Sodium Chloride Stress Induced Morphological Changes in Some Halotolerant Fungi

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ABSTRACT

Materials and methods: Nine fungal isolates namely *Emericill anidulans, Mucor racemosus*, *Alternaria pluriseptata Penicillium canescens, Syncephalastrum racemosum, Aspergillus fumigatus, Alternaria chlamydospora, Aspergillus parasiticus* and *Ulocladium atrum* were isolated from AL – SHEGA area at AL- QASSIM region. **Results:** The influence of different sodium chloride concentrations on the growth rate, morphological and ultrastructure—were studied. Considerable differences in their growth rate and morphology were detected on medium containing different concentrations of sodium chloride (NaCl). Low growth rates were obseved on high NaCl concentrations. At 15 % NaCl, low growth of *Emericill anidulans, Penicillium canescens, Syncephalastrum racemosum, Aspergillus parasiticus* and *Mucor racemosus* was detected, whereas all fungal isolates were failed to grow at 20% NaCl. Scanning Electron Microscope (SEM) revealed that all fungal asexual reproduction organs were metamorphosed at higher NaCl concentration, fungal heads and sporangia were speculated or elongated. Sporangiophores and conidiophores were shortened and dwarfed ,little number of conidia or spores were detected.

Key words: halotolerant fungi, salt stress, SEM. sodium chloride, morphology

INTRODUTION

Follow fungi in desert region the strategy of fungus strain and representing growth of stressful environments, lack of food and low humidity, drought and high temperature or osmophilic [1]. Organisms living in environments with high concentrations of salts are challenged by osmotic stress and by the toxicity of specific ions [2].

Most fungi differ from the majority of halophilic prokaryotes. [3] they tend to be extremely halotolerant rather than halophilic and do not require salt to the survival, [4] and especially the growth of microorganisms in highly saline environments requires numerous adaptations [5]. The fungal cell wall is a supramolecular structure that offers strength to the cells, determines their shape, protects them against mechanical damage and mediates the cell-to-cell communication and their interaction with the environment. Physical and biological properties of fungal cell walls are determined by the composition and the arrangement of their structural components. [6]

The fungal cell wall is the first line of defense against environmental stress; therefore, adaptation at the cell wall level is expected to have one of the most important roles for successful growth at a highly saline [7]

The cell wall is essential for maintaining the osmotic homeostasis of cells, since it protects them against mechanical damage as well as high

concentrations of salts. [8] Its balance between a rigid and a dynamic structure influences the shape of cells . [9] and enables growth and hyphal branching . [10]

Some studies have focused on the role of the fungal cell wall in chemical sensing and processing of environmental signals that control growth and cell morphology of microorganisms and synthesis and secretion of extracellular enzymes. [11], [12], & [13]

Little is known about their mechanisms of adaptation to high salinity. To investigate the effects of low and high NaCl concentrations on cell morphology, with particular emphasis on cell wall ultrastructure. [14]

The dominant representatives and the most thoroughly investigated halophilic fungi in hypersaline waters of the salterns are the black yeasts, [15] and particularly the model organism *Hortaeawerneckii*. [16]

An important level of adaptation of the black yeasts to high salinity is seen in their extremophilicecotype, which is characterized by a special meristematic morphology and changes in cell wall structure and pigmentation. [17]

This work aims at investigating the effect of salt stress on growth of some halotolerant fungal isolates, also representing the effect of salt stress on their morphology by (SEM).

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MATERIALS AND METHODS

The soil samples used for isolation of halophilic fungi were collected from different localities representing different saline habitats at AL – SHEGA area at AL-QASSIM region

Organisms and Cultivation

Fungal isolates were isolated and identified ^[18]. These fungi were grown on Dox's agar medium was used for the isolation of halophilic fungi, dilution plate method was employed for the estimation of soil fungi as described by Johnson et al ,1959,but with some modification as employed by Moubasher *et. al.* and his coworkers (1957).

This medium consists of (g / L): NaNO3, 2; MgSO4.7H2O, 0.5; Kcl, 0.5; KH2PO4, 1; FeSO4.7H2O, traces; sucrose, 20; Agar, 20. All these components were added to 1L distilled water. The pH was set to 7 and then the medium was sterilized at 1.5 atm. for 20 min salt

medium supplemented Dox's agar with different concentrations of sodium chloride, 0, 5, 10, 15 and 20% (W/V). Each plate was inoculated with one disc of fungal isolate and incubated at 28± 2°C for 7 days, replicates were made three each treatment also control plates(medium free of sodium chloride) were prepared. Colony size of each fungal isolates was measured.

Image Analysis System

The fungal isolates were subjected for certain morphological studies by an Image Analysis System at the Regional Center for Mycology and Biotechnology, Al-Azhar University, Cairo, Egypt .

Scanning Electron Microscope (SEM)

Coated specimens of each fungal isolate were examined by using (SEM) model of JEOL JSM-5500LVat the Regional Center for Mycology and Biotechnology, Al-Azhar University, Cairo, Egypt .

RESULTS

Table 1 & 2 represented the description of **a** growth characteristics and image analysis examination of nine fungal isolates (*Emericilla nidulans*; *Mucor racemosus*; *Alternaria pluriseptata*; *Penicillium canescens*; *Syncephalastrum racemosum*; *Aspergillus fumigatus*; *Alternaria chlamydospora*;

Aspergillus parasiticus and Ulocladium atrum) isolated from soil samples of AL- SHEGA area at AL- QASSIM region.

