# Ameliorative Effect of Melissaofficinalison Hepatic Toxicity in Rats

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### Abstract

Due to the industrial revolution in recent days, several air pollutants as carbon tetrachloride (CCL<sub>4</sub>), which has been documented as hepato-degenerative agent; have cumulatively damaged the human health. Therefore, the objective of the current study aimed at investigating the effects of supplementing Carbon tetrachloride CCL<sub>4</sub>-exposed male Sprague Dawley albino rats with four levels 2.5, 5 and 10% of Melissa powderedon liver disease. The experiment was carried out on 30 male Albino rats divided into 5 equal groups (each group contain 6 rats); [1<sup>st</sup>group]fed on basal diet and kept as a control (-).

The other rats were intoxicated by subcutaneous injection of Carbon tetrachloride CCl<sub>4</sub> in paraffin oil (1:1 v/v; 2 ml/kg) twice a week for two consecutive weeks to induce hepatic chronic damage, then those rats were divided into: a group that control positivecontrol(+) [2<sup>nd</sup>group], a group that fed on basal diet + Melissapowder 2.5% [3<sup>rd</sup>group]; a group that fed on basal diet +melissapowder 5% [4<sup>th</sup>group]; a group that fed on basal diet diet+Melissapowder 10% [5<sup>th</sup>group]

The results showed that with increasing the amount of melissa powder, the total cholesterol, (TC) triglycerides, (TG), low density lipoprotein cholesterol,(LDL-c), very low density lipoprotein cholesterol,(VLDL-c), Aspartate transaminase, (AST), alanine transaminase,(ALT), uric acid, urea nitrogen, creatinine, Iron and TIBC levels decreased significantly and the best result was belonged tothe group of rats that fed on melissa powder 10%.From the obtained results it can be concluded that, supplementation of food bakery with Melissapowder exerts a positive impact on the liver and, kidney functions, lipid profile and other biochemical parameters.

### Introduction

Liver diseases, including hepatitis B virus and hepatitis C virus infections, alcoholic liver disease, nonalcoholic fatty liver disease and associated cirrhosis, liver failure and hepatocellular carcinoma, are major causes of illness and death worldwide. In China, liver diseases affect approximately 300 million (*Fu-Sheng et al., 2014*). Liver disease causes serious public health problems because of its high prevalence worldwide and poor long-term clinical outcome, including premature deaths from liver decompensation, cirrhosis and hepatocellular carcinoma (*Lok., 2004*).

Melissa officinalisL. is a plant cultivated in some parts of Iran. The leaves of lemon balm, Melissa officinalisL (Lamiaceae), are used in Iranian folk medicine for their digestive, carminative, antispasmodic, sedative, analgesic, tonic, and diuretic properties, as well as for functional gastrointestinal disorders **(Sepide et al., 2017).** 

Melissaofficinalis L. has a large use in traditional medicine, food industry, and aromatherapy, due to its fresh smell and its medicinal properties including hypoglycemic, hepatoprotective, antimicrobial, antidepressant, hypnotic, and sedative *(Miraj et al., 2016).* In addition, there are studies that pointed out the cytotoxic effect of lemon balm extract on breast cancer and colon carcinoma *(Weidner et al., 2015).* 

Chemical composition of lemon balm contains 15.3% moisture content and 0.94% lipid content total Mineral Substances, 7.37% protein content and 0.94% lipid content, Na 83.34 ppm, Mn 16.41 ppm, Fe 119, 4 ppm, Zn 29, 163 ppm, Ni 1.067 ppm, Cu 6.589 ppm (*Negrea et al., 2017*)

Melissaofficinalis *L*, also known as lemon balm, bee balm, honey balm, *(Rasmussen., 2011)* is a perennial herb. It is a member of the Lamiaceae (mint) family, and lemon balm (*Melissa officinalis L*) belongs to a genus that includes 5 species of perennial herbs native to Europe, Central Asia, and Iran. Although *Melissaofficinalis L* originated primarily in Southern Europe, it is now naturalized around the world, from North America to New Zealand *(Jastrzebska et al., 2013).* 

The leaf of (Melissa officinalisL,) contains flavonoids (quercitrin, rhamnocitrin, luteolin), polyphenolic compounds (rosmarinic acid, caffeic acid, and protocatechuic acid), monoterpenoid aldehyde, monoterpene glycosides, triterpenes (ursolic and oleanolic acids), sesquiterpenes, tannins, and essential oils (citral) (Sofowora et al., 2013).

