Effect of Noise and Crowding Related Stress on Serum Level of Cortisol ACTH, Epinephrine and Insulin in Female Albino Rats.

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Abstract

Background: Noise and crowding are the most stressful factors for human beings.

Study aimed to clarify their effect on cortisol, ACTH, epinephrine, insulin and the amelioration effect of Sulpiride.

Material and Methods: Thirty six female rats were divided into six groups (6/each):

1- Rats served as control, 2- Rats treated with Sulpiride drug, 3- Rats exposed to noise (90db, 3hr. per day) for 45 days. 4- Rats exposed to noise and treated with sulpiride drug, 5- Rats exposed to crowding. 6- Rats exposed to noise and treated with Sulpiride drug.

Results: Noise and crowding stresses caused a significant increase of cortisol, ACTH and epinephrine while there was a significant decrease in insulin hormone. Sulpiride drug ameliorated these parameters.

Conclusion: it is useful to use Sulpiride drug with people who are exposing to noise and crowding stress.

Keywords: Noise, Crowding, Sulpiride drug, Physiological parameters, Cortisol, ACTH, Epinephrine, Insulin.

INTERODUCTION

Stress can be defined as a state of threatened balance, induced by external stressor and appear as the display of somatic and psychic reaction, struggling to regain homeostasis ⁽¹⁾ among stressful stimuli, crowding and noise.

Noise is an environmental pollutant capable of causing hearing impairment ⁽²⁾, behavioral, mental and widespread disturbances at several levels in human organs and apparatus due to chemical and physiological modification of endocrine ⁽³⁾.

Crowding stress is a type of psychosocial stress induced by an increased density of population. Population density may be raised either by increasing the number of species living in the same area and/or by reducing their living space ⁽⁴⁾.

Antidepressant drugs are the most successful drugs in patients with clear vegetative characteristics including retardation, psychomotor stress, sleep disturbance, poor appetite and weight loss. However, a variety of different chemical have structures have been found to antidepressant activity. Their number is constantly growing, but as yet no group has been found to have a clear therapeutic advantage over the others ⁽⁵⁾.

Sulpiride is the most favorite drug which used to tolerate stress symptom ⁽⁶⁾. People who

expose to stress take one or some drugs to avoid its effect. So, the present study deals with the possible protective effect of one of the antidepressant drugs (Sulpiride) against noise and crowding stresses in female albino rats..

Aim of the work: during all the day time Cairo is extra crowded and noise city, where it suffer from movement of huge number of especially in the morning about 30,000,000 persons in the morning. Also it is suffer from a very loud noise. This study amid at clarifying the effect of each stress on some hormonal pattern. The question arise here, could Sulpiride drug (which is a favorable and widely used drug) completely ameliorate stress effects or it would be worsen?

Material and Methods

Thirty six normal white female albino rats weighing 150 ± 30 g were purchased from the Farm of the National Organization for Control and Research. They were kept under observation for one week before the beginning of the experiment acclimation period .The chosen animals were housed in cages and kept under to artificial light for 14 hr. and 10 rodent hr. complete darkness at normal atmospheric temperature. All animals were fed on standard rodent diet contained protein, fibers, fats, ash, carbohydrates and supplied with vitamins and minerals mixture, The animals had free access to water

1-Sulpiride administration:

The drug was adiministrated orally by gastric tube at a dose of 0.28mg/100gm. body weigh/day for 45 days. The dose for the rat was calculated according to the Paget's formula on the basis of the human dose ⁽⁷⁾.

2- Application of Noise:

Noise was applied by 5 different sources of enharmonic and high intensity music.

3- Application of crowding:

A group of 6 rats were put in a cage $(20 \times 15 \times 20 \text{ cm})$.

5- Animal groups:

Thirty six female albino rats were divided into six main groups each group contained six rats.

Group1:

Normal rats served as control group untreated for a duration of 45 days in a cage (20 \times 30 \times 20).

Group 2:

Results

Rats were treated with the Sulpiride drug at a dose of 0.28mg/100mg. body weigh/day for a duration of 45days.

