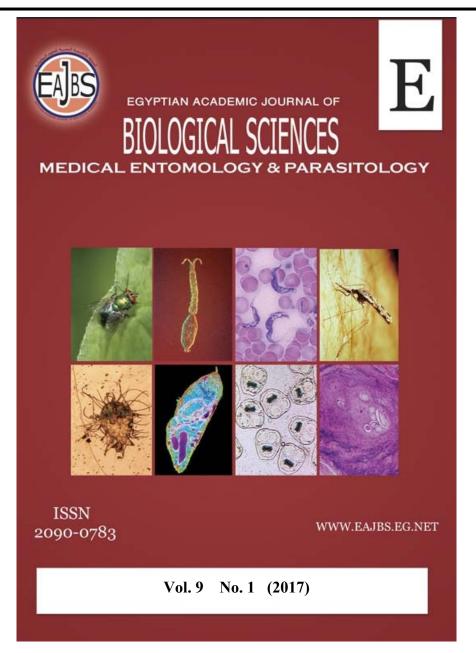
# Provided for non-commercial research and education use. Not for reproduction, distribution or commercial use.



The Journal of Medical Entomology and Parasitology is one of the series issued quarterly by the Egyptian Academic Journal of Biological Sciences. It is an important specialist journal covering the latest advances in that subject.

It publishes original research and review papers on all aspects of basic and applied medical entomology, parasitology and host-parasite relationships, including the latest discoveries in parasite biochemistry, molecular biology, genetics, ecology and epidemiology in the content of the biological, medical entomology and veterinary sciences.

In addition to that, the journal promotes research on the impact of living organisms on their environment with emphasis on subjects such a resource, depletion, pollution, biodiversity, ecosystem.....etc.

## www.eajbs.eg.net

Egypt. Acad. J. Biolog. Sci., 9(1): 21 –32 (2017)



**ARTICLE INFO** 

Accepted: 15/3/2017/1/2017

А

Ultrastructural Study

**Article History** 

Key words:

Eggs,

Schistosoma

Received: 20/1/2016

Egyptian Academic Journal of Biological Sciences E. Medical Entom. & Parasitology

> ISSN: 2090 – 0783 www.eajbs.eg.net



The Effect of A bleaching Solution on *Schistosoma mansoni* Eggs: A scanning Ultrastructural Study

Rafat Afifi<sup>1,2</sup> and Medhat Ali<sup>1,3</sup>

- 1- Biology Department, Faculty of Science, Taibah University, Saudi Arabia.
- 2- Marine Science Department, Faculty of Science, Suez Canal University, Ismalia, Egypt.
- 3- Zoology Department, Faculty of Science, Ain Shams University, Cairo, Egypt.

Medhat-S1@yahoo.com

# ABSTRACT

Schistosoma mansoni is one of the parasites causing schistosomiasis, a disease that threatens millions of people worldwide. The treatment of schistosomiasis is a critical point in research and includes fighting either adult worms or any of the different life cycle stages. Sodium hypochlorite (NaOCl) is the main constituent of the mansoni bleaching solution, in the present study different concentrations of the scanning bleaching solution were used to recognise their effects on the surface topography of Schistosoma mansoni eggs; 1, 5, 10 and 30 ppm of were used at time scale of 15, 30 and 60 minutes for each separate treatment. All of the concentrations experimented in this study affected the eggs if compared to the control ones. The surface topography of the eggs was greatly changed with increasing concentration and time, since, at higher concentration of the bleaching solution, some eggs were shrunk, deformed and egg shells were cracked. So, the bleaching solution may therefore be a useful treatment for *Schistosomiasis mansoni*.

# INTRODUCTION

Schistosomiasis is one of the most common human parasitic diseases in the world, and second most important parasitic infection after malaria in terms of public health and economic impact (Brooker *et al.*, 2009). Schistosomiasis is a major global health problem and principal among a number of neglected tropical diseases for the chronic impairment of both personal and group productivity (Gryseels *et al.*, 2006; Van der Werf *et al.* 2003). Schistosomiasis affects 207 million people in the developing countries, with approximately 800 million, mostly children, at risk of the infection in more than 70 tropical and subtropical countries, particularly in Africa, Asia and Latin America. *Schistosoma mansoni* (one type of intestinal schistosomiasis) represents the dominant species of Schistosoma (Christensen, 1984; Chitsulo *et al.* 2000; Grevelding 2004; Steinmann *et al.*, 2006; Andrade, 2009; El Ridi *et al.*, 2010; and El Ridi & Tallima 2013). There are five species of schistosomes that can infect humans, of which *Schistosoma mansoni* is the most important one. A disease related with schistosomiasis is not directly due to the adult worms but rather to the large numbers of eggs that trapped in tissues (Pearce and MacDonald 2002).

