# FINGER PRINTING FOR SOME MAIZE INBRED LINES THROUGH RAPD-PCR TECHNIQUE <br> Abd El-Hadi, A. H. ; A. M. El-Adl ; Kawther S. Kash and M.Z.M. El-Diasty <br> Genet. Dept., Fac. of Agric., Mansoura University, Egypt. 


#### Abstract

In this investigation 11 inbred lines of maize were used. These inbred lines were crossed among them to obtain $30 \mathrm{~F}_{1}$ hybrids according to factorial mating design. DNA finger printing was made by using RAPD-PCR for all 11 maize inbred lines. The two primers, XD 8 and XD 9 were used in this technique. The results showed similarity between inbred lines. The results also revealed the presence of three common bands between all inbred lines at primer $\mathrm{XD}_{8}$, while five common bands were obtained at primer XD9. Cluster analysis for similarity degree between the 11 parental inbred lines was done and showed similarity degree between the within inbred lines. Phylsgenetic analysis showed high similarity coefficient between the inbred lines 6, 4 and 2, 3. However, there were high genetic distances obtained between inbred lines No: 4, 5, 6, 7 and 11.


## INTRODUCTION

Mackill et al. (1996) found that the 21 RAPD primers produced 103 bands of which 43 were polymorphic. In the same time, Lanza et al. (1997) evaluated the genetic diversity of 18 maize inbred lines. They also determined the correlation between genetic distance and single-cross hybrid performance. They used RAPDOPCR technique with 32 primers. They indicated that cluster analysis divided the samples to three distanict groups Prevast and Mipkinson (1999) concluded that ISSR-PCR provides a quick, reliable and highly informative system for DNA finger printing. Phillip et al. (2000) used of random amplified polymorphic DNA (RAPD) markers for evaluating seed purity. They stated that genomic DNA isolated from single unregimented seed was found to be suitable for RAPD analysis. They illustrated that the RAPD data showed that the parental lines were not very closely related. Shieh and Thseng (2002) evaluated the genetic diversity of 13 maize inbred lines and determined the correlation between genetic distance and single cross hybrid performance. They employed DNA (RAPD), PCR technique. They indicated that 13 inbred lines of maize could be classified into distinct heterotic groups. There was no significant heterosis values of grain dry weight. McGregor et al. (2006) found that several DNA marker systems and associated techniques are available are available for finger printing plant germplasm. They also investigated that PCR based DNA fingerprinting. They also added that techniques differ in the mean number of profiles generated per primer (or primer pair) per cultivar.

## MATERIALS AND METHODS

Genomic DNA extraction from leaves of mays were conducted according to Laroy and Leon (2000). Amplification of genomic DNA was made on perken Elmer DNA cycler using arbitrary December primers, XD8

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and $X_{9}$ which are presented in Table 1. The genomic DNA amplification using RAPD-PCR were made according Changxin et al. (2003). The similarity between parental inbred lines were determined according to Jaccard's (1908) similarity coefficient as follows:

$$
S_{i j}=2 M /\left(\varepsilon_{i}+\varepsilon_{j}\right)
$$

Where:
$M$; number of matching band.
$\varepsilon_{i}$ : total number of band in the first lan.
$\varepsilon_{j}$ : total number of band in the second lan.
Table 1: Oligonucleotide primers used in the study.

| Primer | Sequence (5` \(\longrightarrow \mathbf{3}^{`}\) ) | GC \% |
| :---: | :---: | :---: |
| $X D_{8}$ | GAAGGCATCC | 60 |
| $X D_{9}$ | GAAGTGGTCC | 60 |

## RESULTS AND DISCUSSION

PAPD-PCR protocol plays a major vole in many of the processes that affect many things. Random primers are used in these reactions and they are very useful. When little or no information is known about the species in plant applications the utilization of RPD-PCR reactions would be useful. The obtained data in Tables 2 and 3 and figure 1 showed that with the primer $\mathrm{XD}_{8}$ the total number of band obtained were 15 bands. The molecular size of these bands ranged from 1541.144 b.p. It could be also regarded that many specific bands appeared in inbred line No. 6 with molecular size of 1541 b.p., 1434 b.p. with line No. 3 and 1421 b.p. with line No. 5 . On the other hand, these common bands were obtained with band No. 5 and the two bands No. 7 and No. 8. These bands had molecular size of 1071, 722 and 734, respectively.

