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Impact of nano calcium hydroxide and calcium oxide on some metabolic activities and phenolic compounds of *Lupinus termis* seedlings

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Abstract The effect of 0.5, 1.0 and 2.0 mM nano calcium hydroxide Ca(OH)₂ and calcium oxide CaO or bulk calcium carbonate CaCO₃ on growth and metabolic changes of *Lupinus termis* seedlings was assessed. Root fresh and dry weights were increased by nano CaO at 0.5 and 1.0 mM compared with the control, while they were decreased by increasing nano Ca(OH)₂. Shoot fresh and dry weights were increased by increasing the concentration of nano Ca(OH)₂ and CaO reaching their maximum at 0.5 and 2.0 mM, respectively. Bulk CaCO₃ concentrations increased root and shoot fresh and dry weights at the highest concentration (2mM). The photosynthetic pigments (chl.a, chl.b and carotenoids) were markedly increased with increasing the concentration of nano Ca (OH)₂, nano CaO and bulk CaCO₃ with the highest magnitude recorded for nano Ca(OH)₂, while the lowest was for CaCO₃. The total soluble carbohydrate biosynthesis of both shoot and root was reduced by nano calcium particles, whereas it was enhanced by bulk calcium compared with control. The root soluble protein was decreased by different concentrations of either nano or bulk calcium, while shoot soluble protein was increased by all calcium treatments compared with the control. The nano calcium especially Ca(OH)₂ activated the protein metabolism compared with the bulk. The root and shoot phenolic compounds were decreased by different concentrations of either nano or bulk calcium, except of the shoot phenolic compounds at CaCO₃. Nano CaO at 0.5 mM increased shoot phenolic compounds by 10% compared with control.

Keywords: *Lupinus termis*, Nano calcium, Bulk calcium, Photosynthetic pigment, Growth criteria, Carbohydrates, Protein, Phenolic compounds.

INTRODUCTION

Lupinus termis (Lupin) plant is an herbaceous perennial plant belonging to the order fabales, family

Fabaceae, subfamily papilionoideae (EFSA, 2005). The plant root is a normal tap root. The plant has a simple raceme inflorescence which produces as many as 70

flowers on the main stem. The flowers open in ascending order into cream orbicular or flattened shape pod.

The genus *lupinus* includes over 450 species worldwide (EFSA, 2005) but only the following four major cultivated species have gained agricultural importance viz., white lupin (*Lupinus albus* L.), blue lupin (*L. angustifolius* L.), yellow lupin (*L. luteus* L.) of the “Old World” or Mediterranean area lupin species and one “New World” species namely Andean lupin (*L. mutabilis* L.). Both of the last species have wild forms and cultivated crops (Gladstones, 1998; Erbas *et al.*, 2005) in Mediterranean region, Sudan, Ethiopia, Central and Western Europe, Australia, USA and South America, Tropical and Southern Africa, Russia, and Ukraine (Kwak *et al.*, 2000; Kettel *et al.*, 2003).

Plant seeds are high in protein, lipids, dietary fiber, carbohydrates and calcium (Gabrial and Morcos, 1976; Petterson, 1998 and Bhardwaj *et al.*, 2010). Lupin (quinolizidine alkaloid) is responsible for bitter taste and neurotoxins (Yeheyis *et al.*, 2011). The major *Lupinus albus* seed alkaloids are Lupanine (Huyghe, 1997; Olver, 1998; Getachew, 2009), hydroxyaphylline, albine, multiflorine (Getachew, 2009), anagrain (Yildiz, 2011) and sparteine (Huyghe, 1997; Erbas *et al.*, 2005).

Lupin has food applications, particularly as desirable additives in bakery and pasts products, or as meat, egg and milk replacers, traditional and alcohol production, snack as well as in dietary and functional food products (Loza and Lampart- Szczapa, 2008). Lupin protein with fibers lowers serum cholesterol level, improves glucose tolerance and consequently modifies blood insulin and glucagon (Chango *et al.*, 1998). The seeds are used as carminative, diuretic, emmenagogue, hypoglycemic, pectoral and vermifuge and are used as well as a poultice on ulcers. (Chiej, 1984).

The field of nanotechnology is one of the most active areas of research in modern material science. Nanoparticles exhibit completely new or improved properties based on their specific characteristics such

as size, distribution and morphology affecting their physical and chemical characteristics. Nanoparticles (NPs) or (Nano scale particles= NSPs) are defined as ultrafine particles with lengths in two or three dimensions greater than 1 nm and smaller than 100 nm (ASTM, 2006). The physical and chemical properties of nanoparticles can differ significantly from those of bulk materials of the same composition. Due to NPs unique physical and chemical characteristics, recently they are widely used in many areas such as industry, agriculture, business, medicine, public health among many other (Rao and Shekhawat, 2014).

Ruffini-Castiglione and Cremonini (2009) have identified three types of NSPs: natural, incidental and engineered (can be carbon-based or metal based materials (Peralta-Videa *et al.*, 2011). Metal oxide is one of the metal- based groups. Metal oxide nanoparticles such as $\text{Ca}(\text{OH})_2$ or CaO are important materials due to their widespread applications in various aspects including catalysis, sensors, optoelectronic materials, and environmental remediation (Oskam, 2006).

Calcium is the fifth most abundant element (by mass), usually found in sedimentary rocks in the mineral forms of calcite, dolomite and gypsum. It is one of the main macro essential nutrients for plant growth and development; formation of cellular walls, enzyme activators, metabolic processes, nitrate uptake, biomass accumulation (Savithramma, 2002) and photosynthetic rate (Savithramma, 2004; Savithramma *et al.*, 2007).

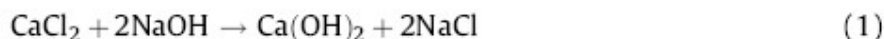
The aim of this study is to evaluate the effect of both nano calcium hydroxide and calcium oxide as well as bulk calcium on the growth and some metabolic activities of *Lupinus termis*

MATERIALS AND METHODS

Synthesis of calcium oxide and calcium hydroxide nanoparticles

Calcium oxide (CaO) nanoparticles were prepared by calcination of $\text{Ca}(\text{OH})_2$ which was synthesized by adding NaOH solution to $\text{CaCl}_2 \cdot 2\text{H}_2\text{O}$ solution without using any surfactant, organic medium

and complicated tools (Mirghiasi *et al.*, 2014), according to the reactions illustrated in equations. (1) and (2).



