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Nutritional Characterizations of Tamarind (*Tamarindusindica*, L.) Pulp Fruits

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Abstract

Chemical composition, minerals, amino acids, fatty acid characteristics and quality to their contents of phytochemicals were determined in tamarind (*Tamarindusindica*, L.) fruits pulp. The results showed that moisture; protein, fat, ash, fiber, carbohydrates and energy value as wet weight of tamarind fruits were 34.25, 6.01, 4.34, 2.27, 4.13, 49.00 % and 259.10 Kcal/100g, respectively. The highest mineral contents of tamarind recorded for calcium, sodium and phosphorus, while the lowest recorded for lead, nickel and zinc. The highest essential amino acids (EAA) content of tamarind fruit pulp recorded for leucine, while the lowest EAA recorded for tryptophan, cysteine and methionine. The highest fatty acids contents of tamarind fruit pulp recorded for linoleic, oleic and palmitic. The values were 3.42, 2.29 and 1.80 $\mu\text{g/g}$, respectively. Total phenols and antioxidants activity (DPPH) content were 21.36 mg/g and 67.90 %, respectively. The highest phenolic compounds of tamarind recorded for pyrogallol, catechin and benzoic acid. The values were 2588.84, 527.55 and 365.85 mg/kg, respectively. On the other hand, the lowest value recorded for cinnamic acid, resveratrol and coumarin. The highest flavonoid compounds of tamarind recorded for luteo.6-arbinose 8-glucose, luteo.6-arbinose 8-arbinose and Apig. 6- glucose 8- rhamnose, while, the lowest value of flavonoid compounds recorded for kampferol, apeginin and rhamnetin. As conclusion, tamarind fruits had high nutritional values due to its contents amino acids, fatty acid, minerals and phenolic compounds, it could be used for improvement human health against some diseases.

Key words: Tamarind fruits, chemical composition, phytochemicals.

Introduction

El-Siddig et al., (1999) reported that tamarind (*Tamarindusindica*, L) of the *Fabaceae*, subfamily *Caesalpinioideae*, is an important food in the tropics. It is a multipurpose tree of which almost every part finds at least some usefulness, either nutritional or medicinal. Tamarind is indigenous to tropical Africa but it has been introduced and naturalized worldwide in over 50 countries. The major production areas are in the Asian countries India and Thailand, but also in Bangladesh, Sri Lanka, Thailand and Indonesia. America, Mexico and Costa Rica are not only the biggest producers. Africa on the whole does not produce tamarind on a commercial scale, though it is widely used by the local people. Minor producing countries in Africa are Senegal, Gambia, Kenya, Tanzania and Zambia.

Tamarind pulp typically contains 20.6% water, 3.1% protein, 0.4% fat, 70.8% carbohydrates, 3.0% fiber and 2.1% ash, thus the pulp has low water content and a high level of protein, carbohydrates and minerals. Nevertheless, the proximate composition of the tamarind fruit depends on locality (**Nordeide et al., 1996**).

Tamarind fruit pulp is used for seasoning, as a food component, to flavour confections, curries and sauces, and is a main component in juices and certain beverages. Tamarind fruit pulp is eaten fresh and often made into a juice, infusion or brine, and can also be processed into jam and sweets. The refreshing drinks are popular in many countries around the world, though there are many different recipes (**El-Siddig et al., 2006**).

Pods contain 1-10 seeds, which are irregularly shaped, flattened or rhomboid. Seeds are very hard, shiny, reddish, or purplish brown. They are embedded in the pulp, lined with a tough parchment resembling a membrane, and joined to each other with tough fibers. There are great differences and variations in fruit size and flavour (**Kumar and Bhattacharya, 2008**).

Fatty acid profile of tamarind fruit pulp is relatively poor in oil, greenish yellow in colour and liquid at room temperature. Saponification values of the oil are high, indicating that it contains a high proportion of low molecular weight fatty acids. With regard to the two essential fatty acids, the fruit pulp contains very little linoleic acid (3.42 mg/g dry

weight) and even lower amounts of α -linolenic acid (0.21 mg/g dry weight) (Glew *et al.*, 2005).

Almeida *et al.*, (2009), reported that the consumption of 100 g tamarind fruit pulp by an adult will cover 10.69% of the recommended daily intake of calcium, 20.49% of magnesium, 14.21% of phosphorous, 12.07% of iron, 2.61% of manganese, 1.29% of zinc, 32.22% of copper and 9.21% of selenium, respectively.

