



Egyptian Journal of Animal Health

P-ISSN: 2735-4938 On Line-ISSN: 2735-4946
Journal homepage: <https://ejah.journals.ekb.eg/>

Antibacterial effect of sodium lactate and thymol on *Salmonella Typhimurium* in fish fillet

Taghreed H. Abbas, Shereen A.Yassin and Manal M. Said

Food Hygiene Department, Animal Health Research Institute, Dokki

Article History

Received in 15/12/2020
Received in revised from 7/2/2021
Accepted in 25/2/2021

Keywords:

Sodium lactate-thymolfish fillet
Salmonella typhimurium

ABSTRACT

The aim of this study was to determine the antibacterial effect of sodium lactate and thymol against *Salmonella Typhimurium* in fish fillet. Fish fillet samples were artificially contaminated with *Salmonella Typhimurium*, then sprayed with thymol (0.1%), sodium lactate (1% & 2%) individually and combination of Sodium lactate (1% & 2%) + thymol (0.1%). The samples were stored at 4°C and analyzed at zero, third and fifth day of storage for sensory analysis, pH, and count of *Salmonella Typhimurium*. The thymol alone had a little antibacterial effect on Salmonella. The initial count of control was $5.65 \pm 0.09 \log_{10}$ cfu/g on zero day which was declined to $4.50 \pm 0.14 \log_{10}$ cfu/g as a result of combined treatment of sodium lactate (2%) + thymol 0.1%, which showed the best results obtained, followed by the treated samples with sodium lactate 1% + thymol 0.1%, as it recorded $4.72 \pm 0.10 \log_{10}$ cfu/g, and lastly $4.63 \pm 0.11 \log_{10}$ cfu/g, which obtained by sodium lactate 2% treated samples. The reduction in the Salmonella count in response to the combined treatment of sodium lactate 2% + thymol 0.1% was greater than the effects of the two compounds applied individually.

INTRODUCTION

Food safety and hygienic standard parameters are the most important issues related to human health in the food industry. Food safety concerns each and every food handling facility; therefore many factors that make food unsafe for consumption. The hazards of food are primarily microbiological agents (Morya *et al.* 2020). As microbial food safety is an increasing public health concern worldwide, foodborne pathogens are a major threat to food safety, especially in developing countries where food hygiene and sanitation facilities are

often poor (Ahmed and Shimamoto 2014). Fish meat is known to be susceptible to undesirable chemical changes and microbial reactions that characterize spoilage (Raeisi *et al.* 2020). During storage the quality of fish degrades due to a complex process in which physical, chemical and microbiological forms of deterioration are implicated (Gonza'lez *et al.* 2005).

Many foodborne outbreaks have been recorded worldwide, and about 30 percent of the population in developed nations is suffering from foodborne disease every year. Foodborne

*Corresponding author: Taghreed Abbas, Food Hygiene Department, Animal Health Research Institute, Dokki
E-mail: taghreed7amdy@gmail.com
DOI: 10.21608/EJAH.2021.166610

diseases are generated due to consumption of contaminated food with the pathogenic micro-organism during the food chain (production, handling, storage, transport and distribution) (Badui 2015 and Junod *et al.* 2015).

Salmonella bacteria have become the major cause of foodborne diseases which have raised a great safety concern to public health (Rabsch *et al.* 2001). *Salmonella typhimurium* is among the most common serovars in the outbreaks of *Salmonella* gastroenteritis. These pathogens are typically associated with infected food and could result in a major economic effect due to illness (Amagliani *et al.* 2012). While most infections cause moderate symptoms to severe death-related infections do occur.

The use of chemical preservatives to inactivate or hinder the growth of pathogenic micro-organisms in foodstuffs is growing fast. Massive numbers of people prefer food items which are processed naturally (Tajkarimi *et al.* 2010). To control foodborne pathogens without affecting quality, foods are subjected to combined treatments using "hurdle technology" which focused on natural antimicrobial food additives (Kim and Rhee 2016, Leistner 2000).

