

**ASSESSMENT OF THE HAZARDOUS EFFECT OF FUNGICIDE
DITHANE ON *CLARIAS lazera* (CATFISH) INCLUDING
HAEMATOLOGICAL, BIOCHEMICAL, AND
IMMUNOLOGICAL PARAMETERS**

[69]

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ABSTRACT

The aim of present work was to study the toxicity of (mancozeb) Dithane fungicide on fish *Claras lazera* (catfish) and consequently to human beings. The fishes were exposed to Dithane in dose of 0.5 ppm /L (equivalent of 1/10 of LD50) for 30 days. Different Haematological, Biochemical, Bacteriological, and Immunological parameters were assessed. The results showed significant increase in Blood level of Sodium (Na), Potassium (K), Cortisol, Urea, Creatinine, Glucose, Insulin as well as Aspartate Amino Transferase (AST) and Alanine Amino Transferase (ALT) in blood. However there was a decrease in blood level of Iron and IgM, accompanied by decrease in Haemoglobin (HB), Macrocytic hypochromic anemia (R.B.Cs) count, Packed cell volume (PCV) which was observed in fish in 7, 15, 30, days after exposure to Dithane. The Haemogram shows reticulocytosis and increase in mean corpuscular volume (MCV). Dithane produces metabolic stress and cell damage with malfunction of haemopoietic system. Microbiological examination revealed a presence of pathogenic bacteria mainly *E. coli*, *Flavobacterium*, and *Staphylococcus aureus*. It was concluded that in catfish reared on low dietary carbohydrate (CHO) diet there was hyperglycemia due to increase in cortisol hormone. However immunological results revealed decrease in the level of IgM in blood; a loss of scales and petichial haemorrhage in parts of skin was observed. Ascitic and erosion due to complication of bacterial infection, was also accorded.

Key words: Dithane (mancozeb), Haematological change, Biochemical change
Microbial changes, Immunoglobulin M (IgM)

INTRODUCTION

Environmental toxicology is the study of how ecological systems, their structure, dynamics and function are

affected by pollutants. A developing subfield of environmental toxicology is ecotoxicology, in which special concern is placed on the release of toxic pollutants

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(Received March 31, 2005)

(Accepted May 16, 2005)

into the environment, especially into aquatic systems, by focusing on how these toxicants may become distributed within food chains and by measuring the toxic responses made within a particular ecosystem to such pollutants. These are by nature interdisciplinary fields drawing their knowledge from ecology, organic chemistry, molecular biology, genetics, soil science, mathematics and so on. The main source of pollution of water bodies with pesticides is in the melt waters, rainwaters and underground waters. Pesticides, may reach water bodies through the air at the time of their application to objects located near by insecticides.

Insecticides are applied to water to prevent the development of aquatic phase or blood sucking insects. Pesticides reach water sources subsequently through the trophic chains and cycles of various substances. As a result of circulation of pesticides in a water body, they sometimes accumulate in fish, silt bottom, zooplankton, algae and aquatic plants. Fish had long been regarded as a highly desirable food due to its content of high quality animal protein, its calcium and phosphorus content as well as its generous supply of vitamins.

Organochlorinated pesticides accumulate in fish mainly in the visceral fat, whereas the gills and muscles retain a lower amount, subsequently with an increase in fat consumption, for example, at the time of migration and hibernation. Pesticides may enter the more sensitive

organs and induce poisoning **Dummer *et al* (1991), Barton and Iwama (1991), Clealand *et al* (1998), Bennett and Wolke (1987), and Mona *et al* (2003).**

Compared to the controls some of the behavioral toxicity of fish showed decreased sheltering and increased horizontal displacements, burst swimming, buccal movements, and antagonistic interactions. The aim of this study is to determine the effect of low CHO diet on *Claras lazara* fish, exposed to Dithane (0.5ppm) as fungicide, on some hematological and clinicopathological, microbiological and immunological parameters.