This method can be used as a simple, cheap and convenient way for producing calcium oxide and hydroxide nanoparticles on a large industrial scale.

Characterization of nano calcium

XRD Analysis

The crystalline structure of Ca(OH)_2 and CaO nanoparticles was characterized by X-ray diffraction (XRD, Shimadzu 6000 system, $\lambda = 1.54 \text{ \AA}$).

TGA Analysis

Thermal behavior for the precursor was studied through thermogravimetric analysis (TGA), which was performed with a Shimadzu DTA- SO under nitrogen flow.

TEM Analysis

The morphology and average particle size of nano-particles were further investigated by a transmission electron microscope (TEM) (JEOL- Jem-2100).

Plant materials

The experimental plant used in the present investigation was *Lupinus termis* (Lupinus) cv. Giza 1. Seeds were obtained from the Egyptian Ministry of Agriculture, Giza, Egypt.

Experimental Design:

During the growth season (November and December, 2014), a fixed number of *Lupinus termis* seeds were sterilised by 0.01% HgCl_2 for one min, washed thoroughly with distilled water and divided into four groups. Each group was sown in plastic pots (15 cm diameter and 10 cm depth) filled with 5 kg clay-sandy soil (2:1 w/w) and 5 pots were used for each treatment. The first group of seeds was sown in untreated soil representing the control. The second one was sown in soil supplemented with different nano calcium hydroxide [Ca(OH)_2] concentrations (0.5, 1.0

and 2mM). The third group was sown in soil provided with different nano calcium oxide (CaO) concentrations (0.5, 1 and 2mM) and the seeds of the fourth one were sown in a soil supplemented with different calcium carbonate concentrations (0.5, 1.0 and 2 mM) and was considered as bulk CaCO_3 . The seeds were left to germinated and grow as the usual practice and irrigated firstly with different concentrations of all calcium treatments to field capacity then irrigated with distilled water under greenhouse conditions. After 15 days of growth the seedlings were executed, washed with distilled water then separated into root and shoot. The growth criteria as lengths, fresh and dry weights of root and shoot were measured. Samples of three fresh leaves were kept frozen immediately for determination of the photosynthetic pigments. The remaining samples were dried in an oven at 60°C for determination of metabolic constituents.

Estimation of Photosynthetic Pigments:

The plant photosynthetic pigments (chlorophyll a, chlorophyll b, and carotenoids) were determined spectrophotometrically as recommended by Arnon, (1949) for chl. and Horváth *et al.* (1972) for carotenoids as adopted by Kissimon (1999). Chlorophyll concentration was calculated as mg g^{-1} dry weight of leaves.

Estimation of total soluble carbohydrates:

Carbohydrate extraction and clarification of plant materials (root and shoot) was performed according to Naguib *et al.* (1968). Total soluble carbohydrates content was estimated by the phenol sulphuric acid method described by Dubois *et al.* (1956) and Krishnaveni *et al.* (1984). The

concentration of total soluble carbohydrates content was calculated as mg/g dry weight.

Determination of total soluble protein

Total soluble protein content was determined in borate buffer extract according to the method described by Bradford (1976) using spectrophotometer (Model RAY LIGHT UV- 9200). The concentration of total soluble protein content was calculated as $\mu\text{g g}^{-1}$ dry weight.

Quantitative estimation of total phenolic compounds

Extraction of total phenolic compounds was carried out according to Velioglu *et al.* (1998) and aqueous methanol extract of total phenolic compounds was determined according to the Folin Ciocalteu's method using spectrophotometer (Model RAY LIGHT UV- 9200). A standard curve was prepared using

different concentrations of gallic acid and results were expressed as mg g^{-1} dry weight.

Statistical analysis:

The obtained results were statistically analyzed using one way of variance (ANOVA) and LSD was determined at 0.05 level. All statistical methods used in this study were according to Bishop (1983), while the analysis was carried out by SPSS statistical package.

Results

Nano particles were analyzed using different techniques to determine their physical nature. Thermogravimetric analysis (TGA) showed two weight losses from 375 to 480 °C and 480 to 650 °C (Figure 1). These losses of weight were due to the decomposition of $\text{Ca}(\text{OH})_2$ to CaO and the decomposition of CaCO_3 to CaO, respectively.

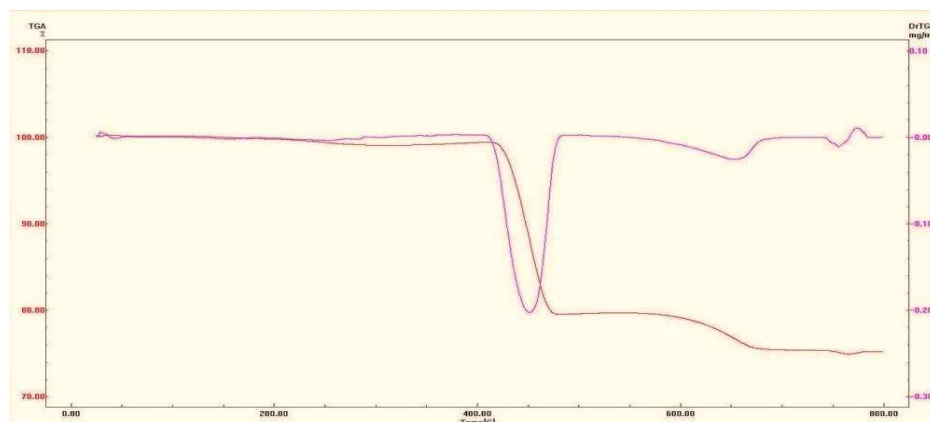


Figure1. TGA curves of the $\text{Ca}(\text{OH})_2$ nano-particles from 25 to 800°C.

