Provided for non-commercial research and education use.

Not for reproduction, distribution or commercial use.



Egyptian Academic Journal of Biological Sciences is the official English language journal of the Egyptian Society for Biological Sciences, Department of Entomology, Faculty of Sciences Ain Shams University.

Physiology & molecular biology journal is one of the series issued twice by the Egyptian Academic Journal of Biological Sciences, and is devoted to publication of original papers that elucidateimportant biological, chemical, or physical mechanisms of broad physiological significance.

www.eajbs.eg.net



Biochemical Effects of Bradykinin Potentiating Factor (BPF) Isolated from Scorpion Venom (*Leiurus quinquestriatus*) against CCl₄- Liver Injury in Male Albino Rats.

Muhammad M. A. Salman

Department of Zoology, Faculty of Science, South Valley University, Qena, Egypt Salman2 52014@yahoo.com

ARTICLE INFO

Article History Received:25/11/2015 Accepted:27/12/2015

Keywords: scorpion venom *L. quinquestriatus* BPF CCl₄ liver injury and Albino rats.

ABSTRACT

The main purpose of this study is to evaluate the ability of bradykinin potentiating factor (BPF) isolated from scorpion venom (Leiurus quinquestriatus) in treatment of liver injuries which induced by injection of CCl₄ in male Albino rats. Male Albino rats (250±20 g. body weight) were divided into four groups. In the control group; Albino rats were interaperitoneally (i,p) injected with 100 µL saline solution. The second group (i.p) injected with BPF in 100 µL saline solutions (1µgm/g. b. w. per 5 days). Third and fourth groups were i.p. injected with 0.5 ml/kg body weight (b. w.) twice weekly of CCl₄ for fifteen days, after that only the fourth group was treated by BPF in 100 µL saline solutions (1µgm/g. b. w. per 5 days). The results indicated that, CCl₄ injection induced a significant decrease in serum catalase (CAT), superoxide dismutase (SOD), reduced glutathione (GSH), total protein and albumin, within thirty days post-injection of CCl₄ as compared to the normal control group. In contrast, CCl₄ induced a significant increase in malondialdehyde (MDA), aspartate amino transferase (AST), alanine amino transferase (ALT), and alkaline phosphatase (ALP) compared to normal control animals. The efficiency of BPF treatment is alleviation the effects of CCl₄ on these parameters. The improvement of these parameters may be attributed to the release antioxidant and cytokines and/or amelioration of the toxic effects of CCl₄ on the liver.

INTRODUCTION

The liver is expected not only to perform physiological functions but also to protect against the hazards of harmful drugs and chemicals. In spite of the tremendous scientific advancement in the field of hepatology in recent years, liver problems are on the rise (Pang *et al.*, 1992 and Al-Jumaily *et al.*, 2014 and Abhilash,*et al.*, 2013 and 2014). It is the key organ regulating homeostasis in the body. It is involved with almost all the biochemical pathways related to growth, fight against disease, nutrient supply, energy provision and reproduction (Wards and Daly, 1999;). Carbon tetrachloride (CCl₄) is widely used for modeling liver injury in rats. Hepatotoxicity is connected with severe impairment of the cell protection mechanisms. CCl₄ is a heavy compound that may act as a nonflammable liquid (Cetin *et al.*, 2011and Mnaa *et al.*, 2015). It is widely used in the dry-cleaning industry although it is a highly toxic chemical agent. Thus, CCl₄ is most widely used for experimental induction of hepatic cirrhosis (Abd-El-Dayem and Moawad, 2001 and Lin *et al.*, 2005).

CCl₄ can induce the oxidative stress beside the inhibition of the activity of antioxidant enzymes in renal tissue (Basu, 2003). The liver injury is induced mainly by the bio-transformation of CCl₄, which is cytchrome p-450 dependent free radicals initiate the process of lipid peroxidation, which is generally caused an inhibition of enzyme activity (Ward and Daly, 1999). Lipid peroxidation is autocatalytic an leading mechanism to oxidative destruction of cell membranes (Wang and Salahudeen, 1995). It is known that the reactive oxygen specious (ROS) would lead to oxidative damage of biological macromolecules, including lipids, proteins, and DNA (Das and Chainy, 2001). Against these types of oxidative injuries, tissues have a variety of defense mechanisms including the non-enzymatic glutathione (GSH), the enzymatic SOD scavenger systems and CAT (Tirkey et al., 2005 and Subudhi et al., 2008). Recently, a few hepatoprotective drugs from natural sources are for the available treatment or ameliorating the liver disorders. The bradykinin potentiating factor **BPF** extracted from Leiurus quinquestriatus venom was shown to enhance the cellular growth of the uterus and development of the ovarian follicle in female mice (Abd-El-Reheim, 1995). Similarly, injection of this BPF enhanced the spermatogenesis .Moreover, topical on burnt application of BPF skin of Guinea pigs accelerated its healing that attributed to a direct effect of a growth like activity or indirectly by stimulating the endogenous prostaglandin E2, both in turn, stimulates collagen and elastin synthesis and skin epithelialization (Salman, 1995). Moreover, Salman (2002) declared that injection of BPF in sublethally-irradiated and non-irradiated Guinea pigs accelerated the generation of thymus and spleen cellularity and completely recovered; the normal platelets, WBCs,

