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Degradative and antimicrobial potentials of *Bacillus* species isolated from water samples and contaminated soil in Ado-Ekiti metropolis, Nigeria

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ABSTRACT

This study investigated the ability of Bacillus species isolated from soil and water samples to degrade some petroleum products and pesticides as well as its ability antimicrobial potency against selected pathogens. Bacillus species were isolated from soil collected from dumping site and water samples from Ado-Ekiti, Nigeria on Luria Bertani agar. The biodegradative and antimicrobial activities of the isolates were conducted using minimal nutrient medium and agar well diffusion methods respectively. Commercial antibiotics were used as positive control. All the Bacillus isolates showed varied degree of degradation on petroleum products and pesticides. Only B. polymyxa showed antimicrobial activity against Staphylococcus aureus with zone of inhibition of 18.00mm. The commercial antibiotics were most inhibitory to most of the test pathogens in varying degrees. The isolates can be used in mitigating pollution caused by oil spillage and overuse or abusive use of herbicides and fungicides. However, these isolates are not good candidate as antibacterial agents for the selected test pathogens.

INTRODUCTION

The release of contaminant to the environment, including petroleum and petroleumderived products, is one of the main causes of global contamination (Rahman *et al.*, 2002) which also constitutes risk to human and animal health because of their toxigenic and carcinogenic properties. Mechanical and chemical methods for remediation of hydrocarbonpolluted environment are expensive, technologically complex and lack public acceptance. The biodegradation of hydrocarbons has a high ecological significance, as it constitutes the major process for remediation of contaminated areas (Prabhu and Phale, 2003). Thus, bioremediation remains the method of choice for effective removal of hydrocarbon pollutants in the environment. Hydrocarbon degrading bacteria and fungi are mainly responsible for the mineralization of oil pollutants and are distributed in diverse ecosystem (Alloway and Ayres 1993). The microbial degradation of hydrocarbon seems to be a promising tool in the control of pollution and is the subject of a large number of current researches on the biochemistry and the genetics involved in activity. Bacteria, yeast and filamentous fungi have been reported as transforming agents (Hester and Mendelssohn, 2000). Screening for new antibiotics from natural sources is becoming increasingly important for the pharmaceutical industry (Schmidt, 2004) as pathogenic bacteria are becoming resistant to commonly quickly therapeutic agents. Secondary used metabolites such as bacitracin, gramycidin, polymyxin and tyrotricidin are produced only by some species of a genus Bacillus (Stachelhaus et al., 1995; Drablos et al., 1999) which are more effective against Gram positive than Gram negative bacteria. Lipopeptides produced by Bacillus also demonstrate anti-fungal, anti-viral. anti-ameobocytic and antimycoplasma activities ((Milner et al., 1995; Peypoux et al., 1999; Steller et al., 1999). The chemical and physical diversity of peptide antibiotics makes them ideal candidates not only for therapeutic applications but also in other areas, especially the agri-food industry (Mukherjee et al., 2004). The present investigation therefore attempts to investigate the biodegradative and antimicrobial potency of Bacillus species isolated from soil and water samples.

MATERIALS AND METHODS Collection of sample

Surface soil sample was collected from a dumping site while water sample were collected in air tight bottles from different regions of Ado-Ekiti, metropolis, Ekiti State, Nigeria. All the samples were brought into the laboratory for further analysis.

Test organisms

Streptococcus pyogenes, Serratia marcescens, Klebsiella pneumoniae, Staphylococcus aureus and Proteus mirabilis were collected from the Microbiology Laboratory, State Teaching Hospital, Akure. Isolation and identification of microbial

Isolation and identification of microbial isolates

Serial dilution technique was used in isolating *Bacillus* species from soil and water samples using Luria Bertani Agar (LBA). The bacterial isolates obtained were characterized using various morphological and biochemical tests described by Fawole and Oso (2001) and identification was carried out using Cowan and Steel (1993) and Holt *et al.* (2000).