Hefnawf and Youssef (1985) mentioned that meat as the main component of beefburger is a suitable medium for the growth of microorganisms starting to be contaminated in the slaughtering house till its manufacture in the beefburger factory. The microbial activity leads to certain changes in either flavor, or color and accumulation of toxins in meats.

During storage, quality attributes of the product deteriorate due to lipid oxidation and microbial growth. Lipids oxidation is responsible for reduction in nutritional quality as well as changes in flavor, while microbial contamination can precipitate major public health hazards and economic loss in terms of food poisoning and meat spoilage. Thus, the application of suitable agents and possessing showed that stopping both antioxidant and antimicrobial activities may be useful for maintaining meat quality, extending shelf-life and preventing economic loss (Yin and Cheng, 2003).

The development of rancidity rapidly occurred especially when the products are exposed to air and cooked in frying oil. In addition to the undesirable quality, the adverse effect of lipid oxidation leads to the development of free radicals which are involved in diseases and a range of disorders including cancer, arthritis, atherosclerosis, Alzheimer's disease, and diabetes. The supplement of synthetic antioxidants is a method of inhibiting lipid oxidation in meat products (Baydar *et al.*, 2007).

The lipid oxidation is one of the major problems in meat industries. Meat products that are constituted of lipid and polyunsaturated fatty acids (PUFAs) tend to deteriorate due to lipid oxidation, leading to development of unpleasant flavors during processing and storage (Mielinket *et al.*, 2008).

Some plant extracts from spices and herbs, however are excellent sources of natural antioxidants that can improve meat shelf-life and

quality mainly by retarding lipid oxidation and microbial growth (Velasco and Williams, 2011).

This work was conducted to study the chemical composition, minerals, amino acids, fatty acid characteristics and phytochemicals quality of tamarind fruit pulp.

Materials And Methods

Materials

The fresh fruit of tamarind (*Tamarindusindica*, L.) was obtained from Herbalist, transferred frozen and stored at -18°C until analysis and processing.

Chemicals

Folin-Ciocalteu reagent and standard substances including gallic acid, sinapic acid, caffeic acid, chlorogenic acid, *p*-coumaric acid and dihydroxy benzoic acid were purchased from SigmaChemical Company (St. Louis, USA), vanillic acid, ferrulic acid, rutin and quercetin from Fluka St. Gallen, Switzerland. All reagents and standards were prepared using Milli-Q deionized water (Millipore, Bedford, USA). All other chemicals and reagents were of analytical reagent grade and purchased from Al-Ghomhoria Company, Egypt.

Methods

Preparation of tamarindfruits

A part of the fresh tamarind fruit pods appropriated has been dried at 45°C for approximately 6 hours in an hot air, then minced by milling using a locally Milling machine and then kept in plastic sackets at room temperature ($25^{\circ}\text{C}\pm 2^{\circ}\text{C}$).

Analytical Methods

Moisture, Protein (N x 6.25 Keldahl method), fat (hexane solvent, Soxhielt apparatus), fiber and ash were determined according to the method recommended by A O A C (2000).

Carbohydrates and energy value

Carbohydrate calculated by differences as follows:

$\% \text{ Carbohydrates} = 100 - (\% \text{ moisture} + \% \text{ protein} + \% \text{ fat} + \% \text{ ash} + \% \text{ fiber}).$

Energy value was estimated by the sum of multiplying protein and carbohydrates by 4.0 and fat by 9.0 according to FAO (1982).

Determination of minerals content

Minerals content (Na, Ca and K) were determined in the diluted solution of ash samples by using emission flame photometer (Model Corning 410). The other minerals (Cu, Zn, Mn, Fe, P and Mg) were determined by Atomic absorption spectrophotometer (PerKin – Elmer Instrument Model 2380, Germany), according to the method described by *Nzikouet al., (2009)*.

Determination of amino acids

Amino acids were determined using amino acid analyzer (LC 3000), according to the methods of *Bassler and Buchholz (1993)*.

Determination of total antioxidant activity of tamarind fruits

Antioxidant activity was determined according to the method described by *Zhang and Hamazu (2004)* as follows: Five grams of tamarind fruits pulp in different parts were extracted by 100 ml 80 % methanol. Different concentrations (10 to 50 μmol) were used to determine the antioxidant activity using 2,2 – diphenyl – 1 – picryl hydroxyl (DPPH).

