

Journal of Plant Protection and Pathology

Journal homepage: www.jppp.mans.edu.eg
Available online at: www.jppp.journals.ekb.eg

Effect of Intercropping of Nematotoxic Plants on Root- Knot Nematode, *Meloidogyne incognita* Infecting some Vegetable Plants in the Screenhouse



Cross Mark

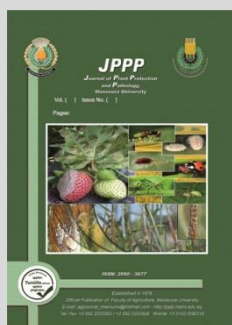
Entsar H. Taha*

Department of Plant Protection, Faculty of Agriculture, Ain Shams University

ABSTRACT

Plant parasitic nematodes (PPNs) especially root-knot nematodes cause significant economic damage and considered as a limiting factor in some vegetables production all over the world. Due to the harmful effect of synthetic nematicides on human health, non-target organisms as well as on environment and agricultural ecosystem, we need to use alternative management practices that are effective, cheap, safe, economically acceptable, and environmentally friendly. Intercropping nematocidal plants may be represented as an alternative efficient practice. Our research was aimed to investigate the suppressive effect of *Crotalaria juncea*, *Tagetes erecta*, and *Brassica juncea* intercropping with tomato (*Solanum lycopersicum* L.), cucumber (*Cucumis sativus* L.), and pepper (*Capsicum annum* L.), on *Meloidogyne incognita* and their effect on the host plant growth. In the screenhouse, the experiment was conducted using 1000 IIs/pot, after 2 months the data were recorded. The highly effective intercropping plant was *Crotalaria* followed by *Tagetes* and *Brassica* in suppressing the nematode galls, egg masses, final population, and rate of reproduction significantly as follow: 153.7, 160.3, 570.0, 0.57 in tomato, and 205.7, 158.3, 692.3, 0.69 in cucumber and 197.0, 189.7, 746.3, 0.75 in pepper, respectively. Moreover, plant growth parameters were enhanced in the three host plants and so, such intercropping system could be of a considerable merit in integrated pest management programs.

Keywords: Root- knot nematodes, Intercropping, *Crotalaria*, *Tagetes*, *Brassica*.



INTRODUCTION

Plant parasitic nematodes (PPNs) are considered one of the most biotic factors which effect negatively on vegetables production. The loss was estimated statistically by US to be \$157 billion worldwide (Singh *et al.*, 2015). One of the most important PPN genus is the root-knot nematode (RKN) (*Meloidogyne*) (Jones *et al.*, 2013), which has more than 3000 host species, affecting vegetables, fruits, and ornamental crops, (Perry *et al.*, 2009; and Khalil, 2013).

Tomato, cucumber, and pepper considered of the most popular versatile vegetables worldwide, because of their diversified use and their high nutritive value such as minerals, vitamins, sugars, essential amino acids, dietary fibers. Moreover, they contain high concentration of anti-carcinogenic and antioxidant substances (Bosland and Votava, 2012). In addition to that the duration crop of these vegetables is relatively short (U.S. Department of Agriculture, 2005).

Because of the pesticide's hazardous effect on human health, as well as non- target organisms and on the environment and their high costs, therefore there is an increased interest for alternative strategies especially the non-chemical nematode management methods (Samaliev, 2009; Wang and Brent, 2009; Nagaraju *et al.*, 2010; and Lamovšek *et al.*, 2013). Each method of plant parasitic nematode management has a limited effect, to achieve a highly efficient management of PPN. So, we need to carefully use a combination of different methods.

Using of intercropping plants, viz, Sunn hemp (*Crotalaria juncea*), Marigold (*Tagetes erecta*), and Mustard

(*Brassica carinata*) offers an alternative to nematicidal chemical (Noling and Becker, 1994; Youssef *et al.*, 2011; Kas, kavalci *et al.*, 2009 and Ploeg, 2008). Intercropping method is an eco-friendly management and has no hazardous effect on plant growth. These plants are a very important source of natural compounds which have nematicidal properties, such as: alkaloids, isothiocyanates, diterpenes, glucosinolates, fatty acids, phenols, thienyls, sesquiterpenes and polyacetylenes (Chitwood, 2002). *Crotalaria*, *Tagetes*, and *Brassica* have been proved to have a reduction effect on nematode population of RKN and can be used as intercropping plants with other plants to reduce the root-knot nematodes population (Bridge, 1996), which may release during decomposition nematicidal plant materials (Halbrendt, 1996).