MATERIAL AND METHODS

Materials

Dithane was obtained from Central Agriculture Pesticide Laboratory, Dokki, Cairo. Dithane M.45 (mancozeb) 80% wp {[1,2-ethanediy]bis(carbamodithioato)} (2-) manganese mixture with {[1,2-ethanediy]bis (carbamodithioato)}(2-)} zinc.



Experimental Conditions

Claras lazara fish (150-200 gram /each) were obtained from the river Nile Rashid branch, at El-Kanater El-Khairiya. Fish were acclimatized to laboratory condition one week before

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(Received March 31, 2005)

(Accepted May 16, 2005)

treatment 115L glass aquaria with a flow system and dechlorinated tap water. Static conditions with water renewal every 3 days were used. Fishes were fed once a day at 10.00 a.m. Three groups 15 fishes each were used. The first group was maintained under the same condition but free from any toxicants, and kept on a balanced diet (Table, 1) that meets its requirements from nutrients as described by **Dixon and Hilton (1981)** and was used as control group. The second group was kept on low carbohydrate diet but free from any toxicants, and maintained on a balanced diet that meets its requirements from nutrients as described by **Dixon and Hilton (1981)** **Hilton and Dixon (1987)** and **Roberts (1989)**. The third group was fed on a mixture of low CHO diet, the constituents of the diet used in the experiment are presented in (Table, 1) and it was exposed to Dithane (0.5 ppm) within 30 days. The diet ingredients included, dissolved oxygen, temperature, pH, ammonia and nitrite, in water tanks. The quality of water was measured daily, while water alkalinity, hardness, carbon dioxide, sodium, potassium and chlorides were analyzed before and after each water renewal using commercial kits of Biochringer, France, as shown in (Table, 2).

Blood Sampling

Blood samples were taken after 7, 15, 30 days. The fish were anaesthetized by 1/1000 aqueous solution of Ms 222 and bled from the caudal vein. Blood samples were taken with heparinized micrhaematocrite tube. The tubes were centrifuged at 3000 r.p.m. for vit. A, 8000U; vit. E, 21U; vit. K, 4mg, vit. B2 3.6mg; niacin 20mg; choline chloride 160mg; pantothenic acid 7mg; vit. B12,

5ug; Mn, 70mg; Zn, 60mg; Fe 20mg; Cu, 2mg; I, 1mg; CO, 0.2mg. 10 min. Serum was separated and stored at 20°C until used. Tested kits supplied from biomerieux (France) were used for determination of the activity of serum Alanine Amino Transferease (ALT) and Aspartate Amino Transferase (AST) as described by **Reitman and Frankel, (1957)**. Serum glucose was assessed according to **Trinder, (1969)**. Enzymatic determination of urea was done according to Patton and Insulin was estimated by radioimmunoassay method using coat A Insulin (**Patton and Crouch 1988**). Kits were obtained from Diagnostic Corporation Co. (DPC) 57700 west 96th street, LosAngeles, U.S.A. (**Pickering and Duston, 1983**).

Haematocrite value was carried out by using microhaematocrite capillary tubes centrifuged at 1200 r.p.m. for 5 min. Mean corpuscular volume (MCV). Reticulocytic count was done according to **Drabkin (1946)**. Serum cortisol level was determined using radio-immunoassay technique according to the method of **Pickering and Pottinger (1983)**. Serum iron was determined using atomic absorption according to **Joseph and Roger (1979)** and **Anderson, (1990)**. Values of sodium and potassium in serum were determined by flamephotometer according to the method described by **Silversmith (1965)**. Serum creatinine was measured according to **Bartels et al (1972)**. Enzymatic determination of urea was done according to Patton and Insulin was estimated by radioimmunoassay method using coat A. Insulin (**Patton and Crouch, 1988**). Kits obtained from Diagnostic Corporation 57700 west 96th street, LosAngeles, U.S.A. (**Pickering and Duston 1983**).