Group 3:

Rats were exposed to noise (90db, 3hr. per day) for a duration of 45days.

Group 4:

Rats were exposed to noise and treated with the drug for a duration of 45days.

Group 5:

Rats were exposed to crowding only for a duration of 45 days.

Group 6:

Rats were exposed to crowding and treated with the drug a duration of 45days.

Serum cortisol was estimated according to method described by **Kytzia** ⁽⁸⁾, adrenocorticotropic hermone (ACTH) was measured according to the method of **Sugiuchi** ⁽⁹⁾, epinephrine was detected using the method of **Burtis** *et al.* ⁽¹⁰⁾ and insulin was measured according to the method of **Junge** *et al.* ⁽¹¹⁾.

6-Data analysis:

The obtained data were statistically analyzed by using the student t-test according to the method of **Snedecor and Cochran**⁽¹²⁾, P ≤ 0.05 considered significant while P ≤ 0.01 highly significant.

Noise + Crowding Group Control Crowding Drug Noise Parameter Drug + Drug 101.7 94.8 149.1 119.8 134 120 Mean Cortisol (µg/ dL) **SE±** 1.3 1.98 1.2 1.6 1.9 1.3 Р N.s < 0.01 < 0.05 < 0.01 < 0.05 % of change -6.8 46.6 17.7 31.7 17.9 39.6 59.3 49.5 39.1 47.3 Mean 62 3.1 1.34 SE± 0.6 2.3 0.8 3 ACTH (pg/mL) <0.01 <0.01 < 0.05 Р N.S < 0.05 % of Change -1.2 56.5 19.4 49.7 25 17.35 20 24.5 32.3 25.5 Mean 30.3 Epinephrine SE± 0.86 0.6 1.37 1.4 1.37 0.6 < 0.05 0.05 (pg/mL) Р N.S < 0.01 0.01< % of Change 15.2 74.6 41.2 86.1 46.9 21.93 22.55 13.5 15.1 12.1 16.3 Mean Insulin **SE**± 0.7 0.22 0.3 0.2 0.31 0.21 (µIU/mL) % of Change 2.8 -38.4 -31.1 -44.8 -25.6

Table (1): Percentage of change of cortisol, ACTH, epinephrine and insulin in female albino rats after exposure to the stressors (noise or crowding) and treated with the Sulpiride.

Table (1) showed a highly significant increase ($P \le 0.01$) of cortisol, ACTH and epinephrine levels in rats exposed to the stressors (noise and crowding) in comparison with those of control group. But Exposure of rats to noise, crowding exposure and treated groups with Sulpiride showed less significant increase ($P \le 0.05$) when compared with control.

However serum insulin level in rats exposed to stress showed a highly significant decrease in comparison with that of control group. Sulpiride supplementation to rats exposed to noise showed an increase in the hormone level from -38.4 to -31.1% while, the rats exposed to crowding and treated with Sulpiride exhibited improvement in insulin level to be 25.6% instead of -44%8%

Discussion

Stress is defined as the state in which the brain interprets the quantity of stimulation as excessive or its quality as threatening ⁽¹³⁾. Stress stimulates several adaptive hormonal responses, prominent among which are the secretion of catecholamines from the adrenal medulla, corticosteroids from the adrenal cortex and adrenocorticotropin from the anterior pituitary ⁽¹⁴⁾.

Results of the present study revealed a significant increase in stress hormones (ACTH, cortisol and epinephrine) levels as compared to those of control group.

When the body is activated by a stressor, the hypothalamus is stimulated by an unknown signal with release of corticotrophin – releasing hormone (CRH) which stimulates the anterior pituitary to secrete adrenocorticotrophic hormone (ACTH) which is considered as the hormone of stress. ACTH stimulates secretion and growth of zona fasciculate and zona reticularis of adrenal gland and stimulates the secretion of cortisol. So, there is an increase in plasma cortisol level and increase in the adrenal weight ⁽¹⁵⁾.