Total phenolics were estimated according to *AOAC (1990)*, by using photometric method with Folin Ciocalteu reagent. Flavonoids were extracted and determined according to *Zhuanget al., (1992)*.

Determination of phenolic compounds

Extraction, separation and quantification of phenolic compounds were carried out according to the method described by *Goupyet al., (1999)*. The HPLC system Perkin Elmer PE200 was composed of a binary pump, a column thermostat and an auto sampler. The mass spectrometer used was a 3200QTRAP MS/MS with ESI ionization (Applied Biosystems / MDSSciex, Foster City, USA). The experimental conditions where: Mobile phase A: 50% acetonitrile, 50% acetic acid (0.5%); mobile phase B: 2% acetic acid, flow rate: 0.7 ml /min; injection volume: 20 μL . Stock solutions of standards were diluted in the mobile phase to obtain working standard solutions. Concentrations of the compounds were calculated from chromatogram peak areas on the basis of calibration curves. The method linearity was assessed by means of linear regression of the mass of compounds injected vs. its peak area. All solvents were of HPLC grade and were filtered and degassed before use.

Statistical analysis

Data were recorded as means and analyzed by (SPSS) (Ver.10.1). One-way analysis of variance (ANOVA) and Duncan comparisons were tested to signify differences between variable treatments of tamarind fruits (SAS 1988).

Results And Discussion

1. Chemical composition of fresh tamarind pulp

The proximate chemical composition of fresh tamarind pulp is shown in table (1). The obtained results indicated that moisture; protein, fat, ash, fiber, carbohydrates and energy value as wet weight of tamarind fruits pulp were 34.25, 6.01, 4.34, 2.27, 4.13, 49.00 % and 259.10 Kcal/100g, respectively. These results are in agreement with **Amoo and Atsie, (2012)** who found that tamarind pulp (*T. indica*) was rich in protein (7.64%). While, crude fat and carbohydrates values in this study were lower than the obtained results. Crude fat content of the samples was (1.03%), carbohydrate values was 56.00%. The high crude fiber content helps to maintain the health of the gastro-intestinal tract (**Ajayiet al., 2006**).

2. Mineral content of tamarind fruit pulp

Data presented in table (2) show the mineral content of tamarind fruit pulp. It is clear to notice that the highest mineral contents of tamarind which recorded for calcium, phosphorus and sodium. The values were 465.75, 97.00 and 76.66 mg/100g, respectively. On the other hand, the lowest mineral contents of tamarind were recorded for lead, nickel and zinc. The values were 0.01, 0.52 and 1.56 mg/100g, respectively. These results are in agreement with **Glewet al., (2005)**, who reported that tamarind fruit pulp is a good source of calcium and phosphorus, but is unfortunately, extraordinarily low in iron while the reverse recorded in present work (table 2). Also, **Almeida et al., (2009)** indicated that tamarind is a rich source of all minerals available, especially magnesium, copper and potassium, in addition to being a good source of calcium, phosphorous and selenium.

3. Amino acids composition of tamarind fruit pulp

Data given in table (3) show the amino acids contents of tamarind fruit pulp. It is clear to notice that the highest non essential amino acids contents of tamarind fruit pulp recorded for glutamic acid, aspartic acid and for essential amino acids (EAA)

isoleucine and lysine. The values were 16.70, 12.00, 8.89, and 8.22 mg/g, respectively. While, the lowest amino acids contents of tamarind fruit pulp was recorded for tryptophan, cysteine and methionine 1.04, 1.35 and 2.48 mg/gdw, respectively. These results are in agreement with **Ishola and collaborators (1990)** mention that the tamarind pulp is a good source of protein. Amino acid profiles of tamarind reveal that the proteins contain fairly balanced essential amino acid levels. Also, **El-Siddiget al., (2006)** reported that in terms of protein content and WHO reference protein, tamarind pulps score well for 3 of the 8 essential amino acids. However, for each of the eight essential amino acid categories baobab leaves score close to or above the 100% mark, except from tryptophan.