Generally, the *Schistosoma* life cycle is complex and needs a snail intermediate host and a mammalian definitive host. Human infection commences with cursorial penetration of the skin. After entrance into the circulatory system, parasites migrate through the lungs and eventually rest in the liver where they grow, differentiate, mate and then migrate to the urogenital system or mesenteric veins of the hepatoportal system to start egg production (Rai *et al.* 2009).

Eggs of Schistosoma mansoni can be targeted to stop the whole life cycle of this important human parasite; the surface topography of Schistosoma eggs was studied by Ford and Blankespoor (1979) who found that numerous microspines covering the surface of S. mansoni eggs and the density of these microspines varied somewhat among eggs. A similar observation was noticed by Eissa *et al.*, (2011) on the egg surface of S. described mansoni. but they these microspines as microspicules like chitinous projections.

studies different Many used compounds to investigate their effect on the such Schistosoma eggs as: calcium cyanamide synthetic drug which had an effect on the eggs of Schistosoma japonicum as well as on their miracidia (Zhou et al., 2015). The herbal extract of Pulsatilla chinensis had a suppressing effect on the hatching rates of eggs of Schistosoma japonicum (Chen et. al, 2013). Many studies also followed the effect of different compounds on the other different stages of Schistosoma such as miracidia, cercaria and adult stages (Magalhães et al., 2009; Ahmed and Rifaat, 2005 and Chen et al. 2013)

Sodium hypochlorite (NaOCl), which is derived from sodium chloride solutions is a convenient source of active oxygen. It is safely used in drinking water (Khan *et al.*, 2008). NaOCl has low molecular weight, so it can be easily penetrating the cellular membranes (Eventov *et al.*, 1998). NaOCl is a potent disinfectant destroying a variety of viruses, bacteria and fungi and can oxidize toxins that are present not only in blood, but also in tissues (Eventov *et al.*, 1998; Gerba and Kennedy, 2007; Abadias *et al.*, 2008; Peeters *et al.*, 2010).

Sodium hypochlorite is also used by Taha et al., (2014) to follow its effect on the eggs of Fasciola gigantica and on the intermediate host snail; NaOCl has an obvious deleterious effect on the surface of eggs and has also increased mortality rate in host snail, which is concentration and time dependent. Concerning schistosomiasis, NaOCl was found to immediately suppress the infectivity of S. mansoni cercariae (Al-Sharkawi, 1997), it also affected the cercarial surface integrity (El-Shaikh, 2001) and caused severe tegumental damage of S. mansoni adult worms (Attia, 2009).

As the sodium hypochlorite is used as a disinfectant destroying a variety of viruses, bacteria and fungi, so the aim of the present study is to evaluate its effect on the eggs of *Schistosoma mansoni* as a strategy to targeting an important stage of the life cycle of *S. mansoni* in a step of toward fighting schistosomaisis.

## MATERIALS AND METHODS

**Obtaining Eggs:** Eggs of Schistosoma mansoni were obtained from Theodor Bilaharz Institute, Giza, Egypt Treatment of eggs with a bleaching solution: A commercial bleaching solution, Clorox<sup>®</sup>, with 5.25% sodium hypochlorite (active ingredient) was used in the present study. The test solutions were prepared from a stock solution of 100 ppm diluted to 1, 5, 10 and 30 ppm. Eggs were divided into five groups; the first group was the control group (untreated eggs) in which eggs were kept in 0.85% saline in dark at 4°C, the other four groups (treated groups) of eggs were kept in dark at 4°C at specific concentrations (1, 5, 10 or 30 ppm) of the bleaching solution. Each of the four eggs' groups was subdivided into three subgroups; the first subgroup was kept for 15 minutes, the second subgroup was kept for 30 minutes and the third subgroup was kept for 60 minutes.

**Examination of eggs using Scanning electron microscopy (SEM):** All egg groups (control and treated) were fixed in 2.5% glutaraldehyde in phosphate buffer (0.1 mol/L, pH 7.4) at 4°C for 24 hours and postfixed in 1% osmium tetroxide in the same buffer for 1 hour. Eggs were then washed in the same buffer then dehydrated in graded series of ethanol (70%, 80% 95% and 100%) and dried using liquid carbon dioxide as a transitional medium. They were transferred on a specimen-holder and coated with a layer of gold in vacuum evaporator to prevent charging of the specimens. Eggs were examined and photographed with a Camscan DV4-scanning electron microscope, Grindel Biocenter. University of Hamburg. Hamburg, Germany.

## RESULTS

**The control group**: Eggs showed normal shape, their surface is covered with tiny projections without any marks, wrinkles or fissures, the spine in its normal position (Fig. 1).

*The first treated group*: The eggs treated with 1 ppm bleaching solution, for 15 minutes, had slightly wrinkled surfaces beside the spine. There was no change in the spine's appearance (Fig. 2A, B and C)., for longer treatment (30 and 60 minutes) eggs were marked with more shrinks and wrinkles (Fig. 2D and E).