X-ray diffraction (XRD) analysis revealed the crystalline nature and the size of both nano calcium (Figure 2). The distinct diffraction peaks at 2θ values of crystalline $\text{Ca}(\text{OH})_2$ and CaO were 3 peaks. The 2θ values of the strongest three peaks of crystalline $\text{Ca}(\text{OH})_2$ were 18.14, 37.19 and 47.20°, whereas the 2θ values of the strongest three peaks of crystalline CaO were 18.09,

34.17 and 37.46°. The average crystallite size of $\text{Ca}(\text{OH})_2$ and CaO nano-particles were calculated using Scherrer's equation (Eq. 3) to be about 73 and 91 nm for $\text{Ca}(\text{OH})_2$ and CaO, respectively .

$$D = \frac{0.9\lambda}{\beta \cos \theta} \quad (3).$$

Where D is the mean crystalline size (nm), λ is the wavelength of Cu K α (0.154 nm), β is the full width at

half maximum intensity (FWHM) in radian and θ is the Bragg angle ($^{\circ}$).

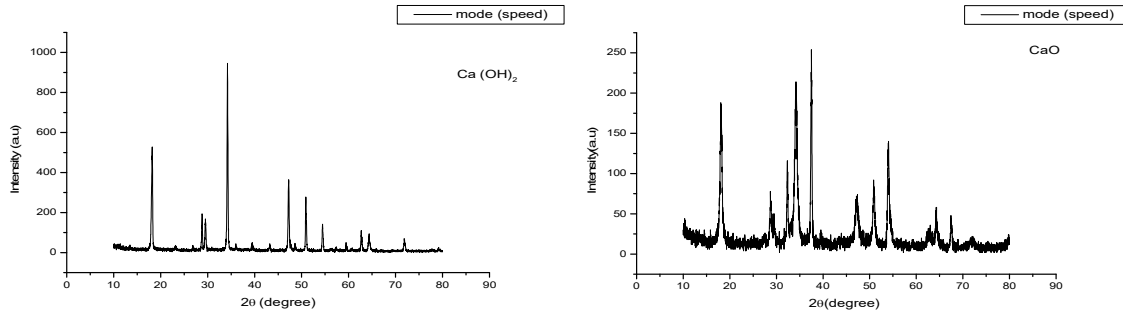


Figure 2. X-ray patterns of $\text{Ca}(\text{OH})_2$ and CaO nanoparticles

The transmission electron microscope (TEM) micrograph of $\text{Ca}(\text{OH})_2$ and CaO describe the nano particles structure (Figure 3). Through the TEM, nano $\text{Ca}(\text{OH})_2$ and CaO had hexagonal and spherical shapes,

respectively. The difference between XRD and TEM is that XRD shows the crystalline size of particles, while TEM shows their grain size of particles.

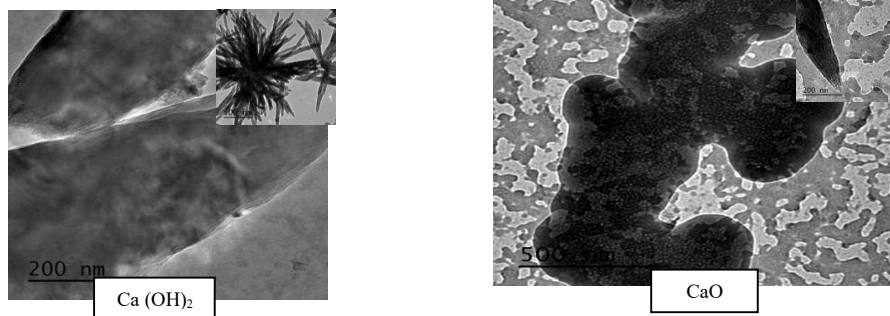


Figure 3. TEM image and distribution of $\text{Ca}(\text{OH})_2$ and CaO nanoparticles

Regarding the effect of different concentrations of both nano $\text{Ca}(\text{OH})_2$ and CaO and bulk CaCO_3 , the results indicated that all treatments decreased the seedling root length in comparison with the control (Figure 4). The decrease was attained by bulk CaCO_3 treatment. Nano $\text{Ca}(\text{OH})_2$ induced the least decrease in root length reaching 26.8, 47.9 and 0.43% respectively at concentrations of 0.5, 1.0 and 2 mM. The shoot length was not significantly affected by increasing nano $\text{Ca}(\text{OH})_2$ concentrations. The greatest decrease by nano CaO in root and shoot lengths was at

2 mM reaching 52.5 and 11.1%, respectively in root and shoot lengths. On the contrary 0.5 and 1.0 mM nano CaO significantly increased shoot length relative to the control. Increasing concentration of bulk CaCO_3 attenuated the decrease in root and shoot length and slightly increased shoot length at 2 m.

The results in Figure 4 indicated that with nano $\text{Ca}(\text{OH})_2$ there was a marked decrease in root fresh weight amounting to 46.3% at 1.0 mM compared with the control, whereas root dry weight was slightly increased by increasing concentration of nano

Ca(OH)₂, with the exception of 11.5% decline at 1.0 mM nano Ca(OH)₂ treatment. It is clear that nano Ca(OH)₂ markedly increased both shoot fresh and dry weights by 34.0 and 29.0% respectively at its highest concentration (2.0 mM) compared with the control.

Application of nano CaO at concentrations of 0.5 and 1.0 mM high significant increased both root and shoot fresh and dry weights, but 2.0 mM nano CaO decreased both root and shoot fresh and dry weights relative to the control.

On the other hand, 0.5 and 2.0 mM bulk CaCO₃ increased both root and shoot fresh weights compared to the control. Root dry weight was gradually decreased by increasing concentration of bulk CaCO₃, with the exception of a slight increase in its dry weight by 2mM bulk CaCO₃ in comparison with the control. Shoot dry weight increased by bulk CaCO₃ at 0.5 and 2 mM, whereas it decreased at 1.0 mM CaCO₃ treatment compared to control.