4. Fatty acids composition of tamarind fruit pulp

Data in table (4) presented the fatty acids contents of tamarind fruit pulp. The obtained results showed that the highest fatty acids contents of tamarind fruit pulp recorded for linoleic, oleic and palmitic. The values were 3.42, 2.29 and 1.80 µg/g, respectively. While, the lowest fatty acids contents of tamarind fruit pulp recorded for C12:0, C14: myristic, C15:0 C20:2n-6, C22:1 and C15:0. The values were 0.01, 0.01, 0.01, 0.01 and 0.01 µg/g, respectively. These results are in agreement with **Glewet al., (2005)** reported that tamarind fruit pulp is relatively poor in oil (25.3 g/kg of crude lipid), greenish yellow in colour and liquid at room temperature. Saponification values of the oil are high, indicating that it contains a high proportion of low molecular weight fatty acids. With regard to the two essential fatty acids, the fruit pulp contained very little linoleic acid (3.42 mg/g dry weight) and even lower amounts of α -linolenic acid (0.21 mg/g dry weight). Also, **Ajayiet al., (2006)**, who found that tamarind, have a higher percentage of unsaturated (55.6%) fatty acids than saturated (44.4%) fatty acids. Linoleic acid, present in tamarind seed oil, is undoubtedly one of the most important polyunsaturated acids in human food because of its association in the reduction or prevention of heart vascular diseases.

5. Total phenols and antioxidant activity of tamarind

Table (5) showed the total phenols and antioxidants activity content of tamarind. It could be observed that the total phenols and antioxidants activity (DPPH) content were 21.36 mg/g and 67.90 %, respectively. These results are in the same line of **Mahmoodet al.,**

(2012), who reported that *Tamarindusindica*, L. contained a large number of polyphenolic compounds with potential for antioxidant activity. However, the quantities of antioxidants may vary with geographical location.

6. Identification of phenolics compounds of tamarind

Data given in table (6) show the identification of phenolic compounds of tamarind. It is clear to mention that the highest phenolic compound of tamarind recorded for pyrogallol, catechin and benzoic acid. The values were 2588.84, 527.55 and 365.85 mg/kg, respectively. On the other hand, the lowest value of phenolic compounds of tamarind recorded for cinnamic acid, reversetrol and coumarin. The values were 2.78, 4.95 and 5.52 mg/kg, respectively. These results are in agreement with **El-Siddiget al., (2006)**, who reported that tamarind fruit being a plant contained a biologically important source of mineral elements and with a high antioxidant capacity associated with high phenolic content that can be considered beneficial to human health. The phenolics include gallic acid equivalent of 626-664 mg per 100g. Also, **Khairunnuuret al., (2009)** reported a wide range of total phenolic content in tamarind parts. Their contents ranged from 1.83 ± 0.02 to 19.21 ± 0.29 mg gallic acid equivalent (GAE)/100g of dried samples, with an average of 9.64 mg (GAE)/100g fresh sample.

7. Identification of flavonoides compounds of tamarind

Data tabulated in table (7) presented the identification of flavonoides compounds of tamarind. It is clear to mention that the highest phenolics compounds of tamarind recorded for luteo.6-arbinose 8-glucose, luteo.6-arbinose 8-arbinose and Apig. 6-glucose 8-rhamnose. The values were 4147.57, 291.40 and 239.58 mg/kg, respectively. On the other hand, the lowest value of flavonoid compounds of tamarind recorded for kampferol, apegnin and quercetin. The values were 1.01, 2.14 and 4.92 mg/kg, respectively. These results are in agreement with **Lamien-Medaet al., (2008)** reported that lower levels of flavonoids (2.18 ± 0.21 mg QE/100 g) in fruit methanolic extracts and (5.68 ± 0.10 mg QE/100 g) in fruit acetone extracts found in tamarind growing in Burkina Faso.

Table (1): Chemical composition of fresh tamarind

Constitutive	% (W/W)	% (D/W)
Moisture	34.25	-----
Protein	6.01	9.15
Fat	4.34	6.60
Ash	2.27	3.45
Fiber	4.13	6.28
Carbohydrates	49.00	74.52
Energy value (Kcal/100g)	259.10	394.08

W/W= Wet weight D/W= Dry weight

Table (2): Mineral content of tamarind fruit pulp

Mineral contents	Concentrations (mg/100g Dw)
Calcium	465.75±0.10
Copper	21.83±0.11
Iron	8.49±0.03
Potassium	62.00±0.13
Magnesium	72.03±0.15
Manganese	21.50±0.22
Sodium	76.66±0.01
Nickel	0.52±0.40
Phosphorus	97.00±0.20
Lead	0.01±0.03
Zinc	1.56±0.12