The second treated group: The incubation of eggs in 5 ppm bleaching solution for 15 minutes, egg shell was slightly folded (Fig. 3A). Wrinkles and cracks were observed on the egg shells (Figs. 3B and C) after 30 minutes' treatment. Eggs exposed to 5 ppm for 60 minutes, were damaged, as many eggs had deformities in their shape and collapsed as well; some eggs were opened through big cracks (Figs. D, E and F). It is also observed that some areas of the eggs were devoid from the microprojections especially at the side near to the spine (3C, E and F).

**The third treated group**: Eggs incubated in 10 ppm bleaching solution for 15 minutes, showed much more wrinkles and shrinks on their surfaces (Fig. 4A), these wrinkles transformed into elevations and depressions (Fig. 4B) after 30 minutes' treatment. Some eggs were devoid from the microprojections that cover the egg surface (Fig. 4B). Deformed miracidium was also extruded from the egg. The eggs treated for 60 minutes were opened, as the egg shells were destroyed and extruded miracidia were affected with the bleaching solution, as some regions of the miracidium were devoid of cilia (Fig. 4C)

The fourth treated group: The largest concentration used in this study was 30 ppm; eggs treated with this concentration for 15 minutes had surfaces with great elevations and depressions, their shape changed and many deformities appeared to these eggs especially at the end opposite to the spin's which was also devoid end of microprojections, egg shells were cracked as well (Fig. 5A). Eggs treated with 30 ppm bleaching solution for 30 minutes were extremely deformed and highly collapsed (Fig. 5B). The last group of eggs which treated for 60 minutes were opened and the miracidium was emerged from the opened egg shell (Fig. 5C).

## DISCUSSION

In the present investigation, the eggs of Schistosoma mansoni were exposed to 1, 5, 10 and 30 ppm of the bleaching solution, for 15, 30 and 60 minutes. The current study showed that the effect of the bleaching solution was a dose and a time dependant. The surface of the egg shell changed gradually with increasing concentration and time within the same treatment. Many studies were performed to recognise the effect of various chemical compounds on different stages of trematodes. Concerning the direct effect of chemicals on the S. mansoni eggs; Zhou et al., (2015) followed the effect of calcium cyanamide (synthetic drug) on eggs of S. mansoni, and found a darkening of eggs colour gradually and the miracidia inside were atrophied as well as thickening in their egg shells. From the severe effects of this synthetic drug, the miracidial embryonic membrane stopped development 3-days after treatment; and the miracidia were severely deformed 7-days after treatment. In the present work,

wrinkles, shrinks were found on the egg shell surface and these changes may be due to increasing rigidity of the egg shell with increasing concentration and time of treatment. The Bleaching solution also affects the miracidium due to its penetration inside the eggs as some areas of the miracidium were devoid from cilia and this effect happened with higher concentrations (10 and 30 ppm) at longer exposure (60 The extrusion of deformed minutes). miracidium is due to the much hardness of the egg shell; the deformation of miracidium may be occurred due to the penetration of the bleaching solution inside the egg.

Eissa et al., (2011) studied the effect of miltefosine on different stages of S. mansoni, and they reported changes on the surface of the eggs, as some microspicules like chitinous projections were devoid from the egg surfaces due to the treatment of miltefosine. In the present work, a similar effect was observed, as the microprojections were devoid from the eggs treated with the bleaching solution at 5ppm for 30 and 60 minutes, at 10 ppm for 30 minutes and at 30 ppm for 15 minutes. Neves et al., (2010) concluded that the imidazolidines derivatives could have a role in destroying eggs through production of nitric oxide. In the present study, the bleaching solution/NaOCl is a source of active oxygen which could possibly damage the eggs of Schistosoma especially at higher concentrations.

Chen et al., (2013) studied the effect of herbal extract of Pulsatilla chinensis on some stages of S. japonicum, and they reported a suppression of hatching rate of eggs at lower concentration (4 microg/ml), They also observed an increase in miracidial death rate with increasing concentration of the extract; Similar results were also observed for the cercarial death rate. In the present work, the hatching of the eggs was observed after 60 minutes' exposure to 10 and 30 ppm of the bleaching solution, morphologically eggs were hatched and miracidia were deformed and emerged and this hatching may be due to the dryness and hardness of egg shells which may lead to

cracking of egg shells and their abnormal hatching.