RBCs and blood globulins picture without noticeable toxic effects in nonirradiated control animals. It is worthy to mention that, the bradykinin-stimulated release of several cytokines important in proliferation and differentiation of blood cell various progenitors, is in achieving implicated the forementioned effects. These cytokines include interleukin-1 (1L-1), IL-3, IL-6, IL-11, IL-12, tumor necrosis factor_{α} (TNF_{α}) and thrombopoietin (Neben et al., 1996). The activation of Kallikarinkinin system (KKS) may regulate the progression of chronic liver diseases by inducing hepatoprotection and reducing fibrogenesis (Sancho-Bru et al., 2007); the KKS also possess anti thrompotic, anti-inflammatory, and anti-apoptotic effects (Kouyoumdjian et al., 2009), which suggesting its beneficial effect in reducing liver damaging cell. Kinin may attenuate inflammatory responses and renal fibrosis by inhibiting oxidative stress and mitogen-activated protein kinase (MAPK) activation (Chao et al., 2007). Therefore, the aim of this study is to investigate the possible prophylactic effect of BPF that isolated from Leiurus quinquestriatus venom against oxidative damage of CCl₄ in male Albino rats.

MATERIALS AND METHODS

Carbon tetrachloride (CCl₄): CCl₄ is a colorless non-flammable pleasant smelling liquid, of molecular weight 153.84 was obtained from El-Nasr pharmaceutical chemical Co., A.R.E. Bradykinin potentiating factor (BPF): BPF was previously isolated from the venom of the scorpion Leiurus quinquestriatus (Salman, 1995; 2002; 2009 and 2010) according to the chemical method of Ferreira (1965). LD₅₀ crude venom was determined as described by Meier and Theakston (1986). The LD 50 of BPF was found to be 1.25 mg/kg b. w. of Albino rats.

Animals: Adult male Albino rats of approximate weight (about 250±20 g. body weight each) were selected from the animal house of the Egyptian organization for Biological products and vaccines (VACSERA), Helwan, Cairo, Egypt. The animals were housed in the animal house of the faculty of science, South Valley University, Qena, Egypt, for two weeks under natural day and night periods and with a balanced diet and water ad libitum. The animals divided into three groups:

Group1: The animals (16 animals) were i.p. injected with 0.9% isotonic saline solution at a dose (100 ml/kg body weight) per 5 days along the experimental period and served as a normal group.

Group2: The animals (16 animals) were (i.p.) injected with CCl_4 (0.5 ml/kg body weight), and left without any treatment.

Group3: The animals (16 animals) were injected with CCl₄ (0.5 ml/kg body weight) and then (i.p.) injected with BPF dissolved in saline solution 1 μ gm/g b.w. per 5 days. Animals were sacrificed after 15 and 30 (8 animals each), when received 3 and 6 successive doses of BPF, respectively.

Sample collection:

Peripheral blood was collected from each animal and divided into two portions; part was taken in EDTA containing tubes for monitoring reduced blood Glutathione (GSH) and the other portion of blood was collected in clean tubes at room temperature. After an hour, serum was separated by centrifugation for 15 minutes at 3000 rpm. (Dacie and Lewis, 1975). The sera were collected in aliquots in labeled Epindorff's tubes and stored at -20° C until used for biochemical assaying.

Prior to dissection, the liver tissue is perfused with a cold BPS (Phosphate buffered saline) solution, pH 7.4 containing 0.16 mg / ml heparin to remove any blood cell and clots. Hardening the dissected tissue by liquid nitrogen then crushed and homogenized in 5-10 ml cold buffer (i. e., 50 mM potassium phosphate, pH 7.5, 1 mM EDTA) per gram tissue. The tissue is centrifuged at 4000 rpm. for 15 minutes and then taken supernatant for assaying or kept frozen at -20°C until assayed.

Assessment of biochemical parameter of blood, serum and tissue of liver:

Biochemical parameters; Alanine transferase, amino aspatate amino transferase (Young et al., 1972), alkaline phosphatase (El-Aaser and El-Merzabani, 1975). total protein (Peters, 1968), albumin (Doumas et al., 1971), were analysed according to the reported method, Malondialdehtde (Ohkawa, et al., 1979), reduced blood Glutathion (Beutler et al., 1963), Catalase (Fossati et al., 1980) and super oxide dismutase (Nishikimi et al., 1972), were analyzed using available kits according the reported method.

Statistical analysis:

The results are expressed as mean \pm S.E. The means comparisons were made by using one-way analysis of variance (ANOVA) using Graph Pad Prism 03n software, where appropriate. Statistical significance was set at p<0.05.

RESULTS

Effect of the bradykinin potentiating factor (BPF) isolated from *Leiurus quinquestriatus* venom on the (ALT), (AST) and (ALP), 1 μ g/g b w per 5 days in Albino rat injected with CCl₄ (0.5 ml/kg body weight) post 15 and 30 days of treatment respectively.