It was revealed that essential oils of *Melissa officinalis L* have good potential for antioxidant activity and can be used in lipidcontaining foods. It is a rich source of antioxidants, in particular from the group of phenolic compounds (*Capecka et al., 2005*). Its activity is comparable with synthetic antioxidants (BHA and BHT), and antioxidant activity is related to phenolic compounds like citronellal and neral (*Meftahizade et al., 2010*).

Lemon balm infusion improves plasma levels of catalase, superoxide dismutase, and glutathione peroxidase and a marked reduction in plasma DNA damage, myeloperoxidase, and lipid peroxidation. Due to its iron chelating activity of the extract, its antioxidant potential was increased (*Dastmalchi et al., 2008*). It was found that flavonoids aglycones were responsible for the free radical scavenging activity and that induced lipid peroxidation in rat cultured hippocampal neurons was significantly inhibited by fractions containing flavonol glycosides, flavonol and biflavone aglycones, or chlorogenic acid type phenolics present in the ethanolic extract (*Ferreira et al., 2006*).

Melissa officinalis L had very high levels of phenolics in 32 plant spices. In another study, it had the highest levels of phenolics and flavonoids (*Dias et al., 2012*).

From the previous concept the present work was a trial to spot the light on the beneficial effects of *(Melissa officinalisL)* powder as hepatoprotective agents in rats.

### Materials and methods

### Material

Fresh plant of Melissa officinalis L wereobtainedfromNational Research Center (NRC) in Dokki, Cairo, Egypt. All chemicals used in this experiment were of analytical grade. Kits used for the quantitative determination of the different parameters were purchased from Bio diagnostic Co., Dokki, Giza, Egypt. Wheat flour (72 % extraction) sucrose, skimmed milk, baking powder, butter, eggs and salt obtained from local market. 30 male Albino rats of Sprague Dawley strain wear obtained from Research Institute of Ophthalmology, Medical Analysis Department, Giza, Egypt.

### Methods:

### Preparation of *MelissaofficinalisL*, Powder:

Melissa officinalisL, were washed with running tap water and dried by solar energy, (10.5 in hours) temperature (50°C) relative humidity % (21) *Ibrahim., (1999).* The whole quantity was distributed on solar cabinet dryer chamber of solar energy laboratory in the National Research Center was used for dehydration after that the dried plantwas ground into fine powder.

### Preparation of cupcake:

Melissa officinalis *L* powder was mixed with the wheat flour (72 % extraction) at 2.5, 5.0, and 10 %. Then, 320 g of the flour mixture was well mixed for 20 min with sucrose (175g), skimmed milk (125g), baking powder (10g), butter (125g), eggs (120g) and salt (2g). The mixture, with and without *Melissa officinalis L* illustrated in powder table (1), baked at 275°C for 30 min. Then, the cupcakes were allowed to cool and chemically evaluated was done according to the methods outlined in *A.A.C. C. (2000).* 

#### Chemical composition (*Melissaofficinalis L*,) and cupcake:

Melissa officinalis Land cupcakewerechemically analyzed todetermine their moisture, protein, fat, ash, fiber and carbohydrate contentaccording to (A.O.A.C. 2005).

#### **Determination of total flavonoids:**

Total flavonoids were estimated using the method of **Ordonez** et al.(2006).

### **Determination of total phenolics:**

Total phenol contents in the extracts were determined by the modified Folin-Ciocalteu method of *Wolfe et al., (2003).* 

Determination of antioxidant activity: were determined by the method of *Re et al.(1999).* 

#### Sensory evaluation of Cupcake:

Sensory evaluation of the prepared cupcakes was carried out according to *Khalifa et al., (2015).* Fresh samples of butter cakes containing, 0.0, 2.5, 5.0, and 10% Melissapowder were sensory evaluated by 20 panelists. All samples were evaluated for color, texture, taste, odor, appearance and overall acceptability using tenpoint hedonic scale, which a score of 10 represented attributes most liked; 5 represented attributes at an unacceptable margin; and 1 represented attributes most disliked.

### **Biological Assay:**

Thirty male albino rats Sprague Dawley strain weight 180+10g were housed in well-aerated cages under hygienic condition and fed on basal diet (*Reeves et al., 1993*), for one week for adaptation.