Table 2: RAPD-PCR analysis of DNA polymorphic using $X_{8}$ primer with different Zea mays inbred lines.

| Band <br> number | $\mathbf{b} . \mathbf{p}$. | $\mathbf{1}$ | $\mathbf{2}$ | $\mathbf{3}$ | $\mathbf{4}$ | $\mathbf{5}$ | $\mathbf{6}$ | $\mathbf{7}$ | $\mathbf{8}$ | $\mathbf{9}$ | $\mathbf{1 0}$ | $\mathbf{1 1}$ |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| $\mathbf{1}$ | 1541 | $\bullet$ | $\bullet$ | $\bullet$ | $\bullet$ | $\bullet$ | 1 | $\bullet$ | $\bullet$ | $\bullet$ | $\bullet$ | $\bullet$ |
| $\mathbf{2}$ | 1434 | $\bullet$ | $\bullet$ | 1 | $\bullet$ | $\bullet$ | $\bullet$ | $\bullet$ | $\bullet$ | $\bullet$ | $\bullet$ | $\bullet$ |
| $\mathbf{3}$ | 1421 | $\bullet$ | $\bullet$ | $\bullet$ | $\bullet$ | $\bullet$ | $\bullet$ | 1 | $\bullet$ | $\bullet$ | $\bullet$ | $\bullet$ |
| $\mathbf{4}$ | 1141 | $\bullet$ | $\bullet$ | $\bullet$ | $\bullet$ | 1 | $\bullet$ | 1 | $\bullet$ | $\bullet$ | $\bullet$ | $\bullet$ |
| $\mathbf{5}$ | 1071 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 |
| $\mathbf{6}$ | 1016 | $\bullet$ | 1 | $\bullet$ | $\bullet$ | 1 | $\bullet$ | 1 | $\bullet$ | $\bullet$ | 1 | $\bullet$ |
| $\mathbf{7}$ | 772 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 |
| $\mathbf{8}$ | 734 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 |
| $\mathbf{9}$ | 529 | 1 | $\bullet$ | $\bullet$ | 1 | $\bullet$ | 1 | $\bullet$ | 1 | 1 | $\bullet$ | 1 |
| $\mathbf{1 0}$ | 513 | 1 | 1 | $\bullet$ | 1 | 1 | $\bullet$ | $\bullet$ | 1 | 1 | 1 | 1 |
| $\mathbf{1 1}$ | 449 | $\bullet$ | $\bullet$ | 1 | 1 | $\bullet$ | $\bullet$ | 1 | $\bullet$ | $\bullet$ | 1 | $\bullet$ |
| $\mathbf{1 2}$ | 144 | 1 | 1 | 1 | $\bullet$ | $\bullet$ | $\bullet$ | $\bullet$ | $\bullet$ | 1 | $\bullet$ | 1 |
| Total |  | 6 | 6 | 6 | 6 | 6 | 5 | 7 | 5 | 6 | 6 | 6 |

The results also revealed that the number of bands obtained in each inbred line ranged from 5 to 7 bands. These bands which have molecular size 513,448 and 144 b.p. were present in more inbred lines.

Table 3: RAPD primer PCR analysis with primer $\mathrm{XD}_{8}$.

| Band <br> number | b.p. | 1 | 2 | 3 | 4 | 5 | 6 | 7 | 8 | 9 | 10 | 11 |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| 1 | 1541 | $\bullet$ | $\bullet$ | $\bullet$ | $\bullet$ | $\bullet$ | 1541 | $\bullet$ | $\bullet$ | $\bullet$ | $\bullet$ | $\bullet$ |
| 2 | 1434 | $\bullet$ | $\bullet$ | 1434 | $\bullet$ | $\bullet$ | $\bullet$ | $\bullet$ | $\bullet$ | $\bullet$ | $\bullet$ | $\bullet$ |
| 3 | 1421 | $\bullet$ | $\bullet$ | $\bullet$ | $\bullet$ | $\bullet$ | $\bullet$ | 1421 | $\bullet$ | $\bullet$ | $\bullet$ | $\bullet$ |
| 4 | 1141 | $\bullet$ | $\bullet$ | $\bullet$ | $\bullet$ | 1141 | $\bullet$ | 1141 | $\bullet$ | $\bullet$ | $\bullet$ | $\bullet$ |
| 5 | 1071 | 1071 | 1071 | 1071 | 1071 | 1071 | 1071 | 1071 | 1071 | 1071 | 1071 | 1071 |
| 6 | 1016 | $\bullet$ | 1016 | $\bullet$ | $\bullet$ | 1016 | $\bullet$ | 1016 | $\bullet$ | $\bullet$ | 1016 | $\bullet$ |
| 7 | 772 | 772 | 772 | 772 | 772 | 772 | 772 | 772 | 772 | 772 | 772 | 772 |
| 8 | 734 | 734 | 734 | 734 | 734 | 734 | 734 | 734 | 734 | 734 | 734 | 734 |
| 9 | 529 | 529 | $\bullet$ | $\bullet$ | 529 | $\bullet$ | 529 | $\bullet$ | 529 | 529 | $\bullet$ | 529 |
| 10 | 513 | 513 | 513 | $\bullet$ | 513 | 513 | $\bullet$ | $\bullet$ | 513 | 513 | 513 | 513 |
| 11 | 449 | $\bullet$ | $\bullet$ | 449 | 449 | $\bullet$ | $\bullet$ | 449 | $\bullet$ | $\bullet$ | 449 | $\bullet$ |
| 12 | 144 | 144 | 144 | 144 | $\bullet$ | $\bullet$ | $\bullet$ | $\bullet$ | $\bullet$ | 144 | $\bullet$ | 144 |



