INSECTS INFESTING STORED GRAIN CROPS AND THEIR BIOLOGICAL CONTROL USING ENTOMOPATHOGENIC NEMATODES

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ABSTRACT: The recent experiments were conducted to classify insects attacking stored grain crops with trail to control Tribolium confusum (Jacquelin du Val) and Bruchidius incarnates (Boheman, 1833) using entomopathogenic nematodes, Heterorhabditis bacteriophora Poinar, and Steinernema feltiae Filipjev. Results indicated that stored grains under study were attacked by 11 insect species. Wheat grains were infested with nine insects where the percentages of infestation varied from 0.2% to 40%. Faba bean seeds were infested with two insects and the percentages of infestation varied from 1.5% to 50%. Rice seeds were infested with only one insect with percentages of infestation varied from 3.6- 26%. Maize seeds were infested with eight insects and the percentages of infestation varied from 0.3% to 48%. Regarding to the pathogenicity of entomopathogenic nematodes to tested insects, the results revealed that 100% mortality of T. confusum occurred with the treatments of 2000 IJs of S. feltiae & 3000 IJs of S. feltiae & 3000 IJs of H. bacteriophora.

Key words: Stored grain insects, Survey, Biological control, Entomopathogenic nematodes.

INTRODUCTION

There are many insects attacking stored grains which in turn led to reduce both quality and quantity of infested grains (Warchalewski and Gralik, 2010). In developing countries, losses caused by pests attacking stored grains ranging between 10 – 50% of the total yield (Fornal et al., 2007, Upadhyay and Ahmed, 2011). Fumigation by chemical pesticides still the main method for control stored grain pests, these chemical pesticides have hazard effects on both the environment and the consumers, on the other hand caused insect resistance (Jovanovic et al., 2007; Lu and Wu, 2010), therefore, scientists work to use other methods to control store insects rather than chemical pesticides.

In recent years, it has seen an increase attention for non-chemical methods of stored-product protection, including biological control of stored-product pests (Arbogast, 1984; Brower *et al.*, 1996; Schoeller *et al.*, 1997; Adler, 1998; Cox & Wilkin, 1998; Schoeller, 1998; Stengård, 2005; Shadia E. Abd El-Aziz, 2011; Traian *et al.*, 2015 and Shaheen *et al.*, 2016).

Entomopathogenic nematodes (EPNs) play as biological control agents for a wide range of insect species as they able to kill their hosts within 24-48 hours, furthermore, EPNs are harmless to plants and livestock and safe for the environment. EPNs characterised by their ability to search for their hosts and staying effective for a long time after application, moreover, it's easy to produce EPNs commercially at low costs (Canhilal, 2016). The use of entomopathogenic nematodes in the control of store product insects is a new field, entomopathogenic nematodes however. have not been previously tested against stored-product insects in natural environments. Recently, there are a few articles were published in this direction i.e. Trdan et al., (2006) who determined the efficacy of four entomopathogenic nematode species, S. feltiae, S. carpocapsae, H.

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bacteriophora, and H. megidis at three different concentrations (500, 1000, and 2000 IJs per adult) and three different temperatures (15, 20, and 25°C) in control of the adults of stored grain pests, Sitophilus granarius and Oryzaephilus surinamensis laboratory conditions. under Ramos-Rodriguez, et al., (2007) tested the efficacy of Steinernema riobrave against Tribolium castaneum, larvae, pupae and adult stages under laboratory coditions. Shahina and Salma (2009) evaluated the virulence of seven Pakistani strains of entomopathogenic nematodes included Steinernema pakistanense Shahina, Anis, Reid and Maqbool (Ham 10 strain); S. asiaticum Anis, Shahina, Reid and Rowe (211 strain); S. abbasi Elawad, Ahmad and Reid (507 strain); S. siamkayai Stock, Somsook and Reid (157 strain); S. feltiae Filipjev (A05 strains); Heterorhabditis bacteriophora Poinar (1743 strain); and H. indica Poinar, Karunakar and David (HAM-64 strain) against last instar and adult stages of the pulse beetle, Callosobruchus chinensis, furthermore, Shahina and Salma (2010 and 2011) tested the same seven Pakistani strains of entomopathogenic nematodes against stored grain insect pest Sitophilus oryzae (L.) and Tribolium castaneum ,respectively.

