K., Resnick, S.M., Jiang, H., Lessner, D.J. and Gibson, D.T.**, Substrate Specificity of Naphthalene Dioxygenase: Effect of Specific Amino Acids at the Active Site of the Enzyme. *Journal of bacteriology*, **182**(6) 1641-1649 (2000)
102. **Pathak, H. and Madamwar, D.**, Biosynthesis of Indigo Dye by Newly Isolated Naphthalene-Degrading Strain *Pseudomonas* Sp. Hob1 and Its Application in Dyeing Cotton Fabric. *Appl Biochem Biotechnol*, **160**(6) 1616-26 (2010)
103. **El-Hennawi, H.M., Ahmed, K.A. and Abd El-Thalouth, I.**, Novel Bio-Technique Using Laccase Enzyme in Textile Printing to Fix Natural Dyes. *Indian Journal of Fibre and Textile Research*, **37** 245 -249 (2012)
104. **Su, J., Noro, J., Silva, S., Fu, J., Wang, Q., Ribeiro, A., Silva, C. and Cavaco-Paulo, A.**, Antimicrobial Coating of Textiles by Laccase in Situ Polymerization of Catechol and P-Phenylenediamine. *Reactive and Functional Polymers*, **136** 25-33 (2019)
105. **Alebeid, O.K. and Zhao, T.**, Review On: Developing Uv Protection for Cotton Fabric. *The Journal of The Textile Institute*, **108**(12) 2027-2039 (2017)
106. **Millington, K.R.**, Photoyellowing of Wool. Part 1: Factors Affecting Photoyellowing and Experimental Techniques. *Coloration Technology*, **122**(4) 169-186 (2006)
107. **Hossain, K., González, M., Juan, A. and Tzanov, T.**, Enzyme-Mediated Coupling of a Bi-Functional Phenolic Compound onto Wool to Enhance Its Physical, Mechanical and Functional Properties. *Enzyme and Microbial Technology*, **46** 326-330 (2010)
108. **Javaid, R., Sabir, A., Sheikh, N. and Ferhan, M.**, Recent Advances in Applications of Acidophilic Fungi to Produce Chemicals. *Molecules*, **24**(4) 786 (2019)
109. **Silva, C., Matamá, T., Kim, S., Padrão, J., Nugroho Prasetyo, E., Kudanga, T., Nyanhongo, G.S., Guebitz, G.M., Casal, M. and Cavaco-Paulo, A.**, Antimicrobial and Antioxidant Linen Via Laccase-Assisted Grafting. *Reactive and Functional Polymers*, **71**(7) 713-720 (2011)
110. **Zhao, X., Noro, J., Fu, J., Wang, H., Silva, C. and Cavaco-Paulo, A.**, " In-Situ " Lipase-Catalyzed Cotton Coating with Polyesters from Ethylene Glycol and Glycerol. *Process Biochemistry*, **66** 82-88 (2018)
111. **Kashima, K., Fujisaki, T., Serrano-Luginbühl, S., Kissner, R., Janošević, A., Bajuk-Bogdanović, D., Cirić-Marjanović, G., Busato, S., Ishikawa, T. and Peter, W.**, Effect of Template Type on the *Trametes Versicolor* Laccase-Catalyzed Oligomerization of the Aniline Dimer P -Aminodiphenylamine (Padpa). *ACS Omega*, **4** 2931-2947 (2019)
112. **Su, J., Noro, J., Fu, J., Wang, Q., Silva, C. and Cavaco-Paulo, A.**, Coloured and Low Conductive Fabrics by in Situ Laccase-Catalysed Polymerization. *Process Biochemistry*, **77** 77-84 (2019)