Influence of different NaCl concentrations on the growth of fungal isolates

Figure (1) shows the growth rate of all investigated fungal isolates cultivated in medium supplemented with various concentrations of NaCl. The results indicate that the isolates exhibited good growth rate at low salt concentration while all the investigated fungi were found to decrease their growth at high salt concentration. The most suitable concentrations for well fungal growth was at 5% NaCl. At 15 % NaCl only low growth of Emericilla nidulans, Penicillium canescens, Syncephalastru mracemosum, Aspergillus parasitic and Mucor racemosus was detected, whereas the investigated fungi failed to grow at 20% NaCl. As shown in figure 1 heavy fungal growth attained at low NaCl concentration and decreased by increasing the salt concentration, so these fungal isolates are considered to be halotolerant fungi. The most tolerant ones were Emericilla nidulans (No.1); Penicillium canescens (No. 4); Syncephalastru mracemosum (No. 5) and

Table 2 shows Morphological characteristics of fungal isolates by Image Analysis System, In this study Nine fungal isolates were isolated and identified from soil samples they are as follows: *Emericilla nidulans* (No.1); *Mucor racemosus* (No.2); *Alternaria pluriseptata* (No. 3); *Penicillium canescens* (No. 4); *Syncephalastru mracemosum* (No. 5); *Aspergillus fumigatus* (No.6); *Alternaria chlamydospora* (No.7); *Aspergillu sparasiticus* (No. 8) and *Ulocladium atrum* (No.9).

Aspergillus parasiticus (No.8) (in descending

order).

To study the effect of salinity on Morphological characteristics of the isolated fungi , they were grown on Dox's agar medium supplemented with different concentrations of sodium chloride, 0, 5 and 10% (W/V)).

The most halotolerant fungal isolates were used to identify the effect of different concentrations of sodium chloride on them by using S E M.

Table 3 shows that No 1 : *Emericilla nidulans* as at NaCl free medium have globosely heads surrounded completely by conidia. As this fungal

was subjected to 5% NaCl; the globosely heads shrunken appeared spatulas' to some extent the globosely conidia changed and reduced in number .At 10 % heads completely shrunken became rode in shape, little conidia appeared.

Number 2: Mucor racemosus at NaCl free medium sporangiophores and sporangia appeared normally, columella appeared obovate. At 5 % sporangia are shrunken, spores reduced in number.; increasing NaCl to 10 % sporangiophores were dwarfed, sporangia were shrunken, and spores were rare.

Number 3: Alternaria pluriseptata this fungus was characterized by long conidiophores with chains of conidia has terminal peach .When Alternaria pluriseptata subjected to 5 % NaCl long conidiophores became shorter with chains of swollen conidia with rounded terminal. By increasing NaCl to 10% conidiophores became drastic with clusters of swollen conidia with rounded terminal or one swollen conidium.

Number 4: *Penicillium canescens* this fungus was characterized by heavily branching conidiophores carrying metulae and phialieds which have chains of conidia. At 5% and 10 % NaCl the fungus has the same appearance to some extent; conidiophores was less branching to some extent metulae and phialieds are carrying smaller number of conidia.

Number 5: Syncephalastrum racemosum this fungus has long sporangiophores with terminal vesicle which forms merosporangia all over its surface on NaCl free medium while on NaCl medium sporangia are smaller in size carrying smaller No of spores to some extent; sporangiospores were less than before; drastic action appeared on sporangiophores.

Number 6: Aspergillus fumigates the halotolerant A. fumigatus has flask-shaped vesicle carrying sterigmata with long chains of conidia .Sodium chloride affected this fungus as the flask-shaped vesicle became longitudinal with swollen sterigmata carrying little No of conidia. By increasing NaCl concentration to 10 %, vesicles became longitudinal and drastic with drastic (to some extent) sterigmata carrying little number of shrunken conidia.

Number 7: *Alternaria chlamydospora* at sodium chloride free medium conidia and conidiophores are appeared normally with some chlamydospores. At 5 % NaCl conidia and

conidiophores were enlarged clusters of conidia appeared while at 10% NaCl conidia are enlarged ; conidiophores were appeared fine covered with sheath of salt, conidia reduced in number.

Number 8: Aspergillus parasiticus flask shape heads covered with sterigmata on long conidiophores were appeared at NaCl free medium. In presence of NaCl flask shape heads became triangle covered with sterigmata on long conidiophores carrying smaller No of conidia. At 10 % NaCl flask shaped heads became triangle or rounded covered with sterigmata or naked carrying smaller No of conidia . All the fungus covered with sheath of salt in presence of all NaCl concentrations.

Number 9 : Ulocladium atrum conidia were varicose, sometimes ellipsoidal with one or more transverse and longitudinal septa but most common spherical or subspherical. In presence of NaCl spherical and ellipsoidal smooth conidia were without septa, by increasing salt concentration ellipsoidal and longitudinal conidia were without septa .Hyphae covered with sheath of salt.