Athanassiou, et al., (2010) examined the insecticidal effect of H. bacteriophora Poinar, S. carpocapsae (Weiser), and S. feltiae (Filipjev) against Mediterranean flour moth, Ephestia kuehniella (Zeller) larvae, lesser grain borer, Rhyzopertha dominica (F.) adults, rice weevil, S. oryzae (L.) adults, and confused flour beetle, T. confusum Jacquelin du Val larvae and adults stages under laboratory conditions in wheat grains. Laznik, and Trdan, (2010) tested the efficacy of three strains (B30, B49 and 3162) of S. feltiae to control adults of rice weevil S. oryzae. Shrestha and Kim (2010) reported that the two entomopathogenic bacteria, Photorhabdus temperata sub sp. temperata (Ptt) and Xenorhabdus nematophila (Xn), are symbiotically associated with the

nematodes, *H. megidis* and *S. carpocapsae*, respectively, and found that a significant difference in pathogenicity was observed between these two bacteria against the red flour beetle, *T. castaneum*. Recently, in Egypt Sweelam *et al.*, (2010) controlled red palm weevil, *Rhynchophorus ferrugineus* Oliver by entomopathogenic nematode species.

From these points of view, this research was conducted to identify insects attacking stored grain and seeds, as well as to throw a possibility light on the of using entomopathogenic nematodes. Н. bacteriophora and S. feltiae in the biological control of the red flour beetle, T. castaneum and the faba bean beetle, Bruchidius incarnatus.

MATERIALS AND METHODS

Experiments were conducted at the laboratories of the Economic Entomology and Agricultural Zoology Department, Faculty of Agriculture, Menoufia University, Shebin Elkom, Egypt.

1- Survey of insects infesting stored grains:

Twenty five kg of each of the tested grains (Wheat, *Triticum astivum*, var. Sakha 93, Faba bean, *Vicia faba*, var. Giza 111, Rice, *Oryzae* spp, Maize, *Zea mayes*, var. Balady) were put in a cloth bag in the open greenhouse, left open to be naturally infested with insects for six months (June – November, 2015).

Every month, randomized three samples each of one kg were taken from each crop seeds and examined for insect infestation. The obtained insects were identified and percentages of infested seeds were calculated.

2- Pathogenicity of entomopathogenic nematodes to two grain insects.

2.1. Propagation of entomopathogenic nematodes:

Two species of entomopathogenic nematodes: Heterorhabditis bacteriophora Poinar (Heterorhabditidae) and Steinernema feltiae Filipjev (Steinernematidae) were obtained from Dr. M.E. Sweelam biological laboratory, in the Econ. Ent. & Agric. Zoology Dept. Fac. Agric. Menoufia Univ. The greater wax moth, Galleria mellonella were used for culturing both entomopathogenic nematodes using the method described by (White, 1927). White traps were used to harvest the infective Juveniles (IJs). Collected IJs were stored in plastic tubes (50 ml) in a refrigerator adjusted to 10 °C until used.

2.2. Procedure of infection:

Two of obtained insects, *Tribolium confusum* and *Bruchidius incarnates* were used in this study. Adults of both insects were subjected to infection at different concentrations of 50, 100, 500, 1000, 2000 and 3000 of *Heterorhabditis bacteriophora* or *Steinernema feltiae* infective juveniles (IJs) /10 insects

Ten adult insects were kept in Petri dish, each 5-cm diameter containing moist filter papers with the same diameter of Petri dishes, and exposed to entomopathogenic nematodes. Every nematode concentration was sprayed on the insects as 1 ml distilled water containing nematodes, to keep filter paper wetted, drops of distilled water were added when needed. In control treatment, insects were sprayed with 1 ml distilled water without nematodes. Each treatment was replicated three times. Mortality was checked after 24, 48, 72, 96 h for all concentrations of the two tested nematode species, and percentages of mortality were calculated for each nematode species at different concentrations using Abbott's formula (1925). Mortality percentage was corrected by Schneider-Orelli's formula (Püntener, 1981).