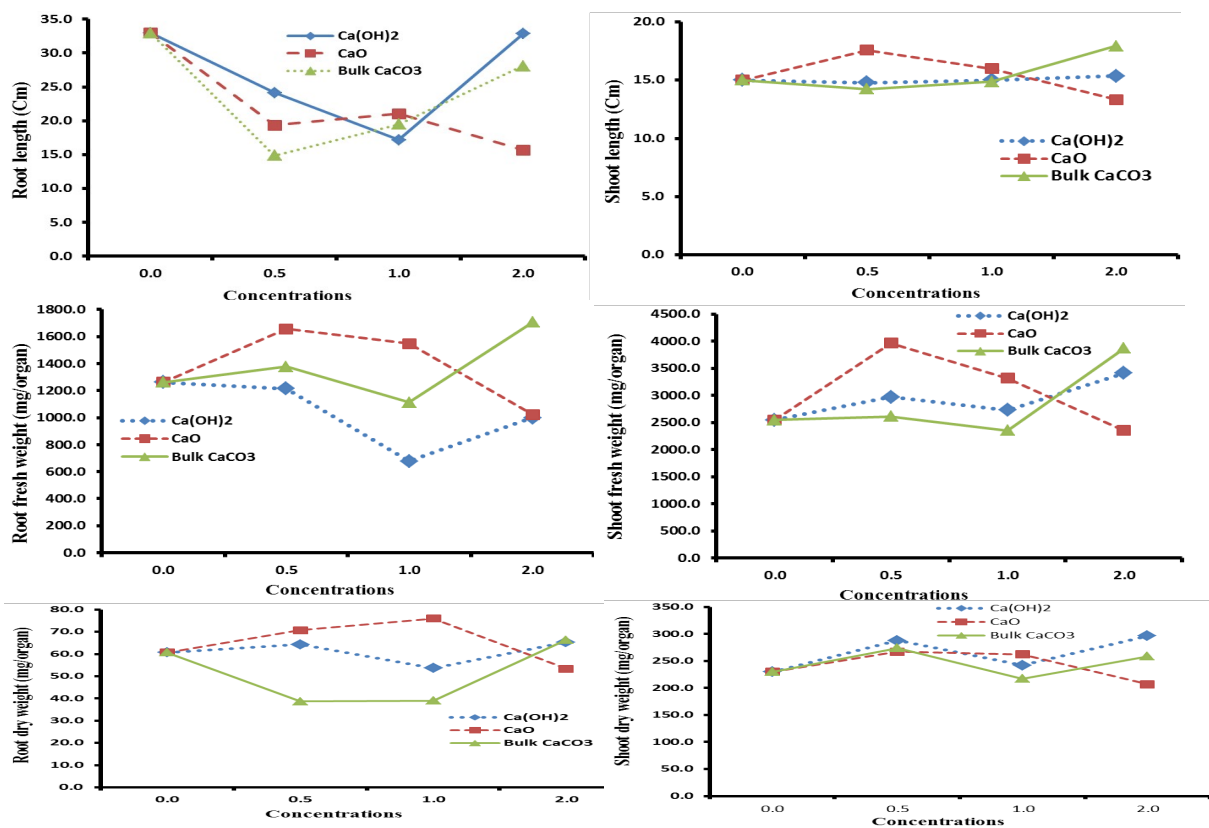


Figure 4. Effect of different concentrations of nano calcium either Ca(OH)₂ or CaO and bulk CaCO₃ (mM) on length (cm) and both fresh and dry weights (mg/organ) of root and shoot of

Lupinus termis seedling.

Application of both nano Ca(OH)₂ and CaO and bulk CaCO₃ induced pronounced increase in chlorophyll a, chlorophyll b and carotenoids with increasing their concentrations compared to control

(Figure 5). The content of Chl.a, Ch.b and carotenoids was increased by nano Ca(OH)₂ reaching their maxima at 2.0mM where it amount to 188.0, 161.7 and 187.6%, respectively. The magnitude of increase by nano CaO

was 152.4, 139.4 and 167.3%, respectively and by bulk CaCO_3 it reached 113.8, 118.7 and 92.2%, respectively compared with the control. This indicated that pigment

biosynthesis was greatest by nano Ca(OH)_2 compared to their nano CaO or bulk CaCO_3 .

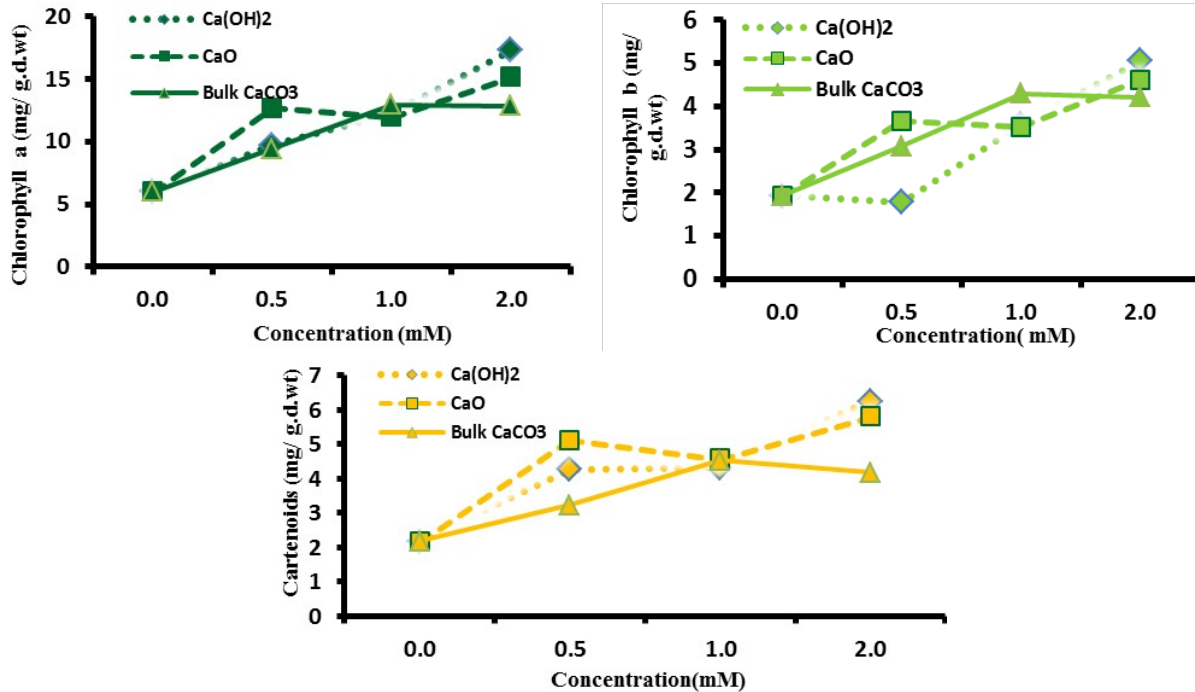


Figure 5. Effect of different concentrations of nano calcium either Ca(OH)_2 or CaO and bulk CaCO_3 (mM) on photosynthetic pigments (mg/g d.wt.) of *Lupinus termis* seedling.

