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Potential protective Effects of okra aquatasextract against the complications of diabetes mellitus in rats

Amira A Sheteewy¹, Neveen A Elwardany¹, Ali A Abdel-Nabey.²

Home Economics Department, Faculty of Specific Education, Alexandria University¹ Food Sciences and Technology Department, Faculty of Agriculture, Alexandria University²

Abstract

The current study aimed to use different concentrations of okra aquatas extract(OAE) (15,30and 45g/100ml), in addition to whole and residues mashed okrain feeding induced diabetes rat groups for six weeks. Such groups were biologically and histopathologically compared with another negative and positive group. The proximate chemical composition as well as fiber fractions (crude, neutral detergent and acid detergent) and some minerals (Ca,Mg and Fe) of okra pods were also studied. Organoleptic evaluation of OAE was also assessed by diabetic patients. The results indicated that significant differences ($P \le 0.05$) were noted between body and organs weight especially heart and spleen of rats fed on OAE diets in comparison with the other groups. In addition, OAE enhanced insulin secretion and lowered glucose level. Also, OAE lowered TG, LDL and VLDL while the HDL, liver and kidney functions improved. Pancreatic histopathological examination showed that groups fed on OAE (30,45g/100ml) seemed to be close to the negative control group with no histopathological changes in relative to the other diabetic rat groups. The results also showed that diabetic patients preferredOAE (30g/100ml). In general addition of OAE in diets could be managed of diabetes through improve liver and kidney functions as welledpancreashistopathological.

Key words: okra aquatas extract, diabetes mellitus, organs weight, serum lipid profile, histopathological examination.

Introduction

Okra (Abelmoschusesculentus, L. Moench.) is known as lady's finger in some countries and bamia in Egypt. It is a plant of the *Malvaceae* family originated in the subtropical and tropical regions of the world (Adetuyi et al., 2011). Recently, it is spread worldwide, while its planting and consumption are more common around the Mediterranean basin, and especially Egypt, Cyprus, Greece, and Turkey. It is the main ingredient in many local and traditional dishes (Calisiret al., 2005 and Kumar et al., 2013).In addition, its upplies common nutrients like minerals, vitamins, dietary fibers and bioactive chemicals. Sabithaet al. (2011) reported that the peel and seed of okra have antidiabetic and antihyperlipidemic affects instreptozotocin-induced diabetic rats. Deters et al., (2005) reported that okra can reduce blood glucose level and lipid level in obese rats, also it may play an important role in the regulation of glucose and lipid metabolisms. Okra is rich in bioactive polysaccharides, which have many biological activities (Panagiotis 2008 and Wittschieret al., 2007). Apart from edible use, extracts from okra fruit have been used for many applications in the food and pharmaceutical industry as emulsifiers, drug tablet formulations or blood plasma replacement, due to their highly content of biopolymers, such as polysaccharides and bioactive compounds such as ascorbic acid and B-carotene (Adetuviet al., 2011, Arlaiet al., 2012 and Ghoriet al., 2014). Fresh okra pods are the most important vegetable source of viscous fiber to lower cholesterol. Seven-days-old fresh okra pods have the highest concentration of nutrients (Gemedeet al., 2014). The different in polysaccharides found in the mucilage are high in okra pods according to Hirose et al. (2004). Okra is reported to have hypolipidemic effect by lowering the absorption of cholesterol from the diet (Huynh et al., 2008). Type 2 diabetes mellitus (DM) is the most common disease of the endocrine system that arises as a result of impaired insulin sensitivity and decreased cellular uptake of glucose (Vetrichelvanet al., 2001). properties of DM is hyperglycemia and dysregulation of carbohydrate, fat and protein metabolism due to destruction or inactivation of pancreatic B- cells, and defective insulin secretion, leading to morbidity which affect more than 100 million people worldwide (Scherbaum 2002), and is gradually emerging as an important health problem in developing countries (Bahmaniet al., 2014). Epidemiological data indicated that 2.8% of the world's population was diabetic in the year 2000 and it may progress to 4.4% of the world's population by 2030. It affects all age groups of

people and ethnic groups (Xing et al., 2009). There is a need to take urgent actions to counter the increase in diabetes through awareness, better detection, prevention continuously, knowledge of the prevalence of diabetes and prediabetes and risk factors could raise awareness of the disease and lead to new policies and strategies for prevention and management. Long-standing diabetes can lead to circulation, kidney and heart problems, including stroke. In traditional world, nutrition and health care have a connection for which many plants are consumed as food in order to benefit health (Pieroni and Price, 2005). The nutraueutical value and the antioxidant activity of wild, semi-cultivated or neglected vegetables are regarded worldwide as an important area of the nutritional and phytotherapic research. Motivation of people towards herbal medicines is increasing to avoid side effects of drugs prepared from synthetic materials (El and Karakaya, 2004). Therefore, the current study was undertaken to investigate the role of fresh OAE in managing the biological parameters of normal and induced diabetic rats as well as the sensory attributes of these extracts by patients suffering from diabetes mellitus.

Materials and Methods

Materials

Chemicals

Alloxan monohydrate was purchased from Sigma Chemicals Company, st Louis US, and all the othermaterials were purchased from EL-Goumhorya Company for trades Drugs Chemicals, and Medical Instruments.

Okra selection

Okra (*Abelmoschusesculentus*) pods were collected from a local market at Alexandria, Egypt. The fruits were selected for uniformity of size and colour, and they were free from visible wounds and rottenness.

Methods

Analytical methods

Preparation of okra pods

The fresh okra was washed with tap water, and cut into small pieces, homogenized and used for analysis. Part of okra pieces was soaked in three concentrations (15, 30 and 45 g fresh pieces of okra /100 ml of potable water). They were soaked overnight. Okra pieces were removed and the liquid was kept in glass bottles and used for sensory evaluation andfeeding rats.

Proximate chemical composition

Crude protein, crude fat and total ash were determined as described in **AOAC** (2000) procedures unless otherwise stated. Total carbohydrates were calculated by difference.Crude fiber, neutral detergent fiber (NDF) and acid detergent fiber (ADF)contents were determined according to the **AOAC** (2000) procedure via filter bags technology (Fiber analyzer, Ankon zoo) USA model no: Azzo. Minerals (Ca, Mg and Fe) were determined in ash solution using Atomic Absorption Spectrometer (AAS) 300Va-50-60H2-100-240V, UK as described by the **AOAC** (2000) procedure.