After the period of adaptation on basal diet, the rats were divided into 5equal groups (each group contain 6 rats), the 1<sup>st</sup>group fed on basal diet as a negative control, the other rats (25 rats) were intoxicated by subcutaneous injection of Carbon tetra chloride CCl4 in paraffin oil (1:1 v/v; 2 ml/kg) twice a week for two consecutive weeks to induce hepatic chronic damage (*Jayasekhar et al.,1997*). The 2<sup>nd</sup>group fed on basal diet as apositive control group, The 3<sup>rd</sup>, 4<sup>th</sup> and 5<sup>th</sup>group fed as the positive control group with

addition of 2.5, 5 and 10% melissa powder, respectively.

At the end of the experimental period (28 days), rats were fasted overnight, then anaesthetize &incised longitudinally and blood samples were collected from the aorta. The blood samples were centrifuged and serum was separated to estimate some biochemical parameters, i.e. Serum cholesterol (*Allain T et al., 1974*),(*TG*):

Triglycerides (Fossati, and Prencipl., 1982),(HDL-c): High density lipoprotein cholesterol, (Lopes-Virella et al., 1977),(LDL-c): Low density lipoprotein cholesterol, and Very low density lipoprotein cholesterol,(VLDL-c), (Friedewald et al., 1972),, Aspartate transaminase, (AST), and Alanine transaminase, (ALT),(Ritman and Frankl., 1957), Urea nitrogen, Creatinine, ,Uric acid.

#### Histopathological Examination:

Specimens from liver and spleen were placed in 10% neutral buffered formalin. The fixed tissues were then trimmed, washed with ice saline and dehydrated in ascending grades of isopropyl alcohol and cleared in xylene. The wax-impregnated tissues were embedded in paraffin blocks using the same grade wax, the paraffin blocks were cut with rotary microtome at 3-5µ thickness. The sections were floated on a tissue floatation bath at 40°C and taken on glass slides. The sections were then melted in an incubator at 60°C and after 5 min., they were allowed to cool and stained with Hematoxylin and Eosin according to *(Bancroft and Cook, 1998),* and examined microscopically at the Faculty of Veterinary Medicine, Cairo University, Egypt.

### Statistical analysis:

Data are presented as means  $\pm$  SD and the analysis was conducted using SPSS program, Version 16.0 (2007).

### Results and discussion

Total phenolic, total flavonoid and antioxidant activity of Melissa officinalis L:

Data in table (2) showed the total phenolic (mg/g), total flavonoid and antioxidant activity (AOA) of *Melissa officinalis*. Our results agree with who demonstrated that the leaf of *Melissa officinalis L* contains 13.2

mg GAE/100 g dw in 32 plant spices (*Koksal et al. 2011*). This study showed that antioxidant activity (%) was higher than that obtained by *Kamdem et al. (2013)* who found that antioxidant activity was 77.55  $\pm$  3.7 %.

#### **Biological assay:**

# Effect of different levels of Melissa powder on FI, BWG, and Relative Organs weight of hepatic rats:

As shown in Table 4, feed intake of control (+) decreased, then that of the negative control group. While feeding rats of diet containing 2.5, 5 and 10% MP induced increase in feed intake, as compared to the positive control group.

BWG % decreed significant in control (+), as compared to the negative control group.

The results in this Table showed non–significant changes in BWG% in the group fed on diet contain 2.5% and 5% MP, as compared to the positive control group.

While treatedrats with 10%MPinduced significant increase P <0.05, ascompared to the positive control group.Injected rats with CCL₄increased liver, kidneys, heart and spleenthan thenegative control rats.However, these increases in vital organs were propagated by MPsupplementation for liver, kidneys, heart and spleen. On the other hand rats given MP show variations in organs weight than control. The body weight is a sensitive indicator that reflects the state of health of experimental animals and the decrease in body weight correlates with defects in body metabolism (*Bhatiaand Khera, 2013*).

# Effect of different levels of Melissa powder on serum lipids fractions of hepatic rats:

Table (3) illustrated the effect of melissa powder on lipid fractions. The values of serum cholesterol, triglycerides, HDL-c,LDL-c and VLDL-c mg/dl) showed significant increase (P<0.05)for positive control group in compared with negative control group while HDL-c value (mg/dl) decreased for positive control group significantly, as compared to the control (-) group. Data in this table showed that, total cholesterol (mg/dl) were increased significantly (P<0.05) for rats

(control +). The statistical analysis showed a significant decrease in total cholesterol of all treated groups with Melissapowder when compared with (control +). The lowest decrease in all treated group in cholesterol was recorded in the group fed on diet containing MP10%. This treatment decreased this parameter by (36%). Also, the best result of serum triglycerides was observed in the group fed on basal diet containing MP10% (13.50%). In this respect **Capecka et al.,** (2005) who found that essential oils of *Melissa officinalis* L have good potential for antioxidant activity and can be used in lipid-containing foods. It is a rich source of antioxidants, in particular from the group of phenolic compounds.