The hypothalamus also stimulates the adrenal medulla to increase secretion of catecholamine ⁽¹⁴⁾. The mechanism by which noise and crowding stresses produced elevation in stress hormones may be explained by activation of sympatho-adreno-medullary (SAM) system with increased secretion of epinephrine and whole body activation that is called fight and flight response. Release of epinephrine from the adrenal medulla and glucocorticoids from the adrenal cortex initiate the biological responses permitting the organism to cope with adverse psychological, physiological and environmental stressors. Following its massive release during stress, epinephrine must be restored to replenish cellular pools and sustain release to maintain the heightened awareness and squeal of responses to re-establish homeostasis survival (16) and ensure Epinephrine is regulated in part through its biosynthesis catalyzed by the final enzyme in catecholamine the pathway, phenylethanolamine N-methyltransferase (PNMT) expression, in turn, is controlled

through hormonal and neural stimuli, which exert their effects on gene transcription through protein stability ⁽¹⁷⁾.

Dirk *et al.* ⁽¹⁸⁾ described the physiological responses to physical or psychological stress as the release of "adrenocorticotropin (ACTH)

from the anterior pituitary, glucocorticoids from the adrenal cortex, epinephrine from the adrenal medulla, and norepinephrine from sympathetic nerves. Together, these hormones enable organisms to biologically adapt to stressors, ranging from mild to severe, thereby ensuring the organism's survival. In cases of prolonged stress, release of epinephrine evokes compensatory elevation of the catecholamine biosynthetic enzymes, tyrosine hydroxylase (TH), dopamine β -hydroxylase (DBH) and replenish PNMT. to depleted neurotransmitter/neurohormone stores, and the magnitude and temporal pattern of change in hormonal and neural activity elicited by a particular stress may differentially affect these enzymes. Variation in plasma catecholamine levels evoked by a variety of stressors points to this possibility ⁽¹⁹⁾.

In noise and crowding stress groups, these results are in agreement with the findings of **Lofgren** *et al.* ⁽²⁰⁾. who reported that, there is an elevation in ACTH and cortisol level in humans and animals that exposed to air craft noise, road traffic noise and density population. **Sylvia** *et al.* ⁽²¹⁾ reported that, exposure of rats to noise and crowding produced increase of ACTH, cortisol and epinepherine level combined with changes of the myocardial ultra-structure.

These results are also compatible with those of **Marks** *et al.* ⁽²²⁾ who demonstrated that, noise and crowding stress induced cortisol secretion based upon the existence of much closed connection between sub cortical structure of central nervous system and part of auditory system can activate HPA axis. The increased volumes of cortex, medulla and total adrenal gland in noise and crowding exposed groups may be also correlated to the hyperactivity of HPA axis causing an increase in the activity and secretion of this gland. The hyperactivity of adrenal gland may be dependent on cell hypertrophy ⁽²²⁾. Some stereological studies have shown that after exposure to noise and crowding stress an increase adrenal gland weight and enlargement of the zona fasciculate cells occurred due to a notable increase in the volume of mitochondria and hypertrophy changes in different layers of adrenal cortex ^(23, 24, 25 and 26). However these results disagreement with Those of **Chrousos** ⁽²⁷⁾ who found that, cortisol and epinephrine level decreased in rat after chronic repeated exposure to noise and crowding stresses due to adaptation of HPA axis.

More over, the present results also disagreement with those of **Animesh** *et al.* ⁽²⁸⁾ who reported that, plasma cortisol and glucose concentration returned to values approximately similar to those prior to stress on the 30th day of the experiment, reflecting possible adaptation of the animals to stress.

The hypothalamus –pituitary –adrenal (HPA) axis; its response to a stimulus is sustained for an appropriate time, and is turned off when the stressor is efficiently inactivated and when stressor inactivation is inefficient, the HPA axis becomes desensitized through its normal adaptation to repeated stressors of the same type and this results in inadequate secretion of cortisol ⁽²⁹⁾.

