

Functional attributes of ethnically edible ectomycorrhizal wild mushroom *Amanita* in India

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Abstract

Functional qualities serve as valuable yardsticks towards consumer acceptability of foods and food products. This study evaluates functional properties of an ectomycorrhizal wild mushroom *Amanita* sp. occurring in the lateritic scrub jungles of southwestern India. Based on the ethnic knowledge, immature cooked fruit bodies of this mushroom are edible. Standard protocols were followed to evaluate functional properties of uncooked and cooked immature fruit bodies (pH-dependent protein solubility; least gelation concentration; water- and oil-absorption capacities; emulsion and foam properties). The protein solubility was significantly higher in uncooked against cooked samples (pH 2-8, $p < 0.05$). Cooking has not altered the least gelation concentration (14%). There was no significant difference between uncooked and cooked samples ($p > 0.05$) despite water-absorption and oil-absorption capacities were higher in cooked samples. The emulsion activity ($p < 0.05$), emulsion stability ($p < 0.05$), foam capacity ($p < 0.01$) and foam stability ($p > 0.05$) were higher in cooked than uncooked samples. The Principal Component Analysis (PCA) between proximal and functional properties reveals that the crude protein, total lipids and crude fibre influenced the protein-solubility, emulsion stability and foam capacity in uncooked samples. In cooked samples, the crude fibre and carbohydrates influenced all the functional properties studied except for protein solubility. It is assumed that the composition and proportion of proximal components influence the functional attributes of *Amanita* sp. The properties like high emulsion activity, emulsion stability and foam capacity in cooked mushroom will be useful in formulation of value-added foods or nutraceutical products.

Key words – Scrub jungles, macrofungi, ethnic food, proximal qualities, nutraceutical source.

Introduction

Functional properties characterize the structural quality, nutritional value and acceptability of food or food products. Generally, food stuffs show several functional properties like hydration properties (protein solubility, protein dispensability, water- and fat- holding capacities), surface properties (emulsion and foam), structural and textural properties (viscosity, gelation and visco-elasticity). The proximal constituents of food (proteins, carbohydrates, fats and fibre) undergo several changes during processing leading to favourable transformation in form, texture and taste of food or food product. Proteins in foods could be enzymatically modified by controlled proteolysis, which may enhance their functional properties over a wide range of pH, ionic concentration and other processing conditions (Panyam and Kilara 1996).

Similarly, water-holding capacity describes the ability in matrix of molecule and physically entrapped water does not flow from tissue of foods when they are cut or minced (Damodaran 1996). Foaming potential contributes to smoothness, lightness, flavour dispersions and palatability of food, while foam formation by proteins in solutions is desirable in many food applications.

A wide variety of edible macrofungi or mushrooms are rich in dietary proteins (Manjunatha *et al.* 2011; Wani *et al.* 2010). They possess several essential amino acids required for children and adults (Bernas *et al.* 2006; Afiukwa *et al.* 2015). Aremu *et al.* (2009) demonstrated that mushroom flours (e.g. *Ganoderma* spp., *Omphalotus olearius* and *Hebeloma mesophaeum*) possess properties useful in the formulation of different food products, where gelling, foaming, emulsification, flavour retention are required for desired palatability. The composition of proteins, carbohydrates, fat and fibre contents in mushrooms make them ideal non-conventional food source for diabetic, cancer and cardiac patients (Usha and Suguna 2014). Edible mushrooms could also be used as food to combat protein malnutrition owing to their capability to convert agriculture wastes into nutritionally rich food source (Lelley 1987).

Among several edible wild mushrooms available in the southwestern India, young fruit bodies of *Amanita* sp. constitute an ethnic food source (Karun and Sridhar 2014). It is an ectomycorrhizal fungus associated with several tree species (e.g. *Acacia* spp., *Anacardium occidentale*, *Hopea ponga* and *Terminalia paniculata*). The local people harvest tender fruit bodies of this mushroom (spherical, beak, dumble and partially opened volva) during early monsoon season (June-July) for consumption. Tender sporocarps of *Amanita* sp. is known for high quantities of proteins and fibre; moderate amount of carbohydrates; low total lipid content; adequate quantity of iron; favourable Na-K ratio (<1); sufficient quantities of essential amino acids; high *in vitro* protein digestibility; endowed with many bioactive components; possess potential antioxidant activities (Greeshma *et al.* 2018a, b). Hence, this study intends to fill the gap on functional qualities of uncooked and cooked tender sporocarps of *Amanita* sp. sampled from the lateritic scrub jungles of southwestern India to utilize for food formulations and nutraceutical products.