Experimental design

Forty-eight healthy adult male western rats weighing 190-200g were obtained from the Animal House, Home Economics Department, Faculty of Agriculture, Alexandria University. Animals were housed under laboratory conditions of temperature at 23 ± 3 °C with a 12 h light–dark cycle and were given standard diet and access to distilled water *ad libitum* according to the American Institute of Nutrition **AIN**, (1980). The protocol conforms to the guidelines of National Institutes of Health (NIH). Rats were acclimatized before the commencement of the experiment for a period of seven days, and then they were divided into eight groups (six animals in each group) as follows:

Group 1: negative control (healthy untreated rats).

- **Group 2:**positive control (diabetic rat) that was treated by administering alloxan monohydrate (150 mg/ kg bodyweight) as described by**Vanitha***et al.* (2013) and were considered diabetic when the fasting blood glucose levels was in the range of 150–200 mg/dL for 5 consecutive days.
- Groups 3, 4 and 5:included diabetic rats which were orally given OAEat dose of 2.25, 4.5 and 6.75g / kg bw/day, respectively using an orogastric tube.
- **Group 6:** diabetic rats which were orally given 10 g /kg bw /day of residues mashed okra after extraction using an orogastric tube.
- **Group 7**: diabetic rats which were given suspended whole okra in distilled water with the dosage of 10g /kg bw /day using an orogastric tube.
- **Group 8:** diabetic ratstreated with 5 mg/kg bw /day glibenclamide as standard drug.

After the end of the treatment period (42 days), the rats were weighed, fasted overnight for 10 h and then sacrificed under ether anesthesia.

Blood samples were collected for biochemical analyses and organs such as liver, kidney, brain, lung, heart, pancreas, testes and spleen were removed; washed with cold saline solution and weighed. After that pancreas samples were stored in 10% formalin for histopathological examination.

Biochemical analysis

Serum samples were separated from blood by centrifugation at 3000 rpm for 15 min and analyzed for the following biochemical parameters: glucose levels, and insulin level were determined according to the methods of Trinder (1969) and Temple et al. (1992), respectively, serum lipid profile such as triglycerides, total cholesterol, high density lipoprotein (HDL) and low-density lipoprotein (LDL) were analyzed enzymatically according to Banchereauet al., (2000), Grundy et al., (1993) and Expert Panel on Detection, (2001), respectively. Serum urea, uric acid andcreatinine were determined as kidney functions byLumeij and Remple (1991), Young(1995), andFossatiet al. (1980), respectively. The activities of the aspartate aminotransferase (AST), alanine aminotransferase (ALT) and alkaline phosphatase (ALP) were estimated according to International Federation of Clinical Chemistry as liver functions (Tietzet al., 1983 and Banchereauet al., 2000). **Organoleptic assessment of OAE**

The three liquids remained of

The three liquids remained after soaking okra overnight (15, 30,45g, / 100 ml water) were subjected to sensory evaluation. Diabetic patients (20 members) were selected from clients with health insurance in Kafr El-Dawar city, Behera Governorate, Egypt. These patients were asked to assess the odour, texture, appearance, taste, colour and overall acceptability according to **Wichchukit and O'Mahony (2015)**. This experiment was approved by the Ethics committee on human research, Alexandria University, Alexandria, Egypt.

Histophathological examination of pancreas

After fixation in 10% formalin, tissues were washed, and dehydrated in ascending concentration of alcohol. The dried organs were cut into thin slices (4 μ m) then stained with hematoxylin and eosin according to **Bancroft (2002)**. Tissue sections were examined and photographed using microscope (H&E stains, X400mag). **.Statistical Analysis**

The results were expressed as mean values and standard deviations (SD), and analyzed using one-way analysis of variance (ANOVA)

followed by Duncan's multiple range Test with $p \le 0.05$ (Kirkpatrick and Feeney2012).

Results and Discussion

Chemical composition, minerals and fiber content of fresh okra

The results in Table (1) showed that crude protein content of fresh okra was 39.60%. This value is in a good agreement with the value reported by Gopalanet al. (2007), varmudy (2011), Benchacri (2012) and Falusiet al. (2012). As it can be noted from Table (1) the crude fat of fresh okra was very low being 2.04%. This value is not in agreement with the value reported by Adetuyiet al. (2011). The percentage of total ash was 13.45%, on the other handcarbohydrate content of fresh okra was 35.05%. The results in Table (1) also showed that fresh okra contained 53.17% neutral detergent fiber and 20.81% acid detergent fiber. Also, the results showed that okra contained 925,447 and 1.97mg/100g for Ca, Mg and Fe, respectively. In accordance with the results obtained in the present study, Adetuviet al. (2011), Dossantoset al. (2013), Kumar et al. (2013), Gemedeet al. (2016) and Petropoulos et al. (2018) found that okra fruits contained from 58.22-382 mg/100g Ca, from 1.08 to 101mg/100g Mg and from 0.29 to 36.68mg/100g Fe. These wide variations are mainly due to variety, origin where the okra was grown.

Constituent	g/ 100g
Crude protein	39.60±0.19
Crudefat	2.04±0.02
Total ash	13.45±0.55
Crude fiber	9.88±0.13
Carbohydrate	35.03±0.34
Neutral detergent fiber	53.17±1.02
Acid detergent fiber	20.81±0.65
Mineral (mg /100g)	
Ca	925±11.0
Mg	447±7.0
Fe	1.97±0.30

 Table (1):Chemical composition,fiber contents of freshokra(on dry weight basis)

Effect of okra and OAE on body weight and organ weightin alloxan - induced diabetic rats