Table (3): Amino acids composition of tamarind fruit pulp

Amino acids	Concentrations (mg/g Dw)
Aspartic acid	12.00
Glutamic acid	16.70
Serine	6.88
Glycine	5.15
Histidine	3.37
Arginine	8.74
Threonine	6.05
Alanine	6.20
Proline	7.61
Tyrosine	4.34
Valine	6.97
Methionine	2.48
Isoleucine	5.20
Leucine	8.89
Phenylalanine	4.78
Lysine	8.22
Cysteine	1.35
Tryptophan	1.04

Table (4): Fatty acids composition of tamarind fruit pulp

Fatty Acid	Concentrations($\mu\text{g/g Dw}$)
C12:0	0.01
C14:0 Myristic	0.01
C15:0	0.01
C16:0 Palmitic	1.80
C16:1 Palmitoleic	0.12
C18:0 Stearic	0.70
C18:1n-9 Oleic	2.29
C18:1n-7	0.55
C18:2n-6 Linoleic	3.42
C18:3n-3 α -linolenic	0.21
C20:0 Arachidic	0.07
C20:1 Gadoleic	0.02
C20:2n-6	0.01
C22:0	0.03
C22:1	0.01
C24:0	0.03
C24:1	0.20

Table (5): Total phenols and antioxidant activity of tamarind

Active compounds	Total phenols(Mg/g)	Antioxidant activity(DPPH)%
Tamarind	21.36 \pm 0.10	67.90 \pm 0.02

Table (6): Level of individual phenolic compounds of tamarind

Phenolics compounds	Dried tamarind (mg/kg)
Galic acid	43.16
Pyrogallol	2588.84
4-Amino benzoic	18.48
Protocatechuic	271.01
Catechein	527.55
Chlorogenic acid	74.45
Catechol	348.87
Epicatachin	219.29
Caffeine	39.92
P-OH-benzoic	106.77
Caffeic acid	27.32
Vanilic acid	161.92
<i>p</i> -cumaric acid	8.09
Ferulic acid	13.80
Isoferulic acid	8.09
Reversetrol	4.95
Ellagic acid	114.03
e-vanilic acid	123.84
Alpha-coumaric	17.60
Benzoic acid	365.85
3,4,5-methoxy cinnamic acid	10.76
Coumarin	5.52
Salycilic acid	40.25
Cinnamic acid	2.78

Table (7): Level of individual flavonoid compounds of tamarind

Flavonoids compounds	Dried tamarind(mg/kg)
Luteo.6-arbinose 8-glucose	4147.57
Luteo.6-arbinose 8-arbinose	291.40
Apig. 6-arbinose 8-glactose	14.43
Apig. 6-rhamnose 8-glucose	89.19
Apig. 6-glucose 8- rhamnose	239.58
Luteo.7- glucose	19.63
Luteolin	80.54
Narengin	25.31
Rutin	48.74
Hespiridin	210.81
Rosemarinic	6.65
Apig.7-O-neohespiroside	18.90
Kamp.3,7-dirhamoside	21.36
Apig.7-glocouse	28.32
Quercetrin	6.18
Quercetin	4.92
Kaemp.3-(2-P-comaroyl) glucose	98.91
Naringenin	8.87
Hespirtin	11.84
Kampferol	1.01
Rhamnetin	3.00
Apegnin	2.14
Acacetin	58.55

References

- Ajayi, I.A.; Oderinde, R.A.; Kajogbola, D.O. and Uponi, J.I. (2006):** Oil content and fatty acid composition of some underutilized legumes from Nigeria. *Food Chemistry*, 99: 115-120.
- Almeida, M.M.B.; de Sousa, P.H.M.; Fonseca, M.L.; Magalhes, C.E.C.; Lopes, M. F.G. and de Lemos, T.L.G. (2009):** Evaluation of macro and micro-mineral content in tropical fruits cultivated in the northeast of Brazil. *Ciência e Tecnologia de Alimentos*, 29: 581-586.
- Amoo, I.A. and Atasie, V.N. (2012):** Nutritional and functional properties of *Tamarindusindicapulp* and *Zizyphusspinachristifruit* and seed, *J. Food, Agric. Environ.*, 10: 16-19.
- AOAC (2000):** Official Methods of Analysis of the Association of Official Analytical Chemists. Arlington, Virginia, U.S.A.
- AOAC. (1990):** Official Methods of Analysis. Association of Official Analytical Chemists 15th Ed., Arlington Virginia.