As shown in (Fig. 1) the ALT, AST, and ALP recorded a significant increase in group 2 post 15 and 30 days of injection when compared with normal animals.

In group 3 when the animals treated with BPF (i. e. 3 doses within 15 days and 6 doses within 30 days), the serum ALT, AST, and ALP decreased significantly compared with group 2, and almost recorded to normal level.



Fig.(1) : Effect of a bradykinin potentialing factor (BFF) isolated from scorpton venom; *Leturus quinquestriatus* (1µgm/g b.w.) treated per 5 days on serum (ALT), (AST) and (ALP), in Albino rats after injection with CCl₄ (0.5 ml/kg b. w.) post of 15 and 30 days from treatment. P < 0.05 = significant different from the control NS=Insignificant different from the control

Effect of the bradykinin potentiating factor (BPF) isolated from *Leiurus quinquestriatus* venom on the albumin and total protein, 1 μ gm/g b.w. per 5 days in Albino rat injected with CCl₄ (0.5 ml/kg body weight) post 15 and 30 days of treatment respectively.

Total protein and albumin levels were significantly decreased in group 2

when compared with normal animals as shown in (Fig. 2). In group 3 when treated with BPF (3 doses within 15 days and 6 doses within 30 days), total protein and albumin recorded a significant decrease, when compared with group 2, and almost recorded the normal level.



Fig. (2): Effect of a bradykinin potentiating factor (BPF) isolated from *Leiurus quinquestriatus* venom on the Serum proteins (T. protein and albumin) 1 μgm/g b.w. per 5 days in Albino rats injected with CCl₄ (0.5 ml/kg body weight) post 15 and 30 days of treatment.
 P < 0.05 = significant different from the control

Lipid peroxidation:

As shown in (Fig. 3), the MDA level was significantly increased in group 2 when compared with normal animals. On the treatment, in group 3 which injected with BPF (3 doses within 15 days and 6 doses within 30 days), MDA recorded a significant decrease, when compared with the group 2 and almost recorded the normal level.

118

Hepatic antioxidant enzyme activities:

GSH, CAT and SOD levels were significantly decreased in group 2 when compared with normal animals as shown in (Fig. 3 and 4). With treating, the

animals which injected with BPF (3 doses within 15 days and 6 doses within 30 days) recorded a significant increase. when compared with group 2, and almost recorded the normal level.



Fig. (3): Effects of a bradykinin potentiating factor (BPF) 1 µgm/gm b.w. per 5 days isolated from Leiurus quinquestriatus on MDA and SOD of liver tissues of Albino rats injected with CCl₄ (0.5 ml/kg body weight) post 15 and 30 days of treatment.

P < 0.05 = significant different from the control **NS**=Insignificant different from the control





P < 0.05 = significant different from the control P < 0.01 = highly significant different from the control NS=Insignificant different from the control

DISCUSSION

The hepatic injury produced by carbon tetrachloride in Albino rats is well-known as hepatotoxic agent (Thrall et al., 2000 and Al-Jumaily et al., 2014).

The changes associated with CCl₄induced liver damage are similar to that of acute viral hepatitis. An obvious sign of hepatic injury is the leaking of cellular enzymes into the plasma (Kumar et al., 2005) due to the disturbance caused in the transport function of hepatocytes. When plasma of liver cell is damaged a variety of enzymes located normally in cytosol is released into the blood. The estimation of enzymes in the serum is a useful quantitative marker of the extent and types of hepatocellular damage (Jadon et al., 2007). In the present investigation, the injection of CCl₄ caused liver injury of Albino rats and developed significant hepatic damage, which was observed through a substantial change in the concentration of serum parameters. Liver enzymes such as ALT, AST and ALP are marker enzymes for liver function and integrity (Adaramoye et al., 2008). Necrosis or membrane damage releases the enzyme into circulation and hence it can be measured in the serum. A high level of AST indicates liver damage (Rosa et al., 2009). AST catalyses the conversion of alanine to pyruvate and glutamate and is released in a similar manner. ALT is more specific to the liver, and is thus a better parameter for detecting liver injury (Palanivel et al., 2008). Elevated levels of serum enzymes are indicative of cellular leakage and loss of functional integrity of cell membrane in liver (Drotman and Lawhan 1978). Serum ALP, albumin and total protein levels on other hand are related to the function of hepatic cell. Increase in serum level of ALP is due to increased synthesis, in presence of increasing billiary pressure (Muriel and Garcipiana, 1992).