Corrected % = (Mortality % in treated plot - Mortality % in control plot) 100 - Mortality % in control plot × 100

RESULTS AND DISCUSSION 1- Survey of insects infesting stored grains:

Data presented in table (1) indicated the presence of 11 insect species attacked the stored grain. These insects were *Sitophilus granaries* (L.), *S. oryzae* (L.), *Sitotroga cerealella* (Olivier), *Tribolium confusum* (Jacquelin du Val), *T. castaneum* (Herbst), *Bruchidius incarnates* (Boheman, 1833), *Bruchus rufimanus* (Boheman, 1833), *Tenebriodes mauritanicus* L., *Rhizopertha dominica* L., *Plodia interpunctella* (Hubner) and *Ephestia kuehniella* Zell.

Results indicated that the wheat grains were infested with nine insects; Granary weevil, S. granaries, Angoumois grain moth, S. cerealella, Rice weevil, S. oryzae, Confused flour beetle, T. confusum, Red flour beetle, T. castaneum, Cadelle beetle T. mauritanicus, Lesser grain borer, R. dominica, Indian Ρ. meal moth, interpunctella, and Mediterranean flour moth, E. kuehniella.

The percentages of infestation varied from 0.2% to 40%, where the highest percentages in wheat grains was 40 % under six months of natural infestation with *T. confusum & T. castaneum*, followed by *S. oryzae* and *S. cerealella* recording 35%, while the lowest infestation was 6% and 2% with *P. interpunctella*, and *E. kuehniella*, respectively.

Results also indicated that the faba bean seeds were infested with the two insects; Faba bean beetle, *B. incarnatus*, and Large broad bean beetle, *B. rufimanus*, where the percentages of infestation varied from 1.5% to 50%, the highest infested seeds percentages were 50 % under six months of natural infestation with *B. incarnatus*, followed by *B. rufimanus* recording 31%.

Insect species	Seed	June, 2015	July, 2015	Aug., 2015	Sept., 2015	Oct., 2015	Nov., 2015
	crops			% infest	ed seeds		
	Wheat	0.0	0.0	1.0	6.0	31.0	33.0
Granary weevil	Faba bean	0.0	0.0	0.0	0.0	0.0	0.0
Sitophilus granaries	Rice	0.0	0.0	0.0	0.0	0.0	0.0
	Maize	0.0	0.0	1.0	6.0	15.0	28.0
	Wheat	0.0	0.3	1.5	6.0	20.0	35.0
Angoumois grain moth	Faba bean	0.0	0.0	0.0	0.0	0.0	0.0
Sitotroga cerealella	Rice	0.0	0.0	0.0	0.0	0.0	0.0
	Maize	0.0	0.3	1.5	6.0	20.0	35.0
	Wheat	0.0	0.3	1.5	6.0	20.0	35.0
Rice weevil	Faba bean	0.0	0.0	0.0	0.0	0.0	0.0
Sitophilus oryzae	Rice	0.0	0.0	0.0	0.0	0.0	0.0
	Maize	0.0	0.3	1.5	6.0	0.0	28.0
Confused flour beetle	Wheat	0.0	0.0	0.0	10.0	25.0	40.0
Tribolium confusum	Faba bean	0.0	0.0	0.0	0.0	0.0	0.0
Red flour beetle	Rice	0.0	0.0	0.0	0.0	0.0	0.0
Tribolium castaneum	Maize	0.0	0.0	0.0	10.0	25.0	40.0
	Wheat	0.0	0.0	0.0	0.0	0.0	0.0
Faba bean beetle	Faba bean	1.5	3.6	6.0	20.0	32.0	50.0
Bruchidius incarnatus	Rice	0.0	0.0	0.0	0.0	0.0	0.0
	Maize	0.0	0.0	0.0	0.0	0.0	0.0
	Wheat	0.0	0.0	0.0	0.0	0.0	0.0
large broad bean beetle	Faba bean	0.0	0.0	0.0	0.0	27.0	31.0
Bruchus rufimanus	Rice	0.0	0.0	0.0	0.0	0.0	0.0
	Maize	0.0	0.0	0.0	0.0	0.0	0.0
	Wheat	0.0	0.0	1.5	3.0	10.0	23.0
Cadelle beetle	Faba bean	0.0	0.0	0.0	0.0	0.0	0.0
Tenebriodes mauritanicus	Rice	0.0	0.0	0.0	0.0	0.0	0.0
	Maize	0.0	0.0	1.5	4.0	12.0	26.0
	Wheat	0.0	2.5	2.6	5.0	22.0	31.0
Lesser grain borer	Faba bean	0.0	0.0	0.0	0.0	0.0	0.0
Rhizopertha dominica	Rice	0.0	0.0	3.6	4.0	12.0	26.0
	Maize	0.0	3.5	5.6	8.0	34.0	48.0
	Wheat	0.0	0.0	0.0	0.3	1.5	6.0
Indian meal moth	Faba bean	0.0	0.0	0.0	0.0	0.0	0.0
Plodia interpunctella	Rice	0.0	0.0	0.0	0.0	0.0	0.0
	Maize	0.0	0.0	0.0	0.3	1.5	6.0
Mediterranean flour moth	Wheat	0.0	0.0	0.0	0.3	2.0	2.0
Enhestia kuehniella	Faba bean	0.0	0.0	0.0	0.0	0.0	0.0
	Rice	0.0	0.0	0.0	0.0	0.0	0.0
	Maize	0.0	0.0	0.0	0.3	1.5	6.0