The results of the present study revealed a significant decrease in the insulin level as compared to control group. Exposure of rats to noise and crowding stress showed activation of (HPA) with subsequent release of adrenocorticotrophic hormone, cortisol and catecholamines leading to increase glucose level and decreased insulin level ⁽³⁰⁾. Significantly higher plasma cortisol level could increase responsiveness of pancreatic cell to glucose of low insulin level ⁽³¹⁾.

These results are also in agreement with those of **Julia** *et al.* ⁽³²⁾ who reported that, plasma cortisol is the stress hormone that affects the way of the body processes insulin. This steroid hormone makes muscle and fat cells resistant to insulin and increases the production of glucose. The body depends on insulin to regulate glucose levels in blood. Insulin works by processing glucose into energy in cells. Diabetes is a condition that affects of ability to produce or process insulin. The increase in glucose due to stressful situations can alter the amount of insulin necessary to provide healthy blood-sugar levels, meaning individuals with diabetes may experience glucose spikes during periods of stress ⁽³³⁾.

Sissel *et al.* ⁽³⁴⁾ reported that, hyperglycemia may also be decreased insulin secretion which may be due to noise or crowding stress or due to the associated food intake.

The hyperglycemia effect of stress may be also due to sympatho adrenal stimulation which may cause a concomitant increase in glucagons level ⁽³⁵⁾. Stress hyperglycemia and decreased insulin may be also due to stimulation of hypothalamic noradrenergic activity and hepatic glucose output. So, if the central noradrenergic activity had been inhibited, the stress hyperglycemia also inhibited ⁽³⁶⁾. Stressed rats treated with Sulpiride drug led to some extent an amelioration in ACTH, cortisol, epinephrine and insulin because antidepressant effect of Stress increased lipid peroxidation, which resulted from increased free radical generation. Dose of Sulpiride and treatment duration have been chosen on the basis of previous studies performed by Wang⁽³⁷⁾ who found that, the long-term administration of Sulipiride at the daily dose 4mg/kg is effective in animal models of depression.

The mechanism by which Sulpiride decreases stress hormones is by lowering responsiveness of HPA axis, which is up regulated by stress and contributes to the alleviation ⁽³⁸⁾. Sulpiride improves the quality of life for patient receiving palliative care, enhancing a sense of comfort and help one to cope with stress; it may also evoke stress, anexiety and intolerance ⁽³⁹⁾.

Sulpiride; a atypical (1-4) neuroleptic from the group of banzamides, is a selective dopaminergic receptor, namely D2 and D3 receptor antagonis. Sulpiride selectively blocks the above–mentioned types of dopaminergic receptors. It is an exception ally hydrophilic drug and not lipophilic as most drugs of this type are. It causes no strong extra pyramidal symptoms which could result from D2 receptor blocked in the corpaus striatum and which are the equivalent of catalepsia in animals ⁽⁴⁰⁾.

According to above mentioned results it is possible to conclude that Sulpiride a administration to stressed animals nearly inhibited the effect of stress, so the functions of the affected glands were nearly improved. This led to the improvement of the hormonal disruption which induced by stress. More studies must be done or other types of antidepressant drugs, and variable stress factors in order to justify the possible ameliorative effects of antidepressant drugs against the screening risks of exposure to environmental stressors which induce several diseases in the body due to the induction of hormonal imbalance stress and combination of different stresses.

Reference

1-Unidad d, Angiología y, Cirugía V (2010): The Role of the Stress-Related Anti- Inflammatory Hormones ACTH and Cortisol in Atherosclerosis. Current Vascular Pharmacology, Bentham Science Publishers Ltd., 66: 516-525.

2-Paul M, Radosevich D, Brooks L, Brown E (**2010**): Effects of insulin induced hypoglycemia on plasma cerebrospinal fluid level of ACTH, cortisol, norepinephrin, insulin and glucose in the dog. Brain Research, 458: 325-338.

3- Knott LJ, Cummins FR, Dunshea BJ (2010): Feed efficiency and body composition are related to cortisol response to adrencorticotropin hormone and insulin–induced hypoglycemia in rams. Domestic Animal Endocrinology, 39:137-146.