Materials and Methods

Mushroom

Developing sporocarps of *Amanita* sp. were sampled from the lateritic scrub jungles of Konaje Village (Dakshina Kannada, Mangalore, India: 12°48'N, 74°55'E; 115 m asl) in consultation with the local dwellers who has experience to collect during monsoon season (June-August 2016) (Fig. 1). Being ectomycorrhizal, its fruit bodies are common below the canopies of several tree species (*Acacia auriculiformis*, *A. mangium*, *Anacardium occidentale*, *Hopea ponga* and *Terminalia paniculata*). Based on the morphological features, this new *Amanita* species roughly matches with that of *Amanita marmorata* (Cleland & E.-J. Glibert) E.-J. Gilbert reported from Hawaii (Miller *et al.* 1996). The tender sporocarps with different shapes (spherical, oval and dumble-shaped) and partially ruptured (Fig. 1a-c) are utilized for consumption purpose by local people. Samples were made from five locations of scrub jungles with about 50 m apart served as replicates. The young sporocarps were rinsed in distilled water to eliminate debris and wiped with paper towel to remove surface water. Each replicate was divided into two parts and the first part was oven dried (50-55°C), while the second part was cooked in a pressure-cooker with distilled water (1:1 v/v) followed by oven drying. The dried samples were milled (mesh #30) and flours were refrigerated in air-tight glass containers assess functional properties.



Fig. 1- Immature (a-c) (suitable for consumption), maturing (d, e) and mature (f, g) stages *Amanita* sp. seen in the scrub jungles of southwestern India.

Protein solubility

The protein solubility (PS) of uncooked and cooked *Amanita* sp. was evaluated by following the method proposed by Were *et al.* (1997). The flour suspension (0.5%) in distilled water was prepared by blending and adjusting pH ranging from 2-10 with HCl (1M) and NaOH (1M). The suspension was mixed using magnetic stirrer ($20\pm 2^{\circ}\text{C}$, 1 h). The contents were centrifuged (12,000 rpm, 4°C , 20 min) and the supernatant was filtered (Whatman # 1). The amount of nitrogen in filtrate was assessed by the micro-Kjedahl method (Humphries 1956). The percentage of soluble protein was calculated as percent nitrogen in sample by multiplying by 6.25.

Gelation

The least gelation concentration (LGC) of the uncooked and cooked mushroom was estimated based on method outlined by Coffman and Garcia (1977). Slurry of mushroom flour ranging from 2-20% (w/v) was prepared. An aliquot (10 ml) was dispensed into the test tubes and incubated in boiling water bath (1 h) followed by cooling (4°C, 2 h). The LGC was ascertained at the concentration where the sample did not slip on inverting the test tube in five replicates.

Water- and oil-absorption

The procedure proposed by Beuchant (1977) was followed to determine the water-absorption (WA) and oil-absorption (OA) capacities of uncooked and cooked mushroom. The flour (1 g) was vortexed with distilled water (10 ml) and incubated at room temperature (30 min). The contents were centrifuged (5000 rpm, 30 min) and the volume of supernatant was measured. To determine OA, the flour (1 g) was homogenised with oil (10 ml) and processed as described for WA. The WA and OA capacities are expressed as ml water or oil absorbed per gram of mushroom flour, respectively.