Table (1): Monthly infestation of different seeds with insects along six months of storage (June – November, 2015)

As for rice seeds, it was infested with only Lesser grain borer, *R. dominica* L., with % infestation of 3.6- 26%.

Regarding to maize seeds, it was infested with the eight insects; Granary weevil, S. granarius, Angoumois grain moth, S. cerealella, Rice weevil, S. oryzae, Confused flour beetle, T. Confusum, Red flour beetle, T. castaneum, Cadelle beetle, T. mauritanicus, Lesser grain borer, R. dominica, Indian meal moth, Р interpunctella and Mediterranean flour moth, E. kuehniella. The infestation varied from 0.3% to 48%, the highest damaged seed percentages of maize stored under six months of natural infestation was 48 % with R. dominica, followed by T. Confusum, T. castaneum 40%, S. cerealella 35 %, while it was 28 % with both S. granaries and S. oryzae, moreover, the infestation with T. mauritanicus recorded 26%. The lowest damage infestation was only 6% with both of P. interpunctella, and E. kuehniella.

2- Pathogenicity of EPN to tested insects:

Results in table (2) show the effect of different doses of entomopathogenic nematode, *Steinernema feltiae* on the mortality percentages of confused flour beetle, *T. confusum* adults, under laboratory conditions. The highest averages of corrected mortality of *T. confusum* adults were recorded with the treatment of 2000 IJs of entomopathogenic nematode, *S. feltiae* giving 100 %, followed by the treatment of 500 IJs of *S. feltiae* nematode as it recorded 81.58 %. The treatment of 100 IJs of *S. feltiae* nematode gave 76.32 %, while the treatment of 1000 IJs of *S. feltiae* gave the least averages of the corrected mortality of the adult stage insects which calculated as 68.42 %.

Results in table (3) show the effect of different doses of entomopathogenic nematode, Heterorhabditis bacteriophora on the mortality percentages of confused flour beetle, T. confusum adults under laboratory conditions. Results indicated that the highest averages of corrected mortality of confused flour beetle. T. confusum adults were registered with the treatments of 1000, 2000. 3000 IJs of entomopathogenic nematode, H. bacteriophora giving 100 %, followed by the treatment of 500 IJs of H. bacteriophora nematode as it recorded 88.57 %, and the treatment of 100 IJs which recorded 82.86 %.