Sodium lactate is the sodium salt of natural lactic acid, it is approved by the Food and Drug Administration (FDA) as widely accepted as healthy, Generally Recognized As Safe (GRAS) and directly applied to various foods to regulate microbial growth and prolong product shelf life (Hwang *et al.* 2011). Studies have shown that lactates exhibit bacteriostatic effects by affecting the bacteria's metabolism including intercellular acidification and proton transfer across the cell membrane. Further, lactates decrease food water activity (Seydim *et al.* 2006).

Sodium lactate can be added directly to food to control the microbial growth and to extend the shelf life of food products (Burfoot and Mulvey 2011, Smaoui *et al.* 2012, Bolton *et al.* 2014).

Herbs and spices, and their constituents have been used as flavoring agents in food since the earliest history, and it is well established that many have antibacterial activity (Bajpai *et al.* 2012 Jayasena and Jo 2013 Prakash *et al.*

2015, Seow *et al.* 2015). Thymol is one of the phenolic compounds obtained from plants like *Origanum vulgare* and *Thymus vulgaris*, and is classified as (GRAS) by (FDA) (Tajkarimi *et al.* 2010, Hyldgaard *et al.* 2012). Thymol as one of the major constituents of thyme oil presents a wide range of functional possibilities in the food industry (Placha *et al.* 2019, Salehi *et al.* 2018). It is believed that the mode of antibacterial action of thymol alters the physical and chemical properties of cytoplasmic membrane of bacteria, and this may change the permeability of the cell membrane and cause the leakage of ions and other cell contents (Hyldgaard *et al.* 2012 Vergis *et al.* 2015, Calo *et al.* 2015). However, their use as preservatives in foods is often limited due to considerations of flavor and aroma and loss of sensory quality. Therefore, combinations of thymol with other preservatives have been used to minimize the concentrations required. There has been conducted much successful research related to the combinations of thymol with the other antimicrobial agents and methods (Oladunjoye *et al.* 2013; Ilhak and Guran, 2014, Kim and Rhee, 2016).

Considering the antibacterial effect of thymol, it can be assumed that thymol can facilitate the diffusion of sodium lactate into the cytoplasm of the cells. In this way, when the two are used in combination a synergistic or additional effect can be achieved (Ilhak and Guran 2014).

Therefore, the main purpose of the present study was to investigate experimentally the combined antibacterial effect of thymol and sodium lactate on *Salmonella Typhimurium* in fish fillets.

MATERIALS AND METHODS

Collection of samples:

One sample weighted 3000g. of fish fillet was randomly purchased from local markets in Giza governorate in Egypt. The sample was transported to the laboratory of Food Hygiene Department, Animal Health Research Institute (AHRI) as quickly as possible under complete aseptic conditions in ice box. A representative sample was taken to be analyzed for the indigenous *Salmonella* spp. according to (ISO 2002).

Bacterial culture:

Salmonella Typhimurium strain (ATCC® 14028) was provided by food hygiene department, Animal Health Research Institute. The strain was grown in 10 mL of Tryptic soy broth (TSB) (Biomark Laboratories) at 37°C for 18 hrs. One milliliter of the strain was then harvested by centrifugation at 4,192 ×g for 10 min, and washed and serially diluted in 0.1% peptone water (PW) to yield cell suspensions of 10⁷ CFU/ml. The bacterial population was determined by plating 0.1 ml of appropriately diluted culture on Xylose lysine deoxycholate agar plate (XLD) (Merk) with incubation at 35°C for 24–36 h. Active culture with bacterial density of 1x10⁷ cfu/ml was used in samples inoculations (Health Protection Agency, 2005).

Inoculation of fish fillet, and treatment:

The fish fillets meat sample was prepared for inoculation and dipped in peptone water containing 1x10⁷ cfu/ml *Salmonella Typhimurium* strain for 15 minutes at room temperature (25°C). After the inoculation, the fish fillet samples were kept at refrigerator for 10 min to allow for bacterial cell attachment. The inoculated Fish fillets with known *Salmonella Typhimurium* load was divided into six equal groups (500 g each) for further treatment. The decontamination treatments were as follows; 1st. control group remained without treatment, 2nd. group was treated with thymol 0.1% solution, 3rd. group was treated with sodium lactate 1% solution, 4th. group was treated with sodium lactate 2% solution, 5th. group was treated with sodium lactate 1% + thymol 0.1% solution and 6th. group was treated with sodium lactate 2% + thymol 0.1% solution.