Bacterial isolation

Aseptic swabs from the skin, gills, base of fins and blood of tested fish were cultivated on blood agar, MacConky agar, Nutrient agar, TSA, Nutrient broth and peptone water and Sabaroud's dextrose

agar. Inoculated media were incubated at 37°C for 48 hour. Bacterial isolates were identified by examination of the colony morphology and biochemical characteristic described by **Bastawrows and Amal (1999)**, **Koneman *et al* (1994)** and **Palumbo *et al* (1985)** and **Nagae *et al* (1993)**. Bacteria were detected by

Table 1. Ingredients and proximate chemical composition of diets used in the present experiments

Diets 2	Diets 1 (control)	Ingredients
30	30	Fish meal
10	8	Meat meal
3	1	Bone meal
4	3	Skimmed milk
7	5	Soybean
20	20	Wheat Bran
5	20	Wheat flour
15	10	Yeast
4	1	Cod liver oil
2	2	Mineral & vitamin premix

Proximate chemical composition

38.89	35.87	Crude protein (C.P) g %
2415.4	2297.21	Metabolizable energy / Kg
2.86	2.78	Ether extract (E.E.) g %
4.27	3.91	Crude fiber (C.F.) g %
10.25	8.735	Ash g %
3.99	3.094	Calcium (Ca) mg %
2.53	2.069	Phosphorous (Ph) mg %
2.29	2.105	Lysine mg %
0.613	0.562	Methionine mg %

*Mineral and vitamin premix per/kg pelleted food.

Table 2. Water quality characteristics in tanks , Initial conditions values are mean \pm SE

6.40 \pm 0.1	PH
20°C \pm 0.8	Temperature ° C
0.022 \pm 0.04	Nitrates mg/l
0.0015 \pm 0.004	Un ionised amonia (mg / l)
4.03 \pm 0.5	Carbonic dioxine (mg / l)
32.8 \pm 2.8	Alkalinity (mg / l)
4.54 \pm 0.53	Permanganate oxidabole matter (mg / l)
34.6 \pm 0.1	Hardness (mg /l)
8.4 \pm 0.6	Chlorides (mg)
0.12 \pm 0.007	Potassium (mg /l)
5.68 \pm 0.01	Sodium (mg / l)

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(Received March 31, 2005)

(Accepted May16, 2005)

counting colonies using surface spread plate technique according to quantitative method described by **Palumbo *et al* (1985)**.

Measurement of serum immunoglobulin M (IgM) -IgM determination

The serum IgM was measured according to **Fuda *et al* (1991)**. Double antibody sandwich Elisa according to the method of **Matsubara *et al* (1985)** for determination of IgM was done.

Statistical analysis

The obtained data were statistically subjected according to **Duncan (1955)**, **Steel and Torrie (1980)**.

RESULTS

Haematological results

Regarding heamatotoxic effect of dithane, the blood analysis of fish revealed significant decrease in R.B.Cs count, HB% and P.C.V. whereas there was significant increase in mean corpuscular volume M.C.V. than control ($P \leq 0.01$) 36.67 versus 31.00, respectively. However there was insignificant change in reticulocyte count compared to control values (Table, 3).

Concerning periods of exposure, the values of R.B.C.s, HB% and P.C.V. after

exposure to dithane for 7 days were significantly higher ($P \leq 0.01$) than the values after exposure for 15 and 30 days respectively. However, exposure for 30 days showed the lowest values (Table, 3). While the M.C.V. and reticulocyte % values after 30 days exposure were significantly higher ($P \leq 0.01$) values than exposure for other periods.

The changes in the treated groups for R.B.Cs, HB%, P.C.V. values start to decrease after exposure from 15 up to 30 days respectively ($P \leq 0.01$) (Table 3). On the other hand, the change in M.C.V. and reticulocyte % values of the treatment after 30 days of exposure showed highly significant increase than other periods. Concerning periods of exposure treatments interaction in (Table 3), there were non significant difference between treatment and control under 7 days, while there was significant difference between control and treatment under the other two periods for R.B.Cs and M.C.V. parameters, whereas HB, P.C.V. and reticulocyte % were significantly different between control and treatment under all period experiment.