Concentration	Mortality percentages					Corrected
of nematode juveniles	24 h	48 h	72 h	96 h	Mean	mortality
100	80	80	60	90	77.5	76.32
500	80	80	80	90	82.5	81.58
1000	60	60	80	80	70	68.42
2000	100	100	100	100	100	100.00
3000	70	70	80	80	75	73.68
Control	0	0	10	10	5	-

Table (2): Mortality percentages of confused flour beetle, *T. confusum* adults as affectedby S. feltiae nematode under laboratory conditions (25 ± 3 °C & 65% RH)

RH)						
Concentration	Mortality percentages					Corrected
of nematode juveniles	24 h	48 h	72 h	96 h	Mean	mortality
100	80	80	90	90	85	82.86
500	80	90	90	100	90	88.57
1000	100	100	100	100	100	100
2000	100	100	100	100	100	100
3000	100	100	100	100	100	100
Control	10	10	10	20	12.5	-

Table (3): Mortality percentages of confused flour beetle, *T. confusum* adults as affected by *H. bacteriophora* nematode under laboratory conditions ($25 \pm 3 \ ^{\circ}C \ \& 65\%$

As for the toxic effect of the entomopathogenic nematode, S. feltiae on faba bean beetle, B. incarnates adults, results in table (4) indicated that the highest averages of corrected mortality of B. incarnatus adults were registered with the 2000 and 3000 IJs of treatments of entomopathogenic nematode, S. feltiae giving 100 %, followed by the treatments of 500, 1000 IJs of S. feltiae nematode as it recorded 62.16 and 51.35 %, respectively, while the least averages of the corrected mortality of B. incarnates adults were recorded with the treatment of 100 IJs which was 35.14 %.

Results in table (5) show the effect of entomopathogenic different doses of nematode, Heterorhabditis bacteriophora against the faba bean beetle, Bruchidius incarnates adults under laboratory conditions. Results indicated that the highest averages of corrected mortality of В. incarnatus adults were registered with the treatment of 3000 IJs of entomopathogenic nematode, H. bacteriophora giving 100 %, followed by the treatment of 2000 IJs of H. bacteriophora nematode giving 94.87 %, then the treatment of 500 IJs of H. bacteriophora, which gave 87.18%. The averages of the corrected mortality of B. incarnatus adults with the treatments of 100 1000 IJs entomopathogenic and of nematode, H. bacteriophora were calculated as 84.62 and 79.49 %, respectively as overall averages.

The obtained results are in harmony with those obtained by Trdan *et al.*, (2006) who proved that the entomopathogenic nematodes were efficient in the control of *S. granarius and Oryzaephilus surinamensis* at 20°C.

Ramos-Rodriguez, *et al.*, (2007) who reported that in laboratory bioassays, *S. riobrave* reduced survival of red flour beetle, *T. castaneum*, stages to 27.4 % in treatments compared to 77.9% in the control which mean *S. riobrave* compete as a biological control agent for stored-product insects.

Shahina and Salma (2009, 2010 and 2011) found that *H. bacteriophora, S. siamkayai*, and *S. pakistanense* showed high virulence to *C. chinensis, S. oryzae* and *T. Castaneum* stages, respectively, as the mortality of adult, larval and pupal stages were higher in the nematode treatments than in the control in laboratory bioassays.

Athanassiou, *et al.*, (2010) who used entomopathogenic nematodes, *H. bacteriophora*, *S. feltiae* and *S. carpocapsae* at different concentrations (10, 50, 100, 150, 500, 1000, and 2000 IJs / insect) in the control of the rice weevil, *S. oryzae*, the red flour beetle, *T. castaneum*, the lesser grain borer, *R. dominica* (F.), the Mediterranean flour moth, *E. kuehniella* (Zeller), and the pulse beetle, *C. chinensis* (L.) under laboratory conditions.