DISCUSSION

The investigated fungal isolates showed distinctly improved growth on different NaCl concentrations as the solute and these results were consistent with results of many other researches, As Vaupotič *et.al.* [19,] Yadav, *etal.* [20] and Nayak *et.al.* [21,] . Their growth across increasing NaCl concentration ranges shown here confirm these previous findings of Zalar *et al.* [22]

Emericilla nidulans has achieved first place in the ability to grow and survive at salt concentration of 15 %, also Penicillium canescens, Syncephalastrum racemosum, Aspergillus parasiticus and Mucor racemosus. So all these fungal isolates are moderately halotolerant.

Alternaria pluriseptata, Aspergillus fumigates, Alternaria chlamydospora and Ulocladium atrum are weak halotolerant, they able to grow and survive at 5 % NaCl and failed to grow at salt concentration of 15%.

Growth at high salinities requires the adaptation of cellular metabolism. The reductions in the final biomass yields indicate the high energy demands of life at high NaCl concentrations. [23]

The morphological characteristic of investigated fungi grow at different salt concentrations were consistent with results of Marjetka KraljKunc ic et al. [14] who demonstrated that NaCl concentrations have an impact on the cell morphology of the Wallemia spp. At a high salinity, the hyphal compartments in both W. muriae and W. sebi were thicker and shorter compared to what have been observed at the low salinity. Such changes of the cell morphology resulted in changed size and shape of the mycelial pellets. The pellets of both W. muriae and W. sebi were small and regularly shaped at the low salinity, whereas at the high salinity, they were larger and variable in shape.

Shortening and thickening of the hyphal compartments and similar changes in pellet morphology have also been seen in less saltadapted fungi, such as in the halotolerant Aspergillu srepens when grown on medium with 12% NaCl (W/V). [24] This showed distinctly improved growth with NaCl as the solute, Their growth across increasing NaCl concentration ranges shown here confirm these previous findings of Zalaret al. [22] Growth at high salinities requires the adaptation of cellular metabolism. The reductions in the final biomass yields indicate the high energy demands of life at high NaCl concentrations, even in such welladapted species.^[23] The common feature of black yeasts at the high salinities, is hypothesized to enhance the ability to survive under conditions of stress, [25] such as a high concentration of NaCl

Halophilic and halotolerant fungi showed a surprisingly rich diversity and abundance in natural saline environments . [27]. This rich surprisingly and abundance is the result of the adjustment and adaptation of fungi in the salty environment and the impact on the fungal cell in one way or another.

At the impact of higher salinity in the elongated hyphal cells were shorter and thicker, which resulted in a changed surface-to-volume ratio. The decreased surface-to-volume ratio was mainly a consequence of decreased surface area, since the volume of the hyphal cells remained unchanged, but the thickening of the cell walls resulted in decreased functional volume of the cells. The thick cell wall might be important as an armor against changes of osmotic pressure, since

it provides a mechanical protection against hyposaline stress at dilution conditions. [14]

The sphere has a minimal surface-to-volume ratio, and this cell shape could be one of the important adaptations for growth of halotolerant fungi in saturated NaCl. At high salinities, Similar multicellular structures have also been seen at high salinities in the phylogenetically distant, extremely halotolerant ascomycetous black yeasts, such as Hortaeawerneckii (T. Kogej and N. Gunde-Cimerman, unpublished data), Phaeothecatriangularis. [28] and *Trimmatostroma* salinum .[17] phylogenetically distant from Wallemia spp., all grow meristematically and have spherical cells, [14] a finding which supports this conclusion. The overall morphological changes seen in Wallemia spp. correspond to the phylogenetics based on the ITS ribosomal DNA gene. [22].

Through the results of this study and which are consistent with results of many other researches, it was concluded that the acquisition of fungi recipe halophilic and halotolerant is affected by the result of change and deformation in the form of cell adapt to these fungi in saline environment.

CONCLUSION

The Scanning Electron Microscope (SEM), and Image Analysis System had demonstrated various morphological changes at high salinity in Nine fungal isolates. At high salinities, the common morphological feature was decreasing in the growth of fungal isolates by increasing NaCl concentration which means that all these fungi are halotolerant fungi.

REFERENCES

- 1) **Abdulnapi M.** (2012):Shoulder of fungi to extreme environmental conditionshttp://kenanaonline.com
- 2) Yancey P.H. (2005): Organic osmolytes as compatible, metabolic and counteracting cytoprotectants in high osmolarity and other stresses. J Exp Biol., 208:2819-30.
- 3) De Hoog, G. S.; Zalar, P.; van den Ende B.G. and Gunde-Cimerman, N. (2005): Relation of halotolerance to human-pathogenicity in the fungal tree of life: an overview of ecology and evolution under stress, p. 373-395. In N. Gunde-Cimerman, A. Oren, and A. Plemenitaš (ed.), Adaptation to life at

