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STUDY OF HEMATOLOGICAL AND BLOOD BIOCHEMICAL PARAMETERS IN CATS EXPERIMENTALY INFECTED WITH TOXOCARA CATI

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ABSTRACT

The present study was undertaken to assess and compare the hematology and serum biochemical parameters of cats experimentally infected with *Toxocara cati* (infected group) and healthy cats (control group). A total of fourteen and six blood samples were collected from infected and control groups respectively. Hematological changes showed a significant decrease of Hemoglobin concentration (Hb), Total Erythrocytes Count (TRBCs), Packed Cell Volume (PCV), and Platelets Count (Pt), furthermore a significant increase in Mean Corpuscular Volume (MCV) and significant decrease in Mean Corpuscular Hemoglobin Concentration (MCHC) which reflects a macrocytic hypochromic type of anemia is present. Results also revealed a significant increase in Total Leukocytes Count (TWBCs) due to significant increase of neutrophils and eosinophils. The results of serum biochemical parameters shown that glucose, Alanine aminotransferase (AST), Alkaline Phosphatase (AP), cholesterol and triglyceride significantly increased, whereas total protein and creatinine levels were significantly decreased in toxocara infected cats compared to healthy cats. But the serum bilirubin and Blood Urea Nitrogen (BUN) levels were not affected by toxocara infection.

Key words: Toxocara cati, blood parameters

INTRODUCTION

Toxocariasis is a public health problem and clinical syndrome caused by infection of zoonotic roundworms of cats (Toxocara cati) and dogs (Toxocara canis). Toxocara species are not only present worldwide in their definitive hosts only; they also frequently occur in other animal species and humans (In those so-called paratenic hosts) (Soulsby, 1986). T. cati is a common parasite of Felidae, and has a cosmopolitan distribution (Yamaguchi et al., 1996). Cats can become infected with T. cati by ingesting embryonated eggs found in contaminated soil or by predation of paratenic hosts - usually small rodents or birds containing live third-stage larvae (L3) in their tissues (Al-Quraishi, 2013). In pregnant queen, dormant tissue larvae are reactivated and migrate across the placenta to infect the fetus. New born kittens also acquire infection through ingestion of larvae in milk (Overgaauw, 1997).

The live cycle of *T. cati* is a complex, adult worm in the intestinal tract of infected cats shed large number of eggs into the environment; these may be ingested by final host as well as paratenic host. The larvas then hatch and migrate through blood vessels all over the body organs (Azizi *et al.*, 2007). Those arrested larvae may lead to severe inflammatory reactions and consequently to a wide range of pathological and clinical manifestations, are possible causes of visceral larva migrants (VLM) or ocular larvae migrants (OLM) (Dubinsky, 1999).

The present study was taken into consideration to know the effect of *toxocara cati* infection on the hematology and blood biochemical parameters of experimentally infected cats.

MATERIALS and METHODS

The present study was carried out at the College of Veterinary Medicine, Duhok University, Kurdistan Region – Iraq. About 5 ml of blood was collected aseptically from the cephalic vein of two groups of cats. The first group consist of 14 cats were experimentally infected with *T.cati* by ingested of embryonated infected eggs from previous work

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(Dishow, 2014) and the second group consist of 6 clinically normal cats which considered as a control group. The collected blood was stored in two separate tubes, one with anticoagulant EDTA for determine hematological parameters and another tubes without anticoagulant from which later serum was separated and it was put into a screw capped plastic vials, then stored at -20 C about one week for determine effects of *T.cati* infection on blood biochemical parameters.

The blood parameters determined by Auto hematology analyzer (model BC- 2800 vet, Germany), includes Total Erythrocytes Count (TRBCs), Packed Cell Volume (PCV), Hemoglobin concentration (Hb), Platelets count (Pt), Mean Corpuscular Volume (MCV), Mean Corpuscular Hemoglobin Concentration (MCHC). Furthermore total and differential leukocytes count was also encountered.

The serum biochemical parameters examined in the present study including glucose, Alanine Amino Transferase (ALT), Alkaline Phosphatase (AP), cholesterol, triglyceride, total protein, creatinine, bilirubin and Blood Urea Nitrogen (BUN) levels. All these parameters were done according to enzymatically measured using a standard enzymatic assay kit (Biolabo reagents, France).

Statistical analysis, the data of the present study were analyzed using unpaired't' test with accepted level of significant was P < 0.05 (Bowers, 2008).

RESULTS

The present study showed that the experimental toxocara infection in cats causes changes of hematological and serum biochemical parameters when compared with control group.

Hematology results revealed a significant decrease of Hb, TRBCs, PCV, Pt, MCHC and a significant increase in MCV when compared with control group. According to these results showing that there is macrocytic hypochromic anemia in infected group. The results also give a significant increase in TLCs which were due to a significant increase of neutrophils, and eosinophils. While, the levels of lymphocytes, basophils and monocytes were not affected by toxocara infection, the results are seen in Table (1).