Generally the content of total soluble carbohydrates tended to decrease in root and shoot parallelly to the increase in the concentration of both nano Ca(OH)_2 and CaO except for 2mM nano CaO which increased root total soluble carbohydrates by 46.4% relative to control (Figure 6). The content of total soluble carbohydrates was gradually decreased in root by increasing the concentration of bulk CaCO_3 , while it was increased in shoot reaching to 53.2% at 1.0 mM CaCO_3 compared with the control.

It is evident from data in Figure 6 that total soluble protein content was decreased in root and shoot

parallelly to the increase in the concentration of both nano Ca(OH)_2 and CaO and bulk CaCO_3 with the exception of the shoot total soluble protein which was increased with nano Ca(OH)_2 at 1.0 and 2.0 mM compared with the control. The percentage of increase in the content of shoot total soluble protein at 1.0 and 2.0 mM nano Ca(OH)_2 was 5.8 and 23% , orderly compared with the control.

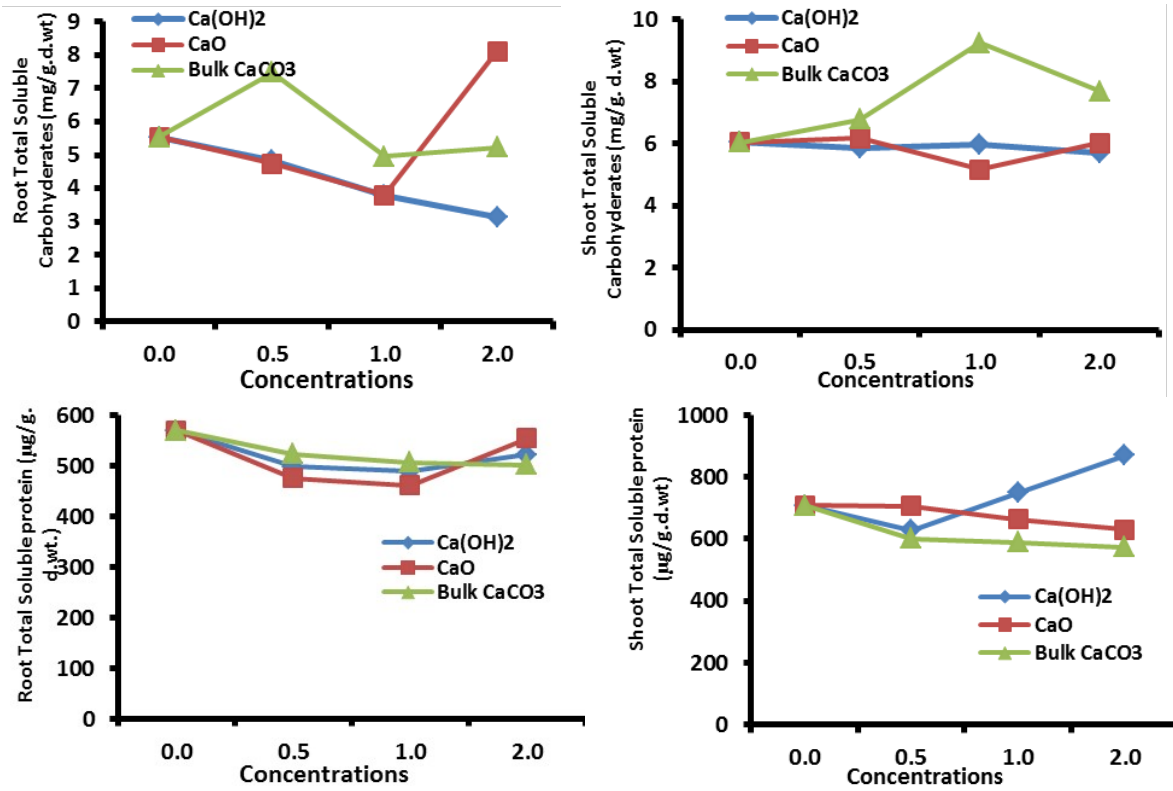


Figure 6. Effect of different concentrations of nano calcium either Ca (OH)₂ or CaO and bulk CaCO₃ (mM) on both root and shoot total soluble carbohydrates and proteins content (mg/g d. wt.) of *Lupinus termis* seedling.

The data in Figure 7 indicated that the content of root phenolic compounds was gradually decreased with increasing the concentration both nano calcium compounds and bulk CaCO₃ compared with the control. The magnitude of decrease was greater by nano compound at 0.5mM than by the bulk CaCO₃, while a reserve situation was observed at 1.0 and 2.0mM.

On the contrary, the content of shoot phenolic compounds was slightly increased at 1mM nano Ca(OH)₂ and at 0.5mM nano CaO and bulk CaCO₃ treatment achieving 4.3, 11.4 and 4.8%, respectively relative to the control.

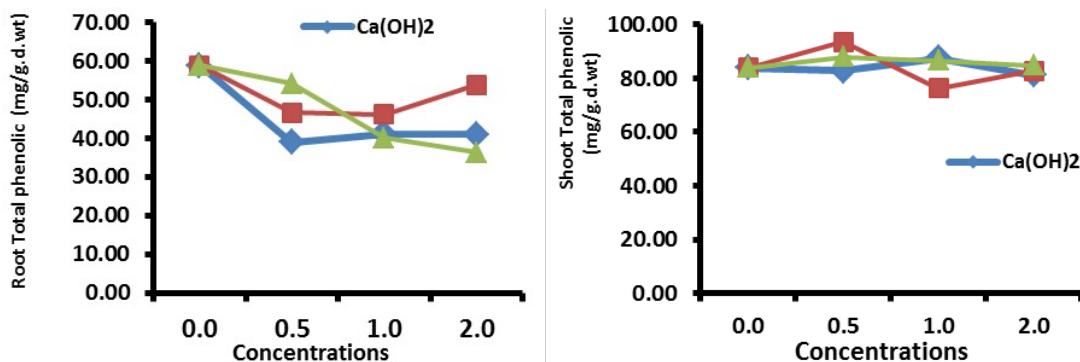


Figure 7. Effect of different concentrations of nano calcium either Ca (OH)₂ or CaO and bulk CaCO₃(mM) on root and shoot total phenolic compounds content (mg/g d. wt.) of *Lupinus termis* seedling.