- high salt concentrations in Archaea, Bacteria, and Eukarya, vol. 9. Springer, Dordrecht, the Netherlands.
- 4) **Blomberg A and Adler L (1992):** Physiology of osmotolerance in fungi. Adv. Microb. Physiol.,33:145-212.
- 5) **Hohmann S (2002)**: Osmotic adaptation in yeast: control of the yeast osmolyte system. Int. Rev. Cytol., 215:149-187.
- 6) Rosemeire A B, Pessoni GF, Rita de Cássia L, Figueiredo-Ribeiro MG and Marcia R B (2005): Cell-wall structure and composition of *Penicillium janczewskii* as affected by inulin.Mycologia,97:2 304-311
- 7) Mager WH and Siderius M (2002): Novel insights into the osmotic stress response of yeast. FEMS Yeast Res., 2:251-257.
- 8) Bowman S M and Free S J (2006): The structure and synthesis of the fungal cell wall. Bioessays ,28:799-808.
- 9) Latgé J P (2007): The cell wall: a carbohydrate armor for the fungal cell. Mol. Microbiol., 66:279-290.
- 10) Chung Y S, Kim J, Han D, Chae K and Jahng K(2003): Ultrastructure of the cell wall of a null pigmentation mutant, npgA1, in Aspergillusnidulans. J. Microbiol., 41:224-231.
- 11) **Terenzi H, Terenzi HF and Jorge JA (1992):** Effect of cyclic AMP on invertase activity of Neurosporacrassa. J Gen Microbiol., 138:2433–2439.
- 12) Silva MM, Polizeli MLT, Jorge JA and Terenzi HF (1994): Cell wall deficiency in "slime" strains of *Neurosporacrassa*: osmotic inhibition of cell wall synthesis and β -D-glucan synthase activity. Braz J Med Biol Res., 27:2843–2857.
- 13) Yi W, Zhenyu L, Kunlai S and Weiming Z (2011): Effects of High Salt Stress on Secondary Metabolite Production in the Marine-Derived Fungus Spicariaelegans. Mar Drugs, 9: 535–542
- 14) Marjetka K, TinaKogej D D and Gunde-Cimerman N (2010): Morphological Response of the Halophilic Fungal Genus Wallemia to High Salinity. Appl. Environ. Microbiol., 76 (1): 329-337.
- 15) Gunde-Cimerman N, Zalar P, de Hoog S and Plemenitaš A (2000): Hypersaline waters in salterns: natural ecological niches for halophilic black yeasts. FEMS Microbiol. Ecol., 32:235-2.
- 16) Kogej T, Gostinčar C,Volkmann M, Gorbushina A A and Gunde-Cimerman N (2006): Mycosporines in extremophilic fungi: novel complementary osmolytes? Environ. Chem., 3:105-110.
- 17) Kogej T, Gorbushina A A and Gunde-Cimerman N (2006): Hypersaline conditions induce changes in cell-wall melanization and colony structure in a halophilic and a xerophilic black yeast species of the genus Trimmatostroma. Mycol. Res., 110:713-724.

- **18) Tamie AL and Mona S (2014):** Effect of Salinity on The Fungal Occurrence of The AL Shega Area at AL- Qassim Saudi Arabia . Research Journal of Microbiology ,9(6): 287-295.
- 19) Vaupotič T, Veranic P, Petrovič U, Gunde-Cimerman N and Plemenitaš A (2008): HMG-CoA reductase is regulated by environmental salinity and its activity is essential for halotolerance in halophilic fungi available online at www.studiesinmycology.org Studies in Mycology, 61: 61–66
- **20) Yadav J, Verma YS K and Tiwari K N (2011):** Effect of salt concentration and pH on soil in habiting fungus *Penicillium citrinumthom*. For solubilization of tricalciumphosphate. Microbiology Journal, 1(1): 25-32.
- 21) Nayak SS, Valerie G and Sarita W N (2012): Isolation and salt tolerance of halophilic fungi mangroes and solar salternsin Goa –India. Indian Journal of Goa marine Sciences, 41(2): 164-172.
- 22) Zalar P, Sybren de Hoog G, Schroers H J, Frank J M and Gunde-Cimerman N (2005): Taxonomy and phylogeny of the xerophilic genus Wallemia (*Wallemiomycetes and Wallemiales*, cl. et ord. nov.). Antonie van Leeuwenhoek, 87:311-328.
- **23**) **Oren A** (**1999**): Bioenergetic aspects of halophilism. Microbiol. Mol. Biol. Rev., 63:334-348.
- **24) Kelavakar U, Rao K S and Chhatpar H S** (1993): Sodium chloride stress induced morphological and ultrastructural changes in Aspergillusrepens. Ind. J. Exp. Biol., 31:511-515.
- 25) Wollenzien U, de Hoog G S, Krumbein W E and Urzì C (1995): On the isolation of microcolonial fungi occurring on and in marble and other calcareous rocks. Sci. Total Environ., 167:287-294.
- **26**) **Zalar P G, de Hoog S and Gunde-Cimerman N** (**1999**): Ecology of halotolerant dothideaceous black yeasts. Stud. Mycol., 43:38-48.
- 27) Butinar L, Santos S, Spencer-Martins I, Oren A and Gunde-Cimerman N (2005): Yeast diversity in hypersaline habitats.FEMS Microbiol. Lett., 244:229-234.
- 28) De Hoog G S, Beguin H and Batenburg-van de Vegte W H (1997): Phaeothecatriangularis, a newmeristematic black yeast from a humidifier. Antonie van Leeuwenhoek, 71:289-295

Table 1: A growth characteristics and image analysis examination of nine fungal isolates from soil samples of AL- SHEGA area at AL-

QASSIM region.