HDL-c is an effective scavenger of cholesterol molecules from several locations, possibly even from some early plaque formation. Therefore, HDL-c has been considered to be a good lipoprotein and the cholesterol associated with HDL has been referred to be good cholesterol. HDL-c among all groups fed on diet containing MP2.5%, MP5%, and MP10%showed significant increase<0.05, compared with (control +).

Low-density lipoprotein cholesterol (LDL-c) of all treated rats with basal diet containing melissa powdered decreased significantly (P<0.05),compared with control (+). Meanwhile these treatments for rats led to increase LDL-c significantly, compared to (control -).

Very low-density lipoprotein cholesterol (VLDL-c) of rats fed on basal diet decreased significantly (P<0.05), compared to control (+) group. Treating rats with melissa powdered (2.5%, 5% and 10%) led to a significant reduction in serum VLDL-c compared with (control +). The lipid lowering effect of the extract might be due to the action of flavonoids and other phenolic compounds, triterpenoids, alkaloids, steroids and glycosides. Normalized rate of lipogenesis is due to the insulin-like activity of triterpenoids (*Sakurai et al., 2002*) or activating normoglycemia by the insulin tropic effect of flavonoids (*Pinent et al., 2008).* Our results agree with who reported that various studies have shown that Melissa officinalis L possesses high amount of antioxidant activity through its chemical compounds including high amount of flavonoids, rosmaric acid, gallic acid, phenolic contents. Many studies confirmed the ant oxidative effects of Melissa officinalis; thus, its effect in preventing and treating oxidative stress-related diseases

might be reliable (*Miraj et al., 2017*). In addition, Antioxidant activity of different fractions from *Melissa officinalis* extract was evaluated, antioxidants are widely used in dietary supplements and have been investigated for the prevention of diseases such as cancer or coronary heart disease (*Sepide et al., 2017*).

# Effect of different levels of melissa powderon serum uric acid, urea nitrogen, and creatininelevels (mg/dl) of hepatic rats:

Uric acid value, urea nitrogen and creatinine (mg/dl) of the control positive group showed highly significant increase, as compared to (control -) group the mean values were  $(2.99\pm0.01, 44.4\pm0.93)$  and  $0.1.65\pm0.06$ ) vs.  $1.46\pm0.05$ ,  $21.28\pm0.99$  and  $0.57\pm0.02$ ) respectively. Results in table (6) indicated that, the mean values of uric acid decreased in the groups fed on diet containing (MP2.5%, MP5% and MP10%), compared with (Control+).

Also, the highest decrease in serum urea nitrogen and creatinine in all treated groups was found in the group fed on diet containing MP10%. While the highest increase in serum urea nitrogen was observed in MP2.5% group after control (+). This result agree with Sepida et al., (2017) who reported that many studies confirmed the ant oxidative effects of Melissa officinalis; thus, its effect in preventing and treating oxidative stress-related diseases might be reliable. The results of numerous studies on antioxidant or radical scavenging effects may be a basis for detailed in vivo research on anti-inflammatory activities of this plant. First traditional usages of Melissa officinalis and second its antioxidant properties were reviewed in detail. Regarding its traditional usage, antimicrobial activity (antiparasitic, antibacterial, antiviral, etc), antispasmodic, insomnia properties were reported. Many studies confirmed the antioxidative effects of Melissa officinalis: thus, its effect in preventing and treating oxidative stress-related diseases might be reliable. The results of numerous studies on antioxidant or radical scavenging effects may be a basis for detailed in vivo research on antiinflammatory activities of this plant (Sepide et al., 2017).

# Effect of different levels of Melissa powder on liver functions (IU/L) of hepatic rats:

Results of AST, ALTALP are presented in Table (7), AST in all treated groups recorded significant decrease (P<0.05), when

compared with (control +). On the other hand, the lowest levels of AST enzymes were found in-group of rats fed on diet containing MP10% (199.31±0.65).

Also, results obtained from this Table showed a significant increase (P<0.05) in the mean values of ALT and ALP enzymes in the group fed on (control +), when compared with all treated groups and the best results were observed in the groups that fed on the MP10% and MP5% respectively. This result agree with **Sofowora et al., (2013)** who reported that a radical scavenging and antioxidant potential of polar extracts from Melissa. These activities have arisen from the content of flavonoids, rosmarinic acid, and the benzodioxole. The antioxidant effects of these compounds are up to 10 times stronger than the effects of those of vitamins B and C.