Statistically, no significant difference was observed in the mean initial body weights between all experimental groups (1-8) as shown in Table (2).Diabetic rats group(positive control) was significantly decreased in the final values of body weightby 26.77% compared to the negative control group. This may be due to injurious effects of alloxan which caused alkylation of DNA and produced hyperglycaemia and necrotic lesions. Oral administration of OAE at dose of 2.25, 4.5 and 6.75g / Kg bw/day (groups3, 4 and 5) caused a significant increased in body weight compared to the positive control, by 25.32.%, 31.34%, and 34.41%, respectively as shown in Table (2). Groups of rats treated with residues mashed okra(6) had no significant decrease in the body weights compared to the positive control group. Also, no significant difference was observed in the final values of the body weights between diabetic rats treated with 5 mg/kg bw /day glibenclamide compared to the negative control group. These results are in agreement with Ahangarpouret al.(2017). This may be due to decrease or insufficient insulin which causes lipolysis and proteolysis that result in weight loss. Treatment male rats with OAE improved the level of insulin and explain the returning of weight gain to nearly the control values (Frier and Fisher, 2006). The results indicated that there were no significant changes in weight of liver, kidney, testes, brain, pancreas and spleen in all groups. However, OAE or okra administration did not cause statistical differences in organ weight of rat except a significant decrease was observed in lungs, when compared to the negative control group (Table2).

m anoxan – muuccu maberie rats										
	Wei	ght	Liver	Kidnev	Testes	Brain	Heart	Lungs	Pancreas	Spleen
Groups	Initial	Final	(g)	(g) [•]	(g)	(g)	(g)	(g)	(g)	(g)
-	(g)	(g)	Ò	Ó	\ ð /	Ó	Ó	Ó	\ 0 /	Ó
(1) Negative	195.3	249.2	6.94	1.39	2.31	1.56	.67	1.79	.39	.83
control (ve ⁻)	$\pm 4.4^{a}$	±4.3 ^e	±.346 ^b	$\pm.245^{a}$	$\pm .373^{a}$	$\pm .204^{ab}$	$\pm.072^{a}$	±.164 ^{bc}	±.073 ^b	$\pm .305^{a}$
(2) Positive	194.5	182.5	4.65	1.16	1.65	1.35	.49	.93	.27	.87
control (ve ⁺)	$\pm 3.7^{a}$	$\pm 5.4^{a}$	$\pm 1.14^{ab}$	±.563 ^b	$\pm .542^{a}$	±.706 ^{ab}	$\pm .231^{e}$	±.373 ^{ab}	±.043 ^{ab}	$\pm.653^{a}$
(3) $ve^+ +$	106.2	<u></u>	6.24	1 47	1.92	1 66	40	1 45	20	80
OAE 2.25g/kg	190.2	220.1	0.24	1.47	$1.03 + 716^{a}$	1.00	.40	1.43 ± 0.45^{ab}	.20	.09 + 200 ^a
bw /day	±3.1	±2.0	±1.00	±.344	±.710	±.410	±.130	±.043	±.070	±.299
$(4) ve^+ + OAE$	103 1	220 7	5 90	1 13	2.02	1 21	62	1.61	38	78
4.5g/kg bw	$+1.0^{a}$	$\pm 1.59.7$	$+1.01^{a}$	+ 1.13	$\pm 760^{a}$	$+ 302^{a}$	$+ 1/8^{de}$	$+ 106^{abc}$	$+ 170^{b}$	$+ 185^{a}$
/day	14.9	14.5	-1.91	1.442	±.700	1.302	140	±. 4 00	1.179	±.105
$(5) ve^+ +$	194 7	245 3	5 38	1 47	2 23	1 76	86	1 41	35	79
OAE 6.75g/kg	$+2.7^{a}$	$+4.6^{e}$	$+784^{a}$	$+074^{a}$	$+ 254^{a}$	$+ 321^{a}$	+ 536 ^{bc}	$+ 156^{\circ}$	$+ 083^{ab}$	$+ 171^{a}$
bw /day	-2.7	± 1.0	1.701	±.071	±.25 T	1.521	2.550	1.150	1.005	<u></u> /1
$(6)ve^+ +$										
residues	195.7	181.5	4.59	1.22	1.70	1.17	.45	1.17	.25	.66
mashed okra	±	± .	±	±	±	± .	±	±	±	±
10 g /kg bw /	5.2ª	4.9 ^a	.595 ^{ab}	.323"	.437ª	.159"	.067°	.176 ^{ab}	.071ª	.242ª
day										
(7) ve ⁺ +										
whole okra	193.2	200.8	5.13	1.08	1.65	1.61	.31	1.07	.23	.54
pods 10 g /kg	$\pm 6.2^{a}$	±5.3°	$\pm 1.70^{a}$	±.236 ^{ab}	$\pm .577^{a}$	$\pm .320^{ab}$	$\pm .191^{ab}$	$\pm.426^{a}$	$\pm .016^{a}$	$\pm .154^{a}$
bw / day.										
$(8)ve^+ + drug$	196.5	246.8	6.34	1.35	1.74	1.51	.56	1.41	.38	.96
(o) it i ulug	$\pm 3.5^{a}$	$\pm 4.3^{e}$	$\pm 1.98^{a}$	±.206 ^a	$\pm .522^{a}$	$\pm .266^{ab}$	$\pm .170^{a}$	±.054 ^{ab}	±.143 ^b	$\pm .350^{a}$

 Table (2): Effect of okra and OAE on body weight and organ weight in alloxan – induced diabetic rats

Values represent mean \pm SD, Means with different superscript letters on the same column were significantly different $p \le 0.05$

Comparison of serum glucose and insulin levelsbetween groups.

Table (3) shows the results of fasting blood glucoseand insulin level of the rats at the end of the experiment. Diabetic rats showed a significant increment in glucose level the serum (221mg/dl) as compared with the control group (106.6 mg/dl). The present data revealed that administration of OAEcaused significant decrement in glucose level of the male rats comparing with the positive control group. The level of glucose was 193.66, 171.17and 150 mg/dl for group 3, 4, and 5, respectively. However, the administration of whole okra or residues mashed okra to the diabetic ratsdid not cause any significant differences

in the level of glucose (Table 3). Highly significant decrease in glucose level was detected in group 8 when compared with group2. The lowering of glucose levels by OAE could be attributed to the high content of insulin which contributes to the control of hyperglycemia in diabetes. Additionally, it could promote glucose uptake in hepatocytes and increase the activities of hepatic hexokinase and glucose-6-phosphate dehydrogenase in the liver of STZ-induced diabetic rats(Nabila, et al., 2018). The present results showed that diabetic rat groups had significant decrement in insulin levels compared to the control group.Treatment with OAE significantly increased insulin level compared to the positive control group (Table 3). Also it can be noted that administration of residues mashed okra showed the lowest insulin value (23.67 U / ml)). On the other hand, treatment with glibenclamideinsignificantly decreased insulin level compared to the control group. The results of insulin is similar to that reported by Fan et al.(2014) who found that insulin levels were slightly increased in highfat diet-induced obese mouse, while the treatment with okra extract (equivalent to 30-g fresh okra pod/kg/ day) reduced insulin levels. Okra polysaccharides possess anticomplementary and hypoglycemic activity in normal mice (Tomodaet al., 1989). Seeds and mucilage of H.esculentus (200 mg okra extract /kg/day,for 19day)exerted positive effects such as increasing serum insulin, improving total antioxidant capacity, reducing MDA, and improving lipid profile by decreasing total cholesterol, LDL-C and triglyceridein pregnant rats (Tianet al., 2015).