In the present study injection with CCl₄ caused a significant elevation of serum enzyme levels such as AST, ALT and ALP, and significant decrease in total protein and albumin, when compared to normal animals. There was a significant restoration of these enzymes and protein levels in animals injected with the BPF as a treatment in injured animals when compared to the group which injected only CCl₄. The reversal of increased serum enzymes in CCl₄-

induced liver damage by the venom fraction (BPF) may be due to the prevention of the leakage of intracellular enzymes by its membrane stabilizing activity. This is in agreement with the commonly accepted view that serum levels of transaminases return to normal with the healing of hepatic parenchyma and the regeneration of hepatocytes (Thabrew et al., 1987and Bekheet, et al., 2013). Additionally, endogenous potentiating by the venom fraction (BPF) on the bradykinin, induces cellular active ation and hence proliferation (Abu-Amra and Abd-El-Rehim, 2000). On the other hand, there are interactions between bradykinin and a classical hormonal transmitter, example of such interactions that bradykinin, stimulates is the synthesis or release of prolactin and growth hormone (Chihara et al., 1982). Furthermore, the growth hormones and growth factors increase protein synthesis stimulate the proliferation and of mammalian cells (Montgomery et al., 1980). Additionally, bradvkinin stimulates the release of several cytokines as important molecules in cellular proliferation and differentiation of various blood cell progenitors (Özotürk, 2001). These cytokines include: interleukin-1 (IL-1), IL-3, IL-6, tumor necrosis factor- α (TNF- α) and interferon- γ (IF- γ), that known to affect recovery from radiation-induced hemopoietic injury (Neta, 1997 a and b and Straub et al., 2002). These findings thus establish a therapeutic role to the venom animals.

Carbon tetrachloride is one of the most commonly used hepatotoxins in the experimental study of liver diseases. The hepatotoxic effects of CCl₄ are largely due to its active metabolite, trichloromethyl radical (Johnson and Kroening, 1998). These activated radicals bind covalently to the macromolecules and induce peroxidative degradation of of endoplasmic membrane lipids reticulum rich in polyunsaturated fatty acids. This leads to the formation of lipid

peroxides. It is well known that MDA is a terminal product of lipid peroxidation (Palanivel et al., 2008). So the content of MDA can be used to estimate the extent of lipid peroxidation. The latter can indirectly reflect the status of the metabolism of free radicals, the degree to which the tissue cells are attacked by free radicals and the degree to which lipid is peroxidated (Messaraha et al., 2010). The hvdrogen superoxide anion $(O_2),$ peroxide $(H_2 O_2)$ and the hydroxyl radical (OH⁻) are the major reactive oxygen species in the body .Free radicals are produced as a consequence of normal metabolism and their activities are controlled by enzymatic defense mechanisms, such as the SOD, GPx and non-enzymatic CAT. and defense mechanisms, such as ascorbic acid, Vitamin E and GSH (Neradilova' et al., 1973; Benzie, 1996 and Subudhi et al., 2008). Furthermore oxidative damage arises when an imbalance occurs in this system, i.e. over-production of free radicals and/or a decrease in antioxidant defenses mechanisms (Favier, 2003). In fact, the increase of some antioxidant enzymes activities such as SOD, GPx and CAT. which are the main antioxidants in the body, may be indicative of the failure of compensating the induced oxidative stress (Fernandez et al., 2005; Iwuanyanwu et al., 2007). These enzymes may scavenge excess O_2^{-1} and H_2O_2 , and peroxides ROOH produced by free radicals. For example, SOD catalyzes the conversion of super oxide anion radical to H_2O_2 . The resulting hydrogen peroxide in turn is decomposed by the enzymes GPx and CAT (Ferna'ndez and Videla, 1989 and Venditti et al., 2003). It is worthy to mintion that, the exogenous bradykinin causes a decrease of hydrogen peroxide and malondialdehyde levels and an increase of antioxidative enzyme activity in hyperglycaemic rats (Mikrut et al., 2001) which indicates the important role of kinins in the development of oxidative

stress. Furthermore, the decreased level of hydrogen peroxide and malondialobserved after bradykinin dehvde. administration, may point to a reduction in free radicals production. Additionally, NADPH is a cofactor required for the resynthesis of reduced GSH. Reduced GSH regulates glutathione peroxidase activity and indirect activity of other antioxidative enzymes (Togashi et al., 1999 and Ramasamy and Agarwal, 2008). Therefore, the increase in NADPH level may lead to the activation of all examined antioxidative enzymes, additionally, the increase of SOD, CAT and GSH-Px activity may also be connected with the increase in kininmediated transport of proteins and amino acids (Mikrut et al., 2001). In the present study, the recorded results indicated that, the levels of GSH and MDA approached the normal in all animals treated with BPF exposed to CCl₄. Restoration of MDA to nearly normal levels by this fraction may be due to an enhancement of antioxidant enzyme, such as SOD, CAT and reduced GSH. Consequently, it could be suggested that the potentiated endogenous bradykinin due to the used venom fraction enhanced the activity of antioxidant enzymes

In conclusion BPF that isolated from *Leiurus quinquestriatus* venom normalized the hepatic injury induced by CCl₄, This normalizing was indicated by the increase of liver GSH content as well as CAT, SOD, total protein and albumin activities, and decrease in ALT, AST, and ALP. Therefore, BPF may have therapeutic values in treatment of CCl₄induced hepatic injury.

REFERENCES

- Abd El Dayem, S. M. and Moawad, K. M. (2001): Toxicity of CCl₄ in rat liver and the effects of antioxidant treatments. J. Egypt. Soc. Zool., 36 (A): 415- 442.
- Abd-El-Rahim, S. A. (1995): Effect of extracted crude venom of the jelly

fish, *Eutonina indicans* on kidney function. J. Egypt. Ger. Soc. Zool., 18 (A): 131 - 147.