From the obtained results, it can be concluded that, supplementation with high percentage of MP10% exerts a positive impact on the liver enzymes and other biochemical parameters in hepatic rats. Administration of Melissa officinalis L. extract increased the content of GSH in liver and blood of hyperlipidemic rats (*Bolkent et al., 2005*). Our result agree with *Spide et al., (2017*) who reported that the anti-inflammatory activities of *Melissa officinalis* L leaves were investigated. The essential oil of *Melissa officinalis* L was shown to have anti-inflammatory activities, supporting the traditional application of this plant in treating various diseases associated with inflammation and pain.

#### Cupcake composition as affected by melissa powder addition:

The cupcake composition of unfortified and fortified cupcakes with 2.5, 5.0, and 10% melissa powder was presented in Table (7). The percentages of moisture, protein, fat, ash, fiber and carbohydrates (control) were 17.73, 7.60, 1.30, 0.71,1.41 and 71.35 %, respectively. Increasing melissa powder addition led to increase the percentage of fiber, ash and fat in the fortified cupcake and decrease the percentage of protein. However, the rest of constituents were slightly changed. Our result agree with *(Ivelina et al., 2018)* who reported that addition of Melissa increased dietary fiber content and antioxidant activity of bread. The sensory evaluation showed that the addition of 5% melissa influenced slightly negatively consumers.

# Sensory evaluation of cupcake as affected by melissa powder addition:

Table (9) illustrates sensory evaluation scores of fortified cupcake with melissa powder. The mean scores for color, taste, texture, odor, appearance and overall acceptability of unfortified cake (control) were 8.64, 8.7, 8.6, 8.82, 8.71 and 8.63, respectively.

The results of the statistical analysis in this Table showed no significant difference between unfortified cupcake (control sample) and cupcake fortified with 2.5% melissa. However, comparing with control, fortification of cupcake with 5%, and 10% melissa resulted in significant decrease (P<0.05) in all sensory evaluation parameters. This may be due to changes in the chemical composition, fiber content in particular. Competition for water between melissa fiber and other ingredients in the mix as flour and sugars resulting in a significant factor in the contribution of these ingredients to overall cupcake flavor in some formulations (*Jasberg, et al., 1989*).

#### Histopathological examination:

Liver of rat fed on basal diet (control-) showing the normal histological structure of hepatic lobule (Fig.3). Meanwhile, liver of injected rat with CCL4 and fed on basal diet (control+) showing portal infiltration with leucocytes (Fig.4 and 5) and Liver of injected rat with CCL<sub>4</sub> and fed on basal diet (control +) showing vacuolations of hepatocytes and focal hepatic necrosis associated with leukocyte cells infiltration.

Moreover, liver of rat suffering from chronic liver diseases and treated daily with Melissa powder 2.5% (MP 2.5%) showing vacuolations of centro lobular hepatocytes (Fig. 6). Liver of rat suffering from chronic liver diseases and treated daily with MP 5% showing slight congestion of central vein (Fig.7). Liver of rat suffering from chronic liver diseases and treated daily with MP 10% showing no histopathological changes (Fig. 8).

**Table (1):** Contents of cupcake with and without (Melissa officinalis L.) powder (g / 100 g).

| Contents(gm)<br>Types of cupcakes    |          | MP  | Flour | Sucrose | Skimmed<br>milk | Baking<br>powder | Butter | Eggs | Salt |
|--------------------------------------|----------|-----|-------|---------|-----------------|------------------|--------|------|------|
| control<br>Cupcake<br>fortified with | -        | 0.0 | 320   | 175     | 125             | 10               | 125    | 120  | 2    |
|                                      | MP 2.5 % | 8   | 312   | 175     | 125             | 10               | 125    | 120  | 2    |
|                                      | MP 5 %   | 16  | 304   | 175     | 125             | 10               | 125    | 120  | 2    |
|                                      | MP 10 %  | 24  | 296   | 175     | 125             | 10               | 125    | 120  | 2    |

**MP:** Mellissa officinalis L powder.

# Table (2): Total phenolic compounds total flavonoid and Antioxidant Activity of Melissa officinalis L.

| Component                    | Value      |
|------------------------------|------------|
| Total phenolic (mg/g)        | 13.57±0.24 |
| Total flavonoid (mg/g)       | 24.06±0.96 |
| Antioxidant Activity (AOA %) | 86.96±0.28 |
| Maar (CD                     |            |