Varma *et al.*, (1978) reported that, Marigold reduced *Meloidogyne* final population and their root galls and increased the plant growth parameters when intercropping with tomato and with eggplants (Ferraz and Grassi de Freitas, 2004) and potato (Chen *et al.*, 2007). So, the usage of mustard can decrease the nematode populations because of its nematicidal activity (Haider *et al.*, 2004; Curto 2008).

According to the previous literatures, this research was conducted to evaluate the nematicidal effect of *Crotalaria*, *Tagetes*, and *Brassica*. as intercropping plants to control the root-knot nematode, *Meloidogyne incognita* activity on tomato, cucumber, and pepper as host plants and their effect on the host growth properties in the screenhouse.

* Corresponding author.

E-mail address: entsar_elsayed@agr.asu.edu.eg

DOI: 10.21608/jppp.2021.58913.1013

MATERIALS AND METHODS

A- Materials:

1- Root-knot nematode culture:

Culture of *Meloidogyne incognita* (Mi) was maintained in the screenhouse on tomato plants (*Solanum lycopersicum* L. cv. Castel Rock) in sterile sandy loam soil. Nematodes were extracted from tomato galled roots; washing, cutting into pieces, and placed in the mist chamber for egg hatching. Hatched J₂s were collected daily and refrigerated at 6 °C for the experimental use.

2- The host plant:

Screenhouse experiments were carried out using seedlings of 21-days-old of tomato (*S. lycopersicum* L. cv. Castel Rock), cucumber (*Cucumis sativus* L.), and pepper (*Capsicum annum* L.). They were transplanted singly in 20-cm-diam pots which filled with sterilized sandy clay soil.

B- Experimental methods:

Effect of intercropping cultivation of nematotoxic plants on the pathogenicity and reproductivity of *M. incognita* on tomato, cucumber, and pepper seedlings in screenhouse:

Seeds of sunn hemp, *Crotalaria juncea* L., marigold, *Tagetes erecta*, and mustard, *Brassica carinata* were planted directly in 20-cm-diam pots filled with sterilized sandy clay soil under outdoor screenhouse conditions. After 10 days from germination, the plants were thinned to two per pot and 21-days-old of tomato, cucumber, or pepper seedlings were transplanted singly in each pot. Forty pots of tomato, cucumber, and pepper were planted and divided into four groups as follow: first group (10 replicates) was intercropped with *Crotalaria* and 1000 2IJs/pot of Mi were added to 5 replicates of them after one week. Second group (10 replicates) was intercropped with marigold and 1000 2IJs/pot were added to 5 replicates of them. Third group was intercropped with mustard and 1000 2IJs/pot were added to 5 replicates of them. The remaining pots were left as control: five replicates received nematodes and the other five without nematodes. The experiment was ended after 2 months.

Determination of plant growth and nematode parameters:

At the end of the experiment, roots were washed, and fresh weight of roots and shoots were measured. Thereafter, roots were stained with acid fuchsin in cold lacto-phenol (McBeth *et al.*, 1941) and stored in it. Stained roots were rinsed in water and cut into pieces to facilitate counting of galls, egg masses, and final population (the final population were extracted from the soil by Burman funnel technique and

from the roots by mist chamber) were recorded, the rate or reproduction was calculated from the ratio of Pf (final population density) to Pi (initial population density).

3- Statistical analysis:

The data of all experiments were statistically analyzed using analysis of variance to check the significance of the differences between treatments using SAS program (2005) and separation between means was applied by Tukey test.

RESULTS AND DISCUSSION

Results of the present study demonstrated that intercropping with selected nematicidal plants recorded significant reduction in nematode parameters and increased the host plant growth parameters.

Tables (1& 2) proved that, intercropping with *Crotalaria* was the highest suppressive effect on nematode parameters. The mean number of nematode galls, egg masses, nematode final population and the rate of reproduction were decreased significantly in the three host plants comparable with the infected plants without intercropping followed by *Tagetes* and *Brassica* intercropping. The data agreed with those obtained by Kowalska and Sonalinska (2001); Kumar and Khanna (2006); Singh and Sitaramaiah (1993) when used *Brassica* intercropping in the greenhouse to decrease the plant parasitic nematode populations. It is worth noting that there was no significant effect of *Brassica* on galls and egg masses numbers when intercropping with pepper, while the effect was significant on the nematode final population and the rate of reproduction (Table 1&2).