No.	ASSIM region.	Character								
110.	Isolated fungi	*Culture Exam.:					* Microscopic Exam.:			
1	Emericellanidul ands var. aurantiobrunn	Growth Characteristics	Conidial heads:	Conidiophore	es:	Vesicles	Sterigmata		Conidia	Hűll cells and Cleistothecia
	ea	Colonies attending 3-4 cm in 7 days at 25°C on both Czapek-dox and malt agar media, central initiate white and velvety and become cream to buff and granular or powdery golden browed reverse	Columnar, 43 µm in diameter	6.5 μm in diameter		10.6 μm in diameter	Sterigmate two series Primary 7.0X3.0 X 6.2X2.5 µ	•	Globose to subglobose 3.5µm in diameter	Hűll cells abundant and cleistothecia usually in margin.
2	Mucorracemos us	Growth Characteristics	Sporangia	μm in Collumellaobovoid, neter, ellipsoidal, slightly pyriform (4 μm in diam).		ella	Sporangiospor es		Chlamydospores	
		Colonies white become brownish – grey; reaching 6 cm colony diameter in 4 days.	65.3 µm in diameter, wall spinulose			7.5 X 5.3 in diamete		Chlamydos in sporangio	spore numerous ophores	
3	Alternariapl uriseptata	Growth Characteristics								
		Colonies grow rapidly, effuse, black.	Conidioph ores 4.3µm thick.	smooth 32.	Conidia obclavate, golden bro smooth 32.0X15.5 µm, with a µm thick, with 2-7 transverse			k 4.0		
4	Penicillimcane	Growth Characteristics	Penicillus	type	Ra	mi	Metulae	Pł	nialides	Conidia
	scens	Light yellow to light orange colonies on CYA with yellow to yellowish brown reverse. Micro colonies at 5°C and 37 °C.	Bivert some			5X4.1	14.2X3.0µm.		8Χ2.4μm.	Conidia globose , smooth walled 3.0 µm
5		Growth characteristics	Sporangiophore Sporangiophore with curl lateral branches each bearing a terminal vesicle which forums			Vesicle Merosporan		angia		Merospores
	Syncephlastru mracemosum	Colonies on white becoming grey with age, 5-6 cm in 7 days.				Vesicle globose; 40.0 µm.	; 25.0X3.0 μm.		lindrical grey	Merosporess ubglobose 5.0 μm.

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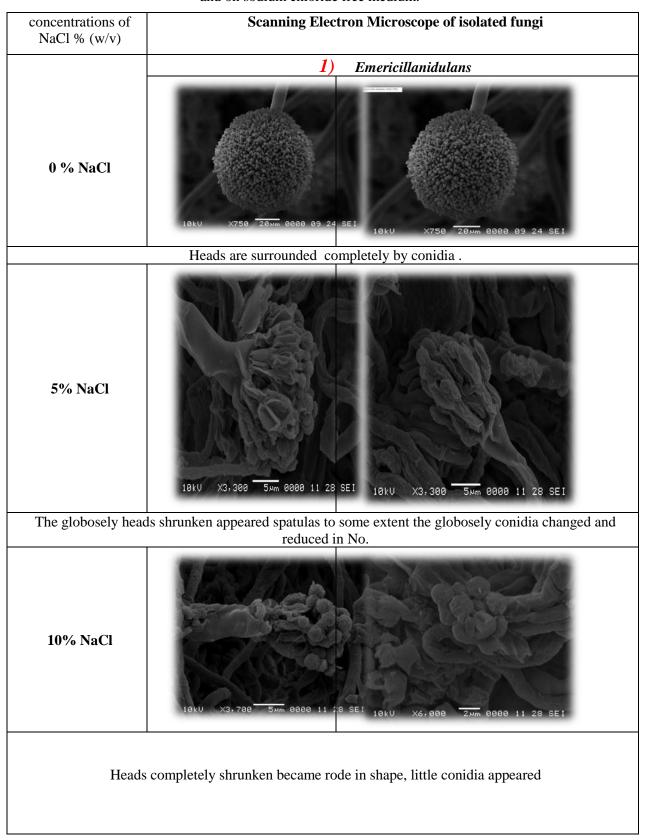
			merosporangia al surface.	l over its					
6	Aspergillus fumigatus	Growth characteristics	Conidiophores		Conidial Vesicles heads		Sterigmata	Conidia	
		Colonies spreeding rapidly on Czapek's agar and MEA at 25°C.			Columnar, 180 µm X 30 µm.	25 µm in diameter, fertile over the upper half only.	Sterigmata in one series, 6.0x2.2 µm.	Globose, echinulate, green- coloured, 2.8 µm in diameter	
7	Alternariachla mydospora	Growth characteristics	Conidiophores	Conidia				Chlamydos pores	
		Colonies floccose, dark olive to blackish brown with numerous chlamydospores.	Conidiophores 5.0 µm thick.		dia obclavate than swelling brown in colour, smooth Κ12.5 μm, with a short, pale beak 4.0 μm thick, with 2-5 werse septa			Abundant	
8	Aspergillusp	Growth characteristics	Conidiophores	Vesicles		Sterigmata	Conidia		
	arasiticus	Colonies fast-growing, reaching 5-7cm diameters in four days at 25°C on malt media; usually consist of a dense felt of yellowish green mycelia, with pale yellow reverse	Conidiophores are coarsely roughened, geenish yellow in color, 10.4 µm in diam.	24.5 μm in one series, 8 diam., 3.33.4 μm.		Sterigmata in one series, 8.6 – 8.33.4 µm.	Conidia globose, 3.5 µm in diam. Yellow-green, conspicuously rough-walled.		
9	Ulocladiuma trum	Growth characteristics Colonies on MEA growing rapidly, golden brown to black. Black reverse.	Conidiophore 5.5 μm. In diameter.	Conidia Conidia golden brown, or dark reddish brown, verrucose, sometimes ellipsoidal 25.6 X 15.8 with 1 – 3 transverse and 1 or more longitudinal septa but most common spherical or subspherical.					