Fig. 1: Agarose gel electrophorasis of PCR amplification f eleven maize inbred lines by $\mathrm{XD}_{8}$ primer.

Data in Figure 3 showed a similarity degree between inbred lines using cluster analysis. The obtained similarity degree was 0.36 in the first group, while sub groups had high similarity degree ( $0.6-0.88$ ) as soon as phylsgenetic diversity between the different 11 inbred lines examined by inserting RAPD data into Jaccard's similarity matrix and analysed by (lq A) to gave phylsgenetic tree.

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The obtained results from RAPD-PCR analysis with XD9 primer are present in Tables 4 and 5 and Figure 2. The results showed the presence of high similarity between studied inbred lines. There were 16 bands had a range of size between 685 to 132 b.p. The number of common bands between all inbred lines of Zea mays, L. were three bands: 610, 414 and 393 b.p. The obtained specific bands were 675 b.p. in the inbred lines 2, 7 and 11. In the same time, specific band with molecular size was 245 b.p. for inbred line No. 2. All of these results wee in agreement with the results obtained by Yu and Nyuyen (1994) who detected similar level between different rice cultivars ( $80 \%$ ) of polymorphism in 9 samples gave 260 RAPD fragments. In this respect, Akimato et al. (1994) and Buso et al. (1998) revealed greater variation between four natural collected using isozyme and RAPD-PCR. Phylsgenetic relationship between and within 11 inbred lines of Zea mays, L. led to classify the inbred lines into three major groups depending on the genetic distance between each inbred line and the others.

RAPD analysis using two primers $X_{8}$ and $X_{9}$ were in agreement with Juff et al. (1993) who classified cultivars using cluster analysis of RAPD. The obtained data showed good similarity with their results, where they obtained. Minor variation within subgroups was observed for two types of markers. The results indicated that RAPD-PCR technique may be used for QTL mapping different types of plant.


Fig. 2: Agarose gel electrophorasis of PCR amplification of eleven miaze inbred lines by $\mathrm{DX}_{9}$ primer.

Table 4: RAPD-PCR analysis of DNA polymorphic using $X_{9}$ primer with different Zea mays inbred lines.