Table 1: Mean value of hematological	parameters of the infected cats and control	group.
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Serial	Hematology parameters	natology parameters Mean + Stander error of mean (M+SE)	
No.	-	Healthy cats (n=6) (control group)	Infected cats (n=14) (infected group)
1.	Total RBCs X 10 ⁶ mL	$7.58 \pm 0.52 \qquad A$	$4.24\pm0.13~B$
2.	Hb (g/dl)	$15.95\pm0.60~A$	$5.86\pm0.27~B$
3.	PCV (%)	35.67 ± 44.09 A	14.14 ± 1.13 B
4.	Pt X 10 ³ μL	$366.67 \pm 44.09 \text{ A}$	$146.43\pm7.76\ B$
5.	MCV (fl)	$42.75\pm2.60 B$	55.86 ± 1.37 A
6.	MCHC (g/100 ml)	$33.20\pm0.46~A$	25.21 ± 1.13 B
7.	Total WBCs X 10 ³ µL	$14.50\pm0.81 B$	$21.86\pm0.75~A$
8.	Neutrophils X10 ³ µL	$65.17\pm2.57 B$	$88.50 \pm 1.05 A$
9.	Eosinophils X10 ³ µL	0.60 ± 0.34 B	6.50 ± 0.94 A
10.	Lymphocytes X10 ³ µL	21.83 ± 2.75 A	25.00 ± 1.94 A
11.	Monocytes X10 ³ µL	3.17 ± 1.56 A	6.07 ± 0.29 A
12.	Basophils X10 ³ mL	0.50 ± 0.22 A	0.64 ± 0.29 A

Different letters are significantly different at P< 0.05

In addition, the present study was carried out by employing various biochemical parameters to detect of the effects of *T.cati* infection on these parameters. The serum biochemical results were showed significant increase in glucose, alkaline phosphatase (AP), Alanine Amino Transferase (ALT), cholesterol and triglyceride levels. Results also indicated a significant decrease of total proteins and creatinine levels. Whereas, the serum bilirubin and BUN concentration were not affected by toxocara infection. The results of serum biochemical parameters are depicted in Table (2).

Serial No.	Serum biochemistry	Mean + Stander error of mean (M+SE)		
		Healthy cats (n=6) (control group)	Infected cats (n=14) (infected group)	
1.	Glucose (g/dl)	102.83 ± 5.94 B	229.23 ± 14.74 A	
2.	AP	$36.00\pm2.38 B$	83.46 ± 3.23 A	
3.	ALT	$71.00\pm3.21 B$	129.2 ± 7.41 A	
4.	Cholesterol (mg/dl)	$76.67\pm2.78 B$	235.23 ± 8.47 A	
5.	Triglyceride (mg/dl)	$48.33 \pm 10.46 \ B$	162.5 ± 8.75 A	
6.	Total proteins (g/dl)	6.50 ± 0.22 A	$1.69\pm0.21\qquad B$	
7.	Creatinine (mg/dl)	6.50 ± 0.22 A	$0.77\pm0.07~B$	
8.	Bilirubin (mg/dl)	0.43 ± 0.08 A	0.16 ± 0.10 A	
9.	BUN (mg/dl)	21.17 ± 0.54 A	21.08 ± 0.24 A	

Table 2: Mean value of serum biochemic	al components in infected and control group.
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Different letters are significantly different at P< 0.05

DISCUSSION

Hematological parameters recorded in this study revealed a significant decrease in Hb, TRBCs, PCV, Pt and MCHC values with only increase in MCV value, that's reflecting macrocytic hypochromic anemia. Similar results were obtained by Bowman (1995) and Al-Obaidi (2012). All these variants may be due to response to migration of the toxocara larvae through different internal organs of cats which causes anemia due to internal bleeding (Dishow, 2014). Leukocytosis occurred in current study were also recorded by Bowman (1995) and Al-Obaidi (2012) which might occur due to stimulation of cellular immunity by specific antigenic structure of the toxocara larva and bone marrow as immune response against the parasite or their toxins (Dishow, 2014). Leukocytosis and eosinophilia may be attributed to immunological mechanisms, Coffman et al. (1993) showed the parasite - derived antigens stimulate Th0 cells to become Th2 cells, leading to the production of IL-4 and IL-5, which stimulate IgE production and eosinophils proliferation and maturation respectively. Leukocytosis and eosinophilia can occur in acute stage of toxocara infection (Sapunar and Ferdella, 1999).

In the current study, various serum biochemical parameters were recorded to evaluate the effect of toxocara infection. Most of them were found affected by the toxocara infection except bilirubin and BUN were not affected.