 Table (3): Serum glucose and insulin levels in alloxan-induced diabetic rats

Experimental	Serum glucose	Serum Insulin
Groups	(mg / dL)	(µIU/mL)
(1) Negative control (ve ⁻)	106.6 ± 0.87^{a}	46.67 ± 2.9^{d}
(2) Positive control (ve ⁺)	221 ± 1.04^{cd}	27.01 ± 2.1^{a}
(3) ve ⁺ + OAE 2.25g/kg bw /day	193.66 ± 4.3^{bcd}	34.16 ± 4.2^{b}
(4) $ve^+ + OAE$ 4.5g/kg bw /day	171.17 ± 5.6^{bc}	36.33 ± 2.2^{bc}
(5) ve ⁺ + OAE 6.75g/kg bw /day	150 ± 3.7^{ab}	38.17±2.6 ^c
(6)ve ⁺ + residues mashed okra 10 g /kg bw / day	235.16 ± 3.18^{d}	23.67 ± 3.3^{a}
(7) ve^+ + whole okra pods 10 g /kg bw / day.	225.8 ± 1.16^{cd}	25.33±1.9 ^a
$(8)ve^+ + drug$	108.33±.31 ^a	46.5 ± 2.4^{d}

Values represent mean \pm SD, Means with different superscript letters on the same column were significantlydifferent $p \le 0.05$

Effect of OAE on serumlipid profile of diabetic rats

The present results showed that the positive group had a significant increase in triglycerides (TG) by 67.44 % and total cholesterol (TC) by 20.76% compared to the negative group as recorded in Table (4). Diabetic groups treated withOAE at three dosage levels showedimprovement in lipid profile levels. Administration of OAEcaused asignificant reduction inTC, and TG compared to the positive groupand significantly increasedHDLcompared to the negative group. The results revealed that the positive group had a significant increment in serum level of VLDL when compared to the negative group. Administration of OAE at 6.75g /kg bw/day produced significant decrement in serum levels of VLDL by 22.07%, while the dose of 2.25 and 4.5g /kg bw/dayshowed no significant change when compared to the positive group as shown in Table (4). The decline in VLDL levels in treated group could be directly correlated to decline in TG levels of these groups, as it is well established that VLDL particles are the main transporters of TG in plasma (Hertoget al., 1993). These results are in agreement with Tianet al. (2015) who reported that the okra extract had effect on serum lipid profile. Also, Alqasoumi (2012) found that administrationethanolic extract of okra (250 and 500 mg/kg bw) decreased TC, LDL and TG levels in rats. Moreover, Hajianet al. (2016) reported that the supplementation with either H. esculentus seed at a daily dose of 2 g/kg bw for 2 weeks or mucilage at a daily dose of 2 g/kg bw for 2 weeks caused a significant reduction in TC, LDL-C, HDL-C and TG compared to the control group.

Exportmontol	Lipid profile [*]						
Crowns	(TG) (TC)		HDL	LDL	VLDL		
Groups	(mg/dl)	(mg/dl)	(mg/dl)	(mg/dl)	(mg/dl)		
(1) Negative control (ve ⁻)	43.3 ± 4.4^{a}	70.8±2.5 ^b	33.3 ± 2.6^{a}	28.8±2.7 ^b	$8.7 \pm .88^{a}$		
(2) Positive control (ve ⁺)	72.5 ± 3.1^{d}	85.5 ± 4.1^{d}	$44.5 \pm 3.6^{\circ}$	26.5 ± 7.5^{b}	$14.5 \pm 1.7^{\text{ d}}$		
(3) $ve^+ + OAE 2.25g/kg bw / day$	48.3 ± 2.6^{b}	$77.2 \pm 3.9^{\circ}$	$42.5 \pm 2.1^{\circ}$	25.0±5.9 ^b	13.9 ± 1.4^{cd}		
(4) $ve^+ + OAE 4.5g/kg bw/day$	47.4 ± 2.1^{b}	63.33±3.4 ^a	38.7 ± 1.1^{b}	15.2 ± 4.1^{a}	12.0 ± 3.3^{cd}		
(5) ve ⁺ + OAE 6.75g/kg bw /day	47.1±1.1 ^b	60.7 ± 2.9^{a}	36.2±2.3 ^{ab}	15.1 ± 2.1^{a}	11.3 ± 1.9^{abc}		
(6)ve ⁺ + residues mashed okra 10 g /kg bw / day	75.6 ± 2.7^{e}	94.7±3.7 ^e	44.3±2.2 ^c	24.2±3.7 ^b	11.7±3.4 ^{bcd}		
(7) ve^+ + whole okra pods 10 g /kg bw / day.	64.3±1.6 [°]	90.8±5.1 ^e	43.7±2.6 ^c	24.4 ± 2.4^{b}	11.7 ± 2.8^{bcd}		
$(8)ve^+ + drug$	45.6±.92 ^{ab}	72.7 ± 3.1^{b}	34.3±2.2 ^a	28.1±2.1 ^b	8.9±.71 ^{ab}		

Table (4): Effect of OAE on serum lipid profile of diabetic rats

Values represent mean \pm SD, Means with different superscript letters on the same column were significantly different $p \le 0.05$

Effect of OAE on Liver functions of diabetic rats

The present results showed that the positive group had significant increment in serum levels of ALP, AST and ALT enzymes in serum when compared to the negative group by 63.38, 59.5and 92.0 %, respectively. Rats given OAE at three dosage levels showed remarkably amelioration in enzymes level and the reduction in AST, ALP and ALT and the increasing dosages of okra water extract produced increase of reduction in enzymes level when compared with those of diabetic rat. Treatment with OAE 6.75g /kg bw/day to rats caused significant decrement in the levels of ALP, AST and ALT enzymes by 35.81,36.69 and23.15%, respectively, when compared to the positive control group. This may be due to the OAE stabilize the cell membrane and prevent the leakage of AST and ALT to the blood stream. In this content**Alqasoumi** (2012) found that ethanolic extract of okra administration(250 and 500 mg/kg bw)affect serum liver functions. On the other hand, glibenclamide treatment showed a mild improvement in the liver function.