- Abhilash, P. A.; Harikrishnan., R. and Indira, M. (2013): Ascorbic acid is superior to silymarin in the recovery of ethanol-induced inflammatory reactions in hepatocytes of guinea pigs. J. Physiol. Biochem. 69(4):785-98.
- Abhilash, P. A.; Harikrishnan, R. and Indira, M. (2014): Ascorbic acid suppresses endotoxemia and NF-κB signaling cascade in alcoholic liver fibrosis in guinea pigs: a mechanistic approach.Toxicol. Appl. Pharmacol. 274(2):215-224.
- Abu-Amra, E. and Abd-El-Rahim, S. A. (2000): Stimulation of some enzymes and cellular proliferation by factors separated from scorpion and jellyfish. J. Egypt. Ger. Soc. Zool., 31: 273 - 287.
- Adaramoye, O. A.; Osaimoje, D. O.;
 Akinsanya, M. A.; Nneji, C. M.;
 Fafunso, M. A. and Ademowo, O. G. (2008): Changes in antioxidant status and biochemical indices after acute administration of artemether, artemether-lumefantrine and halofantrine in rats. J. Compilation: Basic Clin. Pharmacol. Toxicol, 102: 412-418.
- Al-Jumaily, E. F.: Al-Obaidiand R. S. and Abdulla, J. M. (2014): Hepatoprotective Activity of Flavonoids Purified and Ethanolic Extract from Iraqi Propolis Against Carbon tetrachloride-Induced Liver Damage In Male Mice. IOSR Journal Of Pharmacy. 4(3): 22- 27.
- Basu, L., (2003): Carbon tetrachlorideinduced lipid peroxidation: eicosanoid formation and their regulation by antioxidant nutrients, Toxicology, 189: 113-127.
- Bekheet, S. H. M.; Awadalla, E. A; Salman, M, M. A. and Hassan, M. K. 2013): Prevention of hepatic and renal toxicity with bradykinin potentiating factor (BPF) isolated from Egyptian scorpion venom (*Buthus occitanus*) in gentamicin

treated rats. Tissue and Cell 45 (2013) 89–94.

- Benzie, I. F. (1996): Lipid peroxidation: a review of causes, conse- quences, measurement and dietary influences. Int. J. Food Sci. Nutr, 47: 233–261.
- Beutler, E.; Duron, O. and Kelly, B. M. (1963): Improved method for the determination of blood glutathione. J. Lab. Clin. Med., 61:882-888.
- Cetin, E. 1.; Kanbur, M.; Cetin, N.; Eraslan, G. and Atasever, A. (2011): Hepatoprotective effect of ghrelin on carbon tetrachloride-induced acute liver injury in rats. Regul Pept. (1-3):1-5.
- Chao, J.; Li, H.; Yao, Y.; Shen, B.; Gao, L.; Bledsoe, G. and Chao, L. (2007): Kinin infusion prevents renal inflammation, apoptosis, and fibrosis via inhibition of oxidative stress and mitogen-activated protein kinase activity.Hypertension, 49: 490–497.
- Chihara, K.; Iwaski, J.; Minamitani, N.;
 Kaja, H.; Matsukura, S.; Tamaki, N.;
 Matsumooto, S. and Fujita, T. (1982):
 Effect of vasoactive intestinal polypeptide on growth hormone secretion in perfused acromegalic pituitary adenoma tissue. J. clin. Endocrinol. Metab., 54(4): 773-779.
- Dacie, J. V. and Lewis, S. M. (1975): Practical haematology 5th edition. The English Language Book Society and Churchill Livingstone.
- Das, K. and Chainy, G. B. N. (2001): Modulation of liver mitochondrial antioxidant defense system by thyroid hormone. Biochem. Biophys. Acta., 1573:1–13.
- Doumas, B. T.; Watson, W. A. and Biggs, H. G. (1971): Albumin standards and the measurement of serum albumin with bromcresol green. Clin. Chim. Acta.,31(1): 87-96.
- Drotman, R. and Lawhan, G. (1978): Serum enzymes are indications of chemical induced liver damage. Drug Chem. Toxicol., 1: 163–171.
- El-Aaser, A. A. and El-Merzabani, M. M. (1975): Simultaneous determination of

5'-nucleotidase and alkaline phosphatase activities in serum. Z. Klin. Chem. Klin. Biochem., 13(10): 453-459.