- Bassler , N.R. and Buchholz, H. (1993):** Amino acid analysis. In: Methodenbuch Volume. 111, die chemische Untersuchung von Futtermitteln VDLUFA. Verlag, Darmstadt, Germany , Austria , 1-5.
- Baydar, N.G., Özkan, G. and Yasar, S. (2007):** Evaluation of the antiradical and antioxidant potential of grape extracts. *Food Control*, 18: 1131-1136.
- El-Siddig, K.; Ebert, G. and Lüdders, P. (1999):** Tamarind (*Tamarindusindica* L.): A Review on a multipurpose tree with promising future in the Sudan. *Journal of Applied Botany Angewandte Botanik*, 73: 202-205.
- El-Siddig, K.; Gunasena, H.P.M.; Prasa, B.A.; Pushpakumara, D.K.; Ramana, K.V.R.; Vijayanand. P. and Williams, J.T. (2006):** Tamarind – *Tamarindusindica*, L. Fruits for the Future 1. Southampton Centre for Underutilized Crops, Southampton, UK, 188p.
- FAO (Food and Agriculture Organization) (1982):** Food Composition Tables for the Near East, FAO, Food and Nutrition Paper, p. 26.
- Glew, R.H.; VanderJagt, D.J.; Lockett, C.; Grivetti, L.E.; Smith, G.C.; Pastuszyn, A. and Millson, M. (1997):** Amino Acid, Fatty Acid, and Mineral Composition of 24 Indigenous Plants of Burkina Faso. *Journal of Food Composition and Analysis*, 10: 205-217.
- Glew, R.S.; VanderJagt, D.J.; Chuang, L.T.; Huang, Y.S.; Millson, M. and Glew, R.H. (2005):** Nutrient content of four edible wild plants from West Africa. *Plant Foods for Human Nutrition*, 60: 187-193.
- Goupy, P.; Hugues, M.; Boivin, P. and Amoit, M.J. (1999):** Antioxidant composition and activity of barley (*Hordeum vulgare*) and malt extracts and of isolated phenolic compounds. *J. Sci. Food Agric.*, 79: 1625 – 1634.
- Hefnawy, Y. and Youssef, H. (1985):** Microbiological evaluation of some selected spices. *Assiut Veterinary Medical Journal*, 13: 145-166.
- Ishola, M.M.; Agbaji, E.B. and Agbaji, A.S. (1990):** A Chemical Study of *Tamarindusindica* (Tsamia) Fruits Grown in Nigeria. *Journal of the Science of Food and Agriculture*, 51: 141-143.
- Khairunnuur, F. A.; Zulkhairi, A.; Azrina, A.; Moklas, M. A. M.; Khairullizam, S.; Zamree, M. S. and Shahidan, M. A. (2009):** Nutritional composition, *in vitro* antioxidant activity and *Artemiasalina*, L. lethality of pulp and seed of *Tamarindusindica*, L. Extracts. *Malaysia Journal of Nutrition*, 15 (1): 65-75.

- Kumar, C.S. and Bhattacharya, S. (2008):** Tamarind seed: Properties, processing and utilization. *Critical Reviews in Food Science and Nutrition*, 48: 1-20.
- Lamien-Meda, A.; Lamien, C.; Compaoré, M.; Meda, R.; Kiendrebeogo, M.; Zeba, B.; Millogo, J. and Nacoulma, O. (2008):** Polyphenol content and antioxidant activity of four-teen wild edible fruits from Burkina Faso. *Molecules* 13: 581-594.
- Mahmood, T.; Anwar, F.; Abbas, M. and Saari, N. (2012):** Effect of maturity on phenolics (phenolic acids and flavonoids) profile of strawberry cultivars and mulberry species from Pakistan. *International Journal of Molecular Science*, 13: 4591-4607.
- Mielnik, M.B.; Semb, S.; Egeland, B. and Skrede, G. (2008):** Byproducts from herbs essential oil production as ingredient in marinade for turkey thighs. *LWT - Food Science and Technology* 41(1): 93-100.
- Nordeide, M.B.; Harløy, A.; Følling, M.; Lied, E. and Oshaug, A. (1996):** Nutrient composition and nutritional importance of green leaves and wild food resources in an agricultural district, Koutiala, in Southern Mali. *International Journal of Food Sciences and Nutrition*, 47: 455-468.
- Nzikou, I.M.; Matos, L.; Moussounga, J.E.; Ndangu, C.B. and Kimbonguila, A. (2009):** Characterization of *Moringa oleifera* seed oil variety Congo Brazzaville. *J. Food Technol.*, 7 (3): 59 – 65.
- SAS (1988):** SAS Users Guide: Statistics version 5th Ed., SAS. Institute Inc., Cary N.C.
- Velasco, V. and Williams, P. (2011):** Improving Meat Quality Through Natural Antioxidants. *Chilean Journal of Agricultural Research*, 71 (2): 313-322.
- Yin M.C. and Cheng W.S. (2003):** Antioxidant and antimicrobial effects of four garlic-derived organosulfur compounds in ground beef. *Meat Science*, 63: 23–28.
- Zhang, D. and Hamauzu, Y. (2004):** Phenolics, ascorbic acid, carotenoids and antioxidant activity of broccoli and their changes during conventional and microwave cooking. *Food Chemistry*, 88: 503 – 509.
- Zhuang, X.P.; Lu, Y.Y. and Yang, G.S. (1992):** Extraction and determination of flavonoid in ginkgo. *Chinese Herbal Medicine*, 23: 122 – 124.