Clinico pathological results

Regarding treatment effect (Table, 4), there was high significant increase in treatment values of AST, ALT., urea, creatinine, Na, K, Cortisol, Glucose and Insulin ($P \leq 0.01$) compared to control values. On the other hand, iron level

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(Received March 31, 2005)

(Accepted May16, 2005)

values for treated group showed significant lower values ($P \leq 0.01$) than the control as shown in Table (4). Concerning period of exposure, the highest significant values of AST, ALT, Urea, Creatinine, Na⁺, K⁺, Cortisol, Glucose and Insulin were attained after 15 and 30 days exposure, ($P \leq 0.01$) while iron values showed high significant decrease after 30 days exposure compared with 7 days and 15 days.

Concerning periods of exposure by treatment interaction (Table, 4), the treated groups were significantly higher than the control regardless the period of exposure except iron which decreased. On the other hand, the treatment for 30 days of ALT, Urea, Creatinine, Na, K, Cortisol, Glucose and Insulin showed the highest significant increase compared to other groups.

Bacteriological results

At the end of experimental period there was difference on the bacterial counts on the different organs of the fish e.g. mainly external surface, kidneys, livers, and gills in all kinds of detected bacteria (Table, 5).

Immunological results: were shown in (Table, 6). The control values of IgM were significantly higher than mean treated groups ($P \leq 0.01$). However comparison between the values after different periods and the same trend for interaction between treatments and periods of exposure showed insignificant change. The period by treatment interaction for IgM was statistically insignificant ($P \geq 0.05$).

DISCUSSION

Several investigators reported that the application of Dithane a dithiocarbamate compound has been used extensively as fungicide for control of wide range of fungi (**Worthing, 1983 and 1987**). Therefore human and animals may be subjected to such compound via food, water and air and this could affect their health as demonstrated by **Yamanaka et al (1993)**. In the present study, the

toxicity of Dithane was evaluated in *Claras lazera* (catfish) because the fishes were exposed to water pollution by insecticide and fungicide, also fish had long been regarded as a highly desirable food due to its content of high quality animal protein, its calcium and phosphorus content as well as its generous supply of vitamins.

As resulted from hematological examination, iron level decreases also R.B.Cs count, Hb % and P.C.V. with the resulting anemia may be attributed to one consistent effect of Cortisol which causes reduction in the R.B.Cs, Hb%, P.C.V.% and Iron levels as a result of decrease in appetite in the rainbow trout or more likely to be the direct result of a catabolic effect or cortisol of the fish tissues. Also concerning the haemogram it was showed macrocytic hypochromic anemia characterized by significant decrease in R.B.Cs., Hb, P.C.V. and increase in M.C.V. Such animal picture might be attributed to the observed reticulocytosis in fish specially in 15 and 30 days. Such findings are in agreement with those of (**Roberts 1989, Lall (1991) and Mona et al (2003)**). The clinicopathological showed an increase in hepatic enzymes, AST, ALT. The liver is the primary organ of detoxification as well as a major site for detoxification reaction which was due to exposure of dithane in water. Such result agree with **Dummer et al (1991), Yalow and Bawman (1983)** who observed that aquatic pollution with heavy metals cause immunosuppression and contribute to outbreaks of infections, and bacterial diseases in fish. It could be concluded that dithane can affect fish after 7 days but there is some complications with this pesticide. The clinicopathological results of increased

serum level of sodium and potassium kidney impairment, where the concentrations, may be attributed to

Table 5. Bacterial isolates recovered from fish treated with Dithane (0.5 ppm)

Gill	Liver	Kidney	External surface	Bacterial Strain
3 x 2 ³	10 x 3 ³	2 x 7 ³	4 x 10 ³	<i>E-coli</i>
3 x 4 ⁶	4 x 10 ³	7 x 10 ²	3 x 10 ²	<i>Staphylococcus pyogenes</i>
2 x 5 ³	1 x 10 ⁶	2 x 10 ³	2 x 10 ³	<i>Flavobacterium sp.</i>

Table 6. IGM mean values of *Claras lazara* exposed to Dithane (0.5 ppm) and periods