- Favier, A. (2003): Le stress oxidant: inte're't conceptual et expe'rimental dans lacompre'hension des me'canismes des maladies et potentiel the'rapeutique. L'actualite' Chim., 108: 115.
- Fernandez, V.; Tapia, G.; Varela, P.; Romanque, P.; Cartier-Ugarte, D. and Videla, L. A. (2005): Thyroid hormone-induced oxidative stress in rodents and humans: a comparative view and relation to redox regulation of gene expression. Comp. Biochem. Physiol., 142: 213–239.
- Ferna'ndez, V. and Videla, L. (1989): Thyroid hormone, active oxygen, and lipid peroxidation. In: Miquel, J.; Quintanilha, A. T. and Weber, H. editors. Handbook of free radicals and anti- oxidants in biomedicine. Boca. Raton, FL: CRC Press Inc.; p.105– 115.
- Ferreira, S. H. (1965): A bradykininpotentiating factor (BPF) present in the venom of *Bothrops jararaca*. Br.J. Pharmac. Chemother., 24: 163 -169.
- Fossati, P.; Prencipe, L. and Berti, G. (1980): Use of 3,5-dichloro-2hydroxybenzenesulfonic acid/4aminophenazone chromogenic system in direct enzymic assay of uric acid in serum and urine. Clin. Chem., 26(2): 227-231.
- Iwuanyanwu, K. C. P.; Wegwu, M. O. and Ayalogu, E. O. (2007): the protective nature of Garlic, Ginger and Vitamin E on CCl₄-induced hepatotoxicity in rats. Asian J. of Biochemistry, 2 (6): 409-414.
- Jadon, A.; Bhadauria, M. and Shucla, S. (2007): Protective effect of Terminalia belerica Roxb., and gallic acid against carbon tetrachloride induced damage in albino rats. Ethnopharmacol., 109(2): 214-218.

- Kouyoumdjian, M.; Ngaoka, M. R.; Mauricio, R. L. and Durval, R. B. (2009): Portal hypertensive response to kinin. An. Acad. Bras. Cienc., 81 (3): 431-442.
- Johnson, D. E. and Kroening, C. (1998): Mechanism of early carbon tetrachloride toxicity in cultured rat hepatocytes. Pharmacol. Toxicol., 83: 231–239.
- Kumar, R. S.; Sivakumar, T.; Sivakumar, P.; Nethaji, R. Vijayabasker, M.; Perumal, P.; Gupta, M. and Mazumder. U. Κ. (2005): Hepatoprotective and in vivo antioxidant effects of Careva arborea against Carbon tetrachloride induced liver damage in rats. International J. of molecular Medicine and advanced sciences, 1(4): 418-424.
- Lin, W. C.; Wu, Y. W. and Shih, C. C. (2005): Aqueous extract of *Anoectochilus formosanus* attenuated hepatic fibrosis induced by carbon tetrachloride in rats. Phytomedicine, 12 (6/7): 453-460.
- Meier, J. and Theakston, R. D. Q. (1986): Approximate LD₅₀ Determinations of snake venoms using eight to ten experimental animals. Toxicon, 24(4): 395-401.
- Messaraha, M.; Boumendjelb, A.; Chouabiaa, A.; Klibetb. F.: Abdennoura, C.; Boulakouda, M. S. and ElFeki, A. (2010): Influence of thyroid dysfunction on liver lipid peroxidation and antioxidant status in experimental rats. Exp. Toxicol. Pathol., 62(3): 301-310.
- Mikrut, K.; Paluszak, J.; Kozlik, J.; Sosnowski, P.; Krauss, H. and Grzes'kowiak E. (2001): The effect of bradykinin on the oxidative state of rats with acute hyperglycaemia. Diabetes, Res. Clin. Pract., 51(2): 79– 85.
- Montgomery, R. E. X.; Dryer, R. L.; Conway, T. W and Spector, A. A. (1980): Biochemistery, A case oriented approach.Third edition, P.

586-612. Toronto –London: The C.V. Mospy Company.

- Muriel, P. and Garcipiana, T. (1992): Silymarin protects against paracetamol-induced lipid peroxidation and liver damage. J. Appl. Toxicol., 12(6): 439–442.
- Neben, S.; Donaldson, D. and Fitz, L. (1996): Interleukin-4 (IL-4) in combination with IL-11or IL-6 reverses the inhibitory effect of IL-3 on early B lymphocyte development. Exp. Hematol., 24: 783 - 789.
- Neradilova', M.; Hurba', F.; Nova'kova', V. and Blahosova', I. (1973): Investigations of the relationship between thyroid function and α tocopherol concentration of serum and in some organs of the rat. Int. J. Vitam Nutr. Res., 43: 283–90.
- Neta, R. (1997a): Modulation of radiation damage by cytokines. Stem Cells Dayt., 15 (Suppl. 2): 87 94.
- Neta, R. (1997b): Modulation with cytokines of radiation injury: Suggested mechanisms of action. Environ. Health Perspect., 105 (Suppl 6): 1463 - 1465.
- Nishikimi, M.; Appaji, N. and Yogi, K. (1972): The occurrence of superoxide anion in the reaction of reduced phenazine methosulfate and molecular oxygen. Biochem. Bioph. REs. Commun., 46(2): 849-854.
- Ohkawa, H.; Ohishi, N. and Yagi, K. (1979): Assay for lipid peroxides in animal tissues by thiobarbituric acid reaction. Anal. Biochem., 95(2): 351-358.
- Özotürk, Y. (2001): Kinin receptors and their antagonists as novel therapeutic agents. Curr. Pharm., 7(2): 135 - 161.
- Palanivel, M. G.; Rajkapoor, B.; Kumar, R. S.; Einstein, J. W.; Kumar, E. P.; Kumar, M. R.; Kavitha, K.; Kumar, M. P. and Jayakar, B. (2008): Hepatoprotective and Antioxidant Effect of *Pisonia aculeata* L. against CCl₄- Induced Hepatic Damage in Rats. Sci. Pharm., 76: 203–215.