الخصائص التغذوية للب ثمار التمر هندي

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قسم التغذية وعلوم الأطعمة - كلية الاقتصاد المنزلى - جامعة المنوفية - مصر

الملخص العربى

التركيب الكيماوى والاملاح المعدنية والأحماض الأمينية والأحماض الدهنية بالإضافة إلى المركبات الكيميائية الطبيعية تم قياسهم فى لب ثمار التمر هندي. وقد كانت قيم كلا من الرطوبة والبروتين والدهون والرماد والألياف والكربوهيدرات وقيمة الطاقة على أساس الوزن الرطب من ثمار التمر الهندي ٣٤,٢٥، ٦,٠١، ٤,٣٤، ٢,٢٧، ٤,١٣، ٤٩,٠٠٪ و ٢٥٩,١٠ كيلو كالوري / ١٠٠ جم على التوالي. وقد وجد أن أعلى محتوى للأملاح المعدنية فى ثمار التمر الهندي سجلت مع عنصر الكالسيوم والصوديوم والفسفور، في حين كان أقل محتوى هو عنصر الرصاص والنيكل والزنك. كما أعلى محتوى من الأحماض الأمينية الأساسية فى لب ثمار التمر الهندي سجلت مع الحمض اللبوسين، في حين أن أقل محتوى سجل مع الحمض الأميناتريتوفان، السيستين والميثيونين. أما أعلى محتوى من الأحماض الدهنية فلب ثمار التمر الهندي سجلت مع الحمض الدهنيالينوليك، الاوليكوالبالميتيك. حيث كانت القيم ٣,٤٢، ٢,٢٩,١,٨ ميكروجرام / جرام على التوالي. وبلغ محتوى الفينولات الكلية و نشاط مضادات الأكسدة ٢١,٣٦ ملجم / جم و ٦٧,٩٠٪ على التوالي. أعلى قيم للمركبات الفينولية التي تم التعرف عليها في ثمار التمر الهندي سجلت مع حمض البيروجالول، الكاتيشينوحمض البنزويك. حيث كانت القيم ٢٥٨٨,٨٤، ٥٢٧,٥٥ و ٣٦٥,٨٥ ملجم / كجم، على التوالي. من ناحية أخرى، سجلت أقل قيمة مع حمض السيناميك، حمض الريفيرستولوالكومارين. أعلى قيم للفينولات التي تم التعرف عليها فى التمر الهندي سجلت مع مركبليوتيلو ٦- أرابينوز 8 جلوكوز، وليوتيلو ٦- أرابينوز 8 أرابينوز، بينما أقل قيمة من الفلافونيدات سجلت معأبيجينوكامبيفيرولوراميستين. من هنا نستخلص أن لب ثمار التمر الهندي ذات قيمة غذائية عالية نظرا لأحتواءه على الأحماض الأمينية، والأحماض الدهنية والمعادن والمركبات الفينولية ومضادات الأكسدة، وعليه فإنه يمكن استخدامها لحماية و تحسين صحة الإنسان ضد بعض الأمراض.

الكلمات الدالة: ثمار التمر هندي، التركيب الكيماوى، المركبات النباتية الطبيعية.