* IgM Parameters	Periods						Mean Overall of all periods	
	7 days		15 days		30 days		Diet free	Diet With dithane
	Diet free	Diet with dithane	Diet free	Diet with dithane	Diet free	Diet With dithane		
IgM mean value	0.80± 0.15 ^a	0.78± 0.50 ^a	0.80± 0.15 ^a	0.72± 0.55 ^a	0.80± 0.15 ^a	0.65± 0.45 ^a	0.80± 0.15 ^a	0.73± 0.64 ^b
Mean of all treatments	0.79±0.12 ^a		0.76±0.12 ^a		0.74±0.13 ^a			

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(Received March 31, 2005)

(Accepted May16, 2005)

Means having different superscripts within the same row are significantly different at $P \leq 0.05$

* IgM : Immunoglobulin M

kidney is the normal pathway for Na and K. This may explain the main cause for elevation of the serum creatinine and urea in the treated groups. This confirms the previous reports recorded by **Neish and Hughes (1980)**. Also one of the reasons for hyperinsulinaemia was that low CHO diet with dithane causes significant increase of cortisol level which may be due to activation of hypothalamus pituitary internal axis, inducing a significant increase in cortisol level. These results coincide with those observed by **Vargut and Studnieka, (1994)** who observed that serum cortisol increased linearly in salminid fish fed on 5% CHO diet. Microbiologically the results were confirmed by the previous reports of **Neish and Hughes (1980)**, who stated that hyphae of organisms may invade the deep tissue of the fish and penetrate the vital organs such as kidneys, liver, and even the central nervous system.

Such results also agree with that of **Osfor et al (1991 and 1998)**, who demonstrated that the kidneys were the main organs of localization of the lesions, even they reported that yeast cells of the fungus occurred in rat's kidney fed on food diets contaminated with moulds for 30 days. Immunological decrease in level of IgM may be explained by the fact that production of lymphocytes in fish is apparently in the head of kidney, gut-associated tissues and spleen. However only the IgM antibody class has been found not to destroy antigen-bearing

invaders. They instead inactivate antigens and mark them for destruction by macrophages and complement **Jurd, (1985) Willoughy and Pickering (1977)**.

From the results obtained in this study, it was evident that there was a decrease in antibody titer in vaccinated group of fish exposed to the tested pesticides.

REFERENCES

- Anderson. D.P. (1990)**. Immunological indicators: effects of environmental stress on immune protection and disease outbreaks. *Am. Fish Soc. Symp. 8: 38-43*.
- Bartels, H.; M. Bohmer and C. Heierli (1972)**. Determination of serum creatinine *Clinical Chemistry Acta. 37: 193-195*.
- Barton, B.A. and G.K. Iwama (1991)**. Physiological changes in fish from stress in aquaculture with emphasis on the reponse and effects of corticosteroids, *Annual Review of Fish Disease., 1: 43-49*.
- Bastawrows, A.F. and A.M. Amal (1999)**. Some Microbiological Investigations on *Aeromonas hydrophila* grown in *Drechromis nialticus* and *Clarias lazera* *Assiut Vet. Medical. Science, Confernce in Assiut Governorate, Fac. Veterinury, Assiut Univ., Egypt, pp. 465-467*.
- Bennett, R.O. and R.F. Wolke (1987)**. The effect of sublethal endrin exposure on rainbow trout. *Salmon gairdneri*

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(Received March 31, 2005)

(Accepted May16, 2005)