| Band <br> number | b.p. | 1 | 2 | 3 | 4 | 5 | 6 | 7 | 8 | 9 | 10 | 11 |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| 1 | 685 | $\bullet$ | 1 | $\bullet$ | $\bullet$ | $\bullet$ | $\bullet$ | 1 | $\bullet$ | $\bullet$ | $\bullet$ | 1 |
| 2 | 684 | $\bullet$ | $\bullet$ | $\bullet$ | 1 | $\bullet$ | $\bullet$ | $\bullet$ | 1 | $\bullet$ | $\bullet$ | 1 |
| 3 | 653 | 1 | 1 | $\bullet$ | $\bullet$ | 1 | $\bullet$ | $\bullet$ | 1 | $\bullet$ | $\bullet$ | $\bullet$ |
| 4 | 624 | $\bullet$ | $\bullet$ | $\bullet$ | $\bullet$ | $\bullet$ | 1 | $\bullet$ | $\bullet$ | $\bullet$ | $\bullet$ | $\bullet$ |
| 5 | 610 | 1 | $\bullet$ | 1 | 1 | 1 | 1 | 1 | $\bullet$ | $\bullet$ | 1 | $\bullet$ |
| 6 | 593 | 1 | 1 | $\bullet$ | $\bullet$ | 1 | 1 | 1 | 1 | $\bullet$ | 1 | 1 |
| 7 | 561 | 1 | 1 | 1 | 1 | 1 | 1 | $\bullet$ | $\bullet$ | 1 | 1 | 1 |
| 8 | 414 | $\bullet$ | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | $\bullet$ | $\bullet$ |
| 9 | 401 | 1 | $\bullet$ | 1 | $\bullet$ | $\bullet$ | 1 | 1 | 1 | 1 | 1 | 1 |
| 10 | 393 | $\bullet$ | 1 | $\bullet$ | 1 | 1 | 1 | 1 | 1 | 1 | 1 | $\bullet$ |
| 11 | 372 | 1 | $\bullet$ | $\bullet$ | 1 | $\bullet$ | 1 | $\bullet$ | $\bullet$ | 1 | $\bullet$ | $\bullet$ |
| 12 | 295 | $\bullet$ | $\bullet$ | 1 | $\bullet$ | 1 | $\bullet$ | $\bullet$ | $\bullet$ | 1 | $\bullet$ | 1 |
| 13 | 280 | $\bullet$ | $\bullet$ | $\bullet$ | $\bullet$ | $\bullet$ | 1 | 1 | $\bullet$ | $\bullet$ | $\bullet$ | $\bullet$ |
| 14 | 254 | $\bullet$ | 1 | $\bullet$ | $\bullet$ | $\bullet$ | $\bullet$ | $\bullet$ | $\bullet$ | $\bullet$ | $\bullet$ | $\bullet$ |
| 15 | 132 | $\bullet$ | 1 | 1 | 1 | 1 | 1 | $\bullet$ | $\bullet$ | 1 | $\bullet$ | $\bullet$ |
| Total | 132 | 6 | 8 | 6 | 7 | 8 | 10 | 7 | 6 | 7 | 5 | 6 |

Table 5: RAPD primer PCR analysis with primer $\mathrm{XD}_{9}$

| Band <br> number | b.p. | 1 | 2 | 3 | 4 | 5 | 6 | 7 | 8 | 9 | 10 | 11 |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| 1 | 1541 | $\bullet$ | $\bullet$ | $\bullet$ | $\bullet$ | $\bullet$ | 1541 | $\bullet$ | $\bullet$ | $\bullet$ | $\bullet$ | $\bullet$ |
| 2 | 1434 | $\bullet$ | $\bullet$ | 1434 | $\bullet$ | $\bullet$ | $\bullet$ | $\bullet$ | $\bullet$ | $\bullet$ | $\bullet$ | $\bullet$ |
| 3 | 1421 | $\bullet$ | $\bullet$ | $\bullet$ | $\bullet$ | $\bullet$ | $\bullet$ | 1421 | $\bullet$ | $\bullet$ | $\bullet$ | $\bullet$ |
| 4 | 1141 | $\bullet$ | $\bullet$ | $\bullet$ | $\bullet$ | 1141 | $\bullet$ | 1141 | $\bullet$ | $\bullet$ | $\bullet$ | $\bullet$ |
| 5 | 1071 | 1071 | 1071 | 1071 | 1071 | 1071 | 1071 | 1071 | 1071 | 1071 | 1071 | 1071 |
| 6 | 1016 | $\bullet$ | 1016 | $\bullet$ | $\bullet$ | 1016 | $\bullet$ | 1016 | $\bullet$ | $\bullet$ | 1016 | $\bullet$ |
| 7 | 772 | 772 | 772 | 772 | 772 | 772 | 772 | 772 | 772 | 772 | 772 | 772 |
| 8 | 734 | 734 | 734 | 734 | 734 | 734 | 734 | 734 | 734 | 734 | 734 | 734 |
| 9 | 529 | 529 | $\bullet$ | $\bullet$ | 529 | $\bullet$ | 529 | $\bullet$ | 529 | 529 | $\bullet$ | 529 |
| 10 | 513 | 513 | 513 | $\bullet$ | 513 | 513 | $\bullet$ | $\bullet$ | 513 | 513 | 513 | 513 |
| 11 | 449 | $\bullet$ | $\bullet$ | 449 | 449 | $\bullet$ | $\bullet$ | 449 | $\bullet$ | $\bullet$ | 449 | $\bullet$ |
| 12 | 144 | 144 | 144 | 144 | $\bullet$ | $\bullet$ | $\bullet$ | $\bullet$ | $\bullet$ | 144 | $\bullet$ | 144 |



Fig. 3: Phylogenetic analysis of different maize inbred lines using PCR.