- Pang, K. S.; Xin, X. and Stpierre, M. V. (1992): Determinants of metabolic disposition. Ann. Rev. Pharmacol. Toxicol., 32: 625–626.
- Peters, T. J. (1968): Proposals for standardization of total protein assays. Clin. Chem., 14 (12): 1147 - 1159.
- Ramasamy, K. and Agarwal, R. (2008): Multitargeted therapy of cancer by silymarin. Cancer Letters, 269: 352– 362.
- Rosa, M. P.; Gutiérrez, R. M. and Solís, R. V. (2009): Hepatoprotective and Inhibition of Oxidative Stress in Liver of *Prostechea michuacana*. Rec. Nat. Prod., 3: 146-151.
- Salman, M. M. A. (1995): Effect of a bradykinin potentiating factor isolated from scorpion venom, *Buthus* occitanus on burnt skin of Guinea pig in comparison with other drugs. M. S. Thesis, Faculty of science, Ain Shams University.
- Salman, M. M. A. (2002): Serological, Hematological and Biochemical studies on Bradykinin potentiating factor isolated from scorpion venom.M. S. Thesis, Faculty of science, Ain Shams University.
- Salman, M. M. A. (2009): Effect of a single dose of a bradykinin potentiating factor isolated from scorpion venom (*Buthuthus occitanus*) on total protein and albumin in serum of irradiated growing male Guinea pigs. Egypt. Acad. J. biolog. Sci., 1(1): 33-43.
- Salman, M. M. A. (2010): Effect of a single dose of а bradykinin potentiating factor isolated from scorpion venom on red blood cells indices in alloxan-induced diabetic 4^{th} rats. The Conference of Pharmaceutical Sciences. Aleppo. 13-15 April 2010.
- Sancho-Bru, P.; Bataller, R.; Fernandez-Varo, G.; Moreno, M.; Ramalho, L.
 N.; Colmenero, J.; Marí, M.; Clària, J.; Jiménez, W.; Arroyo, V.; Brenner, D. A. and Ginès, P. (2007):

Bradykinin attenuates hepatocellular damage and fibrosis in rats with chronic liver injury. Gastroentero-logy., 133(6): 2019-2028.

- Straub, R. H.; Pongratz, G.; Gunzler, C.; Michna, A.; Baier, S.; Kees, F.; Falk, W. and Scholmerich, J. (2002): Immunoregulation of IL-6 secretion endogenous and exogenous by adenosine and by exogenous purinergic agonists in splenic tissue slices. J. Neuroimmunol., 125 (1 - 2): 73 - 81.
- Subudhi, U.; Dasa, K.; Paital, B.; Bhanja,
 S.; and Chainy, G. B. N. (2008):
 Alleviation of enhanced oxidative stress and oxygen consumption of L-thyroxine induced hyper thyroid rat liver mitochondria by vitamin E and curcumin.Chem. Biol. Interact., 173:105–114.
- Thabrew, M. and Joice, P. (1987): A comparative study of the efficacy of Pavetta indica and Osbeckia octanda in the treatment of liver dysfunction. Planta. Med., 53(3): 239–241.
- Mnaa, S.; Aniess, W.; Yaser, O. and Emad ,S.(2015): Antioxidant Activity of White (Morus alba L.) and Black (Morus nigra L Berries against CCl4 Hepatotoxic Agent. Adv. Tech. Biol. Med. 3:1
- Thrall, K. D., Vucelick, M. E., Gies, R. A., and Benson, J. M. (2000): Comparative metabolism of carbon tetrachloride in

rats, mice, and hamsters using gas uptake and PBPK modeling. J. Toxicol. Env. Health, 60(A): 531–548.

- Tirkey, N.; Pilkhwal, S.; Kuhad, A. and Chopra, K. (2005): Hesperidin, a citrus bioflavonoid, decreases the oxidative stress produced by carbon tetrachloride in rat liver and kidney. BMC., 5(1): 2-6.
- Togashi, H.; Shinzawa, H.; Wakabayashi, H.; Nakamura, T.; Yamada, N.; Takahashi, T. and Ishikawa, M. (1999): Activities of free oxygen radical scavenger enzymes in human liver. J. Hepatol., 11: 200–205.
- Young, D. S.; Thomas, D. W.; Friedman,
 R. B. and Pestaner, L. C. (1972):
 Effects of drugs on clinical laboratory
 tests. Clin. Chem., 8(10):1041-303.
- Venditti, P.; De Rosa, R. and Di Meo, S. (2003): Effect of thyroid state on H2O2 production by rat liver mitochondria. Mol. Cell Endocrinol., 205: 185–192.
- Wang, G. and Salahudeen, A. K. (1995): Lipid peroxidation accompanies cyclosporine ephrotoxicity: effects of vitamin E. Kidney Int., 49: 927-934.
- Ward, F. M. and Daly, M. J. (1999): Hepatic disease, In: Clinical Pharmacy and Therapeutics. Walker, R. Edward, C. editors. New York; Churchill Livingston, p. 195-212.