Mean ±SD

 Table (3): Effect of some levels of melissa powder on FI, BWG% and relative organs weight of hepatic rats.

| Parameters  |             |                              | Organ weight/body weight (%) |                             |                             |                              |  |  |
|-------------|-------------|------------------------------|------------------------------|-----------------------------|-----------------------------|------------------------------|--|--|
| Groups      | FI (gm/day) | BWG%                         | Liver                        | Kidney                      | Heart                       | Spleen                       |  |  |
| Control (-) | 16.92       | 47.46<br>±1.06 <sup>A</sup>  | 3.08<br>±0.01 <sup>D</sup>   | 0.63<br>±0.011 <sup>B</sup> | 0.32<br>±0.01 <sup>CD</sup> | 0.22<br>±0.02 <sup>E</sup>   |  |  |
| Control (+) | 13.58       | -34.26<br>±1.94 <sup>C</sup> | 3.93<br>± 0.04 <sup>A</sup>  | 0.68<br>±0.00 <sup>A</sup>  | 0.38<br>±0.01 <sup>A</sup>  | 0.45<br>±0.0.01 <sup>A</sup> |  |  |
| MP 2.5%     | 14.12       | -33.86<br>±0.01 <sup>C</sup> | 3.51<br>± 0.00 <sup>B</sup>  | 0.65<br>±0.01 <sup>в</sup>  | 0.36<br>±0.02 <sup>AB</sup> | 0.41<br>±0.00 <sup>B</sup>   |  |  |
| MP 5%       | 14.31       | -33.10<br>±0.81 <sup>C</sup> | 3.41<br>± 0.03 <sup>C</sup>  | 0.64<br>±0.01 <sup>в</sup>  | 0.35<br>±0.01 <sup>BC</sup> | 0.33<br>±0.00 <sup>c</sup>   |  |  |
| MP 10%      | 14.75       | -22.38<br>±0.03 <sup>B</sup> | 3.13<br>± 0.02 <sup>D</sup>  | 0.62<br>±0.02 <sup>B</sup>  | 0.31<br>±0.015 <sup>D</sup> | 0.28<br>±0.01 <sup>D</sup>   |  |  |
| LSD         | -           | 1.925                        | 0.051                        | 0.027                       | 0.028                       | 0.018                        |  |  |

Control (-): Control Negative.

Control (+): Control Positive.

MP2.5%: Melissa Powder 2.5 % MP5 %: Melissa Powder 5% MP10 %: Melissa Powder 10 %

Values are expressed as mean  $\pm$  SD.

Significance at p<0.05.

Values, which don't share the same letter in each column, are significantly different.

# Table (4): Effect of different levels of melissa powder on serum lipids fractions of hepatic rats.

| Parameters<br>Groups | Uric acid mg/dl          | Urea nitrogen mg/dl      | Creatinine mg/dl        |  |
|----------------------|--------------------------|--------------------------|-------------------------|--|
| Control (-)          | 1.46 ± 0.05 <sup>C</sup> | 21.28± 0.99 <sup>D</sup> | 0.57± 0.02 <sup>D</sup> |  |
| Control (+)          | 2.99± 0.01 <sup>A</sup>  | 44.40± 0.93 <sup>A</sup> | $1.65 \pm 0.06^{A}$     |  |
| MP 2.5%              | 2.63± 0.00 <sup>B</sup>  | 38.58± 0.67 <sup>B</sup> | 1.35± 0.02 <sup>B</sup> |  |
| MP 5%                | 2.59± 0.14 <sup>B</sup>  | 36.94± 0.23 <sup>B</sup> | 1.32± 0.02 <sup>B</sup> |  |
| MP 10%               | 2.57± 0.01 <sup>B</sup>  | 32.12± 1.89 <sup>C</sup> | 1.16± 0.05 <sup>C</sup> |  |
| LSD                  | 0.128                    | 1.991                    | 0.077                   |  |

Control (-): Control Negative.

Control (+): Control Positive.

MP2.5%: Melissa Powder 2.5 % MP5 %: Melissa Powder 5% MP10 %: Melissa Powder 10 %

TC: Cholesterol TG: Triglycerides HDL: High-density lipoprotein cholesterol

LDL: Low-density lipoprotein cholesterol VLDL: Very low-density lipoprotein cholesterol,

Values are expressed as mean  $\pm$  SD. Significance at p<0.05.