Ahmed and Rifaat, (2005) studied the effect of the water extract of Solanum nigrum leaves on S. mansoni eggs, the number of eggs in hepatic tissues decreased significantly in treated mice. In another study by Magalhães et al., (2009), they found that, the effect of 5 and 10 µM, curcumin reduced the egg production of S. mansoni by 50% compared to control. In another study by Hegazi et al., 2007, they showed the effect of Egyptian propolis from three localities against Fasciola gigantica eggs; in their study, the propolis at specific concentrations lead to a complete failure of development and death of all immature eggs. In the present study, the number and death of eggs were not taken into account as the ultrastructural surface of eggs was the main criterion

El-Shaik et al., (2010) studied the ultra-structural surface of S. mansoni miracidia after their exposure to NaOCl; they reported obvious alteration to the dimensions and shapes of the miracidia and observed excessive damage of the miracidial body cilia. This damage to the miracidial body cilia was increasing while increasing concentration of NaOCl. In the present work, the loss of cilia from some regions on the miracidial body was observed; this loss of cilia may be due to the activity of free oxygen species produced from NaOCl. The same explanation was also mentioned by El-Shaik and his colleagues. NaOCl may also cause biosynthetic alteration in cellular metabolism and phospholipid destruction as suggested by Guida (2006). Hypochlorite is a powerful oxidant agent of biomolecules and NaOCl may cause an oxidative action with irreversible enzymatic inactivation. and degradation for lipid and fatty acid constituents (Krokosz, 2003, Cortezzo et al., 2004; Small et al., 2007). It is also noticed that, exposure of low-density lipoprotein (LDL) to NaOCl resulted in immediate and preferential oxidation of amino acid residues (Hazell and Stocker, 1993). NaOCl. produced by the mammalian host defence to

kill invading bacteria, targeted a select group of redox-regulated proteins, the thiol group (Leichert et al., 2008). The transcription of some bacterial genes is altered upon exposure of cells to high levels of the reactive oxygen species NaOCl (Peeters et al., 2010). Sodium hypochlorite has an effect on cercariae and adult worms of S. mansoni, as NaOCl showed deleterious effects against S. mansoni cercariae (Fripp et al., 1972) as well as an effect on the survival rate of cercariae which varied with its concentrations and the times of exposure (Al-Sharkawi, 1997).

An ultrastructural investigation on S. mansoni cercariae showed that NaOCl caused loss of tail region, vacuolization of the tegument with subsequent disruption of the apical syncytial tegument layer, as well as changes in the head region (El-Shaikh, 2001). NaOCl also decreased the S. mansoni worm motility and caused an extensive destruction of the tegument (Attia, 2009). In a study by Gray et al., (2006), they considered the effect of NaOCl on the diapausing of invertebrate eggs in the sediment introduced with transoceanic vessels and from Lake Erie (North America); as the sediments treated with NaOCl at concentrations 0 - 10,000 mg/l for 24 h, and they found that about 90% reduction in the hatching process at at1000 mg/l; This study performed to prevent species was introduction from foreign habitats. These results confirmed the effect of NaOCl on destruction of eggs which coincide with its present destructive effect on the eggs of S. mansoni. Egg shell of some trematode worms are affected by the treatment with concentrated acids such as sulphuric and hydrochloric acids that weaken the junction between the eggshell and operculum, while the same diluted acids and some alkalis such as potassium and sodium hydroxides do not affect this junction Madhavi (1968) and Balasubramanian et al., (2010)

#### CONCLUSION

In conclusion, this study showed that exposure of eggs of *S. mansoni* to different

concentrations of the bleaching solution for different periods resulted in concentrationdependent changes in their egg shell surfaces. Ultrastructural observations bleaching revealed that the solution markedly altered the topography of the surfaces and even collapsing and cracking shells especially the egg at high concentrations with prolonged treatment time. The alterations appeared on S. mansoni eggs may be linked to oxidative properties of NaOCl, the active ingredient of the bleaching solution. These results may indicate that the bleaching solution/NaOCl could become a cheap and effective molecule that can be used in schistosomiasis control. More work is needed to explore the exact mechanism of how NaOCl affect the biological materials.

#### ACKNOWLEDGMENT

Authors would like to thank Prof. Dr. Kamal El-Shaikh, for his advice and help in photographing the eggs by scanning electron microscope.

#### REFERENCES

- Abadias, M.; Usall, J.; Oliveira, M.; Alegre, I. and Vinas, I. (2008). Efficacy of neutral electrolyzed water (NEW) for reducing microbial contamination on minimally-processed vegetables. Int. J. Food Microbiol., 123:151-158.
- Ahmed, A. H.; Rifaat, M. M. (2005). Effects of *Solanum nigrum* leaves water extract on the penetration and infectivity of *Schistosoma mansoni* cercariae. J. Egypt Soc. Parasitol. 35(1):33-40.
- Al-Sharkawi, I. M. (1997). Chlorinated lime pockets: A model proposed for the control of *Schistosoma mansoni* transmission in irrigation streams and ditches in Egypt. J. Egypt. Ger. Soc. Zool., 23: 209-236.
- Andrade, Z. A. (2009) "Schistosomiasis and liver fibrosis". Parasite Immunol., 31: 656-663.
- Attia, W.Y. (2009). Scanning electron microscopy on the effect of sodium hypochlorite on adult *Schistosoma*

mansoni in vitro. Egypt. J. Exp. Biol. (Zool.), 5: 191-197.