Table (4): Mortality percentages of faba bean beetle, *B. incarnates* adults as affected by *S. feltiae* nematode under laboratory conditions (25 ± 3 °C & 65% RH)

Concentration	Mortality percentages					Corrected
of nematode juveniles	24 h	48 h	72 h	96 h	Mean	mortality
100	20	20	60	60	40	35.14
500	20	80	80	80	65	62.16
1000	20	40	60	100	55	51.35
2000	100	100	100	100	100	100
3000	100	100	100	100	100	100
Control	0	10	10	10	7.5	-

Table (5): Mortality percentages of faba bean beetle, *B. incarnates* adults as affected by *H. bacteriophora* nematode under laboratory conditions (25 ± 3 °C & 65% RH)

Concentration	Mortality percentages					Corrected
of nematode juveniles	24 h	48 h	72 h	96 h	Mean	mortality
100	60	80	100	100	85	84.62
500	50	100	100	100	87.5	87.18
1000	60	60	100	100	80	79.49
2000	80	100	100	100	95	94.87
3000	100	100	100	100	100	100
Control	0	0	0	10	2.5	-

Conclusion

It could be concluded that the use of entomopathogenic nematodes (EPNs), Heterorhabditis bacteriophora and Steinernema feltiae in the control of stored insect products i.e. confused flour beetle, Tribolium confusum, and the faba bean beetle, Bruchidius incarnatus registered good results in laboratory bioassays, but it needs more studies where solutions should be sought to facilitate the application of entomopathogenic nematodes in natural conditions within grain stores, the combination of EPNs with some other biotechnical methods may be one of the solutions in the future.

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REFERENCES

- Abbott, W. S. (1925). A method of computing the effectiveness of an insecticide. J. Econ. Ent., 18, 265-267.
- Adler, C. (1998). What is integrated storage protection? In: C. Adler & M. Schoeller

(eds). Integrated Protection of Stored Products. IOBC WPRS Bulletin 21, 1–8.

- Arbogast, R. T. (1984). Biological control of stored-product insects: status and prospects. In: F. J. Baur (ed.), Insect Management for Food Storage and Processing. Americ. Assoc. Cereal Chem., St. Paul, Minnesota, 225–238.
- Athanassiou, C. G., N. G. Kavallieratos, H. Menti and E. Karanastasi (2010). Mortality of four stored product pests in stored wheat when exposed to doses of three entomopathogenic nematodes. J. Econ. Ent., 103 (3): 977-984.
- Brower, J. H., L. Smith, P. V. Vail and P. W. Flinn (1996). Biological control. In: B. Subramanyam & D.W. Hagstrum (eds), Integrated Management of Insects in Stored Products. Marcel Dekker, Inc., New York, 223–286.
- Canhilal (2016). The use of entomopathogens in the controlling of insect Pests of stored product. Series A. Agronomy, 235- 240.
- Cox, P. D. and D. R. Wilkin (1998). A review of the options for biological control against invertebrate pests of stored grain in the UK. In: C. Adler & M. Schoeller (Eds), Integrated Protection of Stored Products. IOBC WPRS Bulletin 21, 27– 32. search 54, 689–713.
- Fornal, J., T. Jeliński, J. Sadowska, S. Grundas, J. Nawrot, A. Niewiara, J.R. Warchalewski and W. Błaszczak (2007). Detection of granary weevil Sitophilus granarius L. eggs and internal stage in wheat grain using soft X-ray and image analysis. J. of Stored Product Res. 43, 142-148.
- Jovanovic, Z., M. Kostic and Z. Popovic (2007). Grain-protective properties of herbal extracts against the bean weevil *Acanthoscelides obtectus* Say. Indust. Crop and Produc. 26, 100-104.
- Laznik, Z. S. Trdan (2010). Intraspecific variability of *Steinernema feltiae* (Filipjev) (Rhabditida: Steinernematidae) as biological control agent of rice weevil *Sitophilus oryzae* [L.] (Coleoptera,

Curculionidae) adults. Acta Agric. Slovenica, 95 (1): 51-59.