Antimicrobial study

Bacillus species isolated from the different samples were tested against some clinical bacteria isolates such as **Streptococcus** pyogenes, Serratia marcescens, Klebsiella pneumoniae, Staphylococcus aureus and *Proteus* mirabilis using agar well diffusion method. The Bacillus cultures were centrifuged at 6000g for 15mins to remove cell debris. After centrifugation, 0.5ml of the supernatant were transferred carefully into the labelled holes while sterile distilled water was filled into one hole serving as the control and the antibiotic disc served as positive control. The inoculated plates were incubated for 24hrs at 37°C and the diameter of the zone of inhibition was measured (Schillinger and Lucke, 1989)..

Antibiotic sensitivity pattern of the test bacteria

Antibiotic sensitivity test was carried out on *Bacillus* sp. and test pathogens using conventional antibiotic disc. This was done placing the antibiotic disc on solidified Mueller Hinton agar plate already seeded with the organisms and incubated at 30°C for 24hrs. The antibiotic used contained the following commercial antibiotics namely; Pefloxacin (PEF), Gentamycin (CN), Zinnacef Ampiclox (APX), (Z), Amoxacillin (AM), Rocephin (R). Ciprofloxacin (CPX), Streptomycin (S), Septrin Erythromycin (SXT), (E), Chloramphenicol (CH), Augmentin (AU), Tarivid (OFX), Sparfloxacin (SP).

Degradation study

Degradation study was carried out using Minimal salt broth (MSB) (Yin et al., 1998). This was sterilized at 121°C for 15 minutes in the autoclave. About 100ml of minimal salt broth was measured into each Erlenmeyer flask and was supplemented with 0.5ml of the petroleum products (crude oil, petrol, diesel, engine oil and kerosene) and pesticides (herbicides and fungicides). Each Erlenmeyer flask was inoculated with the different *Bacillus* species isolated. All the Erlenmeyer flasks were incubated at 37°C for 5 days; the turbidity was recorded on each day using a spectrophotometer at 540nm optical density.

Statistical analysis

All the experiment carried out was done in triplicates and statistical analyses of data obtained were carried out using analysis of variance (ANOVA) and Duncan's Multiple Range Test for the estimation of means (SPSS windows 16 version).

RESULTS

Biodegradative ability of *Bacillus* species on petroleum hydrocarbons and selected herbicides

Four *Bacillus* species were isolated from contaminated soil and water and they were: *Bacillus subtilis, B. megaterium, B. cereus* and *B. polymyxa*. The degradative abilities of *Bacillus* species on selected petroleum hydrocarbon is shown in Figure 1. All the Bacillus species tested were able to degrade the hydrocarbon to different extent. Bacillus cereus had the highest degradative ability on crude oil, engine oil, kerosene and petrol by having the highest turbidity when compared to the other Bacillus species. This shows the ability of the organism to use these hydrocarbons as a carbon source. *Bacillus megatarium* had the highest degradative ability on diesel. Table 1 shows the degradative abilities of Bacillus species on selected herbicides. Bacillus subtilis had the highest degradative ability on premextra with turbidity increase from 2.50-6.12 as compared to Bacillus megatarium which showed the least degradative ability on premextra with turbidity ranging from 0.56-1.59. Bacillus cereus had the highest degradative ability on glyphosphate as turbidity increased from 0.76 on day1 to 8.90 on day 5 as compared to Bacillus polymyxa, whose turbidity decreased from 4.90 on day2 to 1.91 on day5.



Fig. 1: Degradative abilities of Bacillus species on selected petroleum hydrocarbon

Isolates	Premextra					Glyphosphate				
	Day1	Day2	Day3	Day4	Day5	Day1	Day2	Day3	Day4	Day5
B.cereus	1.80	1.95	2.10	2.80	3.80	0.76	2.55	5.20	6.30	8.90
	± 0.00	±0.10	± 0.00	±0.03	± 0.00	± 0.00	± 0.05	± 0.00	± 0.00	± 0.00
В.	0.56	0.72	1.25	1.32	1.59	2.35	2.90	3.12	3.51	3.80
megatarium	± 0.00	±0.03	± 0.05	± 0.00	± 0.01	±0.05	± 0.00	± 0.00	±0.01	± 0.00
B. subtilis	2.50	3.17	4.60	5.21	6.12	3.00	4.03	4.91	5.05	5.55
	±0.05	±0.03	± 0.00	± 0.00	± 0.00	±0.00	± 0.00	± 0.01	± 0.05	± 0.00
B. polymyxa	1.20	1.60	1.98	1.98	2.40	4.10	4.90	2.00	2.00	1.91
	± 0.00	±0.00	± 0.00	± 0.00	± 0.00	±0.09				
Control	0.60	0.60	0.60	0.60	0.63	0.66	0.67	0.65	0.66	0.67