Emulsion properties

The method followed by Neto *et al.* (2001) was adapted to determine emulsion properties. For emulsion activity (EA), the mushroom flour (50 mg) was suspended in distilled water (5 ml) and vortexed with edible oil (5 ml). The contents were centrifuged (1100 rpm, 5 min). The height of the emulsified layer was measured to calculate the EA.

$$\text{Emulsion activity (EA) (\%)} = \left(\frac{\text{Emulsified layer in ml}}{\text{Total content in ml}} \right) \times 100$$

To determine emulsion stability (ES), the flour suspension was heated in water bath (80°C, 30 min) before centrifugation. The height of emulsified layer was noted to calculate the ES.

$$\text{Emulsion stability (ES) (\%)} = \left(\frac{\text{Emulsified layer after heating in ml}}{\text{Total content in ml}} \right) \times 100$$

Foam properties

The foam properties were determined according to Coffman and Garcia (1977). For foam capacity (FC), the mushroom flour (2%, w/v) in distilled water was whipped vigorously (2 min) in blender and poured into the measuring jar. The volume was recorded before and after homogenization to calculate the FC.

$$\text{Foam capacity (FC) (\%)} = \left(\frac{\text{Volume after whipping in ml} - \text{Volume before whipping in ml}}{\text{Volume before whipping in ml}} \right) \times 100$$

The foam stability (FS) was determined by recording the volume of foam remained after incubation up to 8 h at room temperature to calculate FS.

$$\text{Foam stability (FS) (\%)} = \left(\frac{\text{Foam volume after 8 h in ml}}{\text{Initial foam volume in ml}} \right) \times 100$$

Data analysis

Student *t*-test was employed to ascertain the difference in functional properties between uncooked and cooked mushroom samples by using Statistica version 8.0 (StatSoft Inc. 2008). To find out the relationship between the functional properties with those of proximal attributes (Greeshma *et al.* 2018a), the principal component analysis (PCA) was performed for uncooked and cooked mushroom samples separately (SPSS version 16.0: www.spss.com).

Result and Discussion

Protein solubility

The pH-dependent protein solubility (PS) curves were followed similar pattern in uncooked and cooked mushroom samples (Fig. 2a). The solubility was higher in uncooked than cooked samples throughout the range of pH studied: pH 2, pH 4 ($p < 0.01$), pH 6 ($p < 0.05$), pH 8 ($p < 0.01$) and pH 10 ($p > 0.05$). In uncooked samples, the protein solubility was highest at pH 4 followed by pH 10, while it was highest at pH 10 followed by pH 4 in cooked samples. Generally, protein solubility decreases with increase of pH till it attains isoelectric point and thereafter increases progressively with further raise in pH (Adebowale *et al.* 2005). The uncooked *Amanita* sp. in our study showed considerable protein solubility at acidic (pH 4, 32%) as well as alkaline (pH 10, 27%) range, which is useful in production of value-added protein-rich carbonated beverages as well as infant foods (Idouraine *et al.* 1997; Fasuyi and Aleton 2005).

Gelation

Gelation of foodstuff is one of the important functional properties to design foodstuffs with desired texture. Lower the level of the least gelation concentration (LGC) higher the gelling ability of the protein ingredient (Akintayo *et al.* 2002). The LGC of uncooked as well as cooked mushroom was constant (14%) (Fig. 2b). The LGC of *Amanita* sp. is comparable to *Ganoderma* spp. (14%), lower than *Astraeus hygrometricus* (18-20%) and higher than other mushrooms (*Omphalotus olearius* and *Hebeloma mesophaeum*: 12%) (Aremu *et al.* 2009; Pavithra *et al.* 2017). The LGC of composite flours consisting wheat + mushroom + tapioca was as low as 2-3% (Ekunseitan *et al.* 2016). It is likely the LGC of *Amanita* sp. could be further decreased by mixing its flour with other suitable cereal or legume flour for production of quality foodstuffs.

Water- and oil-absorption

Water-absorption (WA) capacity represents the ability of a product to associate with water under the conditions of water limitation (Singh 2001). It will be usually dependent on starch and fibre contents in the given mushroom flour. The WA ranged from 1.8-3.2 ml g⁻¹ without significant difference between uncooked and cooked *Amanita* sp. ($p > 0.05$) (Fig. 2c). The WA of *Amanita* sp. could be comparable to *Astraeus hygrometricus* (2.2-3.6 ml g⁻¹) and *Pleurotus sajor-caju* (2.4 ml g⁻¹) (Prodhan *et al.* 2015; Pavithra *et al.* 2017). The oil-absorption capacity (OA) was higher in uncooked than cooked samples *Amanita* sp. (2.6 vs. 2.4 ml g⁻¹) without significant difference ($p > 0.05$) (Fig. 2d). The OA capacity *Amanita* sp. is lower compared to *A. hygrometricus* (1.9-2 ml g⁻¹) (Pavithra *et al.* 2017). The WA and OA capacities of protein in foodstuffs are dependent on many intrinsic factors especially amino acid composition, protein conformation and surface polarity (Suresh and Samsheer 2013). The WA and OA capacities influence certain characteristics such as texture and mouth feel of foods (e.g. meat formulations, doughnuts and baked dough) (Alobo 2003). The extent of WA and OA capacities of *Amanita* sp. is favourable in production of food products like soups and baked products.