In addition, Anaya (2006) and Akhtar and Malik (2000); Wilson and Caveness (1980) considered *C. juncea* as a non-host to *M. incognita* and has an antagonistic exudate that suppress *Meloidogyne* in pineapple. Moreover, Esparago *et al.*, (1999) reported that *C. juncea* suppressed the Pf of three species of *Meloidogyne* in vegetables

The effect of *Tagetes* and *Brassica* on the nematode parameters were high and these data agreed with those obtained by Ferraz and Grassi de Freitas (2004); Hooks *et al.* (2010); Evenhuis *et al.* (2004) who reported that *Tagetes* spp. are very efficient in nematode control, especially for *Pratylenchus* and *Meloidogyne* species in intercropping technique. And, agreed with Abid and Maqbool (1990) who reported that the number of root galls due to *M. javanica* in tomato plants grown side by side with *T. erecta* was significantly lower when compared to tomato grown alone. On the other hand, there were no significant differences between intercropping with *Tagetes* and *Brassica* in almost cases.

Table 1. Mean of galls and egg masses in tomato, cucumber, and pepper intercropping with *Crotalaria*, *Tagetes*, and *Brassica* plants in the screenhouse.

Treatments	No. of Galls			No. of Egg masses		
	Tomato	Cucumber	Pepper	Tomato	Cucumber	Pepper
Intercropping with <i>Crotalaria</i>	153.7 c	205.7 c	197.0 c	160.3 c	158.3 c	189.7 c
Intercropping with <i>Tagetes</i>	251.7 bc	311.0 b	305.3 b	233.7 bc	220.7 c	295.0 b
Intercropping with <i>Brassica</i>	332.7 ab	374.0 b	363.7 ab	288.7 b	325.3 b	371.7 a
Without Intercropping	429.7 a	474.0 a	408.7 a	387.7 a	431.7 a	411.0 a
F value	21.83 ** < 0.0003	28.37 ** < 0.0001	31.79 ** < 0.0001	28.94 ** < 0.0001	41.70 ** < 0.0001	102.17 ** < 0.0001
LSD	113.83	95.681	73.48	80.626	84.274	43.687

Means followed by the same letter(s) within a column are not significantly different at 5% level of significance. LSD (5%) = least significant difference, while different letters had a statistically significant differences. *: mean significance. **: mean highly significance.

Table 2. Final population (Pf) and rate of reproduction (Rr) of *Meloidogyne incognita* infecting tomato, cucumber, and pepper intercropping with *Crotalaria*, *Tagetes*, and *Brassica* plants in the screenhouse

Treatments	Pf			Rr		
	Tomato	Cucumber	Pepper	Tomato	Cucumber	Pepper
Intercropping with <i>Crotalaria</i>	570.0 c	692.3 d	746.3 c	0.57 c	0.69 d	0.75 c
Intercropping with <i>Tagetes</i>	875.3 b	1054.7 c	1250.3 b	0.88 b	1.05 c	1.25 b
Intercropping with <i>Brassica</i>	1099.3 b	1180.0 b	1076.3 b	1.10 b	1.18 b	1.08 b
Without Intercropping	2175.7 a	2276.3 a	2179.0 a	2.18 a	2.28 a	2.18 a
F value	180.68**<0.0001	704**<0.0001	90.98**<0.0001	180.68**<0.0001	704**<0.0001	90.98**<0.0001
LSD	235.27	116.46	291.53	0.235	0.116	0.29

Means followed by the same letter(s) within a column are not significantly different at 5% level of significance. LSD (5%) = least significant difference, while different letters had a statistically significant differences. *: mean significance. **: mean highly significance.

Moreover, intercropping *Crotalaria*, *Tagetes*, and *Brassica* increased the plant growth of the main host plants (Figures 1,2,3)

Fig. (1) showed that the different growth parameters of tomato were increased significantly by using the intercropping technique in the case of *Crotalaria* and insignificantly when *Tagetes* and *Brassica* were used.

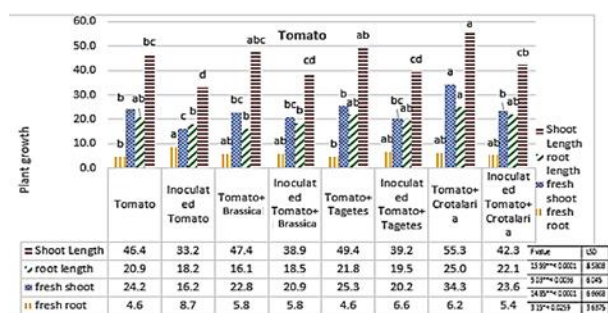


Figure 1. Effect of intercropping *Crotalaria*, *Tagetes*, and *Brassica* with tomato on the plant growth parameters.