Table 1: Degradative abilities of Bacillus species on selected herbicides

Also, B. *subtilis* had the highest degradative ability on benlate and mancozeb as its turbidity increased from 4.50 on day1 to 5.40 on day 5 and from 6.54 on day 1 to 11.54 on day 5 on benlate and mancozeb respectively. Decrease in turbidity was seen in both *B. megatarium* and *B. polymyxa* with

turbidity reducing 2.50 on day1 to 1.90 on day 5 and 3.22 on day1 to 1.90 on day 5 respectively. It was observed that *B. cereus* could not grow well on mancozeb as its turbidity decreased from 2.09 on day1 to 1.41 on day 5 (Table 2).

Table 2: Degradative abilities of Bacillus species on selected fungicides

Isolates	Benlate				Mancozeb					
	Day1	Day2	Day3	Day4	Day5	Day1	Day2	Day3	Day4	Day5
B.cereus	1.90	2.10	2.40	2.57	3.80	02.09	2.00	1.91	1.63	1.41
	± 0.00	±0.10	± 0.00	±0.03	±0.03	±0.01	± 0.00	±0.03	± 0.00	±0.01
B. megatarium	2.50	2.51	2.70	2.40	1.90	1.00	1.00	0.91	0.83	0.69
	± 0.00	±0.03	± 0.00	± 0.00	± 0.00	±0.00	± 0.00	±0.01	±0.01	±0.01
B. subtilis	4.50	4.91	4.99	5.20	5.40	6.54	9.61	9.50	10.01	11.54
	± 0.00	±0.03	± 0.01	± 0.00	± 0.00	±0.02	± 0.01	± 0.00	± 0.05	± 0.04
B. polymyxa	3.22	3.52	3.00	2.10	1.90	4.63	5.21	6.69	7.00	7.68
	± 0.01	±0.00	± 0.00	± 0.00	± 0.00	±0.01	± 0.01	± 0.00	± 0.00	± 0.00
Control	0.70	0.70	0.70	0.73	0.72	0.70	0.70	0.70	0.70	0.70

Antibacterial activities of *Bacillus* species on selected pathogenic bacteria

None of these bacteria was susceptible to *Bacillus* species except *S. aureus* which was susceptible to *B. polymyxa* with zone of inhibition of 18.00mm (Table 3). However, the susceptibility pattern of the test pathogens to commercial antibiotics showed that *Proteus mirabilis* was sensitive to Amoxacillin and *Staphylococcus aureus* was sensitive to Zinnacef while *Streptococcus pyogenes* was very sensitive to Ampiclox, Zinnacef and Amoxacillin while all other isolates were susceptible to the antibiotics in varying degrees as shown in Tables 4 and 5.

Table 3: Antimicrobial activities of Bacillus species against some bacteria pathogens

Isolates	Diameter of zones of inhibition (mm)								
	Proteus	Serratia Staphylococcus		Klebsiella	Streptococcus				
	mirabilis	marcesens	aureus	pneumonia	Pyogenes				
1	00.00±0.00	00.00 ± 0.00	00.00±0.00	00.00±0.00	00.00±0.00				
2	00.00 ± 0.00	00.00 ± 0.00	00.00 ± 0.00	00.00 ± 0.00	00.00±0.00				
3	00.00 ± 0.00	00.00 ± 0.00	00.00 ± 0.00	00.00 ± 0.00	00.00±0.00				
4	00.00 ± 0.00	00.00 ± 0.00	18.00 ± 0.00	00.00 ± 0.00	00.00 ± 0.00				
17 1 D	1								