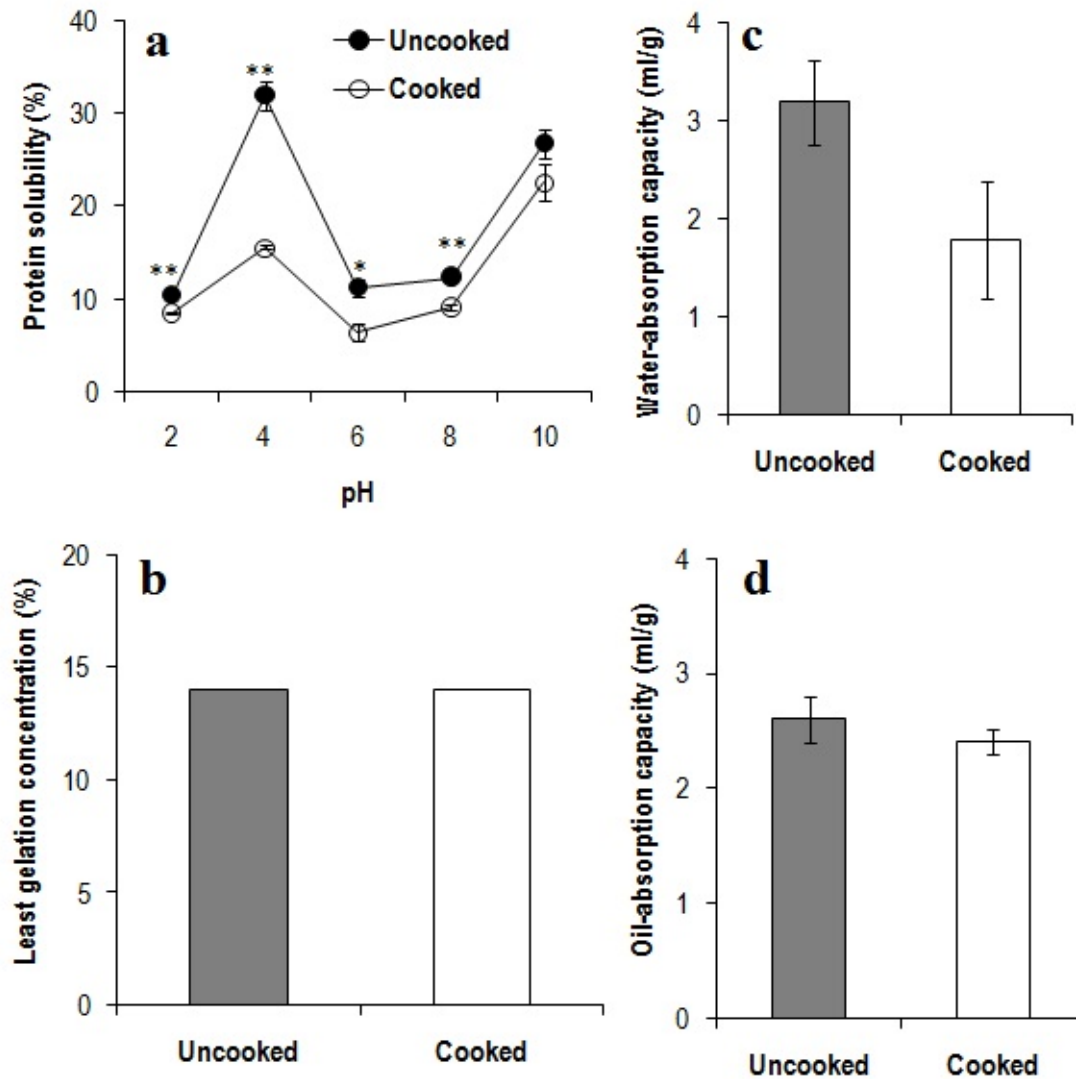


Fig. 2- Protein solubility (a), least gelation concentration (b), water-absorption capacity (c) and oil-absorption capacity (d) of uncooked and cooked tender fruit bodies of *Amanita* sp. (n=5±SD; t-test: *, p<0.05; **, p<0.01).