The groups (except control) sprayed separately (rotating all surface) with approximately 30 ml of solutions sodium lactate 1%, 2 % (Sigma -Aldrich /P code 101044503 and thymol (Lanxess SLC2068, colorless crystals, solubility in water 0.1gm/100ml, Germany) using a spray bottles. Bottles of thymol and sodium lactate 1 and 2% were vigorously shaken to be dispersed in the solution before they were used. After spraying, the inoculated fish fillets samples were separately packed in multiple sterile labeled polyethylene bags, heat sealed,

and stored at refrigerator of 2 - 4°C for 5 days. Sensory, pH and bacteriological analysis were conducted following 0, 3, and 5 days of storage.

Sensory analysis:

Color, odor, texture and overall acceptability were conducted on days 0, 3, and 5 during storage to determine the best treatment. Sensory analysis was performed by a sensory panel team of seven trained assessors (members of food hygiene department. Their opinion was recorded as acceptable or non-acceptable.

Determination of pH of fish fillets samples:

pH of the rinse solution of the samples was performed after bacteriological analysis on each sampling day. It was measured with pH meter (JENEWAY 3310).

Determination of *Salmonella typhimurium* count:

Bacteriological sampling was performed after the spraying treatment on 0, 3 and 5 day. On each sampling day, 25 g. of the fish meat samples were weighed out under aseptic conditions, placed in sterile “Stomacher” bags with 225ml of 0.1% sterile Buffer peptone water (BP) and homogenized for two minutes. 1 ml solution was taken from the solution and serially diluted in 0.1% sterile peptone water and surface plated on xylose lysine deoxycholate agar for enumeration of *Salmonella*. Characteristic colonies were counted after the plates were incubated at 35°C for 24–36 hrs. (ISO 2002).

Statistical Analyses:

Analyses of the microbiological data were carried out using Statistical Package for the Social Sciences (SPSS). One Way ANOVA, The numbers of bacteria were converted to logarithmic values before calculating means and performing statistical analyses. The means were separated using Fisher’s least significant differences according to general linear model procedures. Statistical significant level was expressed as (P ≤ 0.05).

RESULTS

Table (1) Sensory evaluation of fish fillet samples:

Treatment \ Items	control	Thymol 0.1%	Sod lact 1%	Sod lact 2%	Sod lact 1% + Thymol 0.1%	Sod lact 2% + Thymol 0.1%
Color	8	8	8	8	8	8
Odor	7	6	8	8	8	8
Texture	7	8	7	7	8	8
Over all acceptability	7	7	8	8	8	8

Ratings: (scale 10= excellent, 1= unacceptable).

Table (2) Effect of sodium lactate and thymol on pH (mean±SD) values of fish fillet