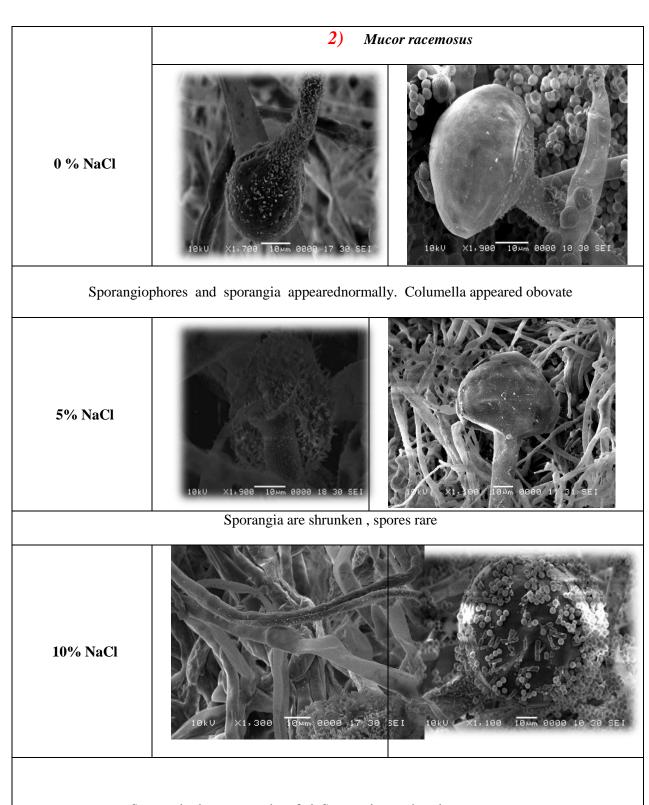
Table 2: Morphological characteristics of fungal isolates by Image Analysis System.

1)Emericell anidulands var. aurantiobrunnea	2) Mucor racemosus	3) Alternaria pluriseptata		
4) Penicillium canescens	5) Syncephlastrum racemosum	6) Aspergillus fumigatus		
7) Alternaria chlamydospora	8) Aspergillus parasiticus	9) Ulocladium atrum		

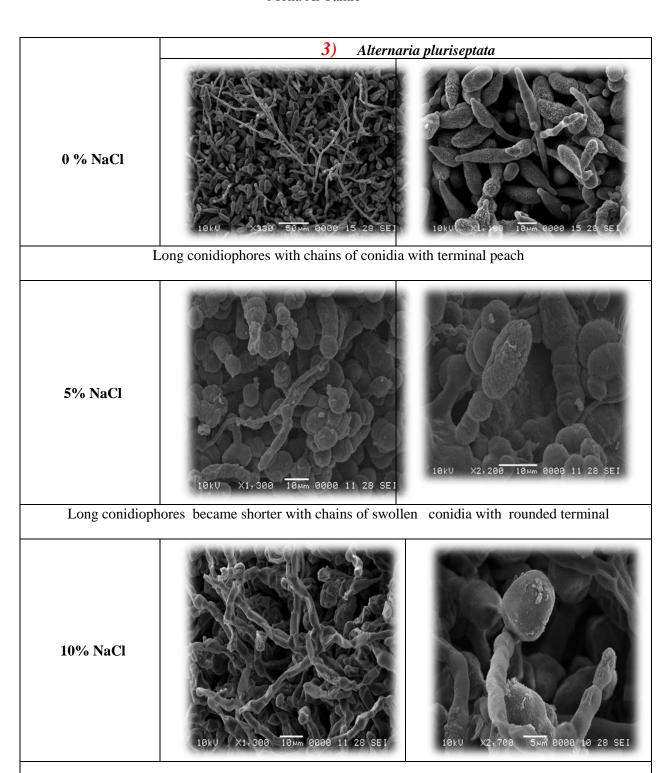
Morphological characteristics of fungal isolates by Image Analysis System, they are as follows: *Emericilla nidulans* (No.1); *Mucor racemosus* (No.2); *Alternaria pluriseptata* (No. 3); *Penicillium canescens* (No. 4); *Syncephalastru mracemosum* (No. 5); *Aspergillus fumigatus* (No.6); *Alternaria chlamydospora* (No.7); *Aspergillu sparasiticus* (No. 8) and *Ulocladium atrum* (No.9).

Table 3: Scanning Electron Microscope of isolated fungi at different concentrations of sodium chloride and on sodium chloride free medium.