4-Eva V, Harald G, Ellen M, Svein O (2010): Cangesin muscle and blood plasma proteomes of Atlantic salmon induced by crowding. Journal Aquaculture, 132: 272-279.

5- Kyohei C, Juhyon K, Kazuo OS (2011): Electrophysiological effects of orexin-B and dopamine on nucleus accumben shell neurons *in vitro*. Pharmacology Biochemistry, 32: 246-252.

6-Otten E, Kanitz M, Tuchscherer KP Brüssow G (2008): Repeated administraton of ACTH hormone during late gestation in pigs: Maternal cortisol response and effects on fetal HPA axis and brain. Theriogenology, 69: 312-322.

7-Paget GE, Barnes JM (1964): Evaluation of drug activity in Pharmaceutics. Laurence and Bacharach eds., Vol.1 Academic press. New York.

8-Kytzia H (2005): Reference intervals for GGY according to the new IFCC 37° C reference procedure. Abstracts: Congress of Clinical Chemistry and Laboratory Medicine, 103:6-8.

9-Sugiuchi (2005): History of development and technical details of homogenous assay for hormones. ENG. J. Med., 1:4-11.

10- Burtis CA, Ashwood ER, Burns DE (2006): Clinical Chemistry and Molecular Diagnostics. Philadelphia, pa: W .B. Sanders. 4th ed. Pp.: 549-587.

11- Junge W, Wilke B, Halabi A, Klein G (2004): Determination of reference intervals for serum insulin and modified method. Clin. Chem. Acta, 344: 137-148.

12- Snedecor GW, **Cochran WG** (**1980**): Statistical Method .United State University Press, Lowa, London, Pp: 59-60.

13-Young EA, Abelson J, Lightman SL (2013): Cortisol pulsatility and its role in stress regulation and health. Front. Neuroendocrinol., 25: 69-76.

14-Broadley AJ, Korszun A, Abdelaal E (**2005**): Inhibition of cortisol production with metyrapone prevents mental stress-induced endothelial dysfunction and baroreflex impairment. J. Am. Coll. Cardiol., 46: 344-350.

15-Broadley AJ, Korszun A, Abdelaal E (2006): Metyrapone improves endothelial dysfunction in patients with treated depression. J. Am. Coll. Cardiol., 48: 170-175.

16-Slominski A, Wortsman J, Luger T, Paus R, Solomon S (2010): Corticotropin releasing hormone and propiomelanocortin involvement in the cutaneous response to stress. Physiol. Rev., 80: 979-1020.

17-Luger TA, Kalden D, Scholzen TE, Brzoska T (2013): Alpha melanocyte stimulating hormone as a mediator of tolerance induction. Pathobiology, 67: 318

18-Dirk H, Hellhammer S, Wüst B (2009): Original salivary cortisol as a biomarker in stress research. Psychoneuroendocrinology, 34: 163-171.

19-Prada TP, Pozzi AO, Coronado MT (2007): Atherogenesis takes place in cholesterol-fed rabbits when circulating concentrations of endogenous cortisol are increased and inflammation suppressed. Atherosclerosis, 27: 333-339.

20-Lofgren R, Sikorski J Sjolander A (2009): Andersson T. Beta 2 integrin engagement triggers actin polymerization and phosphatidylinositol trisphosphate formation in non-adherent human neutrophils. J. Cell. Biol., 123:1597-1605.

21-Sylvia F, Heik B, Norbert, S (2010): Effects of ACTH applications during pregnancy on the female offspring endocrine and behavior in guinea pigs. Physiology Behavior, 70:157-162.

22-Marks N, Kavanaugh AF, Lipsky PE (2013): Inhibition of the transendothelial migration of human T lymphocytes by prostaglandin E2. J. Immunol., 152: 5703-5713.

23-Braun M, Pietsch P, Zepp A, Schror K, Baumann G, Felix SB (2010): Regulation of tumor necrosis factor alpha- and IL-1-beta-induced adhesion molecule expression in human vascular smooth muscle cells by cAMP. Arterioscler. Thromb. Vasc. Biol., 17: 2568-2575.