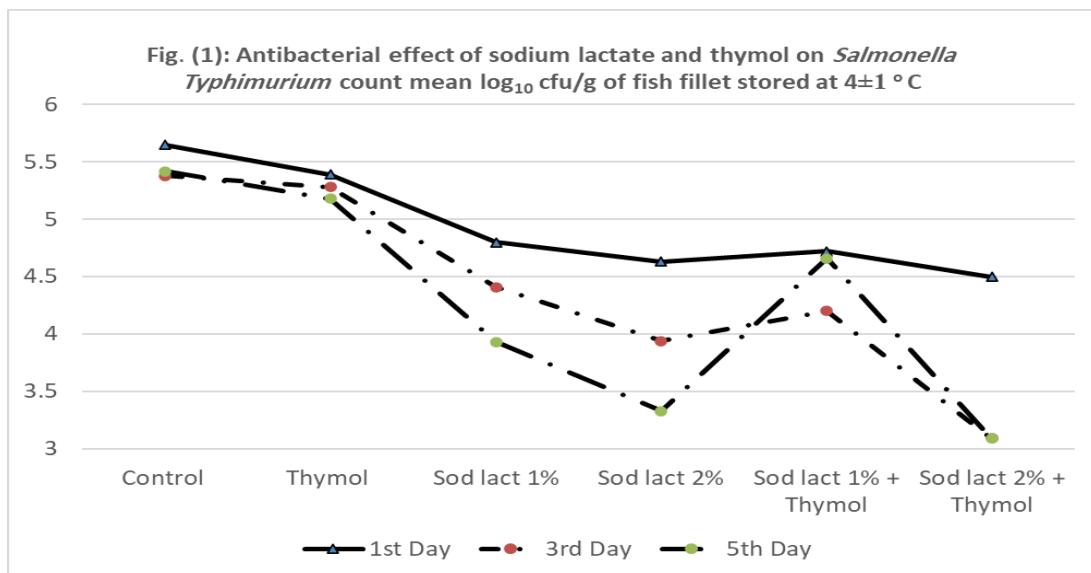
Treatment	Zero Day	3 rd Day	5 th Day
Control			
Thymol 0.1%	6.50 ^a ±0.05	6.60 ^a ±0.02	6.62 ^a ±0.01
Sod lact 1%	6.43 ^b ±0.01	6.41 ^b ±0.01	6.43 ^b ±0.03
Sod lact 2%	6.39 ^b ±0.06	6.41 ^b ±0.03	6.43 ^b ±0.01
Sod lact 1% + Thymol 0.1%	6.31 ^c ±0.03	6.38 ^b ±0.02	6.39 ^b ±0.02
Sod lact 2% + Thymol 0.1%	6.28 ^c ±0.02	6.31 ^c ±0.02	6.32 ^c ±0.03
	6.20 ^c ±0.00	6.30 ^c ±0.01	6.35 ^c ±0.02

There are significance differences ($P < 0.05$) between mean having different super scribed letter in the same day of storage for the same column).

Table (3) Antibacterial effect of sodium lactate and thymol on *Salmonella Typhimurium* Count (mean ± SD log₁₀ cfu/g) of fish fillet stored at 4 ± 1 °C.

Treatment	zero Day	3 rd Day	5 th Day
Control			
Thymol 0.1%	5.65 ^a ± 0.09	5.67 ^a ± 0.15	5.68 ^a ± 0.12
Sod lact 1%	5.39 ^b ± 0.23	5.28 ^a ± 0.05	5.18 ^a ± 0.16
Sod lact 2%	4.80 ^c ± 0.10	4.41 ^b ± 0.09	3.93 ^b ± 0.05
Sod lact 1% + Thymol 0.1%	4.63 ^{cd} ± 0.11	3.94 ^c ± 0.05	3.33 ^c ± 0.35
Sod lact 2% + Thymol 0.1%	4.72 ^{cd} ± 0.10	4.20 ^d ± 0.08	4.16 ^d ± 0.13
	4.50 ^e ± 0.14	3.09 ^e ± 0.16	3.09 ^e ± 0.16

There are significant differences (P<0.05) between mean having different super scribed letter in the same day of storage for the same column).



DISCUSSION

In the present study, the antibacterial effect of sodium lactate and thymol on *Salmonella typhimurium* count was measured to evaluate the most effective compound and also its effect on sensory quality and pH.

In **Table (1)**, the sensory evaluation revealed that only minor differences were noticed for color, odor, texture and overall acceptability of the fish fillet samples. Odor slightly changed in samples which treated with thymol 0.1%. All the treated samples were considered to be accepted by the panelists. **Burt 2004 and Tajkarimi et al. (2010)** mentioned that fish fillet samples which treated with thymol in combi-

nation with sodium lactate 1% and 2% were accepted, which agreed with the results of the present study.