Effect of OAE on Kidney functions of diabetic rats

The results of kidney functions are shown in Table (5). Groups ofdiabetic rats treated withokra (3 to 5)had no significantincrement in the level of uric acid, urea and createninewhen compared with the positive control, whereas significant differences were observed when group 2 was compared with group 1 for the same parameters. Treatment with alloxan monohydrate (150 mg/ kg bw) caused significant increment in the levels of uric acid, urea and createnine by 56.91,36 and 26.2 %, respectively when compared to the negative group. These results are in agreement with the results obtained by **Hajianet al.(2016)** who found that treating rats with okra extract led to insignificant reduction in uric acid, urea and createninecompared to the positive group.

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Table (5):Effect of OAE on Liver and Kidney functions of diabetic rats

Exporimontal	Liv	ver functio	ons	Kidney functions			
Croups	ALP	AST	ALT	Uric acid	Urea	Createnine	
Groups	(U/L)	(U/ml)	(U/ml)	(mg/dl)	(mg/dl)	(mg/dl)	
(1) Negative control (ve ⁻)	82.2 ± 2.5^{a}	36.8±2.3 ^a	22.5±1.9 ^a	34.0 ± 3.6^{a}	0.77 ± 0.18^{a}	$0.67{\pm}0.78^{a}$	
(2) Positive control (ve ⁺)	134.3±3.7 ^d	60.5 ± 3.2^{d}	43.2±3.1 ^f	57.5±3.9 ^e	1.1±0.39 ^b	0.89 ± 0.13^{bc}	
(3) ve ⁺ + OAE 2.25g/kg bw /day	$93.7 \pm 3.7^{\circ}$	41.7±3.1 ^{bc}	35.7 ± 2.2^{cd}	46.7±3.5 ^{cd}	1.2 ± 0.17^{b}	0.83±0.15 ^{abc}	
$\overline{(4) ve^+ + OAE \ 4.5g/kg bw} / day$	$90.2 \pm 2.6^{\circ}$	38.3±3.0 ^{ab}	33.2±2.6 ^c	42.2 ± 3.7^{bc}	1.2± 0.07 ^b	0.73±0.20 ^{abc}	
(5) ve ⁺ + OAE 6.75g/kg bw /day	86.2±1.7 ^b	37.0±2.4 ^a	28.4±2.9 ^b	40.2±5.5 ^b	1.1± 0.13 ^b	0.85±0.11 ^{abc}	
(6)ve ⁺ + residues mashed okra 10 g /kg bw / day	136.3±3.8 ^d	45.2±3.1°	40.5±1.9 ^{ef}	56.3±3.1 ^e	1.2 ± 0.11^{b}	0.79±0.10 ^{abc}	
(7) ve ⁺ + whole okra pods 10 g /kg bw / day	103.5±3.7 ^d	43.2±3.2 ^c	37.8±1.7 ^{de}	49.5±2.9 ^d	0.85±0.13 ^a	0.91±0.13 ^c	
$(8)ve^+ + drug$	85.7±2.2 ^{ab}	38.3 ± 3.5^{ab}	25.2±3.5 ^a	37.5 ± 6.3^{ab}	0.97 ± 0.13^{ab}	0.71 ± 0.16^{ab}	

Values represent mean \pm SD, Means with different superscript letters on the same column were significantly different $p \le 0.05$

Organoleptic assessment of OAE

Table (6) shows sensory evaluation of OAE. It can be noted that there were significant differences between the three OAE from the organoleptic point of view. The scores of the organoleptic attributes given for the three extracts decreased with increasing the amount of okra dissolved in water. No poor or rejected organoleptic attributes were obtained even at the higher amount of okra (30g/100ml water). As a conclusion, the diabetic patients accepted the three samples, but sample (15g/100ml) was well accepted compared with the other samples (30 and 45g/100ml). It has been reported that motivation of people towards herbal medicines is increasing to avoid the side effects of drug prepared from synthetic materials (**El andKarakaya, 2004**).

Table (6):Organoleptic properties of OAE

	Organoleptic attributes									
Sample	Appearance	Taste	Texture	Colour	Odour	Overall acceptability				
15g/100 ml	6.95 ± 1.09^{b}	7.10 ± 1.20^{b}	$6.60{\pm}1.42^{ab}$	7.15±.933 ^a	7.85 ± 1.03^{a}	7.15 ± 1.22^{a}				
30g/100ml	6.90 ± 1.4^{b}	$7.45 \pm .944^{b}$	7.10 ± 1.16^{b}	7.20 ± 1.54^{a}	7.25 ± 1.55^{a}	7.65±1.13 ^a				
45g/100ml	5.25 ± 1.48^{a}	5.95±1.53 ^a	6.05 ± 1.3^{a}	6.85 ± 1.34^{a}	6.90 ± 1.63^{a}	6.80 ± 1.64^{a}				

Values represent mean \pm SD,Means with different superscript letters on the same column were significantly different $p \le 0.05$