Statistical analysis:

The statistical analysis (Table 1) indicated that different concentrations and forms of calcium applied to *Lupinus termis* seeds led to highly significant

variation ($P \leq 0.01$) in the seedling growth criteria, photosynthetic pigments and the investigated primary and secondary metabolites.

Table 1. Analysis of variance (ANOVA) for growth criteria, photosynthetic pigments and other metabolites under different concentrations of calcium forms and their interaction.

Parameters	Calcium		Interactions (Concentrations*Forms)
	Concentrations	Forms	
Root length	90.9**	15.2**	19.9**
Shoot length	1.5**	2.2**	22.3**
Root fresh weight	5.2**	16.4**	9.7**
Shoot fresh weight	22.6**	3.1**	31.6**
Root dry weight	6.6**	22.1**	17.6**
Shoot dry weight	24.6**	11.9**	13.9**
Chlorophyll a content	167219.8**	6033.7**	8898.8**
Chlorophyll b content	5432.6**	183.5**	502.9**
Carotenoids content	353334.0**	56973.4**	25349.6**
Root carbohydrate	16175.2**	27469.1**	24144.5**
Shoot carbohydrate	4592.9**	49311.5**	13749.8**
Root protein	1972.9**	48.9**	272.8**
Shoot protein	4282.3**	27619.0**	12397.2**
Root total phenolic	322258.1**	12.7**	11.2**
Shoot total phenolic	2973939.6**	12.1**	57.0**

* Significant at $P \leq 0.05$ ** highly significant at $P \leq 0.01$

Discussion

Very rare literatures were found about using nano calcium for improving plant growth and production (Liu *et al.*, 2005). Results of the present study showed that calcium in the nano forms was superior in its effects compared with the control. However, both root and shoot lengths of *Lupinus termis* seedlings were significantly decreased with increasing all calcium treatments from (0.5 to 2.0 mM).

These results agreed with those of Yin *et al.* (2011) who showed that Ag nanoparticles (AgNPs) has affected the root elongation and root growth of eleven wetland plants at very low concentrations. The present results are also in accordance with those obtained by Yasur and Usha Rani (2013) who found that using AgNPs (500- 4000 mg/l) did not affect shoot elongation patterns in the castor germinated seedlings. In addition, Yugandhar and Savithramma (2013)

showed that shoot length of *Vigna mungo* was increased by the application of CaCO_3 nanoparticles. On the contrary, treatment of *Bacopa monnieri* by bulk AgNO_3 exhibited a marked retardation with increasing concentration (Krishnaraj *et al.*, 2012). In the present study, the significant increase in both shoot fresh and dry weights at 2 mM nano Ca(OH)_2 and 0.5 and 1.0 mM nano CaO was in agreement with results of Yugandhar and Savithramma (2013) who reported that shoot fresh and dry weights of *Vigna mungo* were increased by CaCO_3 nanoparticles as well as by 10 mM CaCl_2 . In addition, TiO_2 nanoparticles were found to promote growth action of spinach (Yang *et al.*, 2006).

Application of high concentrations of both nano Ca(OH)_2 and CaO and bulk CaCO_3 induced pronounced increases in chlorophyll a, chlorophyll b and carotenoids relative to control. These results were in accordance with those obtained by Morteza (2013) and Tantawy *et al.* (2014) on *Zea mays* and tomato plants respectively. They reported that the content of chlorophyll (chl.a and chl.b) and carotenoids was significantly increased under nano TiO_2 spraying on the first plant and increased total chlorophyll content under 0.5 and 1.0 g/l nano calcium carbonate in the second one.

Total soluble carbohydrates content in root and shoot was significantly decreased by either nano calcium or CaCO_3 . This result was consistent with that of Krishnaraj *et al.* (2012) who found that carbohydrates content of root and leaf of *Bacopa monnieri* were dropped in levels during subsequent exposure to nano silver and bulk silver nitrate. The concomitant increase in carbohydrates content in shoot with the decrease in root may indicate their translocation.

The results indicated that total soluble protein content was decreased in root and shoot parallelly to the increase in concentration of both nano Ca(OH)_2 and CaO and bulk CaCO_3 s. These results agreed with those of Krishnaraj *et al.* (2012) who reported that the

protein content in different organs of *Bacopa monnieri* plants treated with AgNPs was lower than that of control.

Changes in total soluble carbohydrates and protein may be attributed to changes in processes associated with photosystems, starch synthesizing machineries and/or carbohydrates translocation as postulated by Krishnaraj *et al.* (2012).

The biological molecules such as secondary metabolites could possibly play a major role in the synthesis and stabilization of the nanoparticles (Inbakandan *et al.*, 2010). In the present study, increased total phenolic compounds in the seedlings produced from seeds treated with both nano Ca(OH)_2 and CaO and bulk CaCO_3 showed a shift towards secondary metabolism. The content of root phenolic compounds was gradually decreased with increasing concentration of both nano calcium and bulk CaCO_3 compared with control. This result was consistent with Yasur and Usha Rani (2013) who found that the content of total phenolic compounds was decreased at 2000 mg/l of AgNPs in castor seedlings compared with control and all other treatments. The increase in the content of shoot phenolic compounds by 1mM of nano Ca(OH)_2 and by 0.5 mM nano CaO and bulk CaCO_3 may be referred to shoot allocation for such compounds rather than translocation to root. This was in harmony with the finding of Krishnaraj *et al.* (2012) who found that the total phenolic compounds in the leaves of *Bacopa monnieri* was increased by both forms of silver (AgNPs and AgNO_3).

It can be concluded that using calcium nanoparticles in the form of either Ca(OH)_2 or CaO is more beneficial for improvement of plant growth than using it as bulk CaCO_3 .

REFERENCES

- Arnon, D.I. (1949). Copper enzymes in isolated chloroplasts, polyphenoxidase in beta vulgaris. *Plant Physiol.*, 24, 1-15.