Table (2), revealed that the pH mean values of fish fillet samples were increased during storage at 4°C for control and all treated samples. The pH of the control sample was 6.50 and increased to 6.60 and 6.62 on 3rd and 5th days. The results obtained for thymol 0.1%, sodium lactate 1%, sodium lactates 2%, sodium lactate 1% +thymol and for sodium lactate 2%+ thymol were (6.43, 6.39, 6.31, 6.28 and 6.20) on zero day ,(6.41, 6.41,6.38 , 6.31, and 6.30) on 3rd day and finally the 5th day results recorded (6.43,6.43,6.39,6.32, and 6.35), re-

spectively. The results obtained were agreed with Zhou *et al.* 2007a who mentioned that sodium lactate 2% decreased the pH more than sodium lactate 1%, also it was revealed that the inhibitory effect of sodium lactate increases when the pH of food decreased (Nykänen *et al.* 2000 and Mani-López *et al.* 2012). Tiwari *et al.* 2009 reported that the susceptibility of bacteria to the antibacterial effect of essential oil (thymol) increased with the decrease in pH level of food. There was a significant difference ($p < 0.05$) between control group and the other treatments as pH was shown to be reduced.

Table (3) and figure (1), showed that the mean value of *Salmonella Typhimurium* count recorded for control samples were ($\log 5.65 \pm 0.09$, 5.67 ± 0.15 , 5.68 ± 0.12 cfu/g), while thymol results were ($\log 5.39 \pm 0.23$, 5.28 ± 0.05 , 5.18 ± 0.16 cfu/g) on zero, 3rd and 5th day, respectively. On finding obtained, the samples which treated with thymol 0.1% showed little decrease in the *Salmonella* count and it was similar to Tiwari *et al.* 2009. There was a significant difference between the control, and thymol 0.1% ($p < 0.05$).

Treated samples with sodium lactate 1% showed results of ($\log 4.80 \pm 0.10$, 4.41 ± 0.09 , 3.93 ± 0.05 cfu/g) and sodium lactate 2% recorded ($\log 4.63 \pm 0.11$, 3.94 ± 0.05 , 3.33 ± 0.35 cfu/g). Sodium lactate 1% and 2% significantly reduced the count of *Salmonella* than control and thymol ($p < 0.05$).

Sodium lactate 1% + thymol 0.1% treatment recorded ($\log 4.72 \pm 0.10$, 4.20 ± 0.08 , 4.16 ± 0.13) and finally Sod lactate 2% + thymol 0.1% showed ($\log 4.50 \pm 0.14$, 3.09 ± 0.16 , 3.09 ± 0.16) on zero, 3rd and 5th day, respectively. The obtained results revealed that combination of sodium lactate 1, 2% and thymol gave the best inhibitory effect on *Salmonella* count, while the concentration of sodium lactate 2% combined with thymol 0.1% was the most effective one. The same results were obtained by (Ilhak and Guran 2014), who reported that the antibacterial effect of sodium lactate 2% on *Salmonella typhimurium* could be increased when combined with thymol. Meanwhile, no significance difference between treatments including sodium lactate 2%, sod lactate 1%

+thymol and sod lactate 2%+ thymol. Sodium lactate 2%+thymol 0.1% significantly ($p < 0.05$) reduced *Salmonella* count than control (from 5.65 to 4.63); the reduction was more than 1log cfu/g.

CONCLUSION

In the current study, antibacterial compounds, sodium lactate in concentration of 1, 2% and thymol 0.1% had an inhibitory effect on *Salmonella* count. The best results were obtained when using sodium lactate 2% in combination with thymol 0.1% as antibacterial on *Salmonella Typhimurium*, while such used concentration in the current study did not inhibit the organism completely so it is recommended to use a higher concentration and at the same time did not affect the sensory characters of the product.