The same letters above bars have no significant differences, while different letters above bars indicate statistically significant differences in their effects.

The growth of cucumber demonstrated that there was efficient increase in the plant growth without any significant differences between the effect of *Crotalaria* and *Tagetes* followed by *Brassica* (Fig.2).

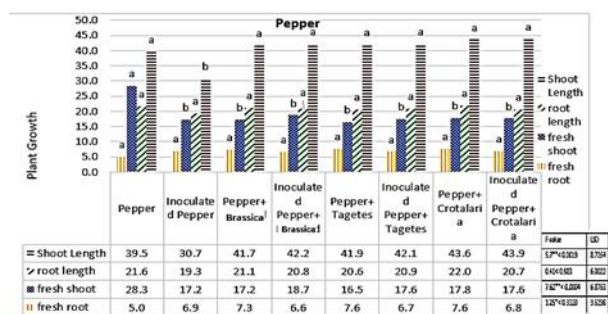


Figure 2. Effect of intercropping *Crotalaria*, *Tagetes*, and *Brassica* with pepper on plant growth parameters.

The same letters above bars have no significant differences, while different letters indicate statistically significant differences in their effects.

Data in Figs. (2&3) recorded that all plant growth parameters of pepper and cucumber were improved with the intercropping of *Crotalaria*, *Tagetes*, and *Brassica* and the differences were insignificant between them.

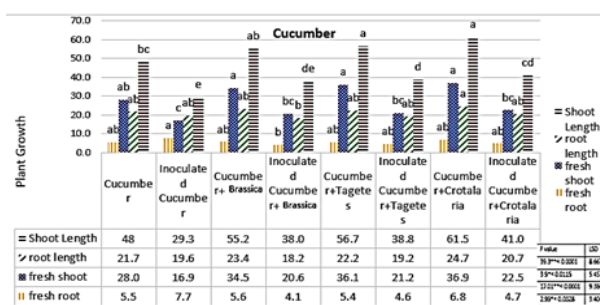


Figure 3. Effect of intercropping *Crotalaria*, *Tagetes*, and *Brassica* with cucumber on plant growth parameters.

The same letters above bars have no significant differences, while different letters above bars indicate statistically significant differences in their effects.

Generally, the previous data stated that, the intercropping of some nematotoxic plants i.e. *Crotalaria*, *Tagetes*, and *Brassica*. decreased the nematode parameters while increased growth. These data were supported by Ferraz and Grassi de Freitas (2004) or at least did not suppress the host plant. Bello *et al.*, (2014) reported that, *T. erecta* and *C. juncea* improved the plant growth and yields when intercropped with tomato plants.

As typified before many researches explained the effect of the plants which have a nematicidal effect on plant parasitic nematodes such as: *Crotalaria*, *Tagetes*, and *Brassica*.

Crotalaria has a nematicidal active ingredient byproduct property on *M. incognita* which can inhibit the growth of the nematodes. This effect may be better than some nematicidal chemicals due to the continued effect in suppression even after the host is planted (Huang *et al.*, 1981; Sharma and Scolari, 1984), as well as safety and improving the host yield. *Crotalaria* effect may be attributed to their nematicidal root exudates (Villar and Zavaleta-Mejia, 1990) which contain allelopathic compounds that called monocrotaline and pyrrolizidine alkaloids (PA) which can affect the nematode (Jourand *et al.*, 2004; and Wang and McSorley, 2004), or penetrate larval body and inhibit their metabolic activity and some enzymes functions such as: acetyl cholinesterase enzyme and respiratory enzymes (Aguillera, *et al.*, 1984). *Crotalaria* can be used as a non-crop plants for some plant parasitic nematodes especially root knot nematodes (Luc *et al.*, 1990). Also, it is a highly resistant plant to some *Pratylenchus* species and *Rotylenchulus reniformis* (Silva *et al.*, 1989a, 1989b; and Ferraz and Grassi de Freitas, 2004).