Histopathologicalexamination of diabetic rats pancreas treated with OAE

Histopathological examination of pancreas sections in the negative control rats showed normal arrangement of the largely islet cells with normal prominent ß-cells which have cytoplasm and rounded dark nuclei and no histopathological changes in the area of eosinophilicacini cells with hyperchromatic nuclei (Fig1, A). While, pancreas of alloxaninduced diabetic rats revealed marked dilation of interlobular and enzyme duct surrounded by infiltrating lymphocyte cells and more necrotic cells were apparent (Fig1, B). The pancreas of the treated rats with OAE at dose 2.25 g/kg bw/day revealed that the islet cells appeared embedded within the acinar cells and surrounded by a fine capsule, normal proportions with eosinophilic deposits in the gland, the islets smaller in volume, more uniform eosinophilic cytoplasm, and round nuclei (Fig1, C). Rat pancreas which was diabetic and treated with OAE at dose 4.5 g/kg bw/day showed the acinar cells which stained strongly and are arranged in lobules with prominent nuclei in their normal proportions. The large vacuolated islet cells with reduction of the β - cells are seen embedded within this acinar cell. There are area necrotic acinar cells (Fig1, D).section of the rat pancreas which was diabetic and treated withOAE at dose 6.75 g/kg bw/day showed the acinar cells which stained pale eosinophilicand are arranged in lobules with pale nuclei (necrotic acinar cells). The enlarged or widely islet cells are seen embedded within the necrotic acinar cells (Fig1, E). Fig (1, F) shows the rat pancreas which was diabetic and fed on the okra residues after water extraction. The cells showed atrophied islet with mild dilation of the capsule space, eosinophilicacini cells with hyperchromatic nuclei and necrotic other cells surrounded the islet and mild dilation of the interlobular duct. Fig (1, G) showed that the rat pancreas which was diabetic and fed whole okra, showed the islets smaller in volumes with more atrophied ß-cells are seen embedded within the eosinophilic cytoplasm acinar cells and surrounded by a fine capsule. Fig (1, H) shows the section of the diabetic rat pancreas treated with 5 mg/kg bw /day glibenclamide. An eosinophilic cytoplasm and necrotic nuclei of the acinar cells are arranged in lobules. The islets were atrophied with few atrophied β -cells, embedded within the acinar cells and surrounded by a fine capsule. There is eosinophilic material deposit in the gland with

mild dilation of blood vessels with no infiltrating lymphocytes. The results are in agreement with Majdet al. (2018)present whodemonstrated that okra may improve glucose homeostasis which are associated with reduced pancreatic tissue damage. The okra administrated showed that the reduction size of islets and population of insulin-producing B-cells were reduced in the pancreas of HFD/STZinduced diabetic rats and these results were in accordance with previous studies that demonstrated that hyperglycemia leads to a progressive decline in β-cells function(Bonora, 2008).



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was ve^+ + whole okra pods 10 g /kg

(E): Rat's pancreas in the group which was ve⁺ + residues mashed okra 10 g /kg bw / day



bw / day

(G): Rat's pancreas in the group which was ve^+ +okra pods 10 g /kg bw / day

(H): Rat's pancreas in the group which was ve^+ + wholedrug

Fig (1):Histopathologicalexamination of diabetic rats pancreas treated with OAE

Conclusion

In general, it could be concluded that OAE at different concentrations showed a beneficial role in the management of many biological factors in diabetic rats. This is because these extracts improved mainly liver and kidney functions. So, we recommended to okra aquatas extracts on our daily dishes.

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- Adetuyi, F.O.,Osagie, A.U. and Adekunle A.T. (2011). Nutrient, antinutrient, mineral and zinc bioavailability of okra *Abelmoschusesculentus*(L) Moench. Am J Food Nutr, 1(2):49-54
- Ahangarpour, A.,Heidari, H.; Oroojan, A.,Mirzavandi, F.,Esfehani,
 K.h. and Mohammadi, D. Z. (2017). Antidiabetic, hypolipidemic and hepatoprotective effects of Arctiumlappa root's hydro-alcoholic extract on nicotinamidestreptozotocin induced type 2 model of diabetes in male mice. Avicenna J Phytomed. 7 (2): 169-179.
- Alqasoumi S.I. (2012). 'Okra' *Hibiscus esculentas* L: a study of its hepatoprotective activity. Saudi Pharm J. 20(2): 135-141.
- American Institute of Nutrition (AIN) (1980). Second report of the ad hoc committee for experimental animals. J. Nutr. 107: 1726.
- Arlai, A.; Nakkong, R.; Samjamin, N. and Sitthipaisarnkun, B. (2012). The effects of heating on physical and chemical constitutes of organic and conventional okra. Procedia Engineering, 32, 38–44.
- Association of Official Analytical Chemists AOAC. (2000). Official methods of analysis of the Association of Official Analytical Chemists: (17th ed.). Washington: AOAC.
- Bahmani, M.,Zargaran, A.,Rafieian-Kopaei, M. and Saki M. (2014). Ethnobotanical study of medicinal plants used in the management of diabetes mellitus in the Urmia, Northwest Iran. Asian Pac J Trop Med. 7:348-54.
- Banchereau, J.,Briere, F.,Caux, C.,Davoust, J.,Lebecque, S., Liu, Y. J. and Palucka, K. (2000). Immunobiology of dendritic cells. Annual Review of Immunology, 18 (1): 767-811.
- **Bancroft JD** (2002). Theory and practice of histological techniques. In: Bancroft JD, Gamble M, editors. 5th ed. 5th ed. Edinburgh: Churchill livingstone;. pp. 125, 130, 175, 206-208.
- **Benchasri, S.(2012)**. Screening for yellow vein mosic virus resistance and yield loss under field conditions in Southern Thailand. J Anim. Plant Sci., 12:1676-1686.
- **Bonora E(2008).** Protection of pancreatic beta-cells: is it feasible? NutrMetabCardiovasc Dis. 18(1):74–83.
- Çalişir, S.,Özcan, M.,Haciseferoğullari, H. and Yildiz, M. U. (2005). A study on some physico-chemical properties of Turkey okra

(*Hibiscus esculenta* L.) seeds. Journal of Food Engineering, 68(1): 73–78.