## REFERECNES

Akimoto, M.; M. Ohara; Y. Shimamoto and H. Morishima (1994). Genetic structure of natural populations of oryza glumaepatula distributed in the Amazon basin. Rice. Res. Newsl., 11: 74-75.
Buso, G.S.C.; P.H. Rangel and M.E. Ferreira (1998). Analysis of genetic variability of South American wild rice populations (Oryza glumaepatula) with iso Zymes and RAPD markers. Mol. Ecol., 7: 107 117.

Chengxin Fu; Q. Yingxiong and K. Hanghui (2003). RAPD analysis for genetic diversity in changium smyrnioides (Apiaceae), an endangered plant. Bot. Bull. Acad. Sin, 44: 13-18.
Jaccard, P. (1908). Nouvelles reserches sur la distribution florale. Bull. Soc. Vaud Sci. Nat., 44: 223 - 270.
Huff, D.R.; R. Peakall and P.E. Smouse (1993). RAPD variation within and among natural populations of out-crossing buffalograss [Buchloe actyloides (Nutt.)]. Theor. Appl. Genet., 86: 927-934.
Lanza, L.L.B.; C.L. de Souza Jr.; L.M.M. Ottoboni; M.L.C. Vieira and A.P. de Souza (1997). Genetic distance of inbred lines and prediction of maize single-cross performance using RAPD markers. J. TAG Theoretical and Applied Gene., 94(8): 1023 - 1030.

Leroy, X.J. and K. Leon (2000). A rapid method for detection of plant genomic instability using unanchored-microsatellite primers. Plant Molecular Reporter 18: 283a-283g.
Mackill, D.J.; Z. Zhang; E.D. Redona and P.M. Colowit (1996). Level of polymorphism and genetic mapping of AFLP markers in rice. J. Genome, 39: 969-977.
McGregor, C.E.; C.A. Lambert; M.M. Greyling; J.H. Louw and L. Warnich (2006). A Comparative assessment of DNA fingerprinting techniques (RAPD, ISSR, AFLP and SSR) in tetraploid potato (Solanum tuberosum, L.) germplasm. J. Euphytica. 113(2): 135-144.
Phillip, A.C.; P.L. Bhalla, C.K. Lee and M.B. Singh (2000). RAPD analysis of seed parity in a commercial hybrid cabbage (Brassica Oleracea var. Capitata) Cultivar. J. Genome, 43: 317-321.
Prevost, A. and M.J. Wilkinson (1999). A new system of comparing PCR primers applied to ISSR fingerprinting of potato cultivars. J. TAG Theoretical and Applied Genetics. 98(1): 107 - 112.
Shieh and Thseng (2002Genetic diversity of Tainal-white maize inbred lines and prediction of single cross hybrid performance using RAPD markers.Euphytica 124 :307-313.
Yu, L.X. and H.T. Nguyen (1994). Genetic variation detected with RAPD markers among upland and lowland rice cultivars (Oryza sativa, L.). Theor. Appl. Genet., 87: 668-672.

RAPD- البصمة الراثية لبصض السلالات النقية فى اللزرة الثـامية باستخدام تكنيك PCR

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تـم عمـل تحليـل البصـمة الور اثيـة لعـدد 1 ( ســلالة مـن الــرة الثــامية باسـتخدام تكنيـك وباستخدام عدد ז برمبير RDPD-PCR
-G/C \% ${ }^{7}$.

- أظهرت النتائج وجود درجـة عاليـة من التماثل بين جميع السـلاتات التى تم استخدامها فى هذه -الدراست
- أظهرت النتائج وجود
-XD9 Primer استخدام
- تم تحليل درجة التماتُل بين السـلاتات المستخدمة فـى الدر اسـة باستخدام التحليل العنقودى لجميع

السلالات، وقد أظهرت النتائج ارتفاع درجة النماتُل بين معظم السلالات المستخدم فى الدراسة.
ـ أظهرت نتائج التحليل الثهرى لدرجة القرابة بين السلاتلات المستخدمة بقسمة السلات إلى ثـلاث
مجمو عات درجة القر ابة.

r، r مع السلالات ع، 0، r، V، I