Intercropping of marigold is very effective (Ferraz and Grassi de Freitas, 2004; and Hooks *et al.*, 2010) on

Pratylenchus pratensis (Oostenbrink, 1960; and Conijn *et al.*, 1996) and *Meloidogyne* by multiple mechanisms; as a poor or a non-host (Hooks *et al.*, 2010), producing allelopathic nematicidal compounds (alpha-terthienyl) (Topp *et al.*, 1998; and Ferraz and Grassi de Freitas, 2004), enhancing some nematode antagonistic fauna (Wang *et al.*, 2001) or using as a trap host (Pudasain *et al.*, 2008). Ferraz and Grassi de Freitas (2004) explained that *T. patula* allowed J2 penetration, but the nematode did not develop beyond the early second stage and some necrotic reaction was observed in the feeding sites of the infective juveniles and may be few and very small giant cells and galls are produced (Suatmadji, 1969). Moreover, the using of marigold may be more effective in nematode control than some fumigants and chemical nematicides in some instances (Hooks *et al.*, 2010), and the effect may have greater depths because of the debris which remained in the soil after the crop rotation that continue to exert its nematicidal effect on PPNs (Evenhuis *et al.*, 2004).

All *Brassica* spp. can release bio-toxic compounds that showed activity against fungi, bacteria, insects, nematodes (Bohinc *et al.*, 2013; Laznik *et al.*, 2014; Grubišić *et al.*, 2018). The nematicidal effect of brassicas is due to their ability to produce glucosinolate compounds which are b-D-thioglucosides, that are accumulated in the different plant parts (roots, stems, and leaves). Brassicas plants attract the juvenile larvae of endoparasitic nematodes as a trap, after root penetration, they are poisoned by hydrolysis products and fail in completing their life cycle during the intercropping time. Consequently, the population density in soil will be decreased (Curto, 2008). Penfold and Collins, (2012) reported that, *Brassica* can be used in rotation crops and the results depend on the geographic regions, and the toxic chemical compounds in *Brassica* species (Chen *et al.*, 2007; Hartz *et al.*, 2005; Monfort *et al.*, 2007; and Edwards and Ploeg 2014), and correlated to the soil moisture at the time of the application (Matthiessen *et al.*, 2004).

In conclusion, the intercropping of the previous plants which have nematotoxic effect on root knot nematode *M.incognita* without any suppressive effect on the plant growth, can be an efficient method in the integrated pest management programs after further studies.

REFERENCES

- Abid M. and Maqbool M. A. (1990). Effects of intercropping of *Tagetes erecta* on root – knot disease and growth of tomato. International Nematology Network Newsletter 7: 41-42
- Aguillera M.M., S.Matsuoka and O.T.Ido, (1984). Efeitos da aduba; Verde Com. *Crotalaria juncea* sobre nematosea produ de canadeaiar, Nematologia Brasileira, 8:25-26.
- Akhtar, M and A. Malik. (2000). Role of organic soil amendments and soil organisms in the biological control of plant-parasitic nematodes: A review. Bioresource Technology, 77 (1): 35-47.
- Anaya, A. L. (2006). Allelopathic organisms and molecules: Promising bioregulators for the control of plant diseases, weeds and other pests. Pp 31-78. In Inderjit and K.G.Mukerji (eds.) Allelochemicals: Biological control of plant pathogens and diseases. Springer. The Netherlands.
- Bello A., Bricka R.M., Gerard P.D., Peebles E.D. (2014). Effects of commercial Inova injection of 25-hydroxycholecalciferol on broiler bone development and mineralization on days 0 and 21 posthatch Poul. Sci., 93, pp. 1053-1058
- Bohinc T; Ban SG; Ban D; Trdan S. (2012). Glucosinolates in plant protection strategies: a review. Arch Biol Sci Belgrade. 64:821–828.
- Bosland, P.W. and Votava, Eric J. (2012). Vegetable and Spice Capsicums, 2nd edition, New Zealand Journal of Crop and Horticultural Science, Vol. 41 (2):102-103
- Bridge J. (1996). Nematode management in sustainable and subsistence agriculture. Annual review of Phytopathology 34 ():201-25
- Chen G., Clark A., Kremen A., Lawley Y., Price A., Stocking L., Weil R. (2007). Brassicas and mustards. In: Managing Cover Crops Profitably, 3rd Edition (Clark A., ed), Sustainable Agriculture Research and Education (SARE) program, Beltsville, MD, USA, pp. 81-90.
- Chitwood D.J. (2002). Phytochemical based strategies for nematode control. Annual Review of Phytopathology 40:221-249.
- Conijn C. G. M., Molendijk L. P. G., Schepman M., Koster A. T., Schenk A. M. E., Kroonen Backbier B., Gommers F. J., Brinkman H., (1996). *Tagetes* and root lesion nematodes. Gewasbescherming 27: 106-110
- Curto G. (2008). Sustainable methods for management of cyst nematodes. In: Integrated Management and Biocontrol of Vegetable and Grain Crops Nematodes. Integrated Management of Plant Pests and Diseases, vol 2. (Ciancio A., Mukerji K. G., eds), Springer, Dordrecht, pp. 221-237.
- Edwards, S. and Ploeg, A. (2014). Evaluation of 31 potential biofumigant brassicaceous plants as hosts for three Meloidogyne Species. J. Nematol., 46(3): 287–295.
- Esparrago, G., Barreiro, J.M., Ruales, C. and Bieche, B.J. (1999). Comparison of the reproduction of *Meloidogyne* populations on roots of *Crotalaria spectabilis* and processing tomato varieties with the Migene. Acta Horticulturae 487: 267–270.
- Evenhuis A., Korthals G. W., Molendijk L. P. G. (2004). *Tagetes patula* as an effective catch crop for long-term control of *Pratylenchus penetrans*. Nematology 6 (6): 877-881.
- Ferraz S.; Grassi de Freitas L. (2004). Use of antagonistic plants and natural products. In: Nematology-Advances and perspectives; volume II, Nematode Management and Utilisation (Chen Z. X., Chen S. Y., Dickson D. W., eds), CABI Publishing, Wallingford, UK, pp. 931-977
- Grubišić Maja;; Roy H.A.van Grunsven; Alessandro Manfrin; Michael T. Monaghan; Franz Hölker. (2018). A transition to white LED increases ecological impacts of nocturnal illumination on aquatic primary producers in a lowland agricultural drainage ditch. Environmental Pollution, 240, :630-638.
- Haider MG, Sinha UP, Dutta K. (2004). Effect of intercropping of mustard (*Brassica campestris* var. toria/*B. campestris* var. sarson) with sugarcane on nematode population and yield of crop. Indian Journal of Nematology 34 (2):160-164.