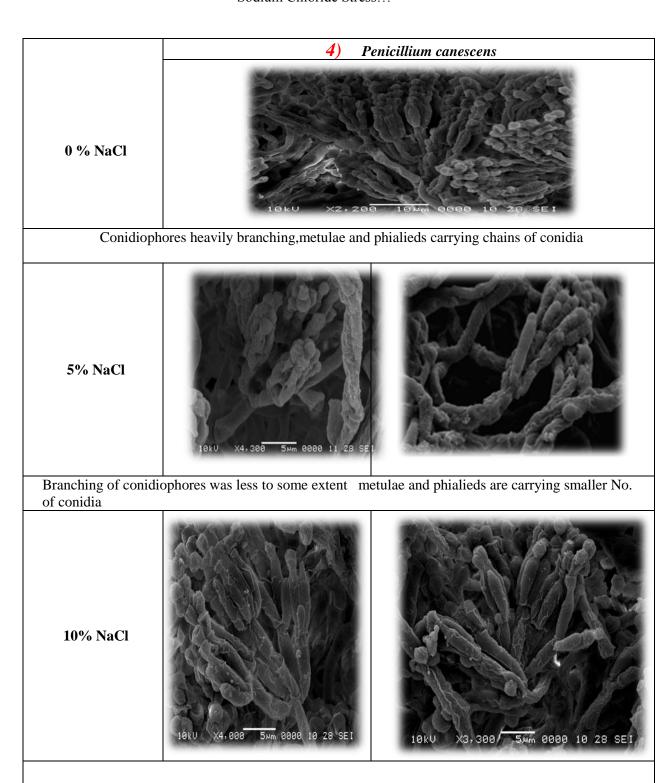




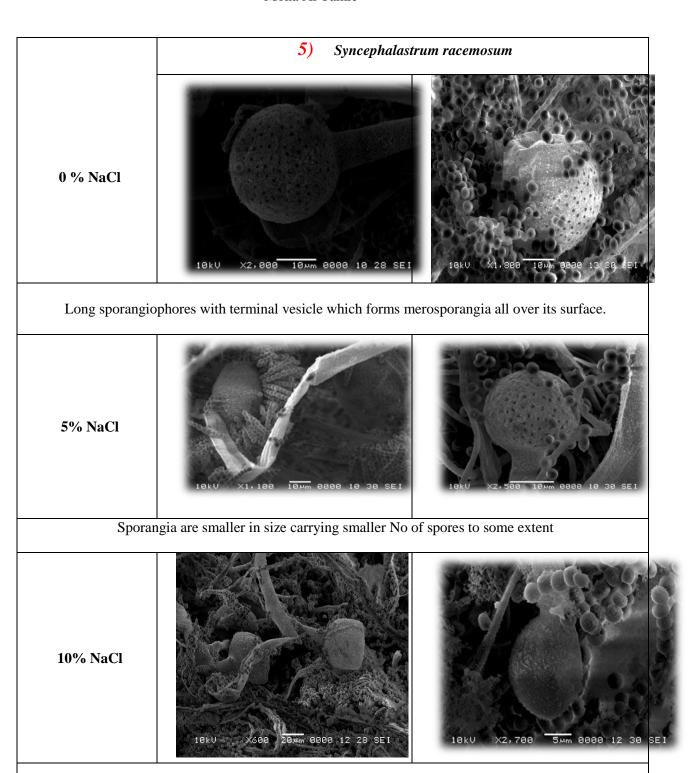
Sporangiophoreswere dwarfed. Sporangia are shrunken, spores rare



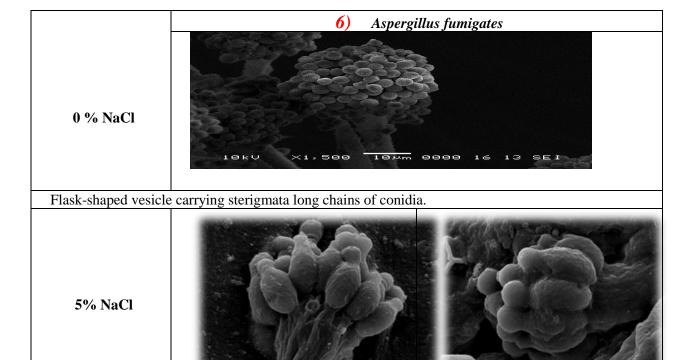
Conidiophores became drastic with clusters of swollen conidia with rounded terminal or one swollen conidium



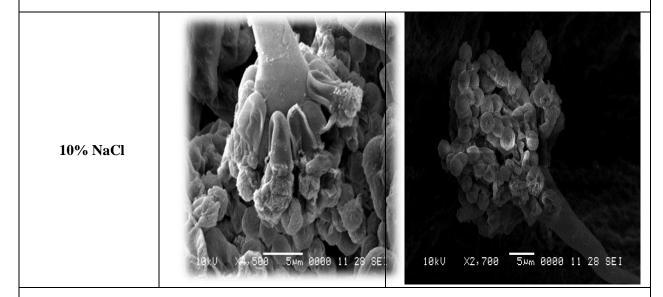
Branching of conidiophores was less to some extent metulae and phialieds are carrying smaller No. of conidia or no conidia



Sporangia size were very small sporangiospores were less than before. Drastic action appeared on sporangiophores



Flask-shaped vesicle became longitudinal with swollen sterigmata carrying little No of conidia