- Richardson. 1. Evaluation of serum cortisol concentrations and immune responsiveness. *J. Fish Boil.* 31: 375-379.
- Clealand, G.B.; P.J. McEtroy and R.A. sonstegard (1988).** The effect of dietary exposure to immune to Aroclor 1254 and/or mirex on humoral immune expression of rainbow. Trout *Salmon gairdneri*. *Aquatic Toxicol.* 2: 241-248.
- Dixon, D.G. and J.W. Hilton (1981).** Nutrition of ranbpwtrout *J. Fish Biol.*, 19: 509-603.
- Drabkin D.J. (1946).** Blood analysis. *Biol. Chem.* 164: 703-705.
- Dummer, M.; A.K. Siwicki and A. Damael (1991).** Effects of organophosphorus insecticides: effects of trichlorfon and dichlorvos on the immune response of carp (*Cyprinus carpio*) III. In vitro effects on lymphocyte proliferation and phagocytosis and in vivo effects on humoral response. *Ecotoxicol. Environ Safty.* 22: 29-38.
- Duncan, D.B. (1955).** Multiple range and multiple F tests. *Biometrics* 11: 1-42.
- Fuda, H.; K. Sayano; F. Yamaji and Haraj (1991).** Serum immunoglobulin M (IgM) during early development of *Masu salmon* on *Corhyrchus masu*. Comparative *Biochem. and Physiol.* 99 A: 637-640.
- Hilton, J.W. and D.G. Dixon (1987).** Nutrition of cat fish. *Journal of Diseases.* 5: 185-189.
- Joseph, A. and W.G. Roger (1979).** For determination of serum iron. *Clinical Chemistry Principles and Procedures.* 4th Ed., pp. 168-197. Boston.
- Jurd, R.D. (1985).** *Specialisation in the Teleost and Anuran Immune Response in Fish Immunologh.* pp. 643-658. Manning, M. J. and F.M. Tatner, (eds.). Academic Press, New York.
- Koneman, E.W.; S.D. Allen; W.M. Janda; P.C. Shreckenberger and W.C. Winn Jr. (1994).** *Introduction to Diagnostic Microbiology.* pp. 117-123. Lippincott Company. Hannover Germany.
- Lall, S.P. (1991).** Salmonid nutrition and feed production. *International; Proceeding of the Special Session on Salmon Aquaculture Society,* pp. 243-248. Los Angeles.,
- Matsubara, A.; S. Mihara and R. Kusuda (1985).** Quantitation of yellow tail immunoglobulin by enzyme-linked immunosorbent assay (Elisa). *Bull. Japan Sac. Sci. Fish,* 51: 921-927.
- Mona, S.Z.; M.H. Osfor; F.S. Bayomi and E.N. Abouel, Gheit (2003).** Impact of low dietary carbohydrate diets on some nutritional and clinicopathological parameters of *Tilapia nilotica* infected with *Saprolegnia parasitica* and exposed to copper sulphate. *Applied Bull. NRC, Egypt,* 28. No(2) : 245-257.
- Nagae, M.; H. Fuda; A. Hara; A. Hamuchi (1993).** Changes in serum immunoglobulin M (IgM) concentrations during early development of churm salmon as determined by sensitive ELISA. *Comp. Biochem. Physiology,* 69: 814-832.
- Neish, G.A. and G.C. Hughes (1980).** *Fungal Disease in Fish.* pp. 602-613. T.F.H. Publications. Inc. Ltd. New Jersey.
- Osfor, M.H.; E.Y. Ismail and M.S. Arbid (1991).** *Effect of Certain Dietary Nutrients on Rats Fed Diets Contained with Molds.* pp. 30-38. Ph.D. Thesis. Fac. Vet. Med. Alex. Univ., Egypt.
- Osfor, M.H.; S. Zaki Mona and A. Saleh Zeinab (1998).** Impact of low dietary CHO diets on some nutritional of