- Halbrendt J.M. (1996). Allelopathy in the management of plant-parasitic nematodes. *Journal of Nematology* 28 (1):8-14.
- Hartz, T.K., Johnstone, P.R.; E.M. Miyao, E.M. and Davis, R.M. (2005). Mustard cover crops are ineffective in suppressing soil-borne disease or improving processing tomato yield. *Hort. Science*, 40 (7): 2016–2019.
- Hooks, C. R. R., Wang, K.-H., Ploeg, A., and McSorley, R. (2010). Using marigold (*Tagetes* spp.) as a cover crop to protect crops from plant-parasitic nematodes. *Applied Soil Ecology* 46 (3):307–320.
- Huang C.S., J.M.Charchar and R.C.V.Tenente (1981), Control of root-knot nematode (*Meloidogyne incognita*) in carrot (*Daucus carota*) through rotation. *Fitopatologia Brasileira* 5:329-336.
- Jones JT, Haegeman A, Danchin EGJ, Gaur HS, Helder J, Jones MGK, Kikuchi T, Rose ML, Palomares-Rius JE, Wesemale WML, Perry RN., (2013). Review of top 10 plant-parasitic nematodes in molecular plant pathology. *Molecular Plant Pathology*, pp. 1-16.
- Jourand, P., S. Rapior, M. Fargette, and T. Mateille, (2004). Nematostatic activity of aqueous extracts of West African *Crotalaria* species. *Nematology* 6 (5):765-771.
- Kas,kavalci, G., Tüzel, Y., Dura, O., Öztekin, G.B. (2009). Effects of alternative control methods against *Meloidogyne incognita* in organic tomato production. *Ekoloji* 18(72): 23–31.
- Khalil M.S. (2013). Alternative approaches to manage plant-parasitic nematodes. *Journal of Plant Pathology and Microbiology* 4 (1): e 105. doi:10.4172/2157-7471.1000e105.
- Kowalska B., Sonalinska U. (2001). Effect of Brassicaceae plants on the nematode *Pratylenchus* spp. and soil microorganisms. *Vegetable Crop Research Bulletin* 55:121-125.
- Kumar S.and Khanna A.S.; (2006). Role of *Trichoderma harzianum* and neem cake separately and in combination against root-knot nematode on tomato. *Indian Journal of Nematology* 36 (2):247-249.
- Lamovsek, J., Urek, G., Trdan, S.; (2013). Biological control of root-knot nematodes (*Meloidogyne* spp.): microbes against the pests. *Acta Agric. Slov.* 101(2): 263–275.
- Laznik Ž., Trdan S., Vučajnk F., Bohinc T., Vidrih M., (2014). Cruciferous plants use as bio-fumigants in potato against wireworms. *Acta. Agric. Scand. B. Soil Plant Sci.* 64 (7): 606-614.
- Luc, M., Sikora R. A. and Bridge J., (1990). *Plant Parasitic Nematodes in Subtropical and Tropical Agriculture*. CAB International, Wallingford, UK. 629 p.
- Matthiessen, J.N., B. Warton B. and Shackleton, M.A. (2004). The importance of plant maceration and water addition in achieving high *Brassica*-derived isothiocyanate levels in soil. *Agroindustria*, 3(3): 277–280.
- McBeth C.W., Taylor A.L., Smith A.L. (1941). Note on staining nematodes in root tissue. *Proceedings of Helminthological Society of Washington*; 8:26.
- Monfort, W.S., A.S. Csinos, J. Desaegeer, K. Seebold, T.M. Webster and J.C. Diaz-Perez, (2007). Evaluating *Brassica* species as an alternative control measure for root-knot nematode (*M. incognita*) in Georgia vegetable plasticulture. *Crop Prot.*, 26 (9): 1359–1368.
- Nagaraju N., Karemegam N., Kadalmani B. (2010). Eco-friendly management of root-knot nematode *Meloidogyne incognita* using organic amendments on tomato. *International Journal of Research Pharmacological Science* 1:530-532.
- Noling J.W., Becker J.O., (1994). The challenge of research and extension to define and implement alternatives to methyl bromide. *Journal of Nematology* 26: 573– 86.
- Oostenbrink M. (1960). *Tagetes patula* L. als voorvrucht van enkele land-en tuinbouwgewassenop zand-en dalgrond. Mededelingen van de Landbouwhoogeschool en Opzoekingsstations Gent. 25:1065–1075.
- Penfold C., Collins C., (2012). Cover crops and plant-parasitic nematodes. *Wine Australia Factsheet* June 2012.
- Perry, R. N., Moens, M., and Starr, J. L., (2009). *Root-knot nematodes*. Wallingford, UK: CAB International.
- Ploeg A. (2008). Biofumigation to manage plant-parasitic nematodes. In: Ciancio A, Mukerji KG (eds.), *Integrated Management and Biocontrol of Vegetable and Grain Crops Nematodes*, Springer Netherlands, pp. 239-248.
- Pudasaini, M. P., Viaene, N., and Moens, M. (2008). Hatching of the root-lesion nematode, *Pratylenchus penetrans*, under the influence of temperature and host. *Nematology* 10: 47–54.
- Samaliev, H. (2009). Solarization of soil for the control of root-knot nematodes (*Meloidogyne* species) vegetable glasshouses in Southern Bulgaria. *Plant Science* 46:361–365.
- Sharma R.D. and Scolari D.D.G. (1984). Efficiency of green manure and crop rotation in the control of nematodes under savannah conditions. *Nematologia Brasileira*. 8: 193-212.
- Silva, G. S., Ferraz, S., Santos, J. M. 1989a. Resistência de espécies de *Crotalaria* a *Pratylenchus brachyurus* e *P. zae*. *Nematologia Brasileira*, 13, 81-86.
- Silva, G. S.; Ferraz, S., Santos, J. M. 1989b. Resistência de espécies de *Crotalaria* a *Rotylenchulus reniformis*. *Nematologia Brasileira*, 13, 87-92.
- Singh R.S., Sitaramaiah K.; (1993). Cultural and land management practices. In: *Plant Pathogens. The Plant Parasitic Nematodes* (Eds. R.S. Singh and K. Sitaramaiah). Oxford and IBH Publishing Co. Pvt. Ltd. New Delhi, pp. 99-109.
- Singh S., Singh B., Singh A.P. (2015). *Nematodes: A Threat to Sustainability of Agriculture*. *Procedia Environmental Sciences* 29:215-216.
- Suatmadji, R. W. (1969). Studies on the effect of *Tagetes* species on plant-parasitic nematodes. Wageningen, The Netherlands. H. Veenman en Zonen. PhD thesis, 132 p
- Topp E., Millar, S., Bork, H., Welsh, M. (1998). Effects of marigold (*Tagetes* sp.) roots on soil microorganisms *Biol Fertil Soils* 27(2):149–154
- U.S. Department of Agriculture, Agricultural Research Service (2005). *Composition of Foods Raw, Processed, Prepared* USDA National Nutrient Database for Standard Reference, Release 18. *Agriculture Handbook*; 1-35.