REFERENCES

- Ahmed AM, Shimamoto T. 2014. Isolation and molecular characterization of *Salmonella enterica*, *Escherichia coli* O157:H7 and *Shigella* spp. from meat and dairy products in Egypt. *Int. J Food Microbiol*, 168-169 (3): 57-62 doi:10.1016/j.foodmicro.2013.10.014.
- Amagliani G, Brandi G, Schiavano GF. 2012. Incidence and role of *Salmonella* in seafood safety. *Food Res. Int.* 45(2): 780-788. <https://doi.org/10.1016/j.foodres.2011.06.022>.
- Badui DS. 2015. Inocuidad en la industria alimentaria. *Industria Alimentaria* 14-25.
- Bajpai VK, Baek KH, Kang SC. 2012. Control of *Salmonella* in foods by using essential oils: A review. *Food Res. Int.* 45(2): 722-734. doi:10.1016/j.foodres.2011.04.052.
- Bolton DJ, Meredith H, Walsh D, McDowel DA. 2014. The effect of chemical treatments in laboratory and broiler plant studies on the microbial status and shelf-life of poultry. *Food Control.* 36(1): 230-237. doi: 10.1016/J Food Cont.2013.08.027.
- Burfoot D, Mulvey E. 2011. Reducing microbial counts on chicken and turkey carcasses using lactic acid. *Food Control*, 22(11): 1729-1735. doi: 10.1016/j.foodcont.2011.04.005.
- Calo JR, Crandall PG, O, Bryan CA, Ricke SC. 2015. Essential oils as antimicrobials in food systems – a review. *Food Con.* 54: 111-

119. Doi:10.1016/j.foodcont.2014.12.040.
- González F, Villarino R, García L, García A, García F. 2005. Microbiological safety and sensory characteristics of salmon slices processed by the sous vide method. *Food Cont.* 16(1): 77–85. Doi: 10.1016/j.foodcont.2003.11.011.
- Health Protection Agency 2005. Recommended Minimum Internal Quality control in food microbiology testing Laboratories. National standard Method QSOP 18 Issue 4. <http://www.hpa-standard-methods.org.uk/pdf>.
- Hwang CA, Sheen S, Jneja V. 2011. Effects of sodium lactate on the survival of *Listeria monocytogenes*, *Escherichia coli* O157:H7 and *Salmonella* spp. in cooked ham at refrigerated and abuse temperatures. *Food Nutr. Sci.* 2(5): 464–470. doi:10.4236/FNS. 2011. 25066.
- Hyltdgaard M, Mygind T, Meyer RL. 2012. Essential oils in food preservation: mode of action, synergies, and interactions with food matrix components. *Front. Microbiol.*, 3 (12): 1–24. Doi:10.3389/fmicb.2012.00012.
- Ilhak OI, Guran HS. 2014. Combined antimicrobial effect of thymol and Sodium lactate against *Listeria monocytogenes* and *Salmonella typhimurium* in fish patty. *J. of Food Safety*, 34(3): 211–217. doi: 10.1111/jfs.12115.
- ISO 6579. 2002. Microbiology of food and animal feeding stuffs -- Horizontal method for the detection of *Salmonella* spp. www.iso.org.
- Jayasena DD, Jo C. 2013. Essential oils as potential antimicrobial agents in meat and meat products: A review. *Trends Food Sci. Tech.* 34(2):96–108. doi:10.1016/j.tifs.2013.09.002.
- Junod T, Lopez Martin J, Gädicke P. 2013. Antimicrobial susceptibility of animal and food isolates of *Salmonella enterica*. *Revista Medica de Chile* 14(3): 298-304. Doi:10.4067/s0034-98872013000300003.
- Kim SA, Rhee MS. 2016. Highly enhanced bactericidal effects of medium chain fatty acids (caprylic, capric, and lauric acid) combined with edible plant essential oils (carvacrol, eugenol, β -resorcylic acid, cinnamaldehyde, thymol, and vanillin) against *Escherichia coli* O157:H7. *Food Cont.* 60: 447-454. doi:10.1016/j.foodcont.2015.08.022.
- Leistner L. 2000. Basic aspects of food preservation by hurdle technology. *inter. J. of food Micro.*, 55 (1-3): 181-186. Doi:10.1016/S0168-1605(00)00161-6.
- Mani-lópez E, Garcia, HS, López-malo A. 2012. Organic acids as antimicrobials to control *Salmonella* in meat and poultry products. *Food Res. Int.*, 45(2):713-721. Doi:10.1016/j.foodres.2011.04.043.
- Morya S, Amoah AD, Snaebjornsson SO. 2020. food poisoning hazards and their consequences over food safety. *Microorganisms for Sustainable Environment and Health*, 383-400. Doi:10.1016/B978-0-12-819001-2.00019-X.
- Nykänen A, Weckman K, Lapveteläinen. 2000. Synergistic inhibition of *Listeria monocytogenes* on cold-smoked rainbow trout by nisin and sodium lactate. *Int. J. Food Microbiol.*, 61(1): 63–72. Doi:10.1016/s0168-1605(00)00368-8.
- Oladunjoye A, Soni KA, Nannapaneni R, Schilling MW, Silva JL, Mikell B, Bailey RH, Mahmoud BSM, Sharma CS. 2013. Synergistic activity between lauric acid and carvacol in reducing *Salmonella* in ground turkey. *Poultry Sci.*, 92(5): 1357–1365. doi:10.3382/PS.2012-02620.
- Placha I, Ocelova V, Chizzola R, Battelli G, Gai F, Bacova K, Faix S. 2019. Effect of thymol on the broiler chicken antioxidant defense system after sustained dietary thyme oil application. *Br. Poult. Sci.* 60 (5):589-596. doi:10.1080/00071668.2019.1631445.
- Prakash B, Kedia A, Mishra P, Dubey NK. 2015. Plant essential oils as food preservatives to control molds, mycotoxin contamination and oxidative deterioration of agri-food commodities – potentials and challenges. *Food Control*, 47: 381–391.
- Raeisi M, Hashemi M, Aminzare M, Bidkopeh FG, Ebrahimi M. 2020. Effect of sodium alginate and chitosan coating combined with three different essential oils on microbial and chemical attributes of rainbow trout fillets. *J. of Aquatic Food Product*, 253-263. Doi:10.1016/j.foodcont.2014.07.023.