Values, which don't share the same letter in each column, are significantly different

### Table (5): Effect of different levels of melissa powder on serum uric acid and creatinine urea nitrogen levels (mg/dl) of hepatic rats.

| Parameters  | Lipid Fraction (mg/dl)    |                           |                          |                           |                          |  |  |  |  |
|-------------|---------------------------|---------------------------|--------------------------|---------------------------|--------------------------|--|--|--|--|
| Groups      | тс                        | TG                        | HDL                      | LDL                       | VLDL                     |  |  |  |  |
| Control (-) | 89.04± 0.82 <sup>C</sup>  | 46.54 ± 0.53 <sup>D</sup> | 55.50± 1.36 <sup>A</sup> | 23.08± 0.79 <sup>D</sup>  | 9.52± 0.61 <sup>C</sup>  |  |  |  |  |
| Control (+) | 226.51±1.35 <sup>A</sup>  | 103.13± 1.90 <sup>A</sup> | 40.56±1.21 <sup>C</sup>  | 167.73± 2.21 <sup>A</sup> | 22.34± 0.49 <sup>A</sup> |  |  |  |  |
| MP 2.5%     | 211.21±1.10 <sup>AB</sup> | 95.04±1.29 <sup>B</sup>   | 45.79± 0.83 <sup>B</sup> | 146.33± 0.79 <sup>B</sup> | 18.40± 0.49 <sup>B</sup> |  |  |  |  |
| MP 5%       | 194.84±0.52 <sup>AB</sup> | 86.40 ±2.92 <sup>B</sup>  | 46.35±0.14 <sup>B</sup>  | 136.44± 2.50 <sup>C</sup> | 17.85± 0.03 <sup>B</sup> |  |  |  |  |
| MP 10%      | 140.05±0.61 <sup>BC</sup> | 86.11 ± 1.19 <sup>c</sup> | 46.75± 1.33 <sup>B</sup> | 131.71± 0.75 <sup>C</sup> | 16.64± 0.59 <sup>B</sup> |  |  |  |  |
| L.S.D       | 85.12                     | 3.21                      | 3.21                     | 1.97                      | 2.93                     |  |  |  |  |

Control (-): Control Negative. Control (+): Control Positive. MP2.5%: Melissa Powder 2.5 % MP5 %: Melissa Powder 5% MP10 %: Melissa Powder 10 %

Values are expressed as mean ± SD. Significance at p<0.05. Values, which do not share the same letter in each column, are significantly different.

### Table (6) :Effect of different levels of Melissa powder on liver functions (IU/L) of hepatic rats.

| Parameters  | AST                       | ΔΙΤ                         | ΔΙΡ                        |
|-------------|---------------------------|-----------------------------|----------------------------|
| Groups      | 701                       |                             |                            |
| Control (-) | 58.07± 0.11 <sup>E</sup>  | 27.38± 0.33 <sup>C</sup>    | 87.29± 1.00 <sup>D</sup>   |
| Control (+) | 238.75± 1.50 <sup>A</sup> | 147.67± 0.51 <sup>A</sup>   | 308.62 ± 2.19 <sup>A</sup> |
| MP 2.5%     | 216.66± 3.71 <sup>B</sup> | 135.73 ± 1.83 <sup>AB</sup> | 293.09 ± 3.51 <sup>B</sup> |
| MP 5%       | 205.2± 2.45 <sup>C</sup>  | 128.51 ± 0.64 <sup>AB</sup> | 284.87± 4.33 <sup>C</sup>  |
| MP 10%      | 199.31± 0.65 <sup>D</sup> | 97.41 ± 0.96 <sup>B</sup>   | 284.69 ± 5.62 <sup>C</sup> |
| L.S.D       | 3.85                      | 49.02                       | 6.73                       |

Control (-): Control Negative. Control (+): Control Positive.

MP2.5%: Melissa Powder 2.5 % MP5 %: Melissa Powder 5% MP10 %: Melissa Powder 10 %

Significance at p<0.05.

Values are expressed as mean ± SD. Values, which don't share the same letter in each column, are significantly different. ALP: Alkaline AST: Aspartate transaminase ALT: Alanine transaminase Phosphatase

| Table (7): | Proximate | cupcakes   | composition  | as | affected | by | melissa |
|------------|-----------|------------|--------------|----|----------|----|---------|
|            | powder ad | dition(wet | weight bases | ). |          |    |         |

| Parameters<br>Samples | Moisture | Protein | Fat  | Ash  | Fiber | Carbohy-drates |
|-----------------------|----------|---------|------|------|-------|----------------|
| Control*              | 17.73    | 7.60    | 1.40 | 0.71 | 1.31  | 71.25          |
| MP 2.5%               | 13.15    | 7.30    | 1.10 | 0.72 | 0.52  | 77.21          |
| MP 5%                 | 14.31    | 6.60    | 1.30 | 0.97 | 0.81  | 77.41          |
| MP 10%                | 16.16    | 6.31    | 1.45 | 0.99 | 1.02  | 74.07          |