- Varma M. K., Sharma H. C., Pathak V. N., (1978). Efficacy of *Tagetes patula* and *Sesamum orientale* against root knot of eggplant. Plant Dis. Rep. 62: 274-275.
- Villar E. M. J; Zavaleta Mejía E.-(1990). Effect of *Crotalaria longirostrata* Hook y Arnott on root galling nematodes (*Meloidogyne* spp.). Rev Mex Fitopatol 8: 166-172
- Wang, K. H. and McSorley, R. (2004). Management of nematodes and soil fertility with sunn hemp cover crop. University of Florida, IFAS Extension, ENY-717. 6 pp. http://edis.ifas.ufl.edu/pd_files/NG/NG04300.pdf.
- Wang, K.H., Brent, S.S. (2009). Solarization and cover cropping as alternatives to soilfumigants for nematode management in Hawaii's pineapple fields. Soil Crop Manag., 1-4.
- Wang, K.-H., Sipes, B. S., and Schmitt, D. P. (2001). Suppression of *Rotylenchulus reniformis* by *Crotalaria juncea*, *Brassica napus*, and *Tagetes erecta*. Nematropica 31(2):235-249.
- Wilson, G.F and Caveness, F.E. (1980). Effects of rotation crops on the survival of root-knot, root lesion and spiral nematodes. Nematropica 10 (1): 56-61.
- Youssef, M.M.A., El-Nagdi, W.M., Ahmed, A.A. (2011). Interaction of cucumber mosaic virus with the root-knot nematode, *Meloidogyne incognita*, and effects of certain medicinal and aromatic plants on infected cucumbers. Nematol. Mediter. 39: 73-80.