- **Deters, A. M.,Lengsfeld, C. and Hensel, A. (2005).**Oligo- and polysaccharides exhibit a structure-dependent bioactivity on human keratinocytes in vitro. Journal of Ethnopharmacology, 102(3): 391–399.
- Dos Santos, I. F., Dos Santos, A. M. P., Barbosa, U. A., Lima, J. S., Dos Santos, D. C., and Matos, G. D. (2013). Multivariate analysis of the mineral content of raw and cooked okra (*Abelmoschusesculentus* L.). Microchemical Journal, 110, 439– 443.
- El S.N. and Karakaya, S. (2004). Radical scavenging and ironchelating activities of some greens used as traditional dishes in mediterranean diet. Int. J. Food SciNutr, 55: 67-74
- **Expert Panel on Detection (2001).** Executive summary of the third report of the National Cholesterol Education Program (NCEP) expert panel on detection, evaluation, and treatment of high blood cholesterol in adults (Adult Treatment Panel III). Jama, 285(19): 2486-97.
- Falusi, O.A., Dangana, M.C., Daudu, O.Y. and Jaime, A. (2012). Studies on morphological and yield parameters of three varieties of Nigerian okra (*Abelmoschusesculentus*(L) Moench). J. Hortic. For., 4(7):126-128.
- Fan, S., Zhang, Y., Sun, Q., Yu, L., Li, M., Zheng, B. and Huang, C. (2014). Extract of okra lowers blood glucose and serum lipids in high-fat diet-induced obese C57BL/6 mice. The Journal of nutritional biochemistry, 25(7), 702-709.
- **Fossati, P.,Prencipe, L. and Berti, G. (1980).** Use of 3, 5-dichloro-2hydroxybenzenesulfonic acid/4-aminophenazone chromogenic system in direct enzymic assay of uric acid in serum and urine. Clinical chemistry, 26 (2): 227-231.
- Frier, B.M. and Fisher, M. (2006).Diabetes mellitus In: Davidson's principles and practices of medicine (20th ed. Eds) Boon NA, Colledge NR and Walker B. Churchill Livingston. Edinburgh.
- Gemede, H. F., Ratta, N., Haki, G. D., Woldegiorgis, A. Z. and Beyene, F (2014). Nutritional Quality and Health Benefits of Okra (*Abelmoschusesculentus*): A Review, Food Science and Quality Management, (33), 87-96.

- Gemede, H. F.,Haki, G. D.,Beyene, F., Woldegiorgis, A. Z. and Rakshit, S. K. (2016).Proximate, mineral, and antinutrient compositions of indigenous Okra (*Abelmoschusesculentus*) pod accessions: Implications for mineral bioavailability. Food Science and Nutrition, 4(2), 223–233.
- Ghori, M. U., Alba, K., Smith, A. M., Conway, B. R. and Kontogiorgos, V. (2014). Okra extracts in pharmaceutical and food applications. Food Hydrocolloids, 42(P3), 342–347.
- Gopalan, C.,Sastri, S.B.V. and Balasubramanian, S. (2007). Nutritive value of Indian foods. National Institute of Niutrition (NIN) ICMR, India.
- Grundy, S. M.,Bilheimer, D.,Chait, A., Clark, L. T.,Denke, M., Havel, R. J. and Kreisberg, R. A. (1993). Summary of the second report of the national cholesterol education program (NCEP) expert panel on detection, evaluation, and treatment of high blood cholesterol in adults (adult treatment panel II). Jama,269 (23): 3015-3023.
- Hajian, S., Akbari, F., Shahinfard, N., Mirhoseini, M., Shirzad,
 H., Heidarian, E. and Kopaei, M. R. (2016). Impacts of *Hibiscus esculentus* extract on glucose and lipid profile of diabetic ratsJNephropharmacol. 5(2): 80–85
- Hertog, M.G.L., Feskens, E.J.M., Hollman, P.C.H., Katan, M.B. and Kromhout, D. (1993). Dietary antioxidant avonoids and risk of coronary heartdisease: the zutphen elderly study. Lancet, 342, 1007 1011.
- Hirose, K., Endo, K. and Hasegawa, K. (2004). A convenient synthesis of lepidimoide from okra mucilage and its mucilage and its growth promoting activity in hypocotyls. Carbohydr. Poly., 339:9-19.
- Huynh, T., Nguyen, Q., Tran, A., Van, T. and Phung, N.(2008).Hypolipidemic effect of extracts from *Abelmoschusesculentus* L. (Malvaceae) on tyloxapol-induced hyperlipidemia on mice. Mahodol. Uni J Pharmacol.Sci, 35 (1-4): 42-46.
- Kirkpatrick, L. A. and Feeney, B. C. (2012). A Simple Guide to IBM SPSS Statistics for Versions 20.0: Nelson Education
- Kumar, K., Tony, E., Kumar, P., Kumar A.K., Rao. B .S. and Nadendla. R. (2013). A review on: Abelmoschusesculentus (okra). Int. Res J. Pharm. App Sci. 3(4):129-132.

- Lumeij, J. and Remple, J. (1991): Plasma urea, creatinine and uric acid concentrations in relation to feeding in peregrine falcons (*Falco peregrinus*). Avian Pathology, 20(1): 79-83.
- Majd, N. E., Tabandeh, M.R., Shahriari, A. and Soleimani, Z. (2018). Okra (*Abelmoscusesculentus*) improved islets structure and down-regulated *ppars* gene expression in pancreas of high-fat diet and streptozotocin-induced diabetic rats. Cell J.; 20(1): 31–40.
- Nabila, M., Damayanthi, E. and Marliyati, S.A. (2018). Extracts of okra (*Abelmoschusesculentus L.*) improves dyslipidemia by ameliorating lipid profile while not affectinghs-CRP levels in streptozotocin-induced rats. Earth and Environmental Science 196(4):1-6
- **Panagiotis, A. (2008).** Identification and quantification of polyphenolic compounds from okra seeds and skins. Food Chemistry, 110 (4): 1041–1045.
- Petropoulos, S., Fernandes, Â., Barros, L. and Ferreira, I.C.F.R.(2018). Chemical composition, nutritional value and antioxidant properties of Mediterranean okra genotypes in relation to harvest stage.Food Chem. 242 (1):466-474. doi: 10.1016/j.foodchem.2017.09.082.
- **Pieroni, A. and Price L.L. (2005):** Eating and healing: Traditional food as medicine. Binghamton: Haworth Press.
- Sabitha, V.,Ramachandran, S., Naveen, K. R. and Panneerselvam,
 K. (2011). Antidiabetic and antihyperlipidemic potential of *Abelmoschusesculentus* (L.) Moench. in streptozotocin- induced diabetic rats. Journal of Pharmacy and Bioallied Sciences, 3 (3): 397–402.
- Scherbaum, W.A. (2002). Insulin therapy in Europe. Diabetes Metab Res Rev., 3:50-6.
- Temple, R. C., Clark, P. M. and Hales, C. N. (1992). Measurement of insulin secretion in type 2diabetes: problems and pitfalls. Diabetic Med., 9:503-512.
- Tian, Z.H., Miao, F.T., Zhang, X., Wang, Q.H., Lei, N. and Guo, L, C. (2015). Therapeutic effect of okra extract on gestational diabetes mellitus rats induced by streptozotocin. Asian Pac J Trop Med. 8(12):1038-42