Emulsion properties

The emulsion activity is the ability of protein in foodstuffs responsible for emulsion formation as well as stability of newly formed emulsion. The emulsion capacity depends on the shape, charge and hydrophobicity of the protein molecules, neutrality of dipoles and hydration of polar groups (Zayas 1997a). The emulsion activity (EA: 12. vs. 18.4%) (Fig. 3a) and emulsion stability (ES: 10.3 vs. 14.5%) in our study were significantly higher in cooked than uncooked mushrooms (p<0.05) (Fig. 3a, b). The EA and ES are lower compared to *Astraeus hygrometricus* (Pavithra *et al.* 2017). The higher EA and ES in cooked *Amanita* sp. could be attributed to the higher concentration of protein in cooked than uncooked sample (Greeshma *et al.* 2018a). However, the EA and ES of *Amanita* sp. are lower than EA (47.1%) and ES (52.2%) of mushroom flours produced by the National Research Centre for Mushroom, Himachal Pradesh, India (Prajapati *et al.* 2015).

Foam properties

Foam is a two-phase system consisting of air cells separated by a thin continuous liquid layer (Zayas 1997b). The optimum foam formation of a material depends on speed of rotation, interval of stirring and pH (Gassmann *et al.* 1987). As seen in emulsion properties, the foam capacity (FC) and foam stability (FS) (Fig. 3c, d) of *Amanita* sp. was higher in cooked than uncooked samples (FC, $p < 0.01$; FS, $p > 0.05$). The FC (7.4-9.3%) and FS (7.6-8.6%) of *Amanita* sp. are comparable to *Astraeus hygrometricus* (Pavithra *et al.* 2017), but lower than *Armillaria mellea*, *Ganoderma* spp., *Hebeloma mesophaeum*, *Omphalotus oleariu*, *Termitomyces heimii* and *Volvariella volvacea* (21.6%, 91.7%) (Aremu *et al.* 2009; Due *et al.* 2016a, b). The extent of FC and FS of uncooked *Amanita* sp. could be useful in developing formulations mainly frozen desserts, cakes, whipped toppings and ice-cream mixes (Niveditha and Sridhar 2017).