- Balasubramanian, P.; Ramasamy, P.; Brennan, G. P. (2010). Fasciola gigantica: eggshell solubility and permeability properties" Indian J. Sci. Technol., 3: 17–20.
- Chen, Y. Q.; Xu, Q. M.; Zhuge, H. X.; Liang, Y. S; Li, X. R.; Yang, S. L. (2013). Effect of *Pulsatilla chinensis* (Bunge) Regel saponins on eggs, miracidia and cercariae of *Schistosoma japonicum*. Zhongguo Xue Xi Chong Bing Fang Zhi Za Zhi., 25(1):24-7.
- Chitsulo L, Engels D, Montreso A, Savioli L (2000). The global status of schistosomiasis and its control. Acta Trop., 77:41–51
- Christensen, N. O; Gotsche, G.; Frandsen, F. (1984). Parasitological technique for use in routine laboratory maintenance of schistosomes and for use in studies on the epidemiology of human and bovine schistosomiasis. Teaching Note, Danish Bilharziasis Laboratory.
- Cortezzo, D. E.; Koziol-Dube, K.; Setlow, B. and Setlow, P. (2004). Treatment with oxidizing agents damages the inner membrane of spores of *Bacillus subtilis* and sensitizes spores to subsequent stress. J. Appl. Microbiol., 97: 838-852.
- Eissa, M. M.; El Bardicy, S. and Tadros, M. (2011). Bioactivity of miltefosine against aquatic stages of *Schistosoma mansoni*, *Schistosoma haematobium* and their snail hosts, supported by scanning electron microscopy. Parasites & Vectors 2011, 4:73.
- El Ridi, R. A. and Tallima, H. A. (2013). Novel therapeutic and prevention approaches for schistosomiasis: review. J. Adv. Res., 4: 467-478.
- El Ridi, R.; Aboueldahab, M.; Tallima, H.; Salah, M.; Mahana, N.; Fawzi, S.; Mohamed, S. H. and Fahmy, O. M. (2010) *In vitro* and *In vivo* Activities of Arachidonic Acid against *Schistosoma mansoni* and *Schistosoma haematobium*. Antimicrob. Agents Chemother., 54 (8): 3383–3389.
- El-Shaikh, K.A. (2001) Scanning electron

microscopic study on the effect of sodium hypochlorite on *Schistosoma mansoni* cercariae. J. Egypt. Ger. Soc. Zool., 35: 1-16.

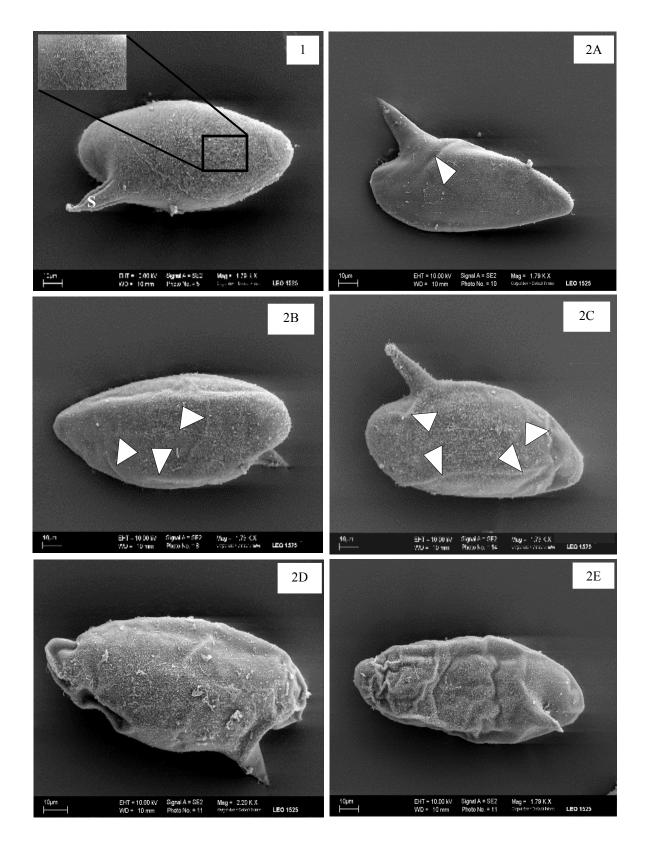
- El-Shaikh, k.; El-naggar, R. E.and Attia, w. y. (2010). Impact of sodium hypochlorite on *Schistosoma mansoni* miracidia and their ability to infect *Biomphalaria alexandrina* snails. proc. 6<sup>th</sup> int. con. biol. sci. (zool)., 6:368-374
- Eventov, V. L.; Andrianova, M. L. and Kukaeva, E. A. (1998). Detoxication and disinfection by sodium hypochlorite. Med. Tekh., 6: 36-39.
- Ford, J. W. and Blankespoor, H. D. (1979). Scanning electron microscopy of the eggs of three human schistosomes. Int. J. Parasitol., 9: 141-5.
- Fripp, P. J.; Armstrong, F. A. and Jaskulla, E. (1972). The cercariacidal properties of commercial hypochlorite preparations. S. Afr. Med. J., 46: 1819-1822.
- Gerba, C.P. and Kennedy, D. (2007). Enteric virus survival during household laundering and impact of disinfection with sodium hypochlorite. Appl. Environ. Microbiol., 73:4425-4428.
- Gray D. K.; Duggan, I. C. and MacIsaac H. J. (2006). Can sodium hypochlorite reduce the risk of species introductions from diapausing invertebrate eggs in non-ballasted ships? Mar. Pollut. Bull., 52(6): 689–695.
- Grevelding, G. C (2004). *Schistosoma*. Institute for Genetics, Heinrich-Heine-University, 40225 Düsseldorf, Germany. Magazine, R545
- Gryseels, B.; Polman, K.; Clerinx, J. and Kestens, L. (2006). Human schistosomiasis. Lancet, 368:1106–1118.
- Guida, A. (2006). Mechanism of action of sodium hypochlorite and its effects on dentin. Minerva Stomatol., 55: 471-482.
- Hazell, L. J. and Stocker, R. (1993). Oxidation of low-density lipoprotein with hypochlorite causes transformation of the lipoprotein into a high-uptake form for macrophages. Biochem. J., 290(1): 165-172.