The blood glucose concentration in infected cats is increased significantly; this increase can be attributed to activation of glycogenolysis in liver due to migration of toxocara larva which causes anoxia, since liver glycogen is relatively unstable in the presence of a deficient oxygen supply (Coles, 1986). The results of the present study were in agreement with Vidal *et al.* (2003) who reported increase in glucose level due to toxocara infection. There is elevation in serum enzyme activity as ALT due to malfunctioning of the liver, this observation is supported by other studies. Coles (1986) and Radostits *et al.* (2007) mentioned that the ALT and AST enzyme were increased due to disruption of hepatic cells as a result of necrosis or altered membrane permeability with visceral larva migrans. Especially, ALT enzyme is present in large quantities in the hepatocytes cytoplasm of these animal species (cats, dogs and primates) only, increased in serum when cellular degeneration or destruction occurs in liver (Coles, 1986).

Experimentally toxocara infection group had higher mean alkaline phosphatase (AP) levels when compared with control group and the difference was statistically significant. These results are in agreement with Azuma *et al.* (2002) who noted a case of hepatic involvement of visceral larva migrans due to toxocara causes increase AP enzyme level. AP enzyme also increased in biliary obstruction in cats or with increase corticosteroid (Coles, 1986).

Serum cholesterol and triglyceride levels are also significantly elevated with the rise of toxocara infection intensity could be due to the higher level of cortisol caused by stress of infection or due to hyperglycemic syndrome which associate with toxocariasis (Stockham and Scott, 2003).

There is a decrease in concentration of serum total protein and creatinine in the toxocara affected cats compared with control group may be due to liver damage and interference with proteins synthesis, not only due to larva migration, but also as a consequence of the development of oxidative stress (Azuma *et al.*, 2002). In addition, the reduction in total protein value is might be due to alimentary tract dysfunction (diarrhea) or loss of appetite (Radostits *et al.*, 2007).

In the present study, the serum bilirubin level does not get altered by toxocara infection. Our study is in agreement with Sapunar and Ferdella (1999) who revealed no change in serum bilirubin level in case of visceral larva migrans. Similarly, Sommerfelt *et al.* (2001) recorded no significant change in serum bilirubin in toxocara infected pigs.

In infected group with toxocara showed no significant change in their BUN, this result is in agreement with Singh *et al.* (1992) who observed no change in the level of BUN by toxocara infection in a case with visceral larva.

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دراسة المعايير الدموية والكيموحيوية في القطط المصابة تجريبيا بداء السهميات القططي

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أستهدفت الدراسة الحالية قياس ومقارنة التغيرات الدموية والكيموحيوية في القطط المصابة تجريبيا بداء السهميات القططي (المجموعة المصابة) ومقارنتها مع القطط السليمة (المجموعة الضابطة). شملت الدراسة فحص ٤ اعينة من دم القطط المصابة و ٢ عينات من الدم من القطط السليمة. أظهرت التغيرات الدموية انخفاضا معنويا في تراكيز الهيمو غلوبين، العدد الكلي لكريات الدم الحمراء، حجم الخلايا المرصوصة وعدد الصفيحات الدموية. فضلا عن زيادة معنوية في معدل حجم الخلية ونقصا في معدل تركيز هيمو غلوبين الخلية وهذا يدل على وجود فقر الدم من النوع خلايا الدموية. فضلا عن زيادة معنوية في معدل حجم الخلية ونقصا في معدل تركيز هيمو غلوبين الخلية وهذا يدل على وجود فقر الدم من النوع خلايا كبيرة قليلة الخضاب. ولوحظت زيادة معنوية في العدد الكلي لكريات الدم الحيان الخلية وهذا يدل على وجود فقر الدم من النوع خلايا ما نتائج الدراسة الكيموحيوية اظهرت زيادة معنوية في تراكيز الكلوكوز، انزيم الاسبارتيت ناقلة الأمين، انزيم الفوسفات. والشحوم الثلاثية بينما تركيز البروتين الكلي والكريات الدم البيضاء بسبب الزيادة المعنوية في كل من الخلايا المتول والشحوم الثلاثية بينما تركيز الموتين الكلي والكريات الم البيضاء مالمصاريات الدراسة الكيوسفوساتين القلوي، الكولستيرول والشحوم الثلاثية بينما تركيز البروتين الكلي والكرياتين انخفضا معنويا في القطط المخمجة عند مقارنتها بالقط السليمة، في حين لم يتأثر اي من والشحوم الثلاثية بينما تركيز الدروتين الكلي والكرياتين انخفضا معنويا في القطط المخمجة عند مقارنتها بالقط السليمة، في حين لم يتأثر اي من تراكيز بليروبين المصل ويوريا الدم بالاصابة.