- ASTM (2006). International Standard E2456-06, Standard Terminology Relating to Nanotechnology. (pdf available at http://www.astm.org/cgi-bin/SoftCart.exe/DATABASE.CART/REDLINE_PAGES/E2456.htm?L+mystore+sfjz7039; accessed 4 Dec 2006).
- Bhardwaj, H. L.; Starner, D. E. and Van Santen, E. (2010). Preliminary evaluation of white lupin (*Lupinus albus* L.) as forage crop in the mid-atlantic region of the United State of America. *Agric. Sci.*, 2, 13-17.
- Bishop, O. N. (1983). *Statistics in Biology*. Longman, Penguin, London., 56-63.
- Bradford, M.M. (1976). A rapid and sensitive method for the quantification of microgram quantities of protein utilizing the principle of protein dye binding. *Anal Biochem* 72, 248-254
- Chango, A.; Villaume, C.; Bau, H.M.; Schwertz, A.; Nicolas, J. and Mejean, L. (1998). Effects of casein, sweet lupin and sweet yellow lupin diet on cholesterol metabolism in rats. *Sci. Food Agric.* 76, 301–309.
- Chiej R. (1984). *The Macdonald Encyclopedia of Medicinal Plants*. Macdonald & Co. (Publishers) Ltd., Maxwell House, London. 447 pp.
- Dubois, M.; Gilles, K.A.; Hamilton, J.K.; Rebers. P.A. and Smith, F. (1956). Colorimetric method for determination of sugars and related substances. *Anal. Chem.*, 28, 350–356.
- EFSA., (2005). Opinion of the scientific panel on dietetic products, nutrition and allergies on a request from the commission related to the evaluation of lupin for labelling purposes. *EFSA J.*, 302, 1-11.
- Erbas, M.; Certel, M. and Uslu, M.K. (2005). Some chemical properties of white lupin seeds (*Lupinus albus* L.). *Food Chem.*, 89, 341-345.
- Gabrial, G.N. and Morcos, S.R. (1976). The use of *Lupinus termis* L. cultivated in Egypt, as a food protein supplement. *Z. Ernährungswiss.* 15, 333-339.
- Getachew, P. (2009). Chemical composition and the effects of traditional processing on nutritional composition of gibto (*Lupinus albus* L.) grown in, Gojam area. M.Sc. Thesis, Addis Ababa University, Addis Ababa, Ethiopia.
- Gladstones, J.S. (1998). Distribution, origin, taxonomy, history and importance. Chapter 1. In: J.S. Gladstones, C. Atkins, J. Hamblin (Eds.), *Lupins as Crop Plants: Biology, Production and Utilization*, CAB International, New York, USA, pp. 1–39
- Horváth, G.; Kissimon, J. and Faludi-Dániel, Á. (1972). Effect of light intensity on the formation of carotenoids in normal and mutant maize leaves. *Phytochem.*, 11, 183–187.
- Huyghe, C. (1997). White lupin (*Lupinus albus* L.). *Field Crops Res.*, 53, 147-160.
- Inbakandan., Venkatesan. and Ajmal Khan. (2010). Biosynthesis of gold nanoparticles utilizing marine sponge *Acanthella elongata* (Dendy, 1905), *Colloids and Surfaces B: Biointerfaces.*, 81(2), 634–639.
- Kettel, K., B.; Tuck, W.A.; Payne, C.; Chen, S. M. and Karow R., (2003). *Narrow-leaf lupin. Dryland Cropping Systems*, Oregon State University Extension Service. <http://ir.library.oregonstate.edu/xmlui/handle/1957/20321>.
- Kissimon, J. (1999). Analysis of the photosynthetic pigment composition. *Proceedings of the International Workshop and Training Course on Microalgal Biol. and Biotech.* Mosonmagyar, Hungary. 13-26.
- Krishnaraj, C.; Jagan, E.G.; Ramachandran, R.; Abirami, S.M.; Mohan, N. and Kalaichelvan, P.T. (2012). Effect of biologically synthesized

- silver nanoparticles on *Bacopa monnieri* (Linn.) Process Biochem., 47, 651–658
- Krishnaveni, S.; Theymoli B. and Sadasivam, S. (1984). Phenol sulphuric acid method. *Food Chem.*, 15, 229.
- Kwak, B.O.; Kim, H.J. and Park H.S. (2000). The effect of different lupin kernel inclusion levels on the growth and carcass composition of growing and finishing pigs. *Asian-Aust. Anim. Sci.*, 13, 207-212.
- Liu, X.; Zhang, F.; Zhang, S.; He, X.; Wang, R.; Feng, Z. and Wang, Y. (2005). Responses of peanut to nano calcium carbonate. *Plant nutrition and fertilizer science*. 11(3), 385-389.
- Loza, A. and Lampart-Szczapa, E. (2008). Allergenicity of lupin proteins – a review. *Polish Food Nutr. Sci.* 58, 283–287.
- Mirghiasi, Z.; Fereshteh, B.; Esmaeel, D. and Esmat E. (2014). Preparation and characterization of CaO nanoparticles from Ca(OH)₂ by direct thermal decomposition method. *J. Indust. and Engin. Chem.* 20, 113–117.
- Morteza, E.; Moaveni, P.; Aliabadi Farahani, H. and Kiyani, M. (2013). Study of photosynthetic pigments changes of maize (*Zea mays* L.) under nano TiO₂ spraying at various growth stages. *Springer Plus*, 2, 247-251.
- Naguib, M.I.; Youssef, E. and Kinawy, M.M. (1968). Effect of kinetin on the nitrogen metabolism of radish plants. *Bull. Fac. Sci. Cairo Univ.*, 42, 13-22.
- Olver, M.D., (1998). Effects of sweet, bitter and soaked micronised bitter lupins on duckling performance. *Br. Poult. Sci.*, 39, 622-626.
- Oskam, G. (2006). Metal oxide nanoparticles: synthesis, characterization and application. *Sol-Gel Sci. Technol.* 37, 161-164.
- Peralta-Videa, J.R.; Zhao, L.; Lopez-Moreno, M.L.; de la Rosa, G.; Hong, J. and Gardea-Torresdey, J.L. (2011). Nanomaterials and the environment: a review for the biennium 2008-2010. *J. Hazar. Mat.* 186, 1-15.
- Petterson ,D.S. (1998). Composition and food uses of lupins. In: Gladstones JS, Atkins C, Hamblin J (eds) *Lupins as crop plants. Biology: production and utilization*. CABI, Oxon, 353–384.
- Rao, S. and Shekhawat, G.S. (2014). Toxicity of ZnO Engineered Nanoparticles and Evaluation of Their Effect on Growth, Metabolism and Tissue Specific Accumulation in *Brassica juncea*. *Environ. Chem.Engin.*2, 105-114.
- Ruffini Castiglione, M. and Cremonini, R. (2009). NPs and higher plants. *Caryol.* 62,161–165
- Savithamma, N. (2002). Influence of calcium supply on biomass production of endemic and endangered tree species of tirumala hills of south Eastern Ghats. *Indian Botani. Soci.*, 81,323-326.
- Savithamma, N. (2004). Influence of calcium supply on photosynthetic rate in relation to calmodulin in endemic and endangered tree saplings of seshachlam hills of South Eastern Ghats of India. *Indian Plant Biol.*, 31(3), 1-6.
- Savithamma, N.; Fareeda, G.; Madhavi, V. and Murthy, S.S.D. (2007). Effect of calcium on photochemical activities of green leafy vegetables. *Plant Biology.* 34 (2), 95-98.
- Tantawy, A.S.; Salama, Y. A. M.; Abdel-Mawgoud, A.M.R. and Ghoname, A.A. (2014). Comparison of Chelated Calcium with Nano Calcium on Alleviation of Salinity Negative Effects on Tomato Plants. *Middle East Journal of Agriculture Research*, 3(4), 912-916.
- Velioglu, Y.S.; Mazza, G.; Gao, L. and Oomah, B.D. (1998). Antioxidant activity and total phenolics in selected fruits, vegetables and grain products. *Agric. Food Chem.* 46, 4113-4117.