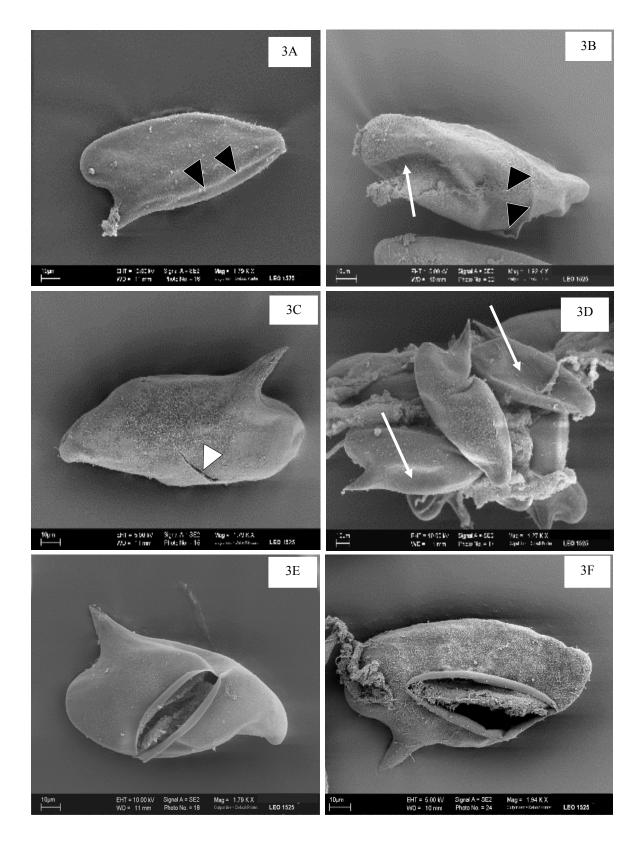
- Hegazi, A. G.; Abd El Hady, F. K.and Shalaby, H. A. (2007).
  Inhibitory effect of Egyptian propolis on *Fasciola gigantica* eggs with reference to its effect on *Clostridium oedematiens* and correlation to chemical composition. Pak. J. Biol. Sci., 10(19):3295-305.
- Khan, A.; Ullah, M. and Khan, M. (2008). Pathological effects of sodium hypochlorite administration through drinking water in male Japanese quails (*Coturnix japonica*). Hum. Exp. Toxicol., 27: 773-780.
- Krokosz, A. (2003). The effect of hypochlorite on human erythrocytes pretreated with X radiation. Cell. Mol. Biol. Lett., 8(1): 215-219.
- Leichert, L. I.; Gehrke, F.; Gudiseva, H. V.; Blackwell, T.; Ilbert, M.; Walker, A. K.; Strahler, J. R.; Andrews, P. C. and Jakob, U. (2008). Quantifying changes in the thiol redox proteome upon oxidative stress *in vivo*. Proc. Natl. Acad. Sci. USA., 105(24): 8197-8202.
- Madhavi, R. (1968). *Diplodiscus mehrai*: Chemical nature of eggshell, Exp. Parasitol., 23: 392-397.
- Magalhães, L. G.; Machado, C. B.; Morais, E. R.; Moreira, É. B. C.; Soares, C. S.; Silva, S. H.; Filho, A. A. S. and Rodrigues, V. (2009) "In vitro schistosomicidal activity of curcumin against *Schistosoma mansoni* adult worms" Parasitol Res., 104:1197–1201
- Neves, J. K. A. L.; Botelho, S. P. S.; Melo, C. M. L.; Pereira, V. R. A.; Lima, M. C. A.; Pitta, I. R.; Albuquerque, M. C. P. A. and S. L. Galdino (2010). Biological and immunological activity of new imidazolidines against adult worms of *Schistosoma mansoni*. Parasitol. Res., 107:531–538
- Pearce, E. J. and MacDonald, A. S. (2002). The immunobiology of schistosomiasis". Nat Rev Immunol., 2:499–511.
- Peeters, E.; Sass, A.; Mahenthiralingam, E.; Nelis, H. and Coenye, T. (2010): Transcriptional response of *Burkholderia cenocepacia* J2315 sessile cells to