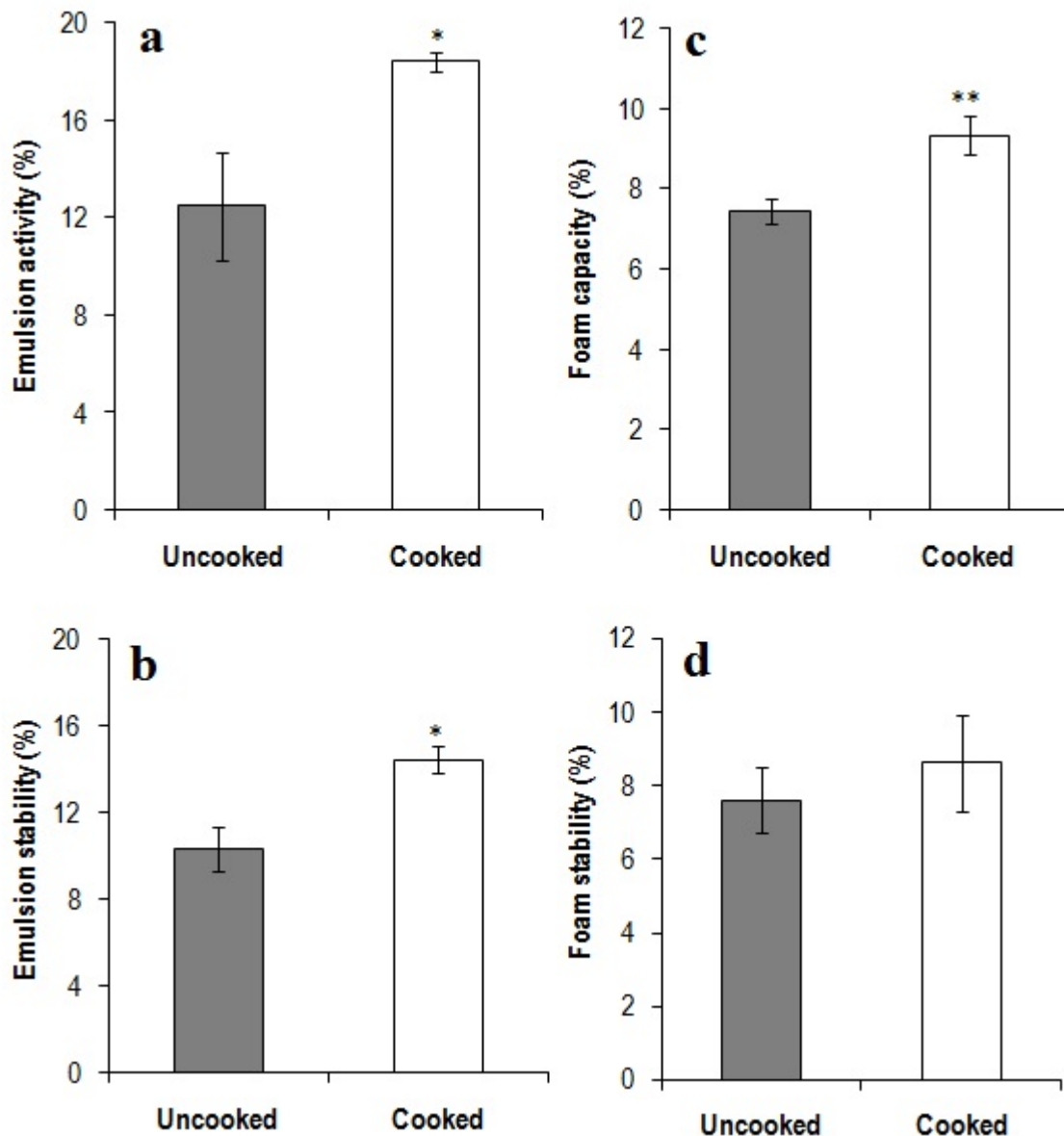


Fig. 3 Emulsion activity (a), emulsion stability (b), foam capacity (c) and foam stability (d) of uncooked and cooked tender fruit bodies of *Amanita* sp. (n=5±SD; t-test: *, $p < 0.05$; **, $p < 0.01$).

Functional vs. proximal properties

The PCA between proximal components and functional properties of uncooked *Amanita* sp. (U) resulted in two components with 100% variance (Fig. 4a). The variance for rotated score plot for component 1 and component 2 was 55.9% and 44.1%, respectively. The proximal components like crude protein (CPU: 16.3±0.49) and crude fibre (CFU: 7.4±0.37) clustered only with protein solubility (PSU) as the first cluster at the right hand region of the plot. The emulsion stability (ESU) and foam capacity (FCU) clustered only with total lipids (TLU: 4.7±0.58) as a second cluster at top of the plot. The PCA between proximal components and functional properties of cooked *Amanita* sp. (C) resulted in two components with 100% variance (Fig. 4b).