تأثير تحميل النباتات التي لها تأثير ابادى على نيماتودا تعقد الجذور فى بعض نباتات الخضر بالصوبة السلكية انتصار حلمي طه قسم وقاية النبات، كلية الزراعة، جامعة عين شمس

تتسبب النيماتودا الممرضة للنبات وخاصة نيماتودا تعقد الجذور فى أضرار اقتصادية بالغة على النباتات وخاصة نباتات الخضر على مستوى العالم. ويسبب الاضرار السلبية لاستخدام المبيدات النيماتودية ذات الاصل الكيمايى على صحة الانسان وعلى الحيوانات وما تسببه لما تسببه من تلوث فى البيئة الزراعية، كانت هناك ضرورة ملحة لايجاد وسائل اخرى غير كيميائية لمكافحة النيماتودا بحيث تكون فعالة وامنة ومقبولة اقتصاديا. يعتبر تحميل النباتات التي لها تأثير مثبط لنشاط النيماتودا احد هذه البدائل المطروحة، وعليه كانت فكرة البحث لدراسة تأثير تحميل نباتات الكروتالاريا *Crotalaria juncea* والقטיפفة *Tagetes erecta* والخردل *Brassica carinata* منفصلة على بعض نباتات الخضر الهامة اقتصاديا مثل الطماطم *Lycopersicon esculentum* L. var. Castel Rock، الخيار *Cucumis sativus* واللفل *Capsicum annum* لدراسة قدرتها على خفض اعداد نيماتودا تعقد الجذور (بتركيز 1000 يرقة نيماتودا/اصيبص مع مراعاة تأثيرها على نمو النباتات، أجريت التجربة فى الصوبة السلكية وانهيت بعد حوالى شهرين وأخذت النتائج وكانت كالتالى: سجلت النباتات الثلاثة المستخدمة فى التحميل تأثر ابادى على النيماتودا حيث تسببت فى خفض أعداد الأورام على الجذور، أعداد البيض، وخفضت أعداد النيماتودا المسجلة فى نهاية التجربة وايضا معدل تكاثر النيماتودا وكانت أشدها فى التأثير الكروتالاريا والتي سجلت: 153.7 - 160.3 - 570.0 - 57.0 فى الطماطم، و 205.7 - 158.3 - 692.3 - 0.69 فى الخيار وأخيرا 197.0 - 189.7 - 746.3 - 0.75 فى الفلفل على الترتيب ثم تلاها نبات القטיפفة فى التأثير ثم نبات الخردل. كما سجلت النباتات الثلاثة المستخدمة فى التحميل تحسین معنوی فى نمو النباتات المنزرعة عند مقارنتها بالنباتات المنزرعة بدون تحميل. من النتائج المتحصل عليها يتضح انه من الممكن استخدام نباتات الكروتالاريا، القטיפفة والخردل فى التحميل كوسيلة جديدة فعالة وأكثر أمان فى مكافحة نيماتودا تعقد الجذور فى نباتات الخضر ضمن برامج مكافحة متكاملة.