- Lu, J. and S.H. Wu (2010). Bioactivity of essential oil from *Ailanthus altissima* bark against 4 major stored-grain insects. African J. Microb. Res. 4,154-157.
- Püntener W. (1981). Manual for field trials in plant protection second edition. Agricultural Division, Ciba-Geigy Limited.
- Ramos-Rodriguez, O., J. F. Campbell and S. B. Ramaswamy (2007). Efficacy of the entomopathogenic nematode *Steinernema riobrave* against the storedproduct insect pests *Tribolium castaneum* and *Plodia interpunctella*. Biol. Cont., 40 (1): 15-21.
- Schoeller, M. (1998). Integration of biological and non-biological methods to control arthropods infesting stored products. In: C. Adler & M. Schoeller (eds.), Integrated Protection of Stored Products. IOBC WPRS Bulletin 21: 13– 25.
- Schoeller, M., S. Prozell, A.G. Al-Kirshi and C.H. Reichmuth (1997).Towards biological control as a major component of integrated pest management in stored product protection. J. Stored Product Res., 33: 81–97.
- Shadia E. Abd El-Aziz (2011). Control Strategies of Stored Product Pests. Journal of Entomology, 8: 101-122.
- Shaheen, F. A., M. W. Akram, M. A. Rashid,
 M. Nadeem, M. Saeed, M. Husain and K.
 Mehmood (2016). Biological control of pulse beetle *Callosobruchus chinensis* L.
 (Bruchidae: Coleoptera) in stored chickpea grains using entomopathogenic fungus *Beauveria bassiana* Balsamo. J.
 Ent. and Zool. Studies, 4(4): 1076-1083.
- Shahina F. and J. Salma (2009). Laboratory evaluation of seven Pakistani strains of entomopathogenic nematodes against a stored grain insect pest, pulse beetle *Callosobruchus chinensis* (L.). J. Nematol., 41 (4): 255-260.
- Shahina, F. and J. Salma (2010). Laboratory evaluation of seven Pakistani strains of

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entomopathogenic nematode against stored grain insect pest *Sitophilus oryzae* L. Pakistan J. Nematol., 28 (2): 295-305.

- Shahina F. and J. Salma (2011). Pakistani Strains of Entomopathogenic Nematode as A Biological Control Agent Against Stored Grain Pest, *Tribolium Castaneum*. Pak. J. Nematol., 29 (1): 25-34.
- Shrestha, S. and Y.G. Kim (2010) Differential pathogenicity of two entomopathogenic bacteria, Photorhabdus temperate subsp. Temperate and Xenorhabdus nematophila against the red flour beetle, Tribolium castaneum. J.Asia-Pacific Ent., 13(3): 209-213.
- Stengård H. L. (2005) Biological Control of Stored-Product Pests. In: Pimentel, D (ed.), Encyclopedia of Pest Management. Marcel Dekker, New York.
- Sweelam, M. E., A. S. Albarrak, A. A. Abd El-All and A. M. Kella (2010). Biological red control of the palm weevil, Rhynchophorus ferrugineus Oliver (Coleoptera: Curculionidae) by entomopathogenic nematode species. Annals of Agric. Sci., Moshtohor, 48(2): 21 - 28.

- Traian, M., L. Carmen, F. Viorel, C. Andrei and C. Florentin (2015). Experimental Model for Biological Control of Stored Grain Pests. J. Agric. Sci. and Techno., 5: 793-798.
- Trdan, S., M. Vidrih and N. Valic (2006). four entomopathogenic Activity of nematode species against young adults of Sitophilus granaries (Coleoptera: Curculionidae) Oryzaephilus and surinamensis (Coleoptera: Silvanidae) under laboratory conditions. J. Plant Diseases and Protec., 113 (4): 168–173.
- Upadhyay, R.K. and S. Ahmad (2011). Management strategies for control of stored grain insect pests in farmer stores and public warehouses. World J. Agric. Sci., 7(5): 527-549.
- Warchalewski, J.R. and J. Gralik (2010). Influence of microwave heating on biological activities and electrophoretic pattern of albumin fraction of wheat grain. Cereal Chemist., 87 (1): 35-41.
- White, G.F. (1927). A method for obtaining infective nematode larvae from cultures. Sci. 66 (1709): 302-303.