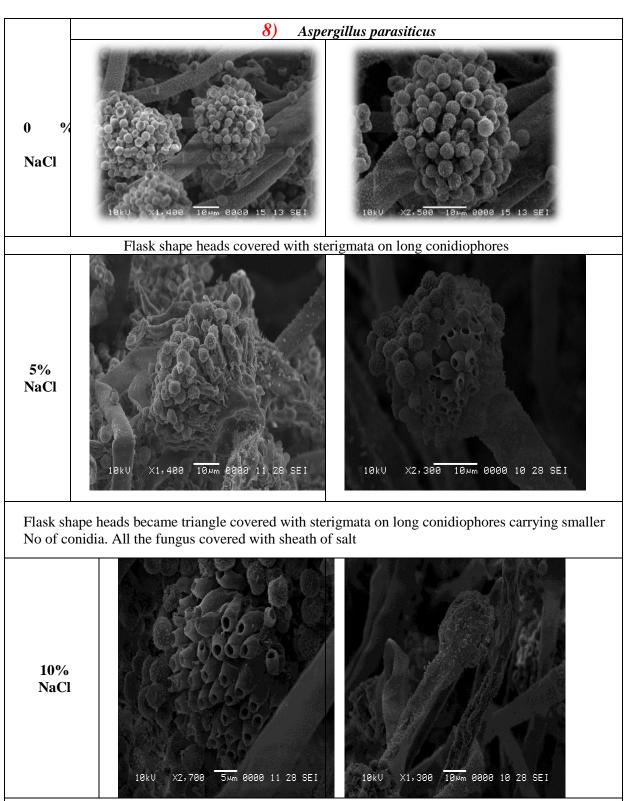


Vesicles became longitudinal and drastic with drastic (to some extent) sterigmata carrying little No of shrunken conidia

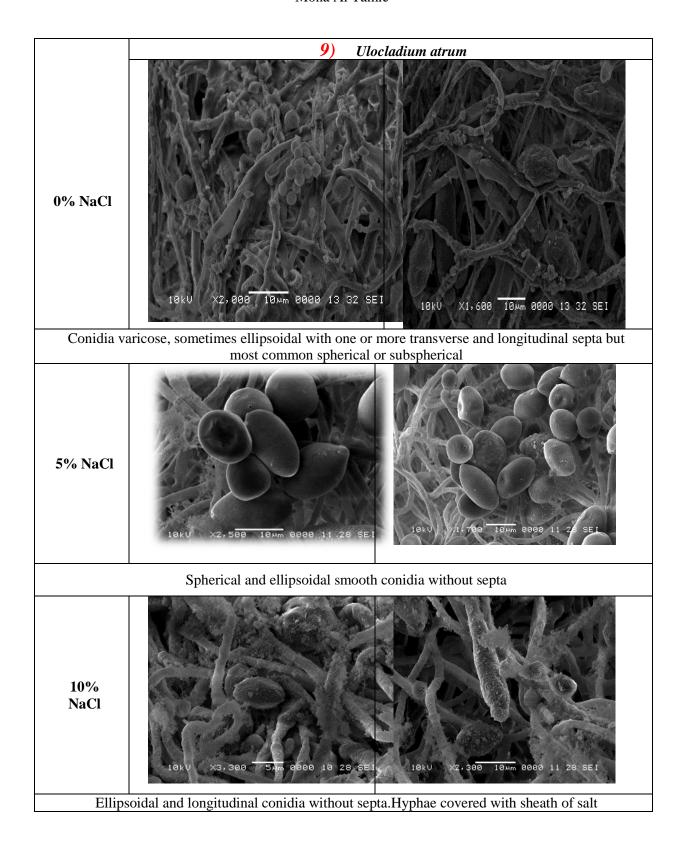
7) Alternaria chlamydospora 0% NaCl 1,400 10 m 0000 16 28 SEI Conidia and conidiophores are appeared normally with some chlamydospores 5% NaCl Conidia and conidiophores are enlarged clusters of conidia appeared. 10% NaCl

Conidia are enlarged while conidiophores are appeared fine covered with sheath of salt, conidia reduced in No.

X2,500 10 m 0000



Flask shape heads became triangle or rounded covered with sterigmata or naked carrying smaller No of conidia .All the fungus covered with sheath of salt.



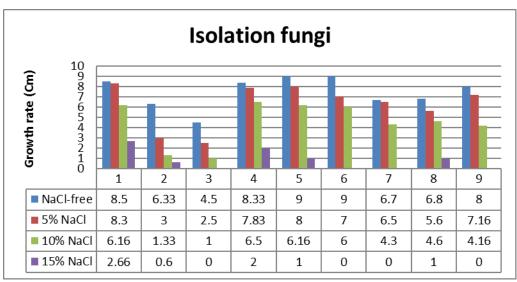


Figure 1: Influence of different NaCl concentrations on the growth of fungal isolates

The fungal isolates as follow: Emericilla nidulans (No.1); Mucor racemosus (No.2); Alternaria pluriseptata (No. 3); Penicillium canescens (No. 4); Syncephalastru mracemosum (No. 5); Aspergillus fumigatus (No.6); Alternaria chlamydospora (No.7); Aspergillu sparasiticus (No. 8) and Ulocladium atrum (No.9)