- Yang, F.; Hong, F.; You, W.; Liu, C.; Gao, F.; Wu, C. and Yang, P. (2006). Influences of nano-anatase TiO₂ on the nitrogen metabolism of growing spinach. *Biological Trace Element Res.*, 110, 179-190.
- Yasur, J. and Usha Rani, P. (2013). Environmental effects of nanosilver: impact on castor seed germination, seedling growth, and plant physiology. *Environmental Science and Pollution Research*. 20 (12), 8636-8648.
- Yeheyis, L.; C. Kijora, Wink, M. and Peters, K.J. (2011). Effect of a traditional processing method on the chemical composition of local white lupin (*Lupinus albus* L.) seed in North-Western Ethiopia. *Z. Naturforsch. C*, 66,403-408.
- Yildiz, S., (2011). Rotational and nematicidal effect of lupine (*Lupinus albus* L.: Leguminosae). *Afr. Biotechnol.*, 10, 13252-13255.
- Yin, L.; Cheng, Y.; Colman, B.P.; Auffan, M.; Wiesner, M.; Rose, J.; Liu, J. and Bernhardt, E.S. (2011). More than the ions: The effect of silver nano particles on *Lolium multiflorum*. *Environ. Sci. and Technol.*, 45, 2360-2367.
- Yugandhar, P. and Savithamma, N. (2013). Green synthesis of calcium carbonate nanoparticles and their effects on seed germination and seedling growth of *Vigna mungo* (L.). *Hepper. Inter. Advanced Res.* 1(8), 89-103.

تأثير نانو هيدروكسيد وأكسيد الكالسيوم على بعض الأنشطة الأيضية والمركبات الفينولية لبادرات نبات الترمس
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فُيِم تأثير تركيزات مختلفه (0.5، 1.0، 2.0 مللى مولر) من هيدروكسيد وأكسيد الكالسيوم فى صورة النانو وكذلك كربونات الكالسيوم فى صورتها العاديه على النمو والتغيرات الأيضية لبادرات نبات الترمس. وأظهرت النتائج أن الوزن الغض والجاف للجذر قد زاد بتركزى 0.5 و 1.0 مللى مولر من نانو أكسيد الكالسيوم مقارنة بالكنترول، بينما كل من الوزن الغض والجاف للجذر قد نقص مع زيادة تركيز نانو هيدروكسيد الكالسيوم. وأزداد الوزن الغض والجاف للمجموع الخضرى مع زيادة تركيز كل من نانو هيدروكسيد وأكسيد الكالسيوم وصولاً الى قيمتهما القصوى بتركيز 0.5 و 2.0 مللى مولر على التوالى. وزادت كربونات الكالسيوم العادية الوزنين الغض والجاف للمجموع الجذرى والخضرى عند أعلى تركيز لها (2.0 مللى مولر). وكان هناك زياده ملحوظه فى أصباغ البناء الضوئى (كلوروفيل ا، كلوروفيل ب والكاروتينيدات) بزيادة تركيز نانو هيدروكسيد وأكسيد الكالسيوم وكربونات الكالسيوم العادية وفقاً لأعلى درجات سجلت للنانو هيدروكسيد الكالسيوم فى حين كان أدنى مستوى لكربونات الكالسيوم. انخفض تخليق الكربوهيدرات الذائبه الكليه للمجموع الجذرى والخضرى بالجزئيات النانويه للكالسيوم بينما تحسن تخليقها بكربونات الكالسيوم العاديه مقارنة بالكنترول. وأدت التركيزات المختلفه من نانو أكسيد وهيدروكسيد أو كربونات الكالسيوم الى انخفاض محتوى البروتينات الذائبه للجذر بينما ازداد محتواها فى المجموع الخضرى بكل أشكال الكالسيوم مقارنة بالكنترول. نانو كالسيوم خاصة نانو هيدروكسيد الكالسيوم نشط أيضاً البروتينات مقارنة بكربونات الكالسيوم العاديه. محتوى المركبات الفينوليه بالمجموع الجذرى والخضرى قد انخفض بالتركيزات المختلفه اما بالنانو كالسيوم أو بكربونات الكالسيوم العاديه بإستثناء محتوى المركبات الفينوليه للمجموع الخضرى المعامله بكربونات الكالسيوم العاديه . وقد ازداد المركبات الفينوليه للمجموع الخضرى بنسبة 10% عند تركيز 0.5 مللى مولر من نانو أكسيد الكالسيوم مقارنة بالكنترول.