- Tietz, N.W.; Rinker, A.D.; and Shaw, L.M. (1983): IFCC methods for the measurement of catalytic concentration of enzymes Part 5. IFCC method for alkaline phosphatase (orthophosphoricmonoester phosphohydrolase, alkaline optimum, EC 3.1. 3.1). Journal of clinical chemistry and clinical biochemistry. Zeitschrift fur klinischeChemie und klinischeBiochemie, 21(11): 731-748.
- Tomoda, M., Shimizu, N.,Gonda, R.,Kanari, M., Yamada H. and Hikino, H.(1989). Anticomplementary and hypoglycemic activity of okra and hibiscus mucilages. Carbohydr Res., 190:323–8.
- **Trinder, P. (1969).** Determination of glucose in blood using glucose oxidase with an alternative oxygen acceptor. Am. Clin. Biochem., 6:24-27.
- Vanitha, M., Pandian, R.S. and Karthikeyan, J. (2013). Evaluation of Aloe vera Gel for its anti inflammatory activity in diabetes mellitus using animal model system. Int. J. Drug Dev. Res. 5(1): 305-309.
- Varmudy, V. (2011). Marking survey need to boost okra exports. Department of Economics, Vivekananda College, Puttur, Karnataka, India. pp. 21-23.
- Vetrichelvan, T.,Jagadeesan, M. and Uma Devi, B.A. (2001). Antidiabetic activity of alcohol of Celosia argentea Linn Seeds in rats. Bio Pharm Bull. 25:526-528.
- Wichchukit, S. and O'Mahony, M. (2015). The 9-point hedonic scale and hedonic ranking in food science: some reappraisals and alternatives. Journal of the Science of Food and Agriculture, 95(11): 2167-2178.
- Wittschier, N.,Lengsfeld, C.,Vorthems, S.,Stratmann, U., Ernst, J. F.,Verspohl, E. J. andHensel, A. (2007). Large molecules as anti-adhesive compounds against pathogens.Journal of Pharmacy and Pharmacology, 59(6): 777–786.
- Xing, X.H., Zhang, Z.M., Hu, X.Z., Wu, R.Q. and Xu, C. (2009). Antidiabetic effects of *Artemisia sphaerocephala*Krasch. gum, a novel food additive in China, on streptozotocininduced type 2 diabetic rats. J Ethnopharmacol. 125:410-6.
- Young, D. S. (1995). Effects of drugs on clinical laboratory tests (Vol. 4): AACC press Washington, DC.

التأثيرات الوقائية المحتملة للمستخلص المائى للباميه تجاه مضاعفات مرض السكر في الفئران أميرة شتيوي, أنفين الورداني, 2علي عبد النبي اقسم الاقتصاد المنزلي، كلية التربية النوعية، جامعة الإسكندرية

"فسم الاقتصاد المنزلي، كليه النربيه النوعيه، جامعه الإسكندريه 22سم علوم وتكنولوجيا الأغذية، كلية الزراعة ،جامعة الإسكندرية

الملخص:

هدفت الدراسة الحالية إلى استخدام تركيز إت مختلفة من المستخلص المائي للباميه (15، 30، 45 جرام /100مل) بالإضافة الى الباميه الكاملة وتلك الناتجة بعد الاستخلاص المائي وذلك لتغذية الفئران المصابة بالداء السكرى والمغذاة لفترة ستة اسابيع. هذه المجاميع تم مقارنتها من الناحية البيولوجية والهستوباثولوجية مع مجموعتين أحدهما تمثل االمجموعة الضابطة الموجبة والأخرى تمثل المجموعة الضابطة السالبة. هذا وقد تم دراسة التركيب التقريبي للباميه بالإضافه إلى تقدير نسب الألياف الخام وكل من الالياف الذاتية الحامضية والمتعادلة وبعض المعادن مثل الكالسيوم والمغنسيوم والحديد. تم اجراء التقييم الحسى للمستخلصات المائية الثلاث باستخدام أفراد مصابة بداء السكرى. وأوضحت النتائج ان هناك اختلافات معنوية (P ≤ 0.05) بين أوزان الجسم والأعضاء الداخلية خصوصا القلب والطحال بالنسبة للفئران التي تم تغذيتها على المستخلصات المائية للباميه بالمقارنة بالمجاميع الآخري. بالاضافة الى ذلك وجد ان المستخلصات المائية تشجع من افراز الأنسولين ومن ثم تقلل من مستوى الجلوكوز . أيضا أثبتت الدراسة ان المستخلص المائي آدى إلى تحسن صورة دهون الدم وذلك لخفض مستوى الجليسريدات الثلاثية والكوليسترول مخفض الكثافة والكلولسترول المنخفض جدا للكثافة في حين حدث تحسن ملحوظ في نسبة الكوليسترول عالى الكثافة وكل الوظائف الخاصة بالكبد والكلي. أثبت الفحص الهستوباثولوجي للبنكرياس ان المجاميع التي تم تغذيتها على المستخلص المائي للبامية (30، 45جرام/100مل) كانت متشابهه الى حد كبير من المجموعة السالبة مع عدم وجود اى تغيرات هستوباثولوجية بالمقارنة مع مجاميع الفئران الآخرى المصابة بالداء السكرى . أوضحت الدراسة أيضا ان مرض الداء السكرى كان تقبلهم للمستخلص المائي (30جرام/100مل) احسن وأفضل من التركيزات الاخرى. وبصفة عامة يمكن القول بان أضافة المستخلص المائي للباميهإلى الوجبات الغذائية ممكن ان يؤدي الى التحكم في مرض الداء السكري وتحسين وظائف الكبد والكلي.

الكلمات المفتاحية: المستخلص المائي للباميه - مرض الداء السكري- صورة دهون الدم – وزن الأعضاء – الفحص الهستولوجي.