Key: 1 - B. subtilis

2-B. cereus

3 – B. megaterium

4 – B. polymyxa

Test	Zones of inhibition (mm)									
pathogen	S	SXT	СН	SP	CPX	AM	AU	PEF	OFX	CN
Proteus	22.00	21.00	17.00	16.00	27.00	00.00	22.00	20.00	19.00	18.00
mirabilis	± 0.60	± 1.00	±0.33	± 0.00	±0.33	± 0.00	±0.03	±0.20	±0.63	±0.67
Serratia	19.00	20.00	15.00	20.00	29.00	19.00	23.00	20.00	21.00	19.00
marcesens	±0.67	±0.33	± 0.00	± 0.00	± 0.00	± 1.20	±0.67	±0.93	±0.67	±0.33
Klebsiella	20.00	19.00	16.00	18.00	26.00	16.00	20.00	18.00	20.00.00	17.00
pneumoniae	± 0.00	±0.33	± 1.10	±1.33	±0.33	± 0.67	± 1.00	± 0.00	± 0.00	±0.33

 Table 4: Antibiotics susceptibility of Gram negative test pathogens

 Table 5: Antibiotics susceptibility of Gram positive test pathogens

Test pathogen	Zones of inhibition (mm)									
	R	СРХ	S	SXT	Ε	PEF	CN	APX	Ζ	AM
Staphylococcus	20.00	20.00	18.00	21.00	18.00	17.00	20.00	16.00	00.00	17.00
aureus	±0.33	±0.67	±0.67	±0.33	± 1.60	± 1.00	±0.67	±0.67	± 0.00	±1.67
Streptococcus	18.00	17.00	19.00	20.00	16.00	17.00	18.00	00.00	00.00	00.00
pyogenes	± 0.00	±0.10	±0.67	±0.33	±0.33	±1.10	± 1.00	±0.00	± 0.00	± 0.00

DISCUSSION AND CONCLUSION

Soil is considered as one of the most suitable environments for microbial growth (Cavalcanti et al., 2006). Four Bacillus namely *Bacillus* species subtilis, *B*. megatarium, B. cereus and B. polymyxa were isolated from contaminated soil and water. The present work shows the degradation of petroleum hydrocarbon, fungicides and herbicides. Most of these Bacillus species live in the soil, multiply and produce enzymes that help in bioremediation (Adesemoye et al., 2006). Generally, the Bacillus species used for this work were all able to degrade all petroleum hydrocarbons. This might be attributed to their ability to produce spores which may shield them from the hydrocarbon (Adesemoye et al., 2006). However, their ability to degrade the selected fungicides was low which may be due to the fact that fungicides contain a little amount of antimicrobial agent (Alloway and Ayres, 1993). The high spectrophotic value obtained in herbicides especially glyphosphate might be as a result of the ability of the organisms either to break or multiply in the presence of the chemical. Increase in spectrophotometer reading indicates that Bacillus species to efficiently utilize petroleum the hydrocarbons, fungicides and herbicides for their growth and establishment (Prabhu and Phale, 2003).

The antibacterial activities of *Bacillus* species against selected test pathogens show

that only *B. polymyxa* could inhibit the growth of S. aureus Yilmaz et al. (2006) also observed that S. aureus was resistant to B. cereus and B. megaterium although Perez et al. (1993) showed the sensitivity of S. aureus to various strains of Bacillus species. Oscariz et al. (1999) isolated and identified a bacteriocin-producing strain of B. cereus from a soil sample which was active against most Gram-positive but not Gram-negative bacteria. The findings of the present study indicate that Bacillus cereus had no antimicrobial effects on both Gram-positive Gram-negative bacteria. and All the commercial antibiotics were more inhibitory to the test pathogens than Bacillus species except zinnacef which was not inhibitory to aureus and S. pneumoniae and S. amoxacillin which was inhibitory to S. pyogenes and P. mirabilis. Also, S. pyogenes was not sensitive to ampiclox.

CONCLUSION

The results obtained in this study have shown that *Bacillus* species used in this work have the ability to degrade the hydrocarbons, fungicides and herbicides. It can therefore be employed in mitigating pollution caused by oil spillage and overuse or abusive use of herbicides and fungicides. However, these isolates are not good candidate as antibacterial agents for the selected test pathogens.