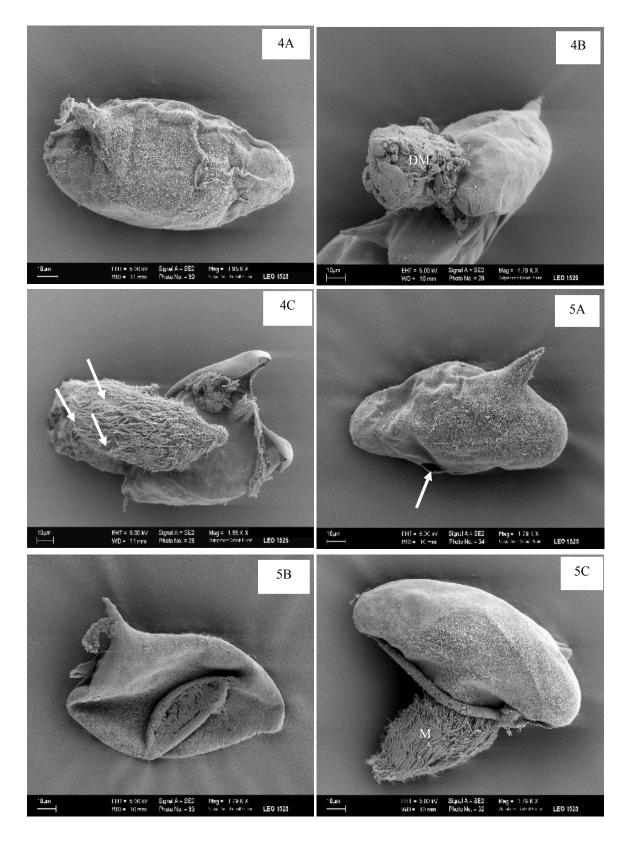
treatments with high doses of hydrogen peroxide and sodium hypochlorite. BMC Genomics, 11: 90-101.

- Rai, G.; Sayed, A. A.; Lea, H.; Wendy, A.; Luecke, F.; Chakrapani, H.; Prast-Nielsen, S.; Jadhav, A.; Leister, W.; Min, I.; James, A.; Christopher, P.; Keefer, L. A.; Elias, S. J.; Simeonov, A.; Maloney, D. J.; Williams, D. L. and Craig, T. J. (2009). Structure mechanism insights and the role of nitric oxide donation guide the development of oxadiazole-2-oxides as therapeutic agents against schistosomiasis. J. Med. Chem., 52:6474-6483
- Small, D. A.; Chang, W.; Toghrol, F. and Bentley, W. E. (2007). Toxicogenomic analysis of sodium hypochlorite antimicrobial mechanisms in *Pseudomonas aeruginosa*. Appl. Microbiol. Biotechnol., 74: 176-185.
- Steinmann, P.; Keiser, J.; Bos, R.; Tanner, M. and Utzinger, J. (2006).
  Schistosomiasis and water resources development: systematic review, metaanalysis, and estimates of people at risk. Lancet Infect. Dis., 6: 411-425.
- Taha, H. A.; El-Shaikh, K. A. and Al-Sadi, M. M. (2014). Effect of sodium hypochlorite on *Fasciola gigantica* eggs and the intermediate host, *Lymnaea natalensis*: A scanning electronmicroscopy study. J. Taibah Uni. Sci., 8:75-83
- Van der Werf, M. J.; de Vlas, S. J.; Brooker, S.; Looman, C. W.; Nagelkerke, N. J.; Habbema, J. D. and Engels, D. (2003). Quantification of clinical morbidity associated with schistosome infection in subSaharan Africa. Acta Trop., 86:125–139.
- Zhou, Y. S.; Peng, G. H.; Hu, Z. H.; Feng, X.
  W.; Zhu, R.; Wei, W. Y and Guo, J. G.
  (2015). Effect of calcium cyanamid synthetic drug on *Schistosoma japonicum* egg morphology. Zhongguo Xue Xi Chong Bing Fang Zhi Za Zhi., 27(1):56-8.