الحشرات التى تصيب حبوب المحاصيل المخزونة ومكافحتها الحيوية باستخدام النيماتودا الممرضة للحشرات

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الملخص العربى

أجريت هذه الدراسة بمعمل البيولوجى – بكلية الزراعة . جامعة المنوفية لحصر أهم الحشرات التى تصيب الحبوب المخزونة تحت الظروف الطبيعية ودراسة إمكانية مكافحة حشرتى خنفساء الدقيق المتشابهة وخنفساء الفول الصغيرة باستخدام نوعين من النيماتودا الممرضة للحشرات Heterorhabditis bacteriophora و Steinernema feltiae وذلك تحت الظروف المعملية.

بالنسبة للدراسة المعملية لتقدير كفاءة نوعين من النيماتودا الممرضة للحشرات فى مكافحة الأطوار الكاملة لكل من حشرتى خنفساء الدقيق المتشابهة وخنفساء الفول الصغيرة وذلك بتعريض الأطوار الكاملة لهذه الحشرات لأعداد مختلفة من يرقات النيماتودا الممرضة للحشرات وعد الحشرات الميتة بعد 24 ،48 ، 72 ، 96 ساعة وحساب نسب الموت المصححة ، قد أثبتت النتائج نجاح كل من نوعى النيماتودا تحت الإختبار فى مكافحة أطوار الحشرات الكاملة تحت الدراسة وتراوحت نسب الموت من 35.14 مع قائد . 20 % مع مع مع النوع . 100 % موت من حشرات خنفساء الدقيق المتشابهه تم تسجيلها مع التركيز 2000 يرقة نيماتودية من النوع . 100 موت من حشرات خنفساء الدقيق المتشابهه تم تسجيلها مع التركيز 2000 يرقة نيماتودية من النوع . 103 موت من حشرات خنفساء الدقيق المتشابهه تم تسجيلها مع التركيز 2000 يرقة نيماتودية من النوع . 104 موت من حشرات موت من خلفساء الدقيق المتشابهه مع التركيز 2000 يرقة نيماتودية من النوع . 105 موت من النوع . 105 موت من خلفي الموت الموت من 1000 يرقة المتركيز 2000 يرقة نيماتودية من النوع . 105 موت نسبة 100 موت من خلفوا الصغيرة مع التركيزات 2000 يرقة نيماتودية من النوع . 105 موت من خلفوا الصغيرة مع التركيزات 2000 يرقه نيماتوديه من النوع . 105 موت النوع . 105 موت من خلفوا الصغيرة مع التركيزات 2000 يرقه نيماتوديه من النوع . 105 موت من النوع . 105 موت من خلفوا الصغيرة مع التركيزات 2000 يرقه نيماتوديه من النوع . 105 موت من خلفوا الصغيرة مع التركيزات 2000 موت من 100 موت من 100 موت من 100 موت من النوع . 105 موت من خلفوا الصغيرة مع التركيزات 2000 موت من 100 موت من 100 موت من 100 موت من 100 موت من خلفوا الصغيرة مع التركيزات 2000 موت من خلفوا الصغيرة مع التركيزات 100 موت من خلفوا الصغيرة مع التركيزات 2000 موت من 100 موت من 100 من النوع . 105 موت التركيز 1000 يرقه نيماتوديه من النوع . 105 موت التركيز 2000 يوقه نيماتوديه من النوع . 105 موت من خلفوا الصغيرة مع التركيزات 2000 موت من خلفوا المول الصغيرة مع التركيزات 2000 موت من خلفوا موت من 100 موت من خلفوا موت من النوع .

ويوصى البحث بإمكانية إستخدام النيماتودا الممرضة للحشرات فى برامج المكافحة البيولوجية لحشرات المخازن تحت الظروف المعملية ، لكننا بحاجه إلى مزيد من الدراسات لإيجاد حلول لتسهيل تطبيق النيماتودا الممرضة للحشرات في الظروف الطبيعية داخل مخازن الحبوب.