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REFERENCES

- Adesemoye, A. O., Opere, B. O. and Makinde, S. C. O. (2006). Microbial content of abattoir wastewater and its contaminated soil in Lagos, Nigeria. Afr. J. Biotechnol., 5: 1963-1968.
- Alloway, B. J. and Ayres, D. C. (1993). Organic pollutants In: Chemical principles or Environmental pollution, 1st edition, Chapman and Hall India. pp. 007-012.
- Cavalcanti, M. A., Oliveira, L. G., Fernandes, M. J. and Lima, D. M. (2006). Filamentous fungi isolated from soil in districts of the Xingó region, Braz. Acta Bot. Bras., 20 (4): 831–837.
- Cowan, S. T. and Steel, K. J. (1993). Cowan and Steel's manual for identification of medical <u>bacteria</u>. Cambridge University press, <u>Cambridge</u>, New York.
- Drablos, F., Nicholson, D. G. and Ronning, M. (1999). EXAFS study of zinc coordination in bacitracin A. Biochem. Biophys. Acta., 1431: 433-442.
- Fawole, M. O. and Oso, B. A. (2001). Laboratory manual for microbiology, spectrum books limited Ibadan., 75.
- Hester, M.W. and Mendelssohn, I. A. (2000).
 Long term recovery of Louisiana Brackish March plant community from oil spill impact: Vegetation response and mitigating effects of marsh surface elevation. Mar. Environ. Res., 49: 233-254.
- Holt, G. J., Krieg, N. R., Sneath, P. H., Staley, J. T. and William, S. T. (2000). Bergey's manual of bacteriology (9th Edition). Lippincott Williams & Wilkins, 530 Walnut St, Philadelphia, PA 19106 USA.

- Milner, J. L., Raffel, S. J., Lethbridge, B. J. and Handelsman, J. (1995). Culture conditions that influence accumulation of zwittermicin A by *Bacillus cereus* UW85. Appl. Microbiol. Biotechnol., 43:685-691.
- Mukherjee, S., Das, P. and Sen, R. (2006). Towards commercial production of microbial surfactants. Trends Biotechnol., 24: 509-515.
- Oscariz, J. C., Lasa, I. and Pisabarro, A. G. (1999). Detection and characterization of cerein 7, a new bacteriocin produced by *Bacillus cereus* with a broad spectrum of activity. FEMS Microbiol. Lett.,178: 337–341.
- Perez C, Suarez C, Castro GR. (1993). Antimicrobial activity determined in strains of *Bacillus circulans* cluster. Folia Microbiol., 38 (1): 25–28.
- Peypoux, F., Bonmatin, J. M. and Wallach, J. (1999). Recent trends in the biochemistry of surfactin. Appl. Microbiol. Biotechnol., 51: 553-563.
- Prabhu, Y and Phale, P. S. (2003). Biodegradation of Phenanthrene by *Pseudomonas* species strain pp 2: novel metabolic pathway, role of biosurfactant and cell surface hydrophobicity in hydrocarbon assimilation. Appl. Env. Microbiol., 61: 342-351.
- Rahman, K. S.M., Banat, I. M., Thahira, J. and Thayumanavan, T. (2002).
 Bioremediation of gasoline contaminated soil by a bacterial consortium amended with poultry litter, coir pith and rhamnolipid biosurfactant. Biores. Technol., 81(1): 25-32.
- Schillinger, U. and Lucke, F. (1989). Antibacterial activities of *Lactobacillus* isolated from meat. Appl. Environ. Microbiol, 55: 1901-1906.
- Schmidt, F. R. (2004). The challenge of multidrug resistance: actual strategies in the development of novel antibacterials. Appl. Microbiol. Biotechnol., 63: 335-343.

- Stachelhaus, T., Schneider, A. and Marahiel, M. A. (1995). Rational design of peptide antibiotics by targeted replacement of bacterial and fungal domains. Sci., 269: 69-72.
- Steller, S., Vollenbroich, D., Leenders, F., Stein, T., Conrad, B., Hofemeister, J., Jacques, P., Thonart, P. and Vater, J. (1999). Structural and functional organization of the fengycin synthetase multienzyme system from *Bacillus subtilis* b213 and A1/3.

Chem. Biol., 6: 31-41.

- Yilmaz, M., Soran, H. and Beyatli, Y. (2006). Antimicrobial activities of some *Bacillus* spp. strains isolated from the soil. Microbiol. Res., 161 (2):127-131.
- Yin, S., Lester, G. S. and Gerrit, V. (1998). Identification of Hydrocarbon-Degrading Bacteria in Soil by Reverse Sample Genome Probing. Appl. Environ. Microbiol., 64(2): 637–645.