# Table (8): Sensory evaluation of cupcakes as affected by melissa powder addition.

| Premaster  | Color  | Tacto  | Toxturo            | Odor   | Appoarance | Overall       |
|------------|--------|--------|--------------------|--------|------------|---------------|
| Samples    | COIOI  | Tasle  | Taste Texture Odor |        | Appearance | acceptability |
| Control*   | 8.64   | 8.7    | 8.6                | 8.82   | 8.71       | 8.63          |
|            | ±0.25a | ±0.15a | ±0.12a             | ±0.09a | ±5.86a     | ±0.29a        |
| MP2.5%     | 8.40   | 8.4    | 8.73               | 8.32   | 8.59       | 8.62          |
|            | ±9.69a | ±0.17a | ±0.35a             | ±0.36a | ±0.15a     | ±0.18a        |
|            | 5.8    | 5.8    | 5.87               | 5.46   | 5.53       | 7.93          |
| IVIP 5%    | ±1.34b | ±1.13b | ±1.54b             | ±1.48b | ±1.17b     | ±0.12ab       |
| MD 10%     | 5.53   | 5.6    | 5.60               | 5.30   | 5.53       | 7.03          |
| IVIF TO /0 | ±0.65b | ±0.46b | ±0.10b             | ±0.97b | ±0.33b     | ±0.76c        |

Control: wheat flour (72% extraction), The average of total score was converted to a descriptive category as follows: Judging scale: very good (9-8), Good (6-7), Fair (4-5), Poor (2-3) and Very Poor (1). All results are expressed as mean $\pm$  SD Means followed by different superscripts within columns are significantly different (P<0.05).

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التأثير المحسن لعشبة المليسيا على التسمم الكبدي في الجرذان

# هيام عبد العزيز الصاوى

مركز البحوث الزراعية -معهد بحوث تكنولوجياالأغذية – قسم الأغذية الخاصة

### الملخص العربى

نظرا للثورة الصناعية في العصر الحديث انتشرت الملوثات الجوية مسببة أضراراً جمة على صحة الإنسان. ومن تلك الملوثات مركب رابع كلوريد الكربون الضار بصحة وسلامة ووظائف الكبد. لذا كان الهدف من تلك الدراسة هو تتبع تأثير إضافة ٢,٥%، ٥٠%، ١٠% ، مسحوق عشبة المليسيا في وجبة ذكور الفئران المصابة بتسمم الكبد على الصحة و مستوى دهون الدم ووظائف الكبد و الكُلي. تم التوزيع العشوائي لعدد ٣٠فأرا ذكر بالغ من سلالة الالبينو على ٥ مجاميع. المجموعة الأولى (المجموعة الضابطة سالبة) حيث تناولت الغذاء الأساسي الخاص بالفئران، أما المجموعات (الثانية والثالثة والرابعة والخامسة) وعددهم ٢٥ فأر فقد حُقنت تحت الجلد لمدة أسبوعين برابع كلوريد الكربون مع زيت البرافين (١:١ ح/ح ٢ ملجم/ كجم) مرتان في الاسبوع، لمدة اسبوعين متتاليين لإحداث خلل كبدى واعتبرت المجموعة الثانية(كونترول موجب) حيث تناولت الغذاء الأساسي فقط، أما المجموعة الثالثة والرابعة والخامسة فقد تناولت الغذاء الأساسي مضاف إليه٥,٢%، ٥%، ١٠% مسحوق عشبة المليسيا على التوالي.ولقد أسفرت النتائج عن حدوث نقص معنوى في صورة الكوليسترول الكلي والجليسريدات الثلاثية وكذلك إنزيمات الكبد ونيتروجين اليوريا وحامض اليوريك والكرياتينين التي تغذت على الغذاء الأساسي والمضاف إلية نسب عالية من مسحوق عشبة المليسيا. وكانت أفضل النتائج للمجموعة التي تغذت على مسحوق عشبة المليسيا ١٠%. وتوصى الدراسة بتدعيم الوجبات والمنتجات الغذائية بمسحوق عشبة المليسيا وذلك لما أظهره من نتائج ايجابية في تحسين وظائف الكبد والكلي وكذلك صورة الدهون في الدم وبعض القياسات البيو كيميائية الأخري.