ARABIC SUMMERY

تأثيرات بيوكيماوية لعامل منشط للبراديكينين معزول من سم العقرب (ليرس كوين كويستراتس) ضد أضرار رابع كلوريد الكربون الكبدية في الجرذان البيضاء

محمد محمود علي سالمان قسم علم الحيوان- كلية العلوم-جامعة جنوب الوادي بقنا- مصر

لقد أكدت البحوث العلمية الحديثة أن استخلاص مستخلصات من السموم الطبيعية مثل سموم العقارب والثعابين وغيرها لها دور واعد في علاج كثير من الأمراض ومن بين تلك المستخلصات عامل منشط للبر اديكينين الذي تم عزله وتعريفه في در اسات عديدة سابقة علي سموم العقارب والثعابين (Bradykinin دور والمتعابين الذي تم علاج أمراض الكبد في الجرذان ذلك المستخلص في علاج أمراض الكبد في الجرذان

و من ثم كان هذا البحث علي دراسة تأثيرت بيوكيماوية لعامل منشط للبراديكينين في علاج الكبد من التسمم برابع كلوريد الكربون في الجرذان البيضاء. ولذا فقد تم تقسيم ذكور الحيوانات الى ثلاث مجموعات كالتالى: المجموعة الأولى اعتبرت مجموعة ضابطة فقد تم حقنها بمحلول فسيولوجي (كلوريد الصوديوم) ٩. % فقط بينما حقنت المجموعة الثانية برابع كلوريد الكربون (٥. مل/كجم) مرتين أسبوعيا. وأما المجموعة الثالثة فقد تم حقنها كالمجموعة الثانية و تم حقنها بجرعات متتالية بعامل منشط البراديكينين كل خمسة ايام (١ ميكروجرام لكل جرام) من وزن الجسم. ثم تم ذبح الحيوانات وكذلك تم جمع عينات الده وجزء من نسيج الكبد طبقا لتصميم التجربة (أي بعد ١٥ يوما ممن العلاج و كذلك بعد ٣٠ يوما من العلاج.)

فلقد أدى الحقن برابع كلوريد الكربون الى زيادة معنوية فى مستويات و أنشطة انزيمات الكبد مثل الألانين تر انسفيريز ، الأسبارتيت تر انسفيريز ، و الألكاين فوسفاتيز . كما ارتفع مستوى البروتين الكلى و الألبومين فى مصل الدم بينما ارتفع تركيز انزيم مالون داى ألداهيد فى أنسجة الكبد للفئر ان البيضاء المجموعة الثانية بالمقار نه بالمجموعة الاولى (الضابطة) . وقد سجلت النتائج السابقة بعد الاسبوع الثانى حتى وصلت مداها بعد الاسبوع الرابع . أما المجموعة الثالثة فقد سجلت انتائج السابقة بعد الاسبوع الثانى حتى وصلت مداها بعد الاسبوع فى مصل الدم وكذلك المالون داى ألداهيد فى أنسجة الكبد مقار نه بالمجموعة الثانية بالمقار نه فى مصل الدم وكذلك المالون داى ألداهيد فى أنسجة الكبد مقار نه بالمجموعة الثانية المسممة برابع كلوريد الكربون . ومن ناحية أخرى، فقد اظهرت المجموعة الكبد مقار نه بالمجموعة الثانية المسممة برابع كلوريد المجموعة الثانية بالمقار داى ألداهيد فى أنسجة الكبد مقار نه بالمجموعة الثانية المسممة برابع كلوريد المربون . ومن ناحية أخرى، فقد اظهرت المجموعة المسممة برابع كلوريد الكربون انخفاضا معنويا لمعدل المجموعة الثانية بالمقار نه الموعة الفهرت المجموعة المسممة برابع كلوريد الكربون انخفاضا معنويا لمعدل المجموعة الثانية بالمقار نه الموعموعة الاولى (الضابطة) . و مما جذب الانتباه أن حقن حيوانات المجموعة الثالثة بعامل منشط البر اديكينين أدي الى تحسن فى تركيز القياسات السالفة الذكر و ذلك عند مقارنتها بالمجموعة الثالثة بعامل منشط البر اديكينين أدي الى تحسن فى تركيز القياسات السالفة الذكر و ذلك عند مقارنتها بالمجموعة ال التالثة بعامل منشط البر اديكينين أدي الى تحسن فى تركيز القياسات السالفة الذكر و ذلك عند مقارنتها بالمجموعة المحابطة) لوحظ أن التحسن فى النتائج يقترب من المعدل الطبيعى. و هذا يدل على أن للمستخلص السمي (الضابطة) لوحظ أن التحسن فى النتائج يقترب من المعدل الطبيعى و هذا يدل على أن المستخلص السمي (الضابطة) لوحظ أن التحسن فى النتائج يقترب من المعدل الطبيعى. وهذا يدل على أن للمستخلص السمي (الضابطة) لوحظ أن الماسين فى السامة لرابع كلوريد الكربون والتي نجم عنها اضرارا حادة للكبد .