- clinicopathological of catfish infected with some pathogens. *Bull N.R.C. Egypt*, **23**: 183-189.
- Palumbo, S.A.; F. Marxino; A.C. Williams; R.L. Buchana and D.W. Thayer (1985)**. Starch-ampicillin agar for the quantitative detection of *Aeromonas hydrophila*. *Japanese Journal of Vet. Science*. **16**: 159-175.
- Patton C.J. and M.I. Crauch (1988)**. Determination of serum insulin. *Endocrinology* **80**: 384-390.
- Pickering, A.D. and J. Duston (1983)**. Analysis of cortisol hormone. *J. Fish Biol.* **23**: 163-172.
- Pickering, A.D. and P. Pottinger (1983)**. Analysis of hormone. *Gen. Com. Endocrinal.*, **49**: 232-239.
- Reitman, S. and S. Frankel (1957)**. Analysis of liver function. *J. Clin. Pathol.* **28**: 56-64.
- Roberts, R.J. (1989)**. Nutritional pathology of teleosts. In: *Fish Pathology* pp. 337-362. (ed. By Roberts R.J.), Bailliere Tindall, London.
- Silversmith, A.B. (1965)**. For Determination of Serum Sodium and Potassium. *Med.* **45**: 175-177.
- Steel, R.G.D. and J.H. Torrie (1980)**. *Principles and Procedures of Statistics; A biometrical Approach 2nd Ed.* pp. 273-275. McGraw-Hill Book Co., Inc., New York, U.S.A.
- Trinder, P. (1969)**. For Determination of Serum Glucose. *Ann. Clin. Biochem* **6**: 24-26.
- Vergut, C. and M. Studnieka (1994)**. Effects of lindane exposure on rainbow trout immunity III. Effect on non-specific immunity and B lymphocyte functions. *Ecotoxicol. Environ. Safty* **27**: 324-328.
- Willoughby, L.G. and A.D. Pickering (1977)**. Viable saprolegniaceae spores on the equidermis of salmonid fish *Salmon trutta* and *Salvelinus alpius*. *Transactions of the British Mycology Society* **68**: 91-97.
- Worthing, CR. (1983)**. *The Pesticide Manual; A World Compendium*, seventh edition. Published by the British Crop Protection Council. London.
- Worthing, CR. (1987)**. *The Pesticide-Manual: A World Compendium*, Eighth Edition, Published by the British Crop Protection Council, London.
- Yalow R. and W.A. Bawman (1983)**. Plasma insulin in health and disease. In: *Diabetes Mellitus; Theory and Practice* pp. 119-150. (eds.). Ellenberg, M. and H. Riking, Excerpta Medica. New York.
- Yamanaka, S.; M. Yashide; Y. Yamamura; M. Nishimura; Y. Ana Takaesu (1993)**. A study on acute organophosphorus poisoning changes in the activity and isozyme pattern of serum cholinesterase in human poisoning. *Nippon Eiseigaku Zasshi*, **48(5)**: 955-965.

13(3)، 1005-1018 مقره اقل ،س مشن ي ع ة عم ا ج ع ا ر ز ل ن و ح ب ل ل ن ا س ا ر د ق ل ي ب ر ع ل ك ت ا ع م ا ج ل د ا ح ت ا ق ل ج م ، 2005

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م ي ك ز د ع س ي ن م - ل م ش و ع ي ا ف م ا ر ك ا

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2- ر ص م - ع ر ه ا ق ل ا ع ي ق د ل ل ن و ح ب ل ل ن ا س ا ر د ق ل ي ب ر ع ل ك ر م ل ا

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ا ر م ح ل ل ي ا ل خ ل ا م ج ط س و ت م و Reticulocytes
ا س ا ر د ل ا ت ر ه ظ ا م ن ي ب ، (M.C.V)

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ا ي ر ك ل ن ل ي ل و ب ل ا و و ل ك و ل ج ن ا ي ل و س ن ا ل ا و
ا س ا ر د ل ا ن ا ن ي ح ي ف ، م د ل ا ي ف
ض ا ر ي ل ت ك ب ل ا د و ن خ م ي ح و ا ل و ي ر ت ك ب ل ا
(*E. coli*, *Flavobacterium* and
Staphylococcus aureus) ح ت ك ا م س ا ل ا ي ف
ف ICM و ت س ج ي ف ص ق ن ت د خ و ي ر ج ت ل ا
ة ي ع ا ن م ل ت ا س ا ر ن ت ل ه ت ب ث ا م ك م د ل ا
و ق س ك ا م س ا ل ل ت د ح د ق ف ا م و م ع و
ن ع ب ي ف ف ي ز ن ع م ة ي و م ت ا ع م ج ن ت و ش ق ل ل
ل ب ل ي ف ل ق س ت س ا ك ل ذ ك ن ل ج ل ا ن م ا ز ج ا
ت ا ح ر ق ت و

د ي ج م ل ل ب ع ي ه ا ر ب ا د م ح م د : ل م ي ك ح ت
ل ي د ن ق ي د ا ه ل ل ب ع د م ح م د ا

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(Received March 31, 2005)

(Accepted May 16, 2005)