- Rabsch W, Tschäpe H, Andreas J, Bäuml A. 2001. Review, non-typhoidal salmonellosis: Emerging problems. *Microbes and Infection*, 3(3):237–247. doi:10.1016/s1286-4579(01)01375-2.
- Salehi B, Mishra AP, Shukla I, Sharifi-Rad M, Contreras, MDM, Segura-Carretero A, Fathi H, Nasrabadi NN, Kobarfard F, Sharifi-Rad J. 2018. Thymol, thyme, and other plant sources: Health and potential uses. *Phytother. Res.*, 32(9): 1688–1706. Doi:10.1002/ptr.6109. Epub 2018 May 22.
- Seow YX, Yeo CR, Chung HL, Yu, k HG. 2015. Plant essential oils as active microbial agents. *Crit. Rev. Food Sci. Nutr.* 54(5): 625–644. doi:10.1080/10408398.2011.599504.
- Seydim AC, GÜzel-Seydim ZB, Acton JC, Dawson PL. 2006. Effects of rosemary extract and sodium lactate on quality of vacuum-packaged ground ostrich meat. *J. Food Sci.*, 71(1):71–76. doi:10.1111/j.1365-2621.2006.tb12409.x
- Smaoui S, Hlima HB, Ghorbel R. 2012. The effect of sodium lactate and lactic acid combinations on the microbial, sensory and chemical attributes of marinated chicken thigh. *Poultry Sci.*, 91(6): 1473–1481. doi:10.3382/ps.2011-01641.
- Tajkarimi MM, Ibrahim SA, Cliver DO. 2010. Antimicrobial herb and spice compounds in food. *Food Cont.* 21(9):1199–1218. doi: 10.1016/j.foodcont.2010.02.003.
- Tiwari BK, Valdramidis VP, O'donnell CP, Muthukumarappan K, Bourke P, Cullen PJ. 2009. Application of natural antimicrobials for food preservation. *J. Agric. Food Chem.*, 57(14): 5987–6000. doi:10.1021/jf900668n.
- Vergis J, Gokulakrishnan P, Agarwal RK, Kumar A. 2015. Essential oils as natural food antimicrobial agents: A review. *Critical Reviews in Food Science and Nutrition*, 55(10):1320-1323. doi:10.1080/10408398.2012.692127.
- Zhou F, Ji B, Zhang H, Jiang H, Yang Z, Li J, Ren Y, Yan W. 2007. Synergistic effect of thymol and carvacol combined with chelators and organic acids against *Salmonella* Typhimurium. *J. Food Prot.*, 70(7): 1704–1709. Doi:10.4315/0362-028x-70.7.1704.
-