- Fig. 1: Scanning Electron Micrograph of the control (untreated) egg of Schistosoma mansoni, the surface and spine (S) are in normal architecture and covered with microprojections (inset). Magnification 1.79KX and Bar 10 μm.
- Fig. 2: Eggs treated with 1 ppm bleaching solution: A, B and C: Eggs treated for 15 minutes; some wrinkles and sutures were observed (white arrow heads) on the surface of the eggs. Magnification 1.79KX and Bar 10  $\mu$ m. D: Eggs treated for 30 minutes; more wrinkles were observed and some areas were shrunk especially on side opposite to the spine. E: Eggs treated for 60 minutes; much more wrinkles were observed and many shrinked areas were noticed especially on side opposite to the spine.
- Fig. 3: Eggs treated with 5 ppm bleaching solution; A: Eggs treated for 15 minutes; eggs showing a large wrinkle (black arrow heads) which is found on the lateral side of the egg beside the spine; Magnification A: 1.79KX and Bar = 10  $\mu$ m. B and C: Eggs treated for 30 minutes; Sutures (black arrow heads) as well as depression (white arrow) on the egg eggshell were observed. For C, a crack on the egg shell was observed (white arrow head); Magnification B: 1.92 KX and C: 1.79KX and Bar for B & C = 10  $\mu$ m. D, E and F: Eggs treated for 60 minutes; D, there is a deformity in the egg shells and eggs were collapsed (white arrows). E and F, egg shells were opened. For E and F egg shells were cracked and opened; Magnification D: 1.27KX, E: 1.79KX and F: 1.94 KX and Bar 10  $\mu$ m.
- Fig. 4: Eggs treated with 10 ppm bleaching solution; A: Eggs treated for 15 minutes; Eggs were greatly wrinkled and the surface is shrunk especially at the opposite side to the spine. Magnification 1.95KX and Bar 10  $\mu$ m. B: Eggs were treated for 30 minutes; the egg's surface is nearly similar to that in 4A, and a deformed miracidium (DM) was released from the egg. Magnification 1.79KX and Bar 10  $\mu$ m. C: Eggs were treated for 60 minutes; eggs were opened, as the egg shell was destroyed and the miracidium was released and some areas of the miracidium surface were devoid from cilia (white arrows); Magnification 1.55KX and Bar 10  $\mu$ m.
- Fig. 5: Eggs were treated with 30 ppm bleaching solution; A: Eggs were treated for 15 minutes; Some eggs were shrunk at opposite side to the spine, there was an opening (white arrow) due to the eggshell cracking. B: Eggs were treated for 30 minutes; eggs were extremely deformed and collapsed. C: Eggs were treated for 60 minutes; eggs were opened and a non-fully developed miracidium (M) was extruded. Magnification 1.79KX and Bar 10 μm.







#### **ABIC SUMMERY**

تأثير محلول مبيض على بيض شيستوسوما مانسونى فى المختبر: دراسة ماسحة للتراكيب الدقيقة

رأفت عفيفي<sup>٢،٢</sup> - مدحت علي<sup>٢،٢</sup> ١- قسم الأحياء، كلية العلوم، جامعة طيبة، المملكة العربية السعودية ٢- قسم علوم البحار، كلية العلوم، جامعة قناة السويس، الإسماعيلية، مصر ٣- قسم علم الحيوان، كلية العلوم، جامعة عين شمس، القاهرة، مصر

دودة البلهارسيا المعوية ( شيستوسوما مانسوني) هي واحدة من الطفيليات التي تسبب مرض البلهارسيا، وهو مرض يهدد ملايين البشر في جميع أنحاء العالم. ويمثل علاج مرض البلهارسيا تحدي للباحثين وقد يكون العلاج إما عن طريق التخلص من الديدان البالغة أو أي من مراحل دورة حياتها المختلفة. وفي هذه الدراسة تم استخدام محلول مبيض -هيبوكلوريت الصوديوم - لمكافحة البلهارسيا من خلال التأثير على بيض البلهارسيا المعوية. وقد استخدمت تركيزات مختلفة من هيبوكلوريت الصوديوم - لمكافحة البلهارسيا من خلال التأثير على بيض البلهارسيا المعوية. وقد أستخدمت تركيزات في هذه الدراسة على مقياس زمني قدره ١٥، ٣٠، ٢٠ دقيقة لكل معاملة منفصلة. وقد أو خلو ان ٢٠ ٢٠، ٢٠ في هذه الدراسة على مقياس زمني قدره ١٥، ٣٠، ٢٠ دقيقة لكل معاملة منفصلة. وقد أو خلو ان جميع التراكيز التي جُربت في هذه الدراسة أثرت على البيض إذا ما قورنت بالمجموعة الضابطة. كما أن سطح البيض قد تغير بشكل ملحوظ مع زيادة التركيز والوقت، حيث أنه عند التركيزات العالية لهيبوكلوريت الصوديوم، أنكمش بعض البيض وتشو هت قشرته. لذلك، فإن هيريكار من المعوديوم قد يكون علاج مفيد المعامية المنون