24-Elenkov IJ, Wilder RL, Chrousos GP, Vizi ES (2000): The sympathetic nerve-an integrative interface between two supersystems: the brain and the immune system. Pharmacol. Rev., 52: 595-638.

25-Singhal PC, Sankaran RT, Nahar N, Shah N, Patel P (2007): Vocative agents modulate migration of monocytes across glomerular endothelial cells. J. Investig. Med., 48: 110-117.

26-Wuyts WA, Vanaudenaerde BM, Dupont LJ, Demedts MG, Verleden GM (2003):

Modulation by cAMP of IL-1beta-induced eotaxin and MCP-1 expression and release in human airway smooth muscle cells. Eur. Respir. J., 22: 220-226.

27-Chrousos G (2013): The hypothalamicpituitary-adrenal axis and immune-mediated inflammation. N. Engl. J. Med., 332: 1351-1362.

28-Animesh S, Paul A, Jean W, Sue G (2013): Gender determines ACTH recovery from hypercortisolemia in healthy older humans Metabolism. Theriogenology, 62: 1819-1829.

29- Simoncini T, **Maffei S, Basta G (2009):** Estrogens and glucocorticoids inhibit endothelial vascular cell adhesion molecule-1 expression by different transcriptional mechanisms. Circ. Res., 87: 19-25.

30-Arana N, Valencia DL, Thompson JR, Mitcham N (2013): Changes in plasma melanocyte –stimulating hormone, ACTH, prolactin, GH, LH, FSH and thyroid- stimulating hormone in response to injection of Sulpiride, thyrotropin- releasing hormone, or vehicle in insulin –sensitive and insensitive. Domestic Animal Endocrinology, 44: 204-212.

31-Kamthong PJ, Wu MC (2010): Inhibitor of nuclear- B induction by cAMP antagonizes interlukin-1-induced human macrophagecolony-stimulating-factor expression. Biochem. J., 356: 525-530.

32- Julia A, Golier K, Caramanica I, Leo Sher R (2014): Cortisol response to cosyntropin administration original research military veterans with or without posttraumatic stress disorder article. Psychoneuroendocrinology, 40: 151-158.

33-Stefan S, Federenko FC, Van Rossum W, Koper D (2005): Habituation of cortisol response to repeated psychosocial stress-further and genetic factors. Psychoneuro endocrinology, 30:199-211.

34-Sissel R, Aastveit A, Peter A, Tortesen C (2005): Effect of stress on growth, cortisol and glucose levels in non- domesticated Eurasian perch and domesticated rain bow trout. Comparative Biochemistry physiology, 141:353-358.

35-Johan P, Schoeman M, Herrtage E (2008): Adrenal response to the low dose ACTH stimulation test and the cortisol–to adrenocorticotrophic hormone ratio in canine. Veterinary Parasitology, 154: 205-213.

36-Liisa K, Järvinen NR, Katri R, Aarno H, Herman A (2010): Relationships between the pituitary adrenal hormones, insulin and glucose in middle-aged men: Moderating influence of stress. Original Research, 37: 440-449.

37- Wang JF, Dyer RA, Wu CY K (2010): Prostaglandin E2 and dexamethasone inhibit IL-12 receptor expression and IL-12 responsiveness. J. Immunol., 161: 2723–2730.

38-Radahmadia M, Shadan F, Seied M, Nasmimi A (2006): Effect of stress on exacerbation of diabetes mellitus, serum glucose and cortisol level and body weight in rats. Pathophysiology, 311: 51-55.

39-Tanck MWT, Clases T, Bobenhuis H, Komen J (2002): Exploring the genetic background of stress using isogentic progenies of common carp selected for high or low stress–related cortisol response. Aquaculture, 204: 419-434.

40-Panzani DI, Zicchino A, Taras P (2011): Sulpiride use of dopamine antagonist sulpiride to advance first ovulation in transitional mares .Theriogenology, 75: 138-143.