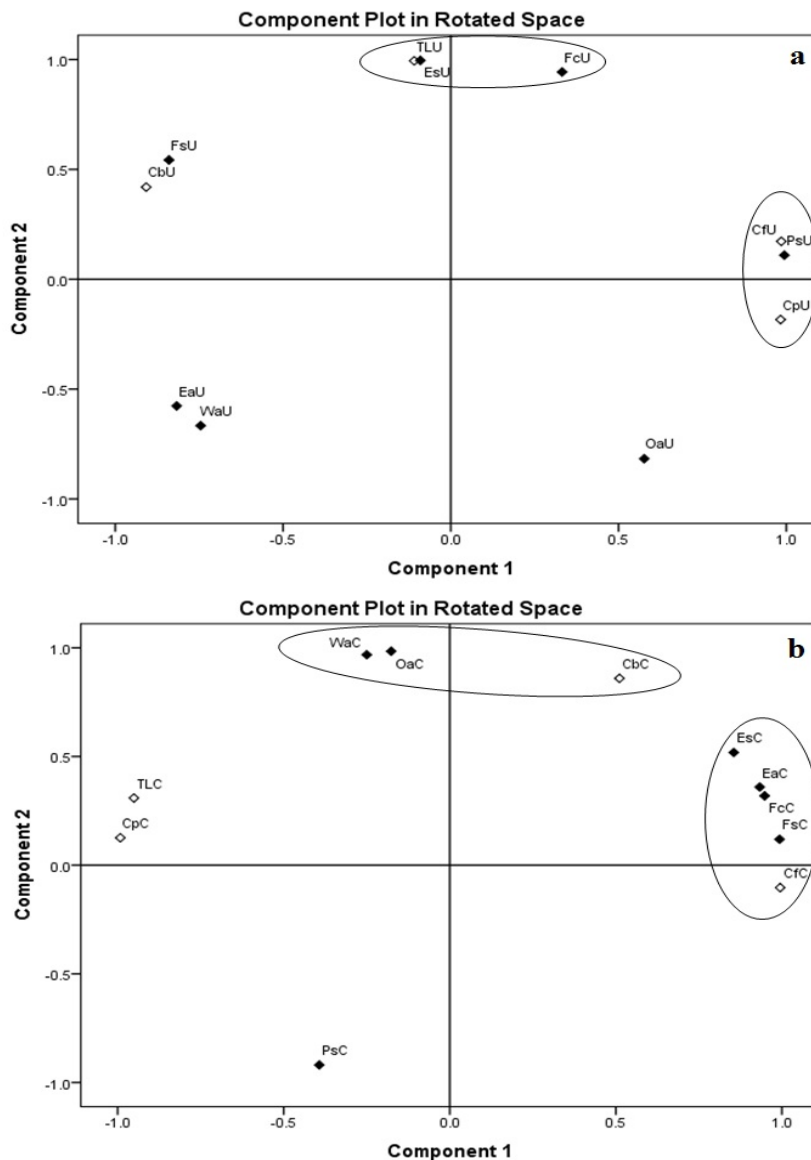


Fig. 4 Principal component analysis of uncooked (with suffix U) (a) and cooked (with suffix C) (b) tender fruit bodies of *Amanita* sp. between proximal components (crude protein, Cp; total lipids, TL; crude fibre, Cf; total carbohydrates, Cb) and functional properties (pH-dependant protein solubility, Ps; water-absorption capacity, Wa; oil-absorption capacity, Oa; emulsion activity, Ea; emulsion stability, Es; foam capacity, Fc; foam stability, Fs).

The variance for rotated score plot for component 1 and component 2 was 65.7% and 34.3%, respectively. The crude fibre (CFC: 6.7±0.43) clustered with four functional properties like

emulsion activity (ECC), emulsion stability (ESC), foam capacity (FAC) and foam stability (FSC) in the first cluster at right hand region of the plot. Water-absorption capacity (WAC) and oil-absorption capacity (OAC) were clustered with only carbohydrates (CBC: 18.5±1.6) at top of the plot as second cluster.

The influence of proximal properties on functional qualities differs between uncooked cooked samples of *Amanita* sp. In uncooked samples crude protein, total lipids and crude fibre influenced three functional properties like protein-solubility, emulsion stability and foam capacity. The crude fibre and carbohydrates of cooked samples influenced all the functional properties except for pH-dependent protein solubility. Hence, it is predictable that instead of high quantities of proximal components, their specific composition or proportion influence the functional attributes in *Amanita* sp.

Conclusions

This study revealed a major difference in functional properties between uncooked and cooked edible mushroom *Amanita* sp. Except for pH-dependent protein solubility, rest of the functional properties in cooked samples either significantly higher or not significantly differed. Significantly high emulsion activity and stability; significantly high foam capacity in cooked samples depicts its suitability in formulation of many value-added foodstuffs. Improved functional properties with desired proximal (protein-rich, low lipid and high fibre) and bioactive potential, cooked *Amanita* sp. could serve individually or additive with other raw materials in production of quality confectionaries and bakery products. It is worth attempting different methods of thermal treatment to choose the best possible combination of functional, proximal and bioactive properties in *Amanita* sp. to employ as a